

Association of Nijmegen Breakage Syndrome 1 Genotypes With Bladder Cancer Risk

MENG CHEN^{1*}, WEN-SHIN CHANG^{2*}, TE-CHUN SHEN^{2*}, CHI-LI GONG³,
MENG-LIANG LIN⁴, ZHI-HONG WANG⁵, YUN-CHI WANG^{2,6},
CHAO-HSUAN CHEN², HSI-CHIN WU^{2,7}, DA-TIAN BAU^{2,6,8} and CHIA-WEN TSAI^{2,6}

¹Department of Clinical Laboratory, National Cancer Center/Cancer Hospital,
Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, P. R. China;

²Terry Fox Cancer Research Laboratory, Department of Medical Research,
China Medical University Hospital, Taichung, Taiwan, R.O.C.;

³Department of Physiology, China Medical University, Taichung, Taiwan, R.O.C.;

⁴Department of Medical Laboratory Science and Biotechnology,
China Medical University, Taichung, Taiwan, R.O.C.;

⁵Department of Food Nutrition and Health Biotechnology, Asia University, Taichung, Taiwan, R.O.C.;

⁶Graduate Institute of Biomedical Sciences, China Medical University, Taichung, Taiwan, R.O.C.;

⁷Department of Urology, China Medical University Hospital, Taichung, Taiwan, R.O.C.;

⁸Department of Bioinformatics and Medical Engineering, Asia University, Taichung, Taiwan, R.O.C.

Abstract. *Background/Aim:* We aimed to examine the association of the genotypes of Nijmegen breakage syndrome 1 (NBS1), a critical gene in DNA double strand break repair machinery, with bladder cancer risk in Taiwan. *Materials and Methods:* NBS1 rs1805794 genotypes among 375 bladder cancer patients and 375 non-cancer healthy controls were determined via the polymerase chain reaction-restriction fragment length polymorphism methodology and their association with bladder cancer risk were evaluated. *Results:* The results showed that the percentages of GG, CG and CC of NBS1 rs1805794 genotypes were 45.4%, 43.7% and 10.9% in the bladder cancer patient group and 47.2%, 43.2% and 9.6% in the non-cancer control group, respectively (p for trend=0.7873). The analysis of allelic frequency distributions showed that the variant C allele of NBS1 rs1805794 does not contribute to an increased bladder cancer susceptibility ($p=0.5066$). *Conclusion:* The genotypes of NBS1 rs1805794 are not closely associated with personal susceptibility to bladder cancer.

Statistically, bladder cancer (BLACA) is worldwide the sixth and seventeenth most prevalent cancer in men and women, respectively, with nearly 500,000 newly diagnosed cases in 2018 (1). In Taiwan, the incidence of BLACA is the ninth and fourteenth most common cancer in men and women, respectively, as recorded in the most updated government annual reports (2). From the viewpoints of epidemiology, the high incidence of BLACA is due to both environmental and genetic factors. As for the environmental factors, it is believed that smoking, exposure to chemicals and PM2.5, prior radiation therapy, and frequent bladder infections contribute to BLACA risk (3, 4). Although family history of the disease has supported the idea that genetic factors also play a role in BLACA risk determination, interestingly, the contribution of genetic factors and the underlying etiology of BLACA remains largely unclear. In the past decades, translational studies were focused into examining useful biomarkers for the prediction of BLACA (5-10), while genetic markers are urgently needed for various populations all over the world.

Among the various kinds of DNA repair proteins, the DNA repair and telomere maintenance protein nbs1, named for its underlying role in the Nijmegen breakage syndrome, plays a critical role in the DNA double strand break (DSB) repair machinery. As an early response to DNA DSBs, histone H2AX in the vicinity of DSBs is phosphorylated by Ataxia telangiectasia mutated (ATM) (11). Then, NBS1 targets the MRE11/RAD50 complex to the sites of DSBs via binding with the FHA/BRCT domain of the phosphorylated gamma-H2AX

*These Authors contributed equally to this study.

Correspondence to: Da-Tian Bau and Chia-Wen Tsai, Terry Fox Cancer Research Laboratory, China Medical University Hospital, 2 Yuh-Der Road, Taichung, 404 Taiwan, R.O.C. Tel: +886 422053366 (Ext. 5805), e-mail: datian@mail.cmuh.org.tw; artbau2@gmail.com

Key Words: Bladder cancer, case-control study, genotype, NBS1, polymorphism.

(12). After that, the NBS1 complex directly binds to the damaged-DNA, initiating the DSB repair (13). In addition, NBS1 also teams up with ATM to control the cell cycle through the phosphorylation of SMC, CHK2 and FANCD2 (14-16). Thus, NBS1 plays a critical role not only in the maintenance of DNA integrity, but also in cell cycle regulation. Furthermore, NBS1 has also been reported to be involved in maintaining the length of telomeres, which have DSB-like structures and in which defects are closely associated with increased genomic instability and aging dysregulation (17, 18).

The *NBS1* gene, also called nibrin and NBN, is located in the human chromosome 8q21 (19, 20). In heterozygous NBS1 (+/–) mice, the tumor formation rates and ionizing radiation sensitivity are both much higher compared to wild-type mice, indicating that *NBS1* plays a critical role in DSB repair and carcinogenesis (21). In the *NBS1* gene, the most commonly investigated polymorphism is rs1805794 (Glu185Gln, E185Q), which has been widely studied regarding its association with susceptibility to several types of cancer including nasopharyngeal cancer (22), lung cancer (23-26), breast cancer (27), colorectal cancer (28), prostate cancer (29), and leukemia (30). Up to now, there is limited literature regarding the contribution of *NBS1* genotypes to bladder cancer risk (31-33). However, the results reported by these studies remained inconclusive, and none of them investigated the *NBS1* genotypes in Eastern countries. In this study, we aimed at examining the association between single nucleotide polymorphisms (SNPs) at the *NBS1* rs1805794 with the risk of bladder cancer in Taiwan.

Materials and Methods

Bladder cancer patients and non-cancer controls. This hospital-based case-control study was approved by the Institutional Review Board of China Medical University Hospital (DMR104-IRB-158) and all participants have provided written-informed consents. All clinical and pathological records were restrictively reviewed according to the principles expressed in the Declaration of Helsinki. Briefly, three hundred and seventy-five cases diagnosed with bladder cancer were recruited in this study, after completing a comprehensive questionnaire and providing 3 to 5 ml of their peripheral blood. An equal number (375) of non-cancer healthy individuals were obtained from the Health Examination Cohort of our hospital from an original pool of 15,000 subjects by matching for age, gender and smoking status. The exclusion criteria of the control subjects were defined as previously published (5-7). In brief, subjects with previous malignancy, metastasized cancer from another site or a tumor of unknown origin, or with any familial or genetic disease were excluded from the control group. As mentioned above, all participants completed a short questionnaire regarding personal characteristics, especially regarding their individual environmental exposures and life styles, such as smoking and alcohol drinking habits. Ever smokers were defined as daily or almost daily smokers who had smoked at least five packs of cigarettes per year in their lifetime for at least one year. The ever alcohol drinkers were defined as those who were twice drunken or

had more than three cups per week for at least one year. The drunken status is defined as the loss of control in straight walking. Overall, the selective demographic characteristics of all the individuals investigated are summarized in Table I.

***NBS1* rs1805794 genotyping conditions.** Genomic DNA from the peripheral blood leucocytes of each patient and control was extracted using the QIAamp Blood Mini Kit, stored and processed as reported in our previous articles (34-36). The specific primer sequences of the forward and reverse primers of *NBS1* rs1805794, were 5'-TGTGCTCTTCTGACCATGAG-3' and 5'-CAGTGA CCAAAGACCGACTT-3', respectively. The specific polymerase chain reaction (PCR) cycling conditions for *NBS1* rs1805794 genotyping were set as one cycle at 94°C for 5 min; followed by 35 cycles of 94°C for 30 sec, 55°C for 30 sec, and 72°C for 30 sec; and finally an extension step at 72°C for 10 min. The PCR products were cut by the restriction enzyme *Hinf*I (New England BioLabs), overnight. The DNA adducts carrying the digestible C allele were cut into 321- and 255-base pair contigs, while those carrying the indigestible G allele remained intact with 576-base pair long contigs. The genotypic process was performed by at least two researchers independently and blindly at least twice. Also, PCR samples from 20 cases and 20 controls were directly sequenced and the results obtained along with those from the PCR–restriction fragment length polymorphism were 100% concordant.

Statistical analysis. The Student's *t*-test was adopted in the age (continuous variable) comparison between the case and control groups. The Pearson's chi-square was used for comparing the distributions of age, gender, personal habits, *NBS1* SNP genotypes and alleles among the subgroups. The associations between *NBS1* genotypes and bladder cancer risk were estimated with individual odds ratios (ORs) and 95% confidence intervals (CIs). A *p*-value less than 0.05 was identified as statistically significant.

Results

The demographic characteristics including age, gender, personal habits of all participants and the stage and grade of the 375 bladder cancer patients are summarized in Table I. First, the average age of the controls and bladder cancer patients were 62.9 and 61.4 years, respectively. The ratio of male *versus* female bladder cancer patients was about 3:1 (Table I). We adopted the matching strategy about their age, gender, smoking and alcohol drinking habits to recruit the same number of non-cancer healthy controls, and there was no difference in age, gender, cigarette smoking and alcohol drinking status between the two groups (*p*=0.7315, 0.5525, 0.3063 and 0.3807, respectively). As for the stage and grade of the bladder cancer patients, the percentages of non-muscle-invasive and muscle-invasive types were 62.7% and 37.3%, respectively; while those with low and high grades were 40.3% and 59.7%, respectively (Table I).

The distributions of the *NBS1* genotypes at rs1805794 among the non-cancer controls and the bladder cancer patients are presented in Table II. The results showed that the genotypes of *NBS1* rs1805794 were not -differentially

Table I. Basic characteristics of the 375 bladder cancer patients and 375 non-cancer controls.

Character	Controls (n=375)			Cases (n=375)			p-Value
	n	%	Mean (SD)	n	%	Mean (SD)	
Age (years)			62.9 (9.8)			61.4 (10.3)	0.7315 ^a
Age group (years)							0.7108 ^b
≤55	152	40.5%		158	42.1%		
>55	223	59.5%		217	57.9%		
Gender							0.5525 ^b
Male	287	76.5%		279	74.4%		
Female	88	23.5%		96	25.6%		
Personal habits							
Cigarette smoking	186	49.6%		201	53.6%		0.3063 ^b
Alcohol drinking	176	46.9%		189	50.4%		0.3807 ^b
Stage							
Non-muscle-invasive				235	62.7%		
Muscle-invasive				140	37.3%		
Grade							
Low				151	40.3%		
High				224	59.7%		

SD: Standard deviation; ^abased on Student's *t*-test; ^bbased on Chi-square test.

Table II. Distribution of NBS1 rs1805794 genotypes among bladder cancer patients and healthy controls.

rs1805794	Controls		Patients		OR (95%CI)	p-Value ^a
	n	%	n	%		
Genotype						
GG	177	47.2%	170	45.4%	1.00 (reference)	
CG	162	43.2%	164	43.7%	1.05 (0.78-1.43)	0.7330
CC	36	9.6%	41	10.9%	1.19 (0.72-1.94)	0.4993
<i>P</i> _{trend}						0.7873
Carrier analysis						
GG+CG	339	90.4%	334	89.1%	1.00 (reference)	
CC	36	9.6%	41	10.9%	1.16 (0.72-1.85)	0.5475
GG	177	47.2%	170	45.3%	1.00 (reference)	
CG+CC	198	52.8%	205	54.7%	1.08 (0.81-1.44)	0.6082

^aBased on chi-square test without Yates' correction; **p*<0.05; OR: odds ratio; CI: confidence interval.

distributed between bladder cancer and non-cancer control groups (*p* for trend=0.7873) (Table II). In detail, *NBS1* rs1805794 heterozygous CG and homozygous CC are not associated with increased bladder cancer risk (OR=1.05 and 1.19, 95%CI=0.78-1.434 and 0.72-1.94, *p*=0.7330 and 0.4993, respectively) (Table II). We further performed a carrier analysis, and the results showed that in both dominant and recessive models, the distributions of *NBS1* rs1805794 genotypes were not significantly different between the bladder cancer and control groups (Table II).

In order to further validate the findings in Table II, we also conducted an analysis of allelic frequency distribution for the *NBS1* rs1805794 among the investigated population, and the results are summarized in Table III. Supporting the findings that neither the heterozygous variant CG nor the homozygous variant CC genotype at *NBS1* rs1805794 is responsible for altered risk of bladder cancer, the C allele was not present at a significantly higher rate in the cases compared to controls (*p*=0.5066) (Table III).

Table III. Distribution of *NBS1* rs1805794 allelic frequencies among bladder cancer patients and healthy controls.

rs1805794	Controls	%	Patients	%	OR (95%CI)	p-Value ^a
Allele G	516	68.8%	504	67.2%	1.00 (reference)	
Allele C	234	31.2%	246	32.8%	1.07 (0.87-1.34)	0.5066

^aBased on chi-square test; **p*<0.05. *Statistically significant; OR: odds ratio; CI: confidence interval.

Discussion

The DNA DSB repair protein *NBS1* encompasses lots of DNA damage sensors, signal transducers, and effectors, which enable our cells to maintain DNA integrity and genomic stability. One of its exonic polymorphisms, rs1805794, has been widely studied in case-control association studies for various types of cancer, however, the results were inconclusive. In the current hospital-based case-control association investigation, we focused on evaluating the contribution of *NBS1* rs1805794 genotype to bladder cancer risk. After examining the bladder cancer cases and non-cancer healthy controls, their genotypic results showed that neither heterozygous CG nor homozygous CC genotype of *NBS1* rs1805794 was significantly associated with risk of bladder cancer (Table II). In addition, the allelic frequency analysis also supported the findings of the genotypic frequency analysis indicating that the variant C allele at *NBS1* rs1805794 was not associated with bladder cancer risk (Table III). This negative finding is not consistent with the previous findings showing that *NBS1* rs1805794 was associated with bladder cancer risk, and the association was limited to ever smokers along was dependent on smoking dose and smoking duration (37). The inconsistency may come from the fact that a different population was investigated, and further validations in larger sample sizes and different populations are needed.

Gender difference is reported to be a risk factor for bladder cancer (38-40), especially for the non-invasive subtype (41-43). Although the underlining mechanism(s) for this gender-specific difference in bladder cancer risk has not been revealed, it is thought that sex steroids play a critical role in the etiology of bladder cancer (42). We are also interested in whether the genotype of *NBS1* rs1805794 contributes to the gender difference in bladder cancer susceptibility. After stratification by gender, it was found that the genotypes of *NBS1* rs1805794 were neither differently distributed among the males, nor among the females (data not shown). Interesting, after considering age and estrogen exposure status, women had a 1.58-fold higher risk of getting bladder cancer than men in the Taiwanese population (data not shown).

The contribution of smoking to bladder cancer risk is not so obvious as in other types of cancer, such as upper urinary tract urothelial carcinoma (44). The joint effects of genetic variation and smoking on bladder cancer is seldom examined. In 2017, Fu and his colleagues have investigated the gene-smoking interaction on bladder cancer risk in a Chinese population (45). They proposed that a haplotype containing the rs2010963-C and rs833052-A alleles of the *vascular endothelial growth factor* (*VEGF*) gene is associated with increased bladder cancer risk, and the genotypes of *VEGF* rs2010963 have a joint effect with smoking status on determining bladder cancer risk (45). In literature, it has been reported that tobacco smoking can induce lots of DNA lesions in the cells and defects in repair of tobacco carcinogen-induced DNA adducts may contribute to carcinogenesis (46). Therefore, in this study, the joint effects of *NBS1* rs1805794 and smoking status were also examined and the results showed that ever smokers who carried the homologous CC genotypes at *NBS1* rs1805794 were of increased risk of bladder cancer after adjusted for age, gender, and alcohol drinking status. On the contrary, there was no significantly elevated bladder cancer risk for those non-smokers with CG or CC genotypes at *NBS1* rs1805794 (data not shown). A phenotypic assay showed that the rs1805794 C allele will attenuate the ability of the *NBS1* protein to repair DNA damage as the cells transfected with a plasmid carrying the rs1805794 C allele had a significantly higher number of DNA breaks than those transfected with a plasmid carrying the rs1805794 G allele after X-ray irradiation (47). To sum up, smoking behavior appears to be more strongly associated with bladder cancer risk in women than in men, which could be related to differences in metabolism, smoking behavior, exposure patterns, and DNA repair mechanisms (48).

In conclusion, our study provides evidence that the C allele of *NBS1* rs1805794 is not associated with an increased lung cancer risk among Taiwanese. Further investigations using the cells of patients with different gender, smoking status and genotypes can help to reveal the phenotypic role of *NBS1* in bladder carcinogenesis.

Conflicts of Interest

All Authors declare no conflict of interest regarding this study.

Authors' Contributions

Research Design: Chen M, Tsai CW and Chang WS; Patient and Questionnaire Summarize: Wu HC and Shen TC; Experiment Performance: Wang YC, Chen CH and Chang WS; Statistical Analysis: Wang CH, Lin ML and Gong CL; Manuscript Writing: Chen M, Tsai CW and Bau DT; Reviewing and Revising: Bau DT, Chang WS and Tsai CW.

Acknowledgements

The Authors thank the Tissue-Bank of China Medical University Hospital for their excellent technical assistance and all the subjects, doctors (under the leadership of Prof. Hsi-Chin Wu), nurses and colleagues. The excellent techniques and efforts from Yu-Chen Hsiao, Tzu-Yu Wang and Tzu-Hsuan Wang are highly appreciated. This study was supported majorly by China Medical University Hospital and Asia University (CMU108-ASIA-02).

References

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 68: 394-424, 2018. PMID: 30207593. DOI: 10.3322/caac.21492
- <https://www.hpa.gov.tw/Pages/List.aspx?nodeid=269>, from Taiwan Ministry of Health and Welfare Clinical Trial and Research Center of Excellence
- Yeh HL, Hsu SW, Chang YC, Chan TC, Tsou HC, Chang YC and Chiang PH: Spatial analysis of ambient PM2.5 exposure and bladder cancer mortality in Taiwan. *Int J Environ Res Public Health* 14: 508, 2017. PMID: 28489042. DOI: 10.3390/ijerph14050508
- Chiu HF, Chen BK and Yang CY: Parity, age at first birth, and risk of death from bladder cancer: a population-based cohort study in Taiwan. *Int J Environ Res Public Health* 13, 2016. PMID: 27918463. DOI: 10.3390/ijerph13121197
- Liao CH, Chang WS, Tsai CW, Hu PS, Wu HC, Hsu SW, Chen GL, Yueh TC, Shen TC, Hsia TC and Bau DT: Association of matrix metalloproteinase-7 genotypes with the risk of bladder cancer. *In Vivo* 32: 1045-1050, 2018. PMID: 30388078. DOI: 10.21873/invivo.11345
- Chen M, Tsai YT, Chang WS, Shih LC, Shen TC, Lin ML, Chao CY, Wang YC, Tsai CW and Bau DT: Association of caspase-8 genotypes with bladder cancer risk. *Anticancer Res* 39: 4767-4773, 2019. PMID: 31519577. DOI: 10.21873/anticancer.13660
- Tsai TH, Wang YM, Chang WS, Tsai CW, Wu HC, Hsu HM, Wang YC, Li HT, Gong CL, Bau DT and Li CY: Association of matrix metalloproteinase-8 genotypes with the risk of bladder cancer. *Anticancer Res* 38: 5159-5164, 2018. PMID: 30194163. DOI: 10.21873/anticancer.12838
- Chang WS, Liao CH, Tsai CW, Hu PS, Wu HC, Hsu SW, Hsiao CL, Hsu CH, Hung YW and Bau DT: Association of enhancer of zeste 2 (EZH2) genotypes with bladder cancer risk in Taiwan. *Anticancer Res* 36: 4509-4514, 2016. PMID: 27630289. DOI: 10.21873/anticancer.10997
- Chang WS, Tsai CW, Ji HX, Wu HC, Chang YT, Lien CS, Liao WL, Shen WC, Tsai CH and Bau DT: Associations of cyclooxygenase 2 polymorphic genotypes with bladder cancer risk in Taiwan. *Anticancer Res* 33: 5401-5405, 2013. PMID: 24324075.
- Chang CH, Chang CL, Tsai CW, Wu HC, Chiu CF, Wang RF, Liu CS, Lin CC and Bau DT: Significant association of an XRCC4 single nucleotide polymorphism with bladder cancer susceptibility in Taiwan. *Anticancer Res* 29: 1777-1782, 2009. PMID: 19443403.
- Burma S, Chen BP, Murphy M, Kurimasa A and Chen DJ: ATM phosphorylates histone H2AX in response to DNA double-strand breaks. *J Biol Chem* 276: 42462-42467, 2001. PMID: 11571274. DOI: 10.1074/jbc.C100466200
- Kobayashi J, Tauchi H, Sakamoto S, Nakamura A, Morishima K, Matsuura S, Kobayashi T, Tamai K, Tanimoto K and Komatsu K: NBS1 localizes to gamma-H2AX foci through Interaction with the FHA/BRCT domain. *Curr Biol* 12: 1846-1851, 2002. PMID: 12419185. DOI: 10.1016/s0960-9822(02)01259-9
- Takeda S, Hoa NN and Sasanuma H: The role of the Mre11-Rad50-Nbs1 complex in double-strand break repair-facts and myths. *J Radiat Res* 57 Suppl 1: i25-i32, 2016. PMID: 27311583. DOI: 10.1093/jrr/rww034
- Kitagawa R, Bakkenist CJ, McKinnon PJ and Kastan MB: Phosphorylation of SMC1 is a critical downstream event in the ATM-NBS1-BRCA1 pathway. *Genes Dev* 18: 1423-1438, 2004. PMID: 15175241. DOI: 10.1101/gad.1200304
- Lee JH and Paull TT: Activation and regulation of ATM kinase activity in response to DNA double-strand breaks. *Oncogene* 26: 7741-7748, 2007. PMID: 18066086. DOI: 10.1038/sj.onc.1210872
- Nakanishi K, Taniguchi T, Ranganathan V, New HV, Moreau LA, Stotsky M, Mathew CG, Kastan MB, Weaver DT and D'Andrea AD: Interaction of FANCD2 and NBS1 in the DNA damage response. *Nat Cell Biol* 4: 913-920, 2002. PMID: 12447395. DOI: 10.1038/ncb879
- Zhang Y, Zhou J and Lim CU: The role of NBS1 in DNA double strand break repair, telomere stability, and cell cycle checkpoint control. *Cell Res* 16: 45-54, 2006. PMID: 16467875. DOI: 10.1038/sj.cr.7310007
- Matsuura S, Kobayashi J, Tauchi H and Komatsu K: Nijmegen breakage syndrome and DNA double strand break repair by NBS1 complex. *Adv Biophys* 38: 65-80, 2004. PMID: 15493328.
- Matsuura S, Tauchi H, Nakamura A, Kondo N, Sakamoto S, Endo S, Smeets D, Solder B, Belohradsky BH, Der Kaloustian VM, Oshimura M, Isomura M, Nakamura Y and Komatsu K: Positional cloning of the gene for Nijmegen breakage syndrome. *Nat Genet* 19: 179-181, 1998. PMID: 9620777. DOI: 10.1038/549
- Carney JP, Maser RS, Olivares H, Davis EM, Le Beau M, Yates JR 3rd, Hays L, Morgan WF and Petrini JH: The hMre11/hRad50 protein complex and Nijmegen breakage syndrome: linkage of double-strand break repair to the cellular DNA damage response. *Cell* 93: 477-486, 1998. PMID: 9590181. DOI: 10.1016/s0092-8674(00)81175-7
- Dumon-Jones V, Frappart PO, Tong WM, Sajithlal G, Hulla W, Schmid G, Herceg Z, Digweed M and Wang ZQ: Nbn heterozygosity renders mice susceptible to tumor formation and ionizing radiation-induced tumorigenesis. *Cancer Res* 63: 7263-7269, 2003. PMID: 14612522.
- Zheng J, Zhang C, Jiang L, You Y, Liu Y, Lu J and Zhou Y: Functional NBS1 polymorphism is associated with occurrence and

- advanced disease status of nasopharyngeal carcinoma. *Mol Carcinog* 50: 689-696, 2011. PMID: 21656575. DOI: 10.1002/mc.20803
- 23 Chuang CL, Wang CH, Hsu CH, Hsiao CL, Chen GL, Yen ST, Li HT, Chang WS, Tsai CW, Wang SC and Bau DT: Contribution of double-strand break repair gene Nijmegen breakage syndrome 1 genotypes, gender difference and smoking status to Taiwanese lung cancer. *Anticancer Res* 37: 2417-2423, 2017. PMID: 28476809. DOI: 10.21873/anticancer.11581
- 24 Ryk C, Kumar R, Thirumaran RK and Hou SM: Polymorphisms in the DNA repair genes XRCC1, APEX1, XRCC3 and NBS1, and the risk for lung cancer in never- and ever-smokers. *Lung Cancer* 54: 285-292, 2006. PMID: 17034901. DOI: 10.1016/j.lungcan.2006.08.004
- 25 Park SL, Bastani D, Goldstein BY, Chang SC, Cozen W, Cai L, Cordon-Cardo C, Ding B, Greenland S, He N, Hussain SK, Jiang Q, Lee YC, Liu S, Lu ML, Mack TM, Mao JT, Morgenstern H, Mu LN, Oh SS, Pantuck A, Papp JC, Rao J, Reuter VE, Tashkin DP, Wang H, You NC, Yu SZ, Zhao JK and Zhang ZF: Associations between NBS1 polymorphisms, haplotypes and smoking-related cancers. *Carcinogenesis* 31: 1264-1271, 2010. PMID: 20478923. DOI: 10.1093/carcin/bgq096
- 26 Zhao JW, Ling XX and Yang L: Association of polymorphism 8360G>C in NBS1 gene and the risk of lung cancer in southern Chinese population. *Acad J Guangzhou Med Coll* 39: 5-8, 2011.
- 27 Forsti A, Angelini S, Festa F, Sanyal S, Zhang Z, Grzybowska E, Pamula J, Pekala W, Zientek H, Hemminki K and Kumar R: Single nucleotide polymorphisms in breast cancer. *Oncol Rep* 11: 917-922, 2004. PMID: 15010895.
- 28 Gil J, Ramsey D, Stembalska A, Karpinski P, Pesz KA, Laczmanska I, Leszczynski P, Grzebieniak Z and Sasiadek MM: The C/A polymorphism in intron 11 of the XPC gene plays a crucial role in the modulation of an individual's susceptibility to sporadic colorectal cancer. *Mol Biol Rep* 39: 527-534, 2012. PMID: 21559836. DOI: 10.1007/s11033-011-0767-5
- 29 Hebring SJ, Fredriksson H, White KA, Maier C, Ewing C, McDonnell SK, Jacobsen SJ, Cerhan J, Schaid DJ, Ikonen T, Autio V, Tammela TL, Herkommer K, Paiss T, Vogel W, Gielzak M, Sauvageot J, Schleutker J, Cooney KA, Isaacs W and Thibodeau SN: Role of the Nijmegen breakage syndrome 1 gene in familial and sporadic prostate cancer. *Cancer Epidemiol Biomarkers Prev* 15: 935-938, 2006. PMID: 16702373. DOI: 10.1158/1055-9965.EPI-05-0910
- 30 Li N, Xu Y, Zheng J, Jiang L, You Y, Wu H, Li W, Wu D and Zhou Y: NBS1 rs1805794G>C polymorphism is associated with decreased risk of acute myeloid leukemia in a Chinese population. *Mol Biol Rep* 40: 3749-3756, 2013. PMID: 23283743. DOI: 10.1007/s11033-012-2451-9
- 31 Broberg K, Bjork J, Paulsson K, Hoglund M and Albin M: Constitutional short telomeres are strong genetic susceptibility markers for bladder cancer. *Carcinogenesis* 26: 1263-1271, 2005. PMID: 15746160. DOI: 10.1093/carcin/bgi063
- 32 Figueroa JD, Malats N, Rothman N, Real FX, Silverman D, Kogevinas M, Chanock S, Yeager M, Welch R, Dosemeci M, Tardon A, Serra C, Carrato A, Garcia-Closas R, Castano-Vinyals G and Garcia-Closas M: Evaluation of genetic variation in the double-strand break repair pathway and bladder cancer risk. *Carcinogenesis* 28: 1788-1793, 2007. PMID: 17557904. DOI: 10.1093/carcin/bgm132
- 33 Sanyal S, Festa F, Sakano S, Zhang Z, Steineck G, Norming U, Wijkstrom H, Larsson P, Kumar R and Hemminki K: Polymorphisms in DNA repair and metabolic genes in bladder cancer. *Carcinogenesis* 25: 729-734, 2004. PMID: 14688016. DOI: 10.1093/carcin/bgh058
- 34 Hsu SW, Gong CL, Hsu HM, Chao CC, Wang YC, Chang WS, Tsai YT, Shih LC, Tsai CW and Bau DT: Contribution of matrix metalloproteinase-2 promoter genotypes to nasopharyngeal cancer susceptibility and metastasis in Taiwan. *Cancer Genomics Proteomics* 16: 287-292, 2019. PMID: 31243109. DOI: 10.21873/cgp.20133
- 35 Yueh TC, Hung YW, Shih TC, Wu CN, Wang SC, Lai YL, Hsu SW, Wu MH, Fu CK, Wang YC, Ke TW, Chang WS, Tsai CW and Bau DT: Contribution of murine double minute 2 genotypes to colorectal cancer risk in Taiwan. *Cancer Genomics Proteomics* 15: 405-411, 2018. PMID: 30194081. DOI: 10.21873/cgp.20099
- 36 Hu PS, Wang YC, Liao CH, Hsia NY, Wu MF, Yang JS, Yu CC, Chang WS, Bau DT and Tsai CW: The association of MMP7 genotype with pterygium. *In Vivo* 34: 51-56, 2020. PMID: 31882462. DOI: 10.21873/invivo.11744
- 37 Stern MC, Lin J, Figueroa JD, Kelsey KT, Kiltie AE, Yuan JM, Matullo G, Fletcher T, Benhamou S, Taylor JA, Placidi D, Zhang ZF, Steineck G, Rothman N, Kogevinas M, Silverman D, Malats N, Chanock S, Wu X, Karagas MR, Andrew AS, Nelson HH, Bishop DT, Sak SC, Choudhury A, Barrett JH, Elliot F, Corral R, Joshi AD, Gago-Dominguez M, Cortessis VK, Xiang YB, Gao YT, Vineis P, Sacerdote C, Guarnera S, Polidoro S, Allione A, Gurzau E, Koppova K, Kumar R, Rudnai P, Porru S, Carta A, Campagna M, Arici C, Park SS, Garcia-Closas M and International Consortium of Bladder C: Polymorphisms in DNA repair genes, smoking, and bladder cancer risk: findings from the international consortium of bladder cancer. *Cancer Res* 69: 6857-6864, 2009. PMID: 19706757. DOI: 10.1158/0008-5472.CAN-09-1091
- 38 Bryan RT, Evans T, Dunn JA, Iqbal G, Bathers S, Collins SI, James ND, Catto JWF and Wallace DMA: A Comparative Analysis of the Influence of Gender, Pathway Delays, and Risk Factor Exposures on the Long-term Outcomes of Bladder Cancer. *Eur Urol Focus* 1: 82-89, 2015. PMID: 28723362. DOI: 10.1016/j.euf.2015.01.001
- 39 Shariat SF, Sfakianos JP, Droller MJ, Karakiewicz PI, Meryn S and Bochner BH: The effect of age and gender on bladder cancer: a critical review of the literature. *BJU Int* 105: 300-308, 2010. PMID: 19912200. DOI: 10.1111/j.1464-410X.2009.09076.x
- 40 Thorstenson A, Hagberg O, Ljungberg B, Liedberg F, Jancke G, Holmang S, Malmstrom PU, Hosseini A and Jahnson S: Gender-related differences in urothelial carcinoma of the bladder: a population-based study from the Swedish National Registry of Urinary Bladder Cancer. *Scand J Urol* 50: 292-297, 2016. PMID: 27002743. DOI: 10.3109/21681805.2016.1158207
- 41 Bilski K, Zapala L, Skrzypczyk MA, Oszczudlowski M and Dobruch J: Review on gender differences in non-muscle invasive bladder cancer. *Transl Androl Urol* 8: 12-20, 2019. PMID: 30976563. DOI: 10.21037/tau.2018.11.06
- 42 Lucca I, Fajkovic H and Klatte T: Sex steroids and gender differences in nonmuscle invasive bladder cancer. *Curr Opin Urol* 24: 500-505, 2014. PMID: 24978392. DOI: 10.1097/MOU.0000000000000092
- 43 Gakis G and Stenzl A: Gender-specific differences in muscle-invasive bladder cancer: the concept of sex steroid sensitivity. *World J Urol* 31: 1059-1064, 2013. PMID: 23397433. DOI: 10.1007/s00345-013-1037-z

- 44 Wang YH, Yeh SD, Wu MM, Liu CT, Shen CH, Shen KH, Pu YS, Hsu LI, Chiou HY and Chen CJ: Comparing the joint effect of arsenic exposure, cigarette smoking and risk genotypes of vascular endothelial growth factor on upper urinary tract urothelial carcinoma and bladder cancer. *J Hazard Mater* 262: 1139-1146, 2013. PMID: 23009795. DOI: 10.1016/j.jhazmat.2012.08.056
- 45 Fu D, Li P, Cheng W, Tian F, Xu X, Yi X, Tang C, Wang Y, Hu Q and Zhang Z: Impact of vascular endothelial growth factor gene-gene and gene-smoking interaction and haplotype combination on bladder cancer risk in Chinese population. *Oncotarget* 8: 22927-22935, 2017. PMID: 28206971. DOI: 10.18632/oncotarget.15287
- 46 Wei Q, Cheng L, Amos CI, Wang LE, Guo Z, Hong WK and Spitz MR: Repair of tobacco carcinogen-induced DNA adducts and lung cancer risk: a molecular epidemiologic study. *J Natl Cancer Inst* 92: 1764-1772, 2000. PMID: 11058619. DOI: 10.1093/jnci/92.21.1764
- 47 Fang W, Qiu F, Zhang L, Deng J, Zhang H, Yang L, Zhou Y and Lu J: The functional polymorphism of NBS1 p.Glu185Gln is associated with an increased risk of lung cancer in Chinese populations: case-control and a meta-analysis. *Mutat Res* 770: 61-68, 2014. PMID: 25771871. DOI: 10.1016/j.mrfmmm.2014.07.009
- 48 Janisch F, Shariat SF, Schernhammer E, Rink M and Fajkovic H: The interaction of gender and smoking on bladder cancer risks. *Curr Opin Urol* 29: 249-255, 2019. PMID: 30888973. DOI: 10.1097/MOU.0000000000000602

Received February 15, 2020

Revised February 25, 2020

Accepted February 26, 2020