

Review

Application of Pharmacometrics of 5-Fluorouracil to Personalized Medicine: A Tool for Predicting Pharmacokinetic–Pharmacodynamic/Toxicodynamic Responses

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Abstract. *Recently, therapeutic drug monitoring of 5-fluorouracil (5-FU), the key chemotherapeutic drug for colorectal cancer, has been applied in daily clinical practice and has contributed towards improving clinical outcomes. However, current dose modifications are based only on values of the area under the plasma concentration–time profile, which are simply calculated from plasma 5-FU concentrations and infusion periods. When dose-limiting toxicities occur, the dosing is empirically reduced or discontinued, leading to treatment failure. To prevent this predictable failure and obtain better clinical outcomes, rational dosage-based strategies are required for 5-FU. Combining therapeutic drug monitoring with a mathematical approach using a pharmacokinetic–pharmacodynamic/toxicodynamic model is expected to help simulate time-course profiles of the efficacy of drugs and the degree of toxicity, thereby contributing towards dose setting for individual patients. Therefore, to facilitate pharmacometric modelling and simulation techniques for optimising current oncology therapies, this review focuses on pharmacometrics approaches for personalizing 5-FU-based chemotherapy.*

5-Fluorouracil (5-FU) has been used to treat patients for approximately 60 years and remains a cornerstone of colorectal cancer chemotherapy. The mechanism underlying the pharmacodynamic (PD) action of 5-FU involves its conversion

to fluoro-deoxyuridine monophosphate (FdUMP), which subsequently inhibits thymidylate synthase via the formation of a ternary complex comprising FdUMP, thymidylate synthase, and 5,10-methylene tetrahydrofolate in tumour cells, consequently inhibiting DNA synthesis. 5-FU is also incorporated into RNA and prevents protein synthesis (1). After it reaches the blood circulation, over 80% of administered 5-FU is metabolized by dihydropyrimidine dehydrogenase (DPD), which is a rate-limiting enzyme, in the liver (2, 3).

An appropriate 5-FU administration schedule has been developed to improve clinical responses in the clinical oncology setting. A meta-analysis showed that the infusion schedule of 5-FU is superior to the bolus administration schedule with respect to response rate and overall survival (4). Currently, bolus plus long-term infusion schedules, such as the folinic acid/5-FU/irinotecan (FOLFIRI) and the folinic acid/5-FU/oxaliplatin (FOLFOX) regimens, are standard approaches for treating adjuvant or metastatic colorectal cancer. However, infusion regimens involve long hospital stays for patients and require catheterization. To overcome these shortcomings of infusion therapy, orally available prodrugs of 5-FU have been developed, such as capecitabine, tegafur/gimeracil/oteracil (S-1), and tegafur/uracil (UFT). The capecitabine plus oxaliplatin (XELOX), S-1 plus oxaliplatin, and UFT plus leucovorin regimens are accepted for clinical chemotherapy of colorectal cancer.

Various administration schedules have been developed for 5-FU-based chemotherapy; however, there are large inter- and intra-individual pharmacokinetic (PK) variabilities which are important contributors to clinical treatment failure, and the method for determining the optimal 5-FU dose is debatable. The standard approach for 5-FU dose determination is based on the body surface area (BSA); however, this approach leads to large, approximately 100-fold variability, inter-individual variability in plasma 5-FU level (5-7). Many clinical studies have shown that PK-guided dose adjustments of 5-FU can improve clinical efficacies and reduce toxicity (8-10). Therapeutic drug

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monitoring (TDM) of the plasma 5-FU level is recommended for personalization of 5-FU dosing to obtain adequate systemic exposure, leading to improved clinical efficacy and lesser adverse effects. In 2018, the academic members of the International Association of Therapeutic Drug Monitoring and Clinical Toxicology reviewed the use of TDM for 5-FU infusion and strongly recommended TDM of 5-FU in clinical practice (11). While TDM of 5-FU is an important tool for determining the optimal dose for individual patients and obtaining appropriate systemic exposure, additional challenges still remain for achieving personalized 5-FU-based chemotherapy and further improved clinical outcomes. During dose adjustments by TDM, for instance, the 5-FU dose in the first cycle must be determined on the basis of BSA, whereas the doses in all subsequent cycles are adjusted on the basis of the plasma 5-FU level achieved. Therefore, the optimum target concentration can only be achieved after some chemotherapy cycles are completed. Moreover, several patient characteristics (*e.g.* sex, age, body weight, tumour type, cancer stage, DPD phenotype and activity level, and co-administered drugs) and circadian fluctuations of plasma 5-FU concentration make it difficult to adequately estimate drug exposure and to determine individual dose setting during TDM. When dose-limiting toxicities are observed, empirical dose reduction or treatment discontinuation is required, which leads to treatment failure. Therefore, an approach that would provide the best quantitative prediction of drug exposure, therapeutic response, and toxicities on the basis of the plasma 5-FU concentration needs to be developed.

In recent years, the pharmacometrics approach has attracted widespread interest in the field of oncology for achieving personalized medicine in clinical practice (12). Pharmacometrics is defined as “the science of developing and applying mathematical and statistical methods to (a) characterize, understand, and predict a drug’s pharmacokinetic and pharmacodynamic behaviour; (b) quantify uncertainty of information about that behaviour; and (c) rationalize data-driven decision making in the drug development process and pharmacotherapy” (13). Pharmacometric modelling and simulation techniques can help understand the relationship between drug exposure and the subsequent effects, which is particularly important for determining the appropriate dose setting for each patient (14). Almost all anticancer agents have narrow therapeutic windows, and there are large inter- and intra-individual variabilities in drug exposure. A balance between drug efficacy and adverse events is required for obtaining the desired clinical outcomes, especially in the field of oncology. A mathematical method such as the use of PK–PD/toxicodynamic (PK–PD/TD) models as part of the pharmacometrics approach would be valuable for predicting the time course of profiles of plasma drug level and PD/TD responses such that the optimal dose schedule can be prescribed for the patient in order to maximize drug efficacy

and minimize toxicity. Close collaboration between oncological physicians and pharmacometricians could lead to better prognosis on application of pharmacometrics in a clinical dose setting.

In this review, recent pharmacometrics approaches for the personalization of 5-FU-based chemotherapy in patients with cancer have been summarized. We have discussed pharmacometrics-related studies describing and simulating both 5-FU exposure and effects in various dose settings. This review focuses only on the PK–PD/TD model approach related to drug responses, including antitumor effects and dose-limiting toxicities, although there are many studies on classic compartmental PK models. Future perspectives on applying pharmacometrics to routinely collected clinical data and personalized medicine in order to optimize drug targeting are also discussed.

PK Model for Evaluating Circadian Variation

A circadian rhythm is observed in plasma 5-FU concentration during long-term infusion in clinical studies (15). Circadian fluctuations of plasma 5-FU level during constant infusion may be a major factor leading to incorrect PK and PD/TD estimation by the pharmacometrics approach; therefore, the current review also focused on the pharmacometrics approach in relation to the circadian rhythm of 5-FU PK. The circadian alterations in 5-FU PK might be derived from circadian variations in DPD activity and contribute to large inter- and intra-individual variations in plasma 5-FU concentration (16, 17). Conflicting results have been reported for the time points for peak and trough levels during the day (15). Harris *et al.* reported that plasma 5-FU levels obtained over a 24-h period reached peak values at 11:00 h and trough values at 23:00 h in patients who had cancer and were receiving continuous 5-FU infusion (300 mg m⁻² d⁻¹) (18). However, Metzger *et al.* reported that peak values were obtained at 04:00 h and trough values at 13:00 h in patients after 5-FU infusion (600 mg m⁻² d⁻¹) (19). Table I shows a previously reported PK model for evaluating and describing circadian changes in plasma 5-FU concentration. To the best of our knowledge, only one study has described a clinical PK model taking into account the circadian rhythm in plasma 5-FU concentration. Bressolle *et al.* defined the circadian model by the sum of two cosine cyclic components of 12- and 24-h periods and described circadian variations in 5-FU clearance (20). This circadian model was developed using 562 5-FU concentration datasets obtained for 65 patients and validated using another 104 datasets obtained for 20 patients. Analysis of this model revealed that the peak period for the plasma 5-FU concentration was approximately 04:00 h, consistent with previous results obtained by Metzger *et al.* (19). Moreover, the model was able to estimate 5-FU PK parameters for

Table I. Summary of pharmacokinetic (PK) model to describe circadian variations in plasma concentration of 5-fluorouracil (5-FU).

Author	Year of study	Administered drug(s)	Regimen	Software	Molecules studied	PK model	Circadian model
Clinical study Bressolle <i>et al.</i> (20)	1999	5-FU during CI	Folinic acid: 200 mg/m ² Bolus: 400 mg/m ² CI: 600 mg/m ² for 22 h Repeated on day 2 and on a 14-day cycle	NONMEM®	5-FU	1-CPT	$CL_{ss}=CL_{av}+CL_{A1}\times\cos\left[\frac{2\pi}{24}\times t-t_{z1}\right]+CL_{A2}\times\cos\left[\frac{2\pi}{12}\times t-t_{z2}\right]$
Animal studies Kobuchi <i>et al.</i> (24)	2015	5-FU	20 mg/kg	Phoenix® NLME™	5-FU	2-CPT	$CL=M+Am\times\cos[2\pi/24\times(t-t_{Ac})]$
Kobuchi <i>et al.</i> (25)	2018	Capecitabine	180 mg/kg	Phoenix® NLME™	Capecitabine 5'-DFCR 5'-DFUR 5-FU	1-CPT 1-CPT 1-CPT	$CL=V\times\{M+Am\times\cos[2\pi/24\times(t-t_{Ac})]\}$
Kobuchi <i>et al.</i> (26)	2018	UFT	15 mg/kg as FT	Phoenix® NLME™	FT 5-FU	1-CPT 1-CPT	$CL=V\times\{M+Am\times\cos[2\pi/24\times(t-t_{Ac})]\}$
Kobuchi <i>et al.</i> (27)	2020	5-FU during CI	Bolus: 60 mg/kg CI: 50 mg/m ² /h for 48 h	Phoenix® NLME™	5-FU	2-CPT	$CL=M+Am\times\cos[2\pi/24\times(t-t_{Ac})]$

Am: Amplitude; CI: continuous infusion; CL: clearance; CL_{ss}: steady-state time-varying clearance; CL_{av}: average clearance; CL_{A1} and CL_{A2}: the amplitude of the first and the second periodic component, respectively; CPT: tegafur; M: mesor; t_{Ac}: acrophase; t_{z1} and t_{z2}: acrophase (peak) times of the first and the second periodic components; UFT: uracil plus tegafur; V: distribution volume; 5'-DFCR: 5'-deoxy-5-fluorocytidine; 5'-DFUR: 5'-deoxy-5-fluorouridine.

individual patients, thereby contributing to optimization of the dose regimen of each patient. On the basis of these findings, a chronomodulated chemotherapy regimen has been proposed (21-23). However, clinical application of chronomodulation of 5-FU dosing is limited in the current standard regimen, which may be attributable to difficulties in estimating the circadian rhythm of each patient.

Various baseline characteristics in patients, including DPD activity level, may also affect circadian changes in plasma 5-FU levels. Recently, the circadian 5-FU patterns in animals were investigated using a PK model with the cosinor method (24-27) to exclude these potential contributing factors (Figure 1). In rats treated with continuous 5FU infusion (50 mg m⁻² h⁻¹) for 48 h, PK model analysis showed that the plasma 5-FU concentration followed a 24-h cosine circadian curve, representing an overall 1.8-fold increase from a nadir to a peak, with a relative amplitude (percentage of mesor) of 28% (27). Additionally, the loading bolus dose before initiating the infusion was found to contribute to circadian variations in plasma 5-FU level (27). These observations from animal studies suggest that in the recently modified regimen that omits bolus 5-FU injection, chronomodulation of dosing may enable sufficient clinical response with minimum toxicities and that timing of blood

sampling during TDM procedures should be determined cautiously. These animal study findings indicate that further clinical evaluations using a PK model that takes into account circadian variations in plasma 5-FU level are required for deciding appropriate dosing schedules and blood sampling times in TDM.

Similar to regimens proposed for continuous 5-FU infusion, chronomodulated regimens using orally available prodrugs of 5-FU, such as capecitabine and UFT, have been proposed for obtaining favourable antitumor effects because they can help avoid 5-FU elimination (28-31). Chronomodulation can easily be achieved in such regimens involving orally available drugs via drug self-administration by patients. However, there are conflicting results regarding the utility of chronomodulated chemotherapy involving oral 5-FU prodrugs. Pilancı *et al.* evaluated the usefulness of capecitabine morning and noon dosing as part of a first-line XELOX regimen in patients with metastatic colorectal cancer (31). Good clinical response with favourable toxicity profiles was obtained, indicating that chronomodulation of capecitabine dosing may provide a valuable therapeutic option. In contrast, Qvortrup *et al.* reported that the chronomodulated XELOX regimen did not improve clinical efficacy or reduce toxicity in their patients

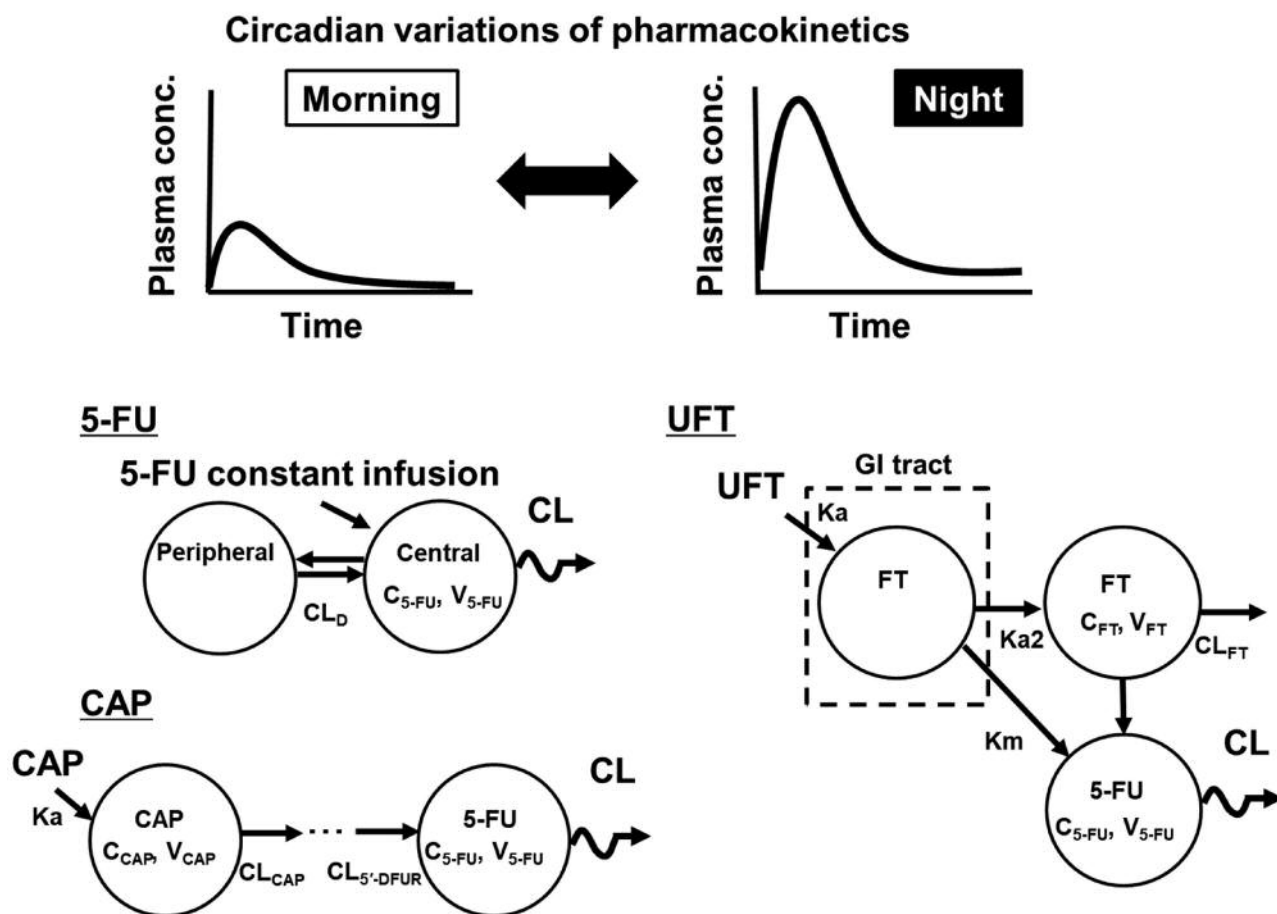


Figure 1. Schematic of the pharmacokinetic (PK) model taking into account the circadian rhythm of 5-fluorouracil (5-FU) after continuous infusion or of 5-FU prodrugs. To determine the circadian variations in plasma 5-FU concentrations, the cosinor method is applied in the PK model and 5-FU clearance (CL) is defined using the cosine curve. These models were developed in animal studies. CAP: Capecitabine; C_{CAP/FT/5-FU}: plasma concentration of CAP/tegafur/5-FU; CL: clearance of 5-FU; CL_{CAP}: CL of CAP; CL_D: intercompartmental clearance; CL_{FT}: clearance of FT; CL_{5'-DFUR}: CL of 5'-deoxy-5-fluorouridine; FT: tegafur; GI: gastrointestinal tract; Ka: absorption rate constant; K_m: conversion rate constant of FT into 5-FU during first-pass metabolism; Ka₂: conversion rate constant of the first pass metabolism of FT; UFT: FT/uracil; V_{CAP/FT/5-FU}: distribution volume of CAP/FT/5-FU.

(29). A recent phase I study showed that there were no circadian variations in exposure to capecitabine and its metabolites (5'-deoxy-5-fluorocytidine and 5'-deoxy-5-fluorouridine), including 5-FU, after continuous chronomodulated administration (32).

To investigate the usability of chronomodulated regimens involving capecitabine, circadian rhythmicity in the PK (*i.e.* chronopharmacokinetics) of capecitabine and its metabolites was evaluated in rats by using a population PK model with the cosinor method (25). Significant circadian variations were observed in the plasma concentration profiles of capecitabine, 5'-deoxy-5-fluorocytidine and 5'-deoxy-5-fluorouridine, and 5-FU according to the dosing time. Similar to the model for capecitabine, a population PK model taking into account the

circadian rhythm showed a circadian pattern of 5-FU clearance after UFT administration to rats (26); such a circadian pattern has not been clearly obtained in clinical studies (33, 34). These animal studies using PK models with circadian rhythm showed that circadian variations in the absorption of prodrugs and their sequential metabolism to 5-FU would also contribute to variations in plasma 5-FU level. These observations suggest that the administration time point for 5-FU prodrugs is a critical factor for achieving appropriate clinical outcomes in patients. A PK model that can help determine circadian variations by the cosinor method can provide evidence to support the development of a suitable dosing strategy for improving antitumor efficacy and minimizing severe toxicities.

PK-PD/TD Model of 5-FU

PK-PD model for antitumor effects. The PK-PD model for determining tumour size profiles after treatment with antitumor agents is a valuable tool for drug development and pre-clinical and clinical studies for establishing personalized medicine. Simeoni *et al.* successfully developed a PK-PD model of 5-FU for tumour growth dynamics in *in vivo* animal studies using xenograft models (35); this model is the standard model for evaluating and comparing the degree of antitumor effects in pre-clinical drug development. In this tumour growth model, an exponential tumour growth pattern is described; it assumes that the anticancer treatment inhibits the proliferation of some cells and eventually leads to their death. Inhibition of the tumour growth rate by 5-FU is described as a factor proportional to an index of 5-FU efficacy. A transit compartment model has been applied for describing the delayed antitumor effects of 5-FU; this model allows for prediction of time delay between drug administration and the observed effects. The tumour growth model has been modified according to the cell death mechanism noted after 5-FU exposure. On the basis of Simeoni *et al.*'s model (35), Sung *et al.* proposed new PD models connected to the physiologically based pharmacokinetic model in order to describe tumour cell growth after UFT administration: The cell cycle phase-specific model, and the dual-transit compartment model where two cell death pathways exist (36). They found that the dual-transit compartment model explained the tumour growth curves in animals well, suggesting that it can be used to develop dosing strategies and patient-specific 5-FU therapies. The tumour growth model developed by Simeoni *et al.* is a platform for investigating responses to drug exposure in the oncology field (35); therefore, this model has been used for analysing combination chemotherapy with other antitumor agents (37) and for translational research on 5-FU (38). Daryani *et al.* scaled a pre-clinical PK-PD model of 5-FU to children and simulated various 5-FU dosing strategies and tumour-growth inhibition in order to determine an appropriate 5-FU dosage for use in a clinical study involving children with ependymoma (38). This translational PK-PD approach is preferable for bridging pre-clinical and clinical studies and can be applied to developing new or optimizing existing dosing strategies for 5-FU.

PK-PD model along with biomarkers for PK and PD estimation for 5-FU. The endogenous DPD substrate uracil is metabolized to dihydrouracil (UH₂) in the liver. Because 5-FU is also metabolized by the same pathway, pre-therapeutic assessment of the plasma concentration ratio of UH₂ to uracil was proposed as a biomarker for estimating 5-FU clearance before its administration (16). Many clinical and animal studies have shown that the pre-therapeutic

UH₂/uracil ratio represents a valuable indirect biomarker that shows good correlation with hepatic DPD activities (16, 39, 40), 5-FU clearance (41, 42), and 5-FU-related toxicity (43-45). On the basis of the tumour-growth model developed by Simeoni *et al.* (35), a PK-PD model involving the UH₂/uracil ratio has been developed to analyse plasma 5-FU concentrations and tumour-growth inhibition in a rat model of 5-FU-treated colorectal cancer (46). In this model, the elimination rate constant of 5-FU was estimated using the plasma UH₂/uracil ratio before 5-FU treatment, and the estimated values were applied to the PK-PD model. A combination strategy involving predictive biomarkers and model-based estimations of the drug response may aid in determining individual 5-FU dosage.

PK-TD model for myelosuppression. Severe treatment-related toxicity occurs in approximately 10-30% of 5-FU-treated patients (47). Myelosuppression is one of the most frequent dose-limiting toxicities related to this treatment (48). Predicting time-course alterations in blood cell counts after 5-FU treatment in patients helps establish the dosing schedule for each patient, thereby preventing treatment discontinuation. Mathematical PK-PD modelling can help determine the relationship between drug exposure and myelotoxicities, thereby predicting the onset and degree of myelosuppression due to 5-FU treatment. Semi-physiological PK-PD models of 5-FU for myelosuppression were developed for both rats and humans (Table II) to determine the time course of alterations in blood cell counts after 5-FU administration.

The Friberg model is a standard and versatile model for investigating anticancer drug-induced myelosuppression (49). The original model was developed to determine time-course alterations in white blood cell counts due to 5-FU. The model consists of three types of compartments: Proliferative cell compartment, transit compartments with maturing cells, and circulating blood cell compartment. To explain the delay in onset of myelosuppression due to 5-FU, a transit compartment model was applied to the original model, that is, the Friberg model. Another feature of the Friberg model is the explanation of regulation of the haematological system by endogenous growth factors and cytokines as a feedback mechanism. This feedback was modelled as the ratio of circulating blood cell counts at baseline divided by the cell counts at time 't' raised to a feedback factor, which allows for description of the rebound of cells (overshoot compared with the baseline) after drug exposure. In clinical practice, the subsequent treatment course is generally initiated before the blood cell counts return to baseline, which makes it difficult to observe the rebound of cells after anticancer drug therapy and limits PD model development from clinical data sets. Therefore, Friberg *et al.* stated that this myelosuppression model should

Table II. Summary of pharmacokinetic-pharmacodynamic/toxicodynamic (PK-PD/TD) model of 5-fluorouracil (5-FU).

Author	Year of study	Administered drug(s)	Regimen	PD/TD data	Software	Molecules studied	Characteristics of the model
Clinical studies Zandvliet <i>et al.</i> (51)	2008	Capecitabine plus indisulan	Capecitabine/indisulan: 1000-1250 mg/m ² <i>b.i.d.</i> / 350-800 mg/m ²	Neutrophil and thrombocyte counts	NONMEM®	Indisulan Capecitabine 5'-DFCR 5'-DFUR 5-FU	PK-TD model for myelosuppression in the combination treatment
Friberg <i>et al.</i> (50)	2010	5-FU	Bolus 1500 mg/m ²	White blood cell count	NONMEM®	5-FU	Prediction of myelosuppression in patients by semi-physiological PK-TD model from rat data
Sáez-Belló <i>et al.</i> (57)	2020	Capecitabine	1250-2500 mg/m ² /24 h	Neutrophil counts	NONMEM®	Capecitabine 5'-DFUR 5-FU	Population PK-TD model of capecitabine for neutropenia based on the polymorphisms of the ATP-binding cassette gene and combination chemotherapy
Arshad <i>et al.</i> (58)	2020	5-FU	CI 650 or 1000 mg/m ² / day for 5 days	White blood cell count	NONMEM®	5-FU 5-Dihydro-5,6-dihydrouracil	Population semi-physiological PK-TD model in 5-FU monotherapy and covariate analysis on PK and myelotoxicity
Animal studies Friberg <i>et al.</i> (49)	2000	5-FU	3×49 mg/kg, 2×63 mg/kg, or 1×127 mg/kg Injection	Leucocyte count	NONMEM®	5-FU	First semi-physiological PK-PD model for leucopenia
Simeoni <i>et al.</i> (35)	2004	5-FU	50 mg/kg Every week	Tumour volume	WinNonlin®	5-FU	First PK-PD model of tumour growth kinetics in xenograft models
Sung <i>et al.</i> (36)	2008	UFT	15 or 150 mg/kg as FT	Timor volume	lsqnonlin	FT Uracil 5-FU	PK-PD model combined with physiologically based PK (PBPK) model
Terranova <i>et al.</i> (37)	2013	5-FU plus developed anticancer compound	5-FU: 50 mg/kg Every 4 days Drug C2: 45 mg/kg for 3 days and 2 cycle	Tumour volume	WinNonlin®	5-FU	PK-PD model of tumour growth kinetics in xenograft models after co-administration of two anticancer agents
Kobuchi <i>et al.</i> (46)	2013	5-FU	20 mg/kg For 7 days	Tumour volume	WinNonlin®	5-FU Uracil UH ₂	PK-PD model with plasma UH ₂ /uracil ratio, which is a possible surrogate biomarker of hepatic DPD activity

Table II. Continued

Table II. *Continued*

Author	Year of study	Administered drug(s)	Regimen	PD/TD data	Software	Molecules studied	Characteristics of the model
Kobuchi <i>et al.</i> (52-55)	2014 2015 2017	5-FU	5, 10, or 20 mg/kg For 4 days	Erythrocyte, thrombocyte, leucocyte, lymphocyte, and neutrophil counts	WinNonlin® Phoenix® NLME™	5-FU	Semi-physiological PK-PD model for myelosuppression using various blood cell counts
Daryani <i>et al.</i> (38)	2016	5-FU	75 mg/kg <i>i.v.</i> Bolus weekly for 4 weeks 75 mg/kg Subcutaneous infusion over 3 or 5 days every 3 weeks 37.5 or 75 mg/kg <i>i.v.</i> Bolus weekly for 4 weeks	Tumour volume	NONMEM®	5-FU (plasma and tumour extracellular concentration)	Translational PK-PD modelling and simulation from preclinical model to paediatric model
Krzyzansk <i>et al.</i> (56)	2019	5-FU	3×49 mg/kg, 2×63 mg/kg, or 1×127 mg/kg injection	White blood cell count	Phoenix®	5-FU	Application of the distributed delay model for the semi-physiological PK-PD model by Friberg <i>et al.</i> (49)

CI: Continuous infusion; DPD: dihydropyrimidine dehydrogenase; *i.v.*: intravenous; UFT: uracil plus tegafur; UH₂: dihydrouracil; 5'-DFCR: 5'-deoxy-5-fluorocytidine; 5'-DFUR: 5'-deoxy-5-fluorouridine.

preferably be developed from animal data (49). The authors tried to extrapolate the time course of alterations in leucocyte counts from rats to those in patients by using the myelosuppression model, while accounting for the differences in drug potency in the two species (50). To determine the alterations in different blood cell counts (thrombocytes and erythrocytes), several modified models were reported for both animals and patients after these studies by Friberg *et al.* (Figure 2) (51-58). The Friberg model (49) provides a robust platform for analysing myelotoxicity due to chemotherapy involving a prodrug of 5-FU or their combination with other drugs (59).

In recent research involving the semi-physiological PK-PD model, a distributed delay approach was applied to model delayed responses in PK-PD studies (56, 60), instead of the traditional transit compartment model approach (49). This traditional model includes a number of different equations and has been widely accepted in describing delayed PD/TD effects, including tumour regression and myelosuppression. Although this classic model approach can adequately capture features of drug effect data, it has some disadvantages, such as the requirement for manual analysis of the preferable number of transit compartments. Moreover, many differential equations in the transit compartment model also need to fit the observed data and are not preferable for use in complex compartment models.

To overcome these disadvantages, a distributed delay approach has been proposed (56, 60). This approach utilizes

an ordinary differential equation approximation of the convolution integral with gamma distribution for modelling the delay in drug absorption and the effects of drugs on myeloid cells, thereby avoiding the time-consuming process required for estimating appropriate model equations and parameters. Krzyzansk *et al.* (56) successfully applied the distributed delay model to previously reported myelosuppression data for FU-treated rats to which the Friberg model had been applied (49). Considering these advantages, instead of the transit compartment model, the distributed delay model should be applied as a standard model in oncology research for analysing the delays in pharmacological effects frequently observed after drug exposure in PK and PD/TD data.

Discussion and Future Perspectives

The current review discusses some modelling and simulation approaches for 5-FU that can be used to analyse drug responses after 5-FU treatment in patients; classic PK models have not been discussed. PK-PD modelling and simulation using clinical data have limitations because of the difficulties faced in collecting tumour size or drug response data from patients, whereas PK-TD model analysis uses routinely collected clinical data (*i.e.* blood cell counts) and is therefore relatively easy to perform. Moreover, 5-FU is administered along with other anticancer agents such as irinotecan and oxaliplatin and combination chemotherapy

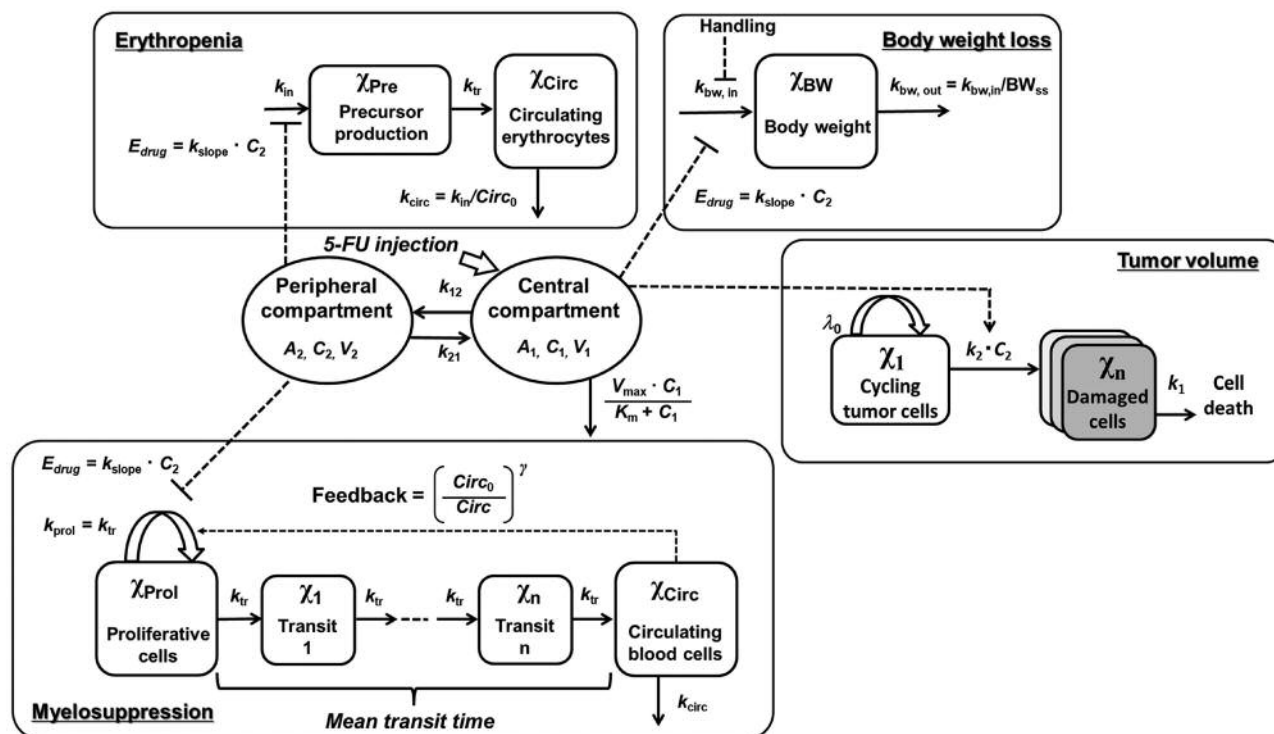


Figure 2. Reference schematic of the pharmacokinetic-pharmacodynamic/toxicodynamic (PK-PD/TD) model of 5-fluorouracil (5-FU) for analysing the tumour volume, myelosuppression, erythropenia, and body weight loss after 5-FU treatment. These basic models have been developed using animal data. A_1 : Amount of 5-FU in the central compartment; A_2 : amount of 5-FU in the peripheral compartment; BW_{SS} : maximal body weight; C_1 : 5-FU concentration in the central compartment; C_2 : 5-FU concentration in the peripheral compartment; $Circ$: circulating blood cell count; $Circ_0$: baseline value of circulating blood cells; E_{drug} : drug effects; k_1 , first-order rate constant of transit; k_{12} : rate constant of central compartment to peripheral compartment; k_2 , measure of drug potency; k_{21} : rate constant of peripheral compartment to central compartment; $k_{bw,in}$: rate constant describing the rate of body weight increase; $k_{bw,out}$: first-order rate constant describing the rate of body weight decrease; k_{circ} : degradation rate of circulating blood cells; k_{in} : precursor production rate; K_m : concentration of 5-FU when the rate of nonlinear elimination is at half its maximum value; k_{prol} : proliferation rate constant determining the rate of cell division; k_{slope} : slope of linear function in drug effect; k_{tr} : first-order rate constant of transit; V_1 : central volume of distribution of 5-FU; V_2 : peripheral volume of distribution of 5-FU; V_{max} : maximal rate of saturable metabolism; XBW : one compartment of observed body weight; X_{Circ} : one compartment of observed circulating blood cells; X_n : some transit compartments; X_{Pre} : precursor production compartment; X_{Pro} : one compartment that represented proliferative cells such as stem cells and progenitor cells; γ : power which describes a feedback mechanism from the circulating blood cells; λ_0 : the rate of exponential tumour growth.

can also make it difficult to isolate the PD/TD data for 5-FU. To develop these types of PK-PD/TD models that can be used to analyse tumour size, drug response and combination chemotherapy data, modelling and simulation with animal data are preferable and have been reported as summarized in Table II. However, models developed using animal data cannot be directly extrapolated to clinical practice. To achieve personalized dosing strategies, approaches involving translational PK-PD/TD modelling and simulation across species and model analysis using clinical data based on the results of the animal PK-PD/TD model are required. The approach used by Daryani *et al.*, who performed translational PK-PD modelling and simulation from a pre-clinical to paediatric model (as described in the PK-PD model for the subsection on

antitumor effects), may provide a framework for future studies on the translational PK-PD/TD modelling approach for optimal dosing strategies that can reduce toxicity while maintaining the chemotherapeutic effects of 5-FU (38).

Many clinical studies have shown that there are large inter- and intra-individual variabilities in plasma 5-FU level, which contribute to clinical treatment failure (10). Contributors to inter-individual variations include differences in chemotherapeutic regimens, for example, whether bolus dosing was used, and patient characteristics. PK analysis of clinical data revealed that the area under plasma concentration-time profile (AUC: 5-FU exposure) is strongly associated with clinical outcomes, including toxicity and efficacy (11). To date, a target AUC range of 20-30 mg h l⁻¹ in the infusion regimen has been proposed in the TDM

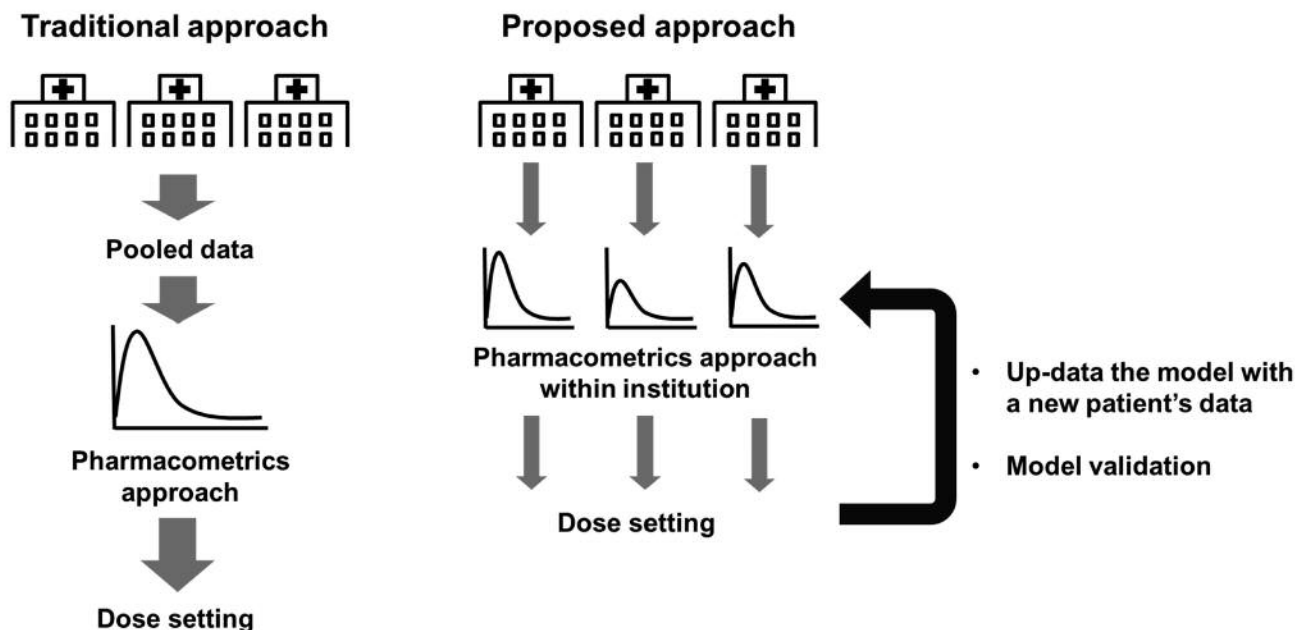


Figure 3. Summary of traditional and proposed prospective pharmacometrics-based approach for realizing personalized dose setting.

procedure, which is simply determined from the steady-state plasma concentration and infusion period of 5-FU (11, 61). However, using this simple method for calculating AUC can lead to over- or underestimation of the values because the circadian concentration is not considered. Simulating the time profiles of both plasma 5-FU concentration and clinical responses from blood sampling data and mathematical approaches, which would enable appropriate dosing modifications for each patient, remains a critical challenge. A PK-PD/TD model that can describe the circadian rhythm of 5-FU PK may aid in realizing this strategy.

Although TDM of 5-FU can reduce toxicity and improve clinical efficacy in long-term infusion regimens, a TDM strategy has not yet been established for 5-FU prodrugs. To elucidate the relationship between exposure to the drug and its toxic properties, some studies performed PK-TD model analysis of 5-FU prodrugs (51, 57). Recently, Oyaga-Iriarte *et al.* successfully developed a multicompartmental PK model for capecitabine and its metabolite in patients and determined optimal sampling times for capecitabine during TDM procedures (62). These proposed sampling times will help predict the PK of capecitabine in new patients, enabling dose adjustment. Chronomodulated chemotherapy of 5-FU prodrugs has also been proposed (28-31). To determine the diurnal cycle of PK properties, some circadian models were used in animal research (25, 26). These pharmacometrics data indicate that further clinical studies on treatment efficacy and toxicity during individualized treatment with

TDM need to be performed with large patient populations in order to realize the goal of personalized medicine using 5-FU prodrug chemotherapy.

Combination chemotherapy such as the FOLFIRI and FOLFOX regimens are standard approaches for treating colorectal cancer. Recently, the frequency of co-administration of drugs has increased; the folinic acid, 5-FU, irinotecan, and oxaliplatin (FOLFIRINOX) regimen has been used in chemotherapy for colorectal and pancreatic cancer (63). Moreover, supportive therapy for nausea and vomiting requires additional drugs, which complicates drug-drug interaction (64). A PK-PD/TD model that links drug exposure and drug response has been developed for 5-FU, but it has not been used for combination chemotherapy involving 5-FU (63). Developing a PK-PD/TD model for each combination chemotherapeutic regimen remains challenging.

Future studies should apply the PK-PD/TD model as a tool for determining 5-FU dosing in clinical practice. Although the currently used procedure for TDM of 5-FU therapy leads to improvement in efficacies and reduction in toxicity, drug responses cannot be predicted from the plasma 5-FU level. The current dose-adjustment method requires completion of a number of cycles of therapy to achieve a narrow target therapeutic range, and the 5-FU dosage to be used in the first cycle is determined on the basis of an empirical index, namely the BSA (10, 65). PK-PD/TD modelling and simulation can facilitate more appropriate clinical dose setting for each patient via close co-operation between physicians and

pharmacometricians. The traditional PK-PD/TD model has been developed using clinical data for large populations in multiple multi-institutional joint studies (Figure 3). However, there is bias with respect to patient background in the data from each clinical hospital (*i.e.* regionality, chronic disease, or hospitals specializing in dialysis, paediatrics, or transplantation). These different hospital characteristics may generate variabilities in the PK of 5-FU and affect dose setting. Therefore, PK-PD/TD models should be developed using routinely collected medical record data from clinical organizations. Use of a specific model for each clinical organization can help realize the goal of personalized medicine within the hospital. The model should be routinely updated with new patient information. Although the pharmacometrics technique has limitations such as model validations and education of clinical pharmacometricians, we believe that the clinical pharmacometrics approach can aid in determining the appropriate 5-FU dose to improve clinical outcomes.

Conclusion

The current review promotes the understanding of the PK-PD/TD model of 5-FU. Personalized medicine involving 5-FU dose setting by using the PK-PD/TD model has not yet been applied in clinical chemotherapy. The review discusses models and current pharmacometrics approaches for personalized medicine, and provides fundamental information for further development of the PK-PD/TD model and platform. This information can help establish rational dosage-based 5-FU dose setting for each combination chemotherapy regimen, which would enable improved prognosis in patients with cancer.

Conflicts of Interest

The Authors declare no competing interests.

Authors' Contributions

S.K.: Conception and study design, analysis, and interpretation of data, drafting of the article; Y.I.: Collection and interpretation of data, and revision of the article. All Authors approved the final version of the article.

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References

1 Longley DB, Harkin DP and Johnston PG: 5-Fluorouracil: Mechanisms of action and clinical strategies. *Nat Rev Cancer* 3(5): 330-338, 2003. PMID: 12724731. DOI: 10.1038/nrc1074

2 Pinedo HM and Peters GF: Fluorouracil: biochemistry and pharmacology. *J Clin Oncol*. 6(10): 1653-1664, 1988. PMID: 3049954. DOI: 10.1200/JCO.1988.6.10.1653

3 Naguib FNM, El Kouni MH and Cha S: Enzymes off Uracil Catabolism in Normal and Neoplastic Human Tissues. *Cancer Res* 45(11 pt 1): 5405-5412, 1985. PMID: 3931905.

4 Piedbois P: Efficacy of intravenous continuous infusion of fluorouracil compared with bolus administration in advanced colorectal cancer. *J Clin Oncol* 16(1): 301-308, 1998. PMID: 9440757. DOI: 10.1200/JCO.1998.16.1.301

5 Fety R, Rolland F, Barberi-Heyob M, Hardouin A, Campion L, Conroy T, Merlin JL, Rivière A, Perrocheau G, Etienne MC and Milano G: Clinical impact of pharmacokinetically-guided dose adaptation of 5-fluorouracil: results from a multicentric randomized trial in patients with locally advanced head and neck carcinomas. *Clin Cancer Res* 4(9): 2039-2045, 1998. PMID: 9748117.

6 Baker SD, Verweij J, Rowinsky EK, Donehower RC, Schellens JH, Grochow LB and Sparreboom A: Role of body surface area in dosing of investigational anticancer agents in adults, 1991-2001. *J Natl Cancer Inst* 94(24): 1883-1888, 2002. PMID: 12488482. DOI: 10.1093/jnci/94.24.1883

7 Undevia SD, Gomez-Abuin G and Ratain MJ: Pharmacokinetic variability of anticancer agents. *Nat Rev Cancer* 5(6): 447-458, 2005. PMID: 15928675. DOI: 10.1038/nrc1629

8 Gamelin E, Boisdron-Celle M, Delva R, Regimbeau C, Cailleux PE, Alleaume C, Maillat ML, Goudier MJ, Sire M, Person-Joly MC, Maigre M, Maillart P, Fety R, Burtin P, Lortholary A, Dumesnil Y, Picon L, Geslin J, Gesta P, Danquechin-Dorval E, Larra F and Robert J: Long-term weekly treatment of colorectal metastatic cancer with fluorouracil and leucovorin: Results of a multicentric prospective trial of fluorouracil dosage optimization by pharmacokinetic monitoring in 152 patients. *J Clin Oncol* 16(4): 1470-1478, 1998. PMID: 9552054. DOI: 10.1200/JCO.1998.16.4.1470

9 Milano G, Etienne MC, Renée N, Thyss A, Schneider M, Ramaioli A and Demard F: Relationship between fluorouracil systemic exposure and tumor response and patient survival. *J Clin Oncol* 12(6): 1291-1295, 1994. PMID: 8201391. DOI: 10.1200/JCO.1994.12.6.1291

10 Saif MW, Choma A, Salamone SJ and Chu E: Pharmacokinetically guided dose adjustment of 5-fluorouracil: A rational approach to improving therapeutic outcomes. *J Natl Cancer Inst* 101(22):1543-1552, 2009. PMID: 8201391. DOI: 10.1200/JCO.1994.12.6.1291

11 Beumer JH, Chu E, Allegra C, Tanigawara Y, Milano G, Diasio R, Kim TW, Mathijssen RH, Zhang L, Arnold D, Muneoka K, Boku N and Joerger M: Therapeutic drug monitoring in oncology: International Association of Therapeutic Drug Monitoring and Clinical Toxicology Recommendations for 5-Fluorouracil Therapy. *Clin Pharmacol Ther* 105(3): 598-613, 2019. PMID: 29923599. DOI: 10.1002/cpt.1124

12 Buil-Bruna N, López-Picazo JM, Martín-Algarra S and Trocóniz IF: Bringing model-based prediction to oncology clinical practice: A review of pharmacometrics principles and applications. *Oncologist* 21(2): 220-232, 2016. PMID: 26668254. DOI: 10.1634/theoncologist.2015-0322.

13 Ete EI and Williams PJ: Pharmacometrics: The Science of Quantitative Pharmacology. John Wiley & Sons, Hoboken, USA, pp. 6, 2007.

- 14 Solans BP, Garrido MJ and Trocóniz IF: Drug exposure to establish pharmacokinetic-response relationships in oncology. *Clin Pharmacokinet* 59(2): 123-135, 2020. PMID: 31654368. DOI: 10.1007/s40262-019-00828-3
- 15 Fleming GF, Schumm P, Friberg G, Ratain MJ, Njiaju UO and Schilsky RL: Circadian variation in plasma 5-fluorouracil concentrations during a 24 hour constant-rate infusion. *BMC Cancer* 15: 69, 2015. PMID: 25885822. DOI: 10.1186/s12885-015-1075-6.
- 16 Jiang H, Lu J and Ji J: Circadian rhythm of dihydrouracil/uracil ratios in biological fluids: A potential biomarker for dihydropyrimidine dehydrogenase levels. *Br J Pharmacol* 141(4): 616-623, 2004. PMID: 14744810. DOI: 10.1038/sj.bjp.0705651
- 17 Kuwahara A, Yamamori M, Nishiguchi K, Okuno T, Chayahara N, Miki I, Tamura T, Kadoyama K, Inokuma T, Takemoto Y, Nakamura T, Kataoka K and Sakaeda T: Effect of dose-escalation of 5-fluorouracil on circadian variability of its pharmacokinetics in Japanese patients with stage III/IVa esophageal squamous cell carcinoma. *Int J Med Sci* 7(1): 48-54, 2010. PMID: 20151048. DOI: 10.7150/ijms.7.48
- 18 Harris BE, Song R, Soong SJ and Diasio RB: Relationship between dihydropyrimidine dehydrogenase activity and plasma 5-fluorouracil levels with evidence for circadian variation of enzyme activity and plasma drug levels in cancer patients receiving 5-fluorouracil by protracted continuous infusion. *Cancer Res* 50(1): 197-201, 1990. PMID: 2293556.
- 19 Metzger G, Massari C, Etienne MC, Comisso M, Brienza S, Touitou Y, Milano G, Bastian G, Misset JL and Lévi F: Spontaneous or imposed circadian changes in plasma concentrations of 5-fluorouracil coadministered with folinic acid and oxaliplatin: Relationship with mucosal toxicity in patients with cancer. *Clin Pharmacol Ther* 56(2):190-201, 1994. PMID: 8062496. DOI: 10.1038/clpt.1994.123
- 20 Bressolle F, Joulia JM, Pinguet F, Ychou M, Astre C, Duffour J and Gomeni R: Circadian rhythm of 5-fluorouracil population pharmacokinetics in patients with metastatic colorectal cancer. *Cancer Chemother Pharmacol* 44(4): 295-302, 1999. PMID: 10447576. DOI: 10.1007/s002800050980
- 21 Altinok A, Lévi F and Goldbeter A: Identifying mechanisms of chronotolerance and chronoefficacy for the anticancer drugs 5-fluorouracil and oxaliplatin by computational modeling. *Eur J Pharm Sci* 36(1): 20-38, 2009. PMID: 19041394. DOI: 10.1016/j.ejps.2008.10.024
- 22 Lévi F, Karaboué A, Etienne-Grimaldi MC, Pintaud G, Focan C, Innominato P, Bouchahda M, Milano G and Chatelut E: Pharmacokinetics of irinotecan, oxaliplatin and 5-fluorouracil during hepatic artery chronomodulated infusion: A translational European OPTILIV study. *Clin Pharmacokinet* 56(2): 165-177, 2017. PMID: 27393140. DOI: 10.1007/s40262-016-0431-2
- 23 Hill RJW, Innominato PF, Lévi F and Ballesta A: Optimizing circadian drug infusion schedules towards personalized cancer chronotherapy. *PLoS Comput Biol* 16(1): e1007218, 2020. PMID: 31986133. DOI: 10.1371/journal.pcbi.1007218
- 24 Kobuchi S, Ito Y, Nakano Y and Sakaeda T: Population pharmacokinetic modelling and simulation of 5-fluorouracil incorporating a circadian rhythm in rats. *Xenobiotica* 46(7): 597-604, 2016. PMID: 26503235. DOI: 10.3109/00498254.2015.1100767
- 25 Kobuchi S, Yazaki Y, Ito Y and Sakaeda T: Circadian variations in the pharmacokinetics of capecitabine and its metabolites in rats. *Eur J Pharm Sci* 112: 152-158, 2018. PMID: 29175408. DOI: 10.1016/j.ejps.2017.11.021
- 26 Kobuchi S, Ito Y, Takamatsu D and Sakaeda T: Circadian variations in the pharmacokinetics of the oral anticancer agent tegafur-uracil (UFT) and its metabolites in rats. *Eur J Pharm Sci* 123: 452-458, 2018. PMID: 30077713. DOI: 10.1016/j.ejps.2018.08.004
- 27 Kobuchi S, Matsumura E, Ito Y and Sakaeda T: Population pharmacokinetic model-based evaluation of circadian variations in plasma 5-fluorouracil concentrations during long-term infusion in rats: A comparison with oral anticancer prodrugs. *J Pharm Sci* 109(7): 2356-2361, 2020. PMID: 32311368. DOI: 10.1016/j.xphs.2020.04.005
- 28 Santini D, Vincenzi B, Schiavon G, Di Seri M, Virzì V, Spalletta B, Caricato M, Coppola R and Tonini G: Chronomodulated administration of oxaliplatin plus capecitabine (XELOX) as first line chemotherapy in advanced colorectal cancer patients: phase II study. *Cancer Chemother Pharmacol* 59(5): 613-620, 2007. PMID: 16944151. DOI: 10.1007/s00280-006-0302-x
- 29 Qvortrup C, Jensen BV, Fokstuen T, Nielsen SE, Keldsen N, Glimelius B, Bjerregaard B, Mejer J, Larsen FO and Pfeiffer P: A randomized study comparing short-time infusion of oxaliplatin in combination with capecitabine XELOX(30) and chronomodulated XELOX(30) as first-line therapy in patients with advanced colorectal cancer. *Ann Oncol* 21(1): 87-91, 2010. PMID: 19622596. DOI: 10.1093/annonc/mdp272
- 30 Akgun Z, Saglam S, Yucel S, Gural Z, Balik E, Cipe G, Yildiz S, Kilickap S, Okyar A and Kaytan-Saglam E: Neoadjuvant chronomodulated capecitabine with radiotherapy in rectal cancer: A phase II brunch regimen study. *Cancer Chemother Pharmacol* 74(4): 751-756, 2014. PMID: 25102935. DOI: 10.1007/s00280-014-2558-x
- 31 Pilancı KN, Saglam S, Okyar A, Yucel S, Pala-Kara Z, Ordu C, Namal E, Ciftci R, Iner-Koksall U and Kaytan-Saglam E: Chronomodulated oxaliplatin plus Capecitabine (XELOX) as a first line chemotherapy in metastatic colorectal cancer: A phase II Brunch regimen study. *Cancer Chemother Pharmacol* 78(1): 143-150, 2016. PMID: 27270460. DOI: 10.1007/s00280-016-3067-x
- 32 Roosendaal J, Jacobs BAW, Pluim D, Rosing H, de Vries N, van Werkhoven E, Nuijen B, Beijnen JH, Huitema ADR, Schellens JHM and Marchetti S: Phase I pharmacological study of continuous chronomodulated capecitabine treatment. *Pharm Res* 37(5): 89, 2020. PMID: 32382808. DOI: 10.1007/s11095-020-02828-6
- 33 Muggia FM, Wu X, Spicer D, Groshen S, Jeffers S, Leichman CG, Leichman L and Chan KK: Phase I and pharmacokinetic study of oral UFT, a combination of the 5-fluorouracil prodrug tegafur and uracil. *Clin Cancer Res* 2(9): 1461-1467, 1996. PMID: 9816321.
- 34 Etienne-Grimaldi MC, Cardot JM, François E, Renée N, Douillard JY, Gamelin E and Milano G: Chronopharmacokinetics of oral tegafur and uracil in colorectal cancer patients. *Clin Pharmacol Ther* 83(3): 413-415, 2008. PMID: 17637782. DOI: 10.1038/sj.clpt.6100297
- 35 Simeoni M, Magni P, Cammia C, De Nicolao G, Croci V, Pesenti E, Germani M, Poggesi I and Rocchetti M: Predictive pharmacokinetic-pharmacodynamic modeling of tumor growth kinetics in xenograft models after administration of anticancer agents. *Cancer Res* 64(3): 1094-1101, 2004. PMID: 14871843. DOI: 10.1158/0008-5472.can-03-2524

- 36 Sung JH, Dhiman A and Shuler ML: A combined pharmacokinetic-pharmacodynamic (PK-PD) model for tumor growth in the rat with UFT administration. *J Pharm Sci* 98(5): 1885-1904, 2009. PMID: 18803264. DOI: 10.1002/jps.21536
- 37 Terranova N, Germani M, Del Bene F and Magni P: A predictive pharmacokinetic-pharmacodynamic model of tumor growth kinetics in xenograft mice after administration of anticancer agents given in combination. *Cancer Chemother Pharmacol* 72(2): 471-482, 2013. PMID: 23812004. DOI: 10.1007/s00280-013-2208-8
- 38 Daryani VM, Patel YT, Tagen M, Turner DC, Carcaboso AM, Atkinson JM, Gajjar A, Gilbertson RJ, Wright KD and Stewart CF: Translational pharmacokinetic-pharmacodynamic modeling and simulation: Optimizing 5-fluorouracil dosing in children with pediatric ependymoma. *CPT Pharmacometrics Syst Pharmacol* 5(4): 211-221, 2016. PMID: 27104090. DOI: 10.1002/psp4.12075
- 39 Kobuchi S, Ito Y, Okada K, Imoto K, Kuwano S and Takada K: Pre-therapeutic assessment of plasma dihydrouracil/uracil ratio for predicting the pharmacokinetic parameters of 5-fluorouracil and tumor growth in a rat model of colorectal cancer. *Biol Pharm Bull* 36(6): 907-916, 2013. PMID: 23575271. DOI: 10.1248/bpb.b12-00819
- 40 Kobuchi S, Kuwano S, Imoto K, Okada K, Nishimura A, Ito Y, Shibata N and Takada K: A predictive biomarker for altered 5-fluorouracil pharmacokinetics following repeated administration in a rat model of colorectal cancer. *Biopharm Drug Dispos* 34(7): 365-376, 2013. PMID: 23836081. DOI: 10.1002/bdd.1851
- 41 Gamelin E, Boisdrón-Celle M, Guérin-Meyer V, Delva R, Lortholary A, Genevieve F, Larra F, Ifrah N and Robert J: Correlation between uracil and dihydrouracil plasma ratio, fluorouracil (5-FU) pharmacokinetic parameters, and tolerance in patients with advanced colorectal cancer: A potential interest for predicting 5-FU toxicity and determining optimal 5-FU dosage. *J Clin Oncol* 17(4): 1105, 1999. PMID: 10561167. DOI: 10.1200/JCO.1999.17.4.1105
- 42 Jiang H, Lu J, Jiang J and Hu P. Important role of the dihydrouracil/uracil ratio in marked interpatient variations of fluoropyrimidine pharmacokinetics and pharmacodynamics. *J Clin Pharmacol* 44(11): 1260-1272, 2004. PMID: 15496644. DOI: 10.1177/0091270004268911.
- 43 Sistonen J, Büchel B, Froehlich TK, Kummer D, Fontana S, Joerger M, van Kuilenburg AB and Largiadèr CR: Predicting 5-fluorouracil toxicity: DPD genotype and 5,6-dihydrouracil:uracil ratio. *Pharmacogenomics* 15(13): 1653-1666, 2014. PMID: 25410891. DOI: 10.2217/pgs.14.126
- 44 Zhou ZW, Wang GQ, Wan de S, Lu ZH, Chen YB, Li S, Chen G and Pan ZZ: The dihydrouracil/uracil ratios in plasma and toxicities of 5-fluorouracil-based adjuvant chemotherapy in colorectal cancer patients. *Chemotherapy* 53(2): 127-131, 2007. PMID: 17308379. DOI: 10.1159/000099984
- 45 Boisdrón-Celle M, Remaud G, Traore S, Poirier AL, Gamelin L, Morel A and Gamelin E: 5-Fluorouracil-related severe toxicity: A comparison of different methods for the pretherapeutic detection of dihydropyrimidine dehydrogenase deficiency. *Cancer Lett* 249(2): 271-282, 2007. PMID: 17064846. DOI: 10.1016/j.canlet.2006.09.006
- 46 Kobuchi S, Ito Y, Okada K, Imoto K, Kuwano S and Takada K: Pharmacokinetic/pharmacodynamic modeling of 5-fluorouracil by using a biomarker to predict tumor growth in a rat model of colorectal cancer. *J Pharm Sci* 102(6): 2056-2067, 2013. PMID: 23592368. DOI: 10.1002/jps.23547
- 47 Meulendijks D, Henricks LM, Jacobs BAW, Aliev A, Deenen MJ, de Vries N, Rosing H, van Werkhoven E, de Boer A, Beijnen JH, Mandigers CMPW, Soesan M, Cats A and Schellens JHM: Pretreatment serum uracil concentration as a predictor of severe and fatal fluoropyrimidine-associated toxicity. *Br J Cancer* 116(11): 1415-1424, 2017. PMID: 28427087. DOI: 10.1038/bjc.2017.94
- 48 Garg MB, Lincz LF, Adler K, Scorgie FE, Ackland SP and Sakoff JA. Predicting 5-fluorouracil toxicity in colorectal cancer patients from peripheral blood cell telomere length: A multivariate analysis. *Br J Cancer* 107(9): 1525-1533, 2012. PMID: 22990653. DOI: 10.1038/bjc.2012.421
- 49 Friberg LE, Freijs A, Sandström M and Karlsson MO: Semiphysiological model for the time course of leukocytes after varying schedules of 5-fluorouracil in rats. *J Pharmacol Exp Ther* 295(2): 734-740, 2000. PMID: 11046112.
- 50 Friberg LE, Sandström M and Karlsson MO: Scaling the time-course of myelosuppression from rats to patients with a semi-physiological model. *Invest New Drugs* 28(6): 744-753, 2010. PMID: 19711011. DOI: 10.1007/s10637-009-9308-7
- 51 Zandvliet AS, Siegel-Lakhai WS, Beijnen JH, Copalu W, Etienne-Grimaldi MC, Milano G, Schellens JH and Huitema AD: PK/PD model of indisulam and capecitabine: Interaction causes excessive myelosuppression. *Clin Pharmacol Ther* 83(6): 829-839, 2008. PMID: 17851564. DOI: 10.1038/sj.clpt.6100344
- 52 Kobuchi S, Ito Y, Hayakawa T, Kuwano S, Baba A, Shinohara K, Nishimura A, Shibata N and Takada K: Semi-physiological pharmacokinetic-pharmacodynamic modeling and simulation of 5-fluorouracil for the whole time course of alterations in leukocyte, neutrophil and lymphocyte counts in rats. *Xenobiotica* 44(9): 804-818, 2014. PMID: 24650147. DOI: 10.3109/00498254.2014.900588
- 53 Kobuchi S, Ito Y and Sakaeda T: Population pharmacokinetic-pharmacodynamic modeling of 5-fluorouracil for toxicities in rats. *Eur J Drug Metab Pharmacokinet* 42(4): 707-718, 2017. PMID: 27889876. DOI: 10.1007/s13318-016-0389-3
- 54 Kobuchi S, Ito Y, Hayakawa T, Nishimura A, Shibata N, Takada K and Sakaeda T: Semi-physiological pharmacokinetic-pharmacodynamic (PK-PD) modeling and simulation of 5-fluorouracil for thrombocytopenia in rats. *Xenobiotica* 45(1): 19-28, 2015. PMID: 25050790. DOI: 10.3109/00498254.2014.943335
- 55 Kobuchi S, Ito Y, Hayakawa T, Nishimura A, Shibata N, Takada K and Sakaeda T: Pharmacokinetic-pharmacodynamic (PK-PD) modeling and simulation of 5-fluorouracil for erythropenia in rats. *J Pharmacol Toxicol Methods* 70(2): 134-144, 2014. PMID: 25072509. DOI: 10.1016/j.vascn.2014.07.007
- 56 Krzyzanski W: Ordinary differential equation approximation of gamma distributed delay model. *J Pharmacokinet Pharmacodyn* 46(1): 53-63, 2019. PMID: 30617672. DOI: 10.1007/s10928-018-09618-z
- 57 Sáez-Belló M, Mangas-Sanjuán V, Martínez-Gómez MA, López-Montenegro Soria MÁ, Climente-Martí M and Merino-Sanjuán M: Evaluation of ABC gene polymorphisms on the pharmacokinetics and pharmacodynamics of capecitabine in colorectal patients: Implications for dosing recommendations. *Br J Clin Pharmacol*, 2020. PMID: 32559325. DOI: 10.1111/bcp.14441
- 58 Arshad U, Ploylearmsaeng SA, Karlsson MO, Doroshenko O, Langer D, Schömig E, Kunze S, Güner SA, Skripnichenko R, Ullah S, Jaehde U, Fuhr U, Jetter A and Taubert M: Prediction

- of exposure-driven myelotoxicity of continuous infusion 5-fluorouracil by a semi-physiological pharmacokinetic-pharmacodynamic model in gastrointestinal cancer patients. *Cancer Chemother Pharmacol* 85(4): 711-722, 2020. PMID: 32152679. DOI: 10.1007/s00280-019-04028-5
- 59 Fornari C, O'Connor LO, Yates JWT, Cheung SYA, Jodrell DI, Mettetal JT and Collins TA: Understanding hematological toxicities using mathematical modeling. *Clin Pharmacol Ther* 104(4): 644-654, 2018. PMID: 29604045. DOI: 10.1002/cpt.1080
- 60 Hu S, Dunlavey M, Guzy S and Teuscher N: A distributed delay approach for modeling delayed outcomes in pharmacokinetics and pharmacodynamics studies. *J Pharmacokinet Pharmacodyn* 45(2): 285-308, 2018. PMID: 29368268. DOI: 10.1007/s10928-018-9570-4
- 61 Kaldate RR, Haregewoin A, Grier CE, Hamilton SA and McLeod HL: Modeling the 5-fluorouracil area under the curve *versus* dose relationship to develop a pharmacokinetic dosing algorithm for colorectal cancer patients receiving FOLFOX6. *Oncologist* 17(3): 296-302, 2012. PMID: 22382460. DOI: 10.1634/theoncologist.2011-0357
- 62 Oyaga-Iriarte E, Insausti A, Bueno L, Sayar O and Aldaz A: Mining small routine clinical data: A population pharmacokinetic model and optimal sampling times of capecitabine and its metabolites. *J Pharm Pharm Sci* 22(1): 112-121, 2019. PMID: 30964613. DOI: 10.18433/jpps30392
- 63 Deyme L, Barbolosi D and Gattacceca F: Population pharmacokinetics of FOLFIRINOX: A review of studies and parameters. *Cancer Chemother Pharmacol* 83(1): 27-42, 2019. PMID: 30446786. DOI: 10.1007/s00280-018-3722-5
- 64 Schoffelen R, Lankheet AG, van Herpen CML, van der Hoeven JJM, Desar IME and Kramers C: Drug-drug interactions with aprepitant in antiemetic prophylaxis for chemotherapy. *Neth J Med* 76(3): 109-114, 2018. PMID: 29667586.
- 65 Gamelin E, Delva R, Jacob J, Merrouche Y, Raoul JL, Pezet D, Dorval E, Piot G, Morel A and Boisdron-Celle M: Individual fluorouracil dose adjustment based on pharmacokinetic follow-up compared with conventional dosage: results of a multicenter randomized trial of patients with metastatic colorectal cancer. *J Clin Oncol* 26(13): 2099-2105, 2008. PMID: 18445839. DOI: 10.1200/JCO.2007.13.3934

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