Review

Drug Delivery Systems for Phthalocyanines for Photodynamic Therapy

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Abstract. The focus of this review is to describe the state-ofart in the development of innovative drug delivery systems for phthalocyanines as photosensitizers for photodynamic therapy (PDT). PDT is a medical treatment combining photosensitizers (PSs) activated by visible light of a specific wavelength to selectively destroy targeted cells, tumor tissues and its surrounding vasculature. In the last decades, PDT has been under intense investigation, first as a promising alternative approach for improved cancer treatment, later against microbial infection and nowadays, mainly in aesthetic medicine, against age-related degeneration. The success of PDT is restricted because of difficulties with administration and skin permeation of PSs. As PDT importance raises, there is high interest for advanced formulations and delivery systems (DDS) for PS, especially formulations based on nanotechnology. Accordingly, this review deals with the innovations pertaining to DDS for PDT as disclosed in recent patents and literature.

The photodynamic therapy (PDT) was originally developed as a tumor therapy with improved selectivity towards diseased tissues in comparison to conventional cancer treatments (surgery, chemo- or radiotherapy). In addition to its primary use to destroy tumors without causing damage to surrounding healthy tissue, PDT has been successfully employed for the photosterilization of the tumor bed after surgical resection of a large neoplasm to support surgical treatment (1-3). Although PDT has multiple clinical applications, it is mainly used in oncology for the treatment of various types of solid tumors. The

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benefits of treating superficial oncologic lesions (tumor thickness <2-3 mm) using PDT are the low level of invasiveness and the excellent cosmetic results after treatment (4, 5). In dermatology, topical PDT is effective for dermatooncological or precancerous skin conditions like squamous cell carcinoma (6, 7) and superficial basal cell carcinoma (8-10), actinic keratosis (11, 12), Bowen's disease (4, 5, 13-15), mycosis fungoides (an indolent subtype of cutaneous T cell lymphoma) (9, 16-19), Kaposi's sarcoma, extramammary Paget's disease, and cutaneous B cell lymphoma (20) as well as for other proliferative disorders, such as vascular malformations (21) or keloid scars (22-24). In aesthetic dermatology PDT is extensively used for the treatment of inflammatory dermatoses that have a high psychological impact, like localized scleroderma (25, 26), acne vulgaris (27-29), rosacea (30, 31) and granuloma annulare (32-34), as well as for aesthetic indications like photo aged skin or sebaceous gland hyperplasia (4, 35, 36). PDT is useful for the treatment of various viral diseases such as warts (human papilomavirus) (37-39) or viral skin lesions (molluscum contagiosum and herpes simplex) (4, 5, 40), various mycotic diseases (41-43) or parasite diseases such as leishmaniasis (44-46) or bovine trichomoniasis (47). Currently, PDT is successfully used in ophthalmology for the treatment of age-related macular degeneration (48, 49).

PDT is used mainly for dermatology, however, light sources such as lasers can be coupled with fibre optic systems allowing acces to inaccessible locations, such as urinary bladder, digestive tract, brain or deep-seated tumors (3). PDT can be used in association with other therapeutic techniques such as surgery or chemotherapy. In the case of surgery for tumour resection, PDT can be used to help to destroy any remaining cancer cells after surgery. In the case of chemotherapy, several trials have demonstrated synergistic effects of the combination of PDT with low doses of chemotherapeutic drugs. The combination destroys cancer cells more efficiently and also reduces the side-effects of chemotherapy, due to the lower doses of chemotherapeutic drugs required to obtain the desired effect (3, 50, 51).

Photodynamic Therapy (PDT)

PDT uses visible light of a specific wavelength, for killing cells and tissues though activation of photosensitizers (PSs), that in the presence of molecular oxygen, generate reactive oxygen species (ROS) leading to oxidative cell damage and cell death (52, 53). Each component is harmless by itself, but, in combination they induce severe toxicity towards targeted cells and tissues. The mechanism of action of PDT is generally nonspecific, but there are two mechanisms leading to greater selectivity towards the selected targets. First, enhanced permeability and retention (EPR) effect causing preferential uptake and accumulation of PSs in rapidly dividing cells and diseased tissues. Second, restricted illumination to specific region/volume of tissue. Only cells that are exposed simultaneously to PS, light and oxygen are subjected to the cytotoxic ROS produced during PDT. Additionally, healthy cells are much more resistant against oxidative stress than cancerous cells. Combination of all the above allows destruction of tumor tissue while sparing surrounding healthy cells from damage. PDT usually damages surrounding blood vessels and thus prevents tumor from receiving nutrients, and may trigger an immune response towards cancer cells as well.

Basic Principles of PDT

The mechanism of PDT has been discussed in detail elsewhere (54-56). Briefly, when the drug (PS or its metabolic precursor) is administered (and metabolized to form PS in the case of the precursor), PS is activated by exposing the tissue that has accumulated PS to the light of a specific wavelength, usually in the maximum absorption band of the dye (1). The PS is excited from its ground state (54) (singlet state, no unpaired electron spins (57, 58)) and reaches a short-lived excited singlet state (54, 55). In a timescale of nanoseconds, PS releases its excess of energy by emitting a photon (fluorescence) or by internal conversion of energy (heat) and decay back to the ground state. Another mechanism involves the conversion to the excited triplet state via intersystem crossing including inversion of the spin of one electron (59) which has longer lifetime due to higher stability than the excited singlet state. Decay from the excited triplet state to the ground state is possible by emitting a photon (phosphorescence) or by internal conversion of energy (heat) or by transfer of energy to surrounding molecules. Interactions with the surroundings can follow two pathways named Type I and Type II reactions (Figure 1).

In Type I, the PS in its excited triplet state interacts with an organic cellular substrate transferring or acquiring one electron/hydrogen *via* the radical mechanism. It generates free radicals and radical ions and thus initiates chain reactions. After interaction with oxygen, PS can produce

highly reactive oxygen species (ROS), such as the superoxide anion O₂⁻, hydrogen peroxide H₂O₂, peroxide anions and highly reactive hydroxyl radical OH•, which then attack cellular targets and damage various cellular components (56, 60-62). However, oxygen is not necessarily required because original free radicals can cause cellular damage directly.

In Type II, the PS in its excited triplet state transfers its energy directly to molecular oxygen in its triplet ground state through energy transfer and yields highly reactive and cytotoxic singlet oxygen (63). This phenomenon is called triplet–triplet annihilation. Singlet oxygen has a short lifetime in a time-scale of microseconds, but a sufficient concentration of highly cytotoxic singlet oxygen induces irreversible cell damage (54, 55).

Both types of reactions take place at the same time and balance between them depends on the PS itself, however, it is considered that the prevailing mechanism during PDT is a Type II process (54, 56, 60-62, 64-66). After generating of cytotoxic ROS the molecule of PS can be destroyed by photobleaching due to oxidation (67) or can return to its ground state without chemical alteration and prepared to repeat excitation-energy transfer process multiple times (1, 59, 68-70).

Mechanism of Action on Cells

Increased levels of ROS exert their affects only in a small area around the region where they were generated, reported radius of action ranging within a 20-200 nanometers (2, 3, 71, 72), and ultimately causes cell death *via* necrosis, apoptosis or autophagy (73-76). The predominant mechanism depends on PS and light dose, cell type, and subcellular localization (site where ROS is generated determines which subcellular target is attacked due to radii of action in the order of dozens nanometres compared to diameters of human cells ranging from 10 to 100 μ m) (2, 77). PS usually act in the plasma membrane, mitochondria, endoplasmic reticulum, and lysosomes (3, 78-80). Produced ROS attack mainly DNA, protein thiol groups, and membrane lipids (3, 81).

Mechanism of Action on Tumor Tissues

The mechanisms of cell death caused by PDT are necrosis or apoptosis. The mechanism of tissue damage is more complex and depends on applied PS agent and treatment conditions. In general, molecules of certain sizes, especially lipophilic macromolecular drugs, as most PS agents, tend to selectively accumulate in tumor tissues. As tumor cells grow quickly, they stimulate the production of new blood vessels. These are usually abnormal in form and architecture, and they are formed by poorly aligned defective endothelial cells.

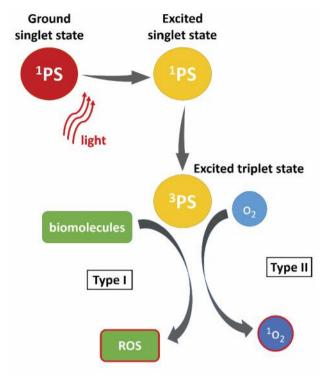


Figure 1. The mechanism of action of photodynamic therapy.

This leads to leaky vasculature. Other particular characteristics of tumors such as lack of effective lymphatic drainage, expression of specific enzymes and receptors, and pH variation play also role. All of these factors lead to accumulation of PS in tumor tissues and this phenomenon is called enhanced permeability and retention (EPR) effect. Disproportionately high numbers of low-density lipoprotein (LDL) receptors of tumour cell membranes and the fact that lipophilic PS agents are transported in the bloodstream bound to lipoproteins (such as LDLs) could enhance the accumulation of PS at close proximity to tumour cells (59, 82-84). PDT-induced changes in the vasculature may include vascular stasis, vascular leakage, or vessel collapse leading to ischaemic necrosis (3, 85-87).

Light Sources

PS is activated by exposing to the light of a specific wavelength, usually in the maximum absorption band of the PS (1). The stronger light absorption of PS at used wavelengths the higher quantum yields of excited states and potentially higher yields of ROS. However, for *in vivo* application PS requires a maximum of optical absorption at wavelengths higher than 650 nm due to strong absorption of endogenous pigments (such as hemoglobin, melanin). Absorption of endogenous pigments results in limited

penetration depth (usually 1-3 mm) of light of wavelengths in the region of 400-600 nm (3, 87, 88). To enhance penetration into tissue to access deep-seated tumors, it is necessary to use high-energy light sources (lasers) that can easily burn surface tissues. With increasing wavelength, penetration depth increases as well. In the red visible region (650-780 nm) and near-infrared region (>780 nm) light of low intensity can be used to achieve depths of several centimeters without healthy tissue damage (3, 87). Energy of singlet oxygen corresponds to 1,270 nm, however, compounds able to absorb in the region higher than 800 nm are in fact extremely rare. For such high absorption wavelengths, it is necessary to conjugate dozens of aromatic and/or incorporate some uncommon uncompatible) metal ions into the structure of the compound. Such compounds suffer of low solubility/bioavailability (unpolar structure) and toxicity (metal ions). Moreover light of these wavelengths tends to highly scatter in tissue (3). Practically, only the red region of the visible spectrum (650-780 nm) is available and is called "therapeutic transparency window" for PDT.

Porphyrin-based structures with extended aromatic systems such as phthalocyanines, naphthalocyanines, and benzoporphyrins have high absorption coefficients in the therapeutic transparency window and yield high levels of ROS. These, in combination with selective accumulation in tumor tissue (EPR effect), are the main advantages of these second-generation PSs.

Optionally lasers can be coupled with fiber optic systems allowing access to inaccessible locations, such as urinary bladder, digestive tract, brain or deep-seated tumors (3).

Photosensitizers

Although the first utilization of phototherapy for the treatment of diseases can be traced back over 4,000 years to the ancient Egyptians (89), contemporary PDT came first in the early twentieth century in the form of the first photosensitiser haematoporphyrin (Hp) which was isolated from blood and lately in the form of the purified haematoporphyrin derivative (HpD) used under the brand name Photofrin. The main disadvantages of this first-generation photosensitisers were weak absorption in the therapeutic transparency window and prolonged patient photosensitivity. This led to the development of improved PSs designed to minimize the drawbacks of the first generation PSs (90, 91).

Most of the second-generation PSs are porphyrin-like molecules (such as chlorins and bacteriochlorins), expanded porphyrins (such as texaphyrins), and structures with aromates fused to pyrrole rings (such as benzoporphyrins, phthalocyanines, and naphthalocyanines) or metabolic precursors of porphyrins (such as 5-aminolevulinic acid and

its esters). Although there exist numerous dyes and pigments displaying photosensitivity and effectively generate ROS, the PSs that are mostly used are only cyclic tetrapyrroles or structural derivatives of this chromophore. Cyclic tetrapyrrolic derivatives have little or no toxicity in the absence of light because of an inherent similarity to endogenous structures naturally occurring in human body. These compounds also display high absorption coefficients in the region of therapeutic transparency window, high yields in production of ROS, short serum half-life and selective tissue accumulation due to EPR effect (3, 92).

5-aminolevulinic acid (ALA) is not a photosensitizer by itself but is naturally occurring precursor in the biosynthetic pathway to photosensitive protoporphyrin IX (PpIX) (Figure 2). ALA is an endogenous metabolite that is synthesized usually in mitochondria from succinate-CoA and glycine. Conjugation of eight ALA molecules results of creation of PpIX which is subsequently metabolised and finally it results in formation of haem. Conversion of PpIX to its subsequent substrates requires activity of the enzyme ferrochelatase and this is the limiting factor concerning the rate of metabolism of ALA to haem. In case of external administration, the conversion of ALA to PpIX is quite fast but PpIX cannot be quickly converted by ferrochelatase to the final product haem and thus, PpIX accumulates in cells. Because PpIX is a strong photosensibilizer, this metabolic path has been frequently used in PDT. Kinetics of skin penetration and localisation can be modulated by using esters of ALA (such as methylester, hexylester or benzylester) which hydrolyze to form ALA and PpIX (93). This compound shows low toxicity and is rapidly cleared from the body by the existing mechanism. In addition, PpIX-induced clearance fluorescence can be visualized under blue light and can be employed in diagnosis (3, 94-98).

Porphyrins (Figure 3) are naturally-occurring intensely purple compounds with a porphine skeleton. Porphine frame consists of four pyrrolic sub-units linked on opposing sides through methine bridges resulting conjugated planar macrocycle. If substituted, compounds are known as porphyrins. The inner core can be deprotonated to form tetradentate dianionic chelators which can readily form complexes with most metal cations. Porphyrins display an intense absorption at around 400 nm (Soret band) followed by four weaker absorptions referred to as the Q bands (450-700 nm). Intensity of absorption in the region of the therapeutic transparency window can be modulated by exoskeleton substitution or by complexation with metal ligands into the centre of the macrocycle, however all porphyrins suffer relatively low extinction coefficients.

Porphyrins with reduced one exocyclic double bond are chlorins and after reduction of another exocyclic double bond bacteriochlorins are formed. Reduced bonds decrease the symmetry of the conjugated macrocycle which causes red-shift (650-680 nm for chlorins and 730-800 nm for bacteriochlorins) and 10-fold stronger absorption in the therapeutic transparency window for chlorins and 40-fold for bacteriochlorins. It is worth to mention that synthesis of them is difficult. Nowadays there are only representatives of chlorines (*e.g.* temoporfin under brand name Foscan, verteporfin under Visudyne and tin complex of etiopurpurin named Purlytin) commercially available or under clinical evaluation (91).

Expanded porphyrins with penta-aza core, called texaphyrins or motexafins, display strong absorption in the 730-770 nm region. These are usually in the form of metal complex with lutetium (motexafin lutetium known as texaphyrin, marketedas Lutex, Lutrin or Antrin) or gadolinium (motexafin gadolinium or gadolinium texaphyrin, marketed as Xcytrin) as potential radiosensitisers.

Porphyrin-like structures with aromates fused to pyrrole rings are mainly benzoporphyrins, phthalocyanines (PCs), and naphthalocyanines (NPCs) (Figure 3). PCs are synthetic macromolecules related to tetra-aza (porphyrazines) with fused benzene ring to each of four pyrrole subunits, which are linked by four nitrogen atoms (like porphyrazines macrocycle) instead of four bridging carbon atoms in porphyrin macrocycle. NPCs are extended PC derivatives with naphthalene rings fused to pyrrole subunits instead of benzene rings. These modifications lead to highly conjugated skeleton of PCs and NPCs and subsequently to a red-shift (680-780 nm for PCs and 740-780 nm for NPCs) and very high extinction coefficients (significantly higher with respect to all PSs described above). Since PCs and NPCs absorb long-wavelength light strongly, they can be used in small doses. PCs and NPCs are usually prepared in the form of complexes with metal cations coordinated to the centre of macrocycle (Figure 4). This is due to the fact that during synthesis transition, metal cation helps to close the ring to easily form a macrocycle. The photophysical properties of PCs and NPCs are strongly influenced by the presence and nature of the central metal ion. Co-ordination of diamagnetic transition metal ions (such as zinc, aluminium, and gallium) usually results to metallophthalocyanines (MPCs) with high singlet oxygen quantum yields (99-105).

Strong absorption in the near infrared (NIR) region makes NPCs candidates for highly pigmented tumours, including melanomas, however, problems associated with lower stability (decomposition in the presence of light and oxygen) and a tendency to form photoinactive aggregates in solution prohibit their clinical use. Contrary, PCs are chemically stable, resistant to chemical and photochemical degradation and easy to prepare (1). Nowadays only members of MPCs family are on the list of drugs in clinical trials (a sulphonated aluminum PC derivative named Photosense, a liposomal formulation of zinc phthalocyanine CGP55847, and silicon

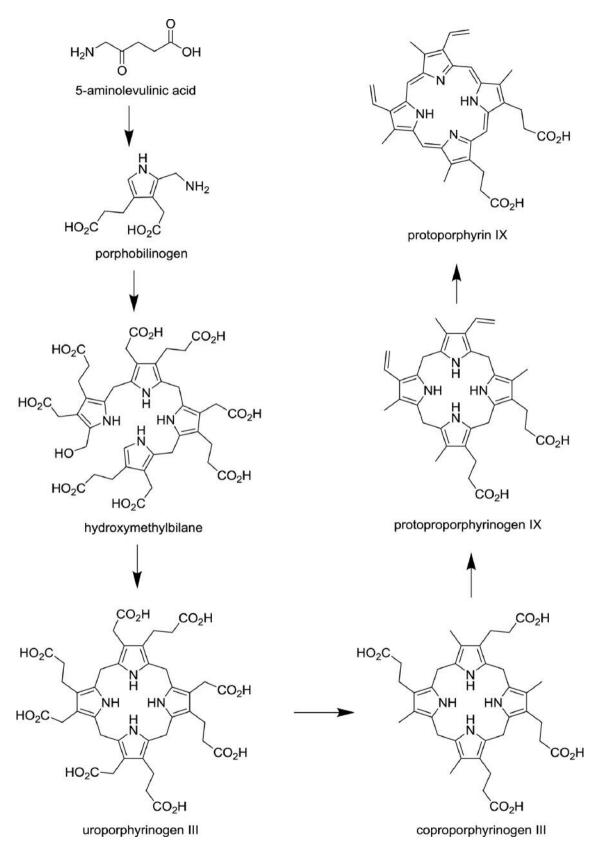


Figure 2. Biosynthetic pathway from ALA to protoporphyrin IX.

Figure 3. Structures of porphyrin-like compounds (chlorin, bacrteriochlorin, porphyrin, tetraazaporphyrin, tetrabenzoporphyrin, phthalocyanine and naphthalocyanine).

complex of PC known as PC4) but not PCs. General tendency of MPCs to aggregate in solution resulting in decrease of their bioavailability and photochemical activity can be overcome (minimized) using appropriate chemical modifications (sulphonated MPCs such as Photosense) or drug formulations (liposomal formulation of CGP55847).

Our interest and scope of this review is focused on the class of PCs and MPCs, their formulations and advanced drug delivery systems (DDSs).

Formulations and DDS of PCs and MPCs

PCs are composed of four isoindole units linked by nitrogen atoms forming a large planar aromatic system. They have the ability to form stable chelates with about sixty metal and metalloid ions inserted into central ring replacing two hydrogens of PC. The properties of these MPCs vary according to the nature of central ion and thus selection of central ion and synthetic modifications offer numerous options to control their physical properties. Closed shell diamagnetic PCs have higher yields and longer lifetimes of triplet states (such as Zn²⁺, Al³⁺, Ga³⁺) in comparison to paramagnetic PCs and therefore, are more appropriate for PDT (99). The type of chelated central ion influences tumor retention as well. Some of the central ions are used for specific treatment such as copper and uranyl (brain tumor accumulation) or radioactive Technecium-99 (Tc-99) MPCs (1, 106-110).

Delocalization of the π -electrons gives them their characteristic intensive blue, cyan or greenish color. These metalocomplexes are valuable as colorants in ink jet printing (CI Pigment Blue 15), laser printing (titanyloxy-PC) (111, 112), color filters for LCD screens (111, 113), in catalysis, organic photovoltaic cells, as semiconductors, optical data storage materials and rewritable optical media (CD-RW) (87, 103).

Unsubstituted PCs and their metalocomplexes suffer poor solubility in water and thus introduction of various peripheral (macrocycle) and axial (coordination to central metal ion) substitutions have been applied to alter their properties, mainly and increase their solubility, consequently hydrophilicity/lipophilicity (1, 114-118). Nevertheless, these PCs tend to dimerize and aggregate in aqueous media and thus lose their photoactivity in solution (only monomers are photoactive). If successfully administrated, PCs and MPCs bind to proteins and membranes and thus result in a complex photophysical behavior in the biological system (1). The advantage of solubilization through appropriate substitution allows direct biological administration without requiring an additional vehicle. In biological systems, PCs are diluted in the blood stream where they tend to disaggregate and bind to transport proteins (such as serum albumin) or lipoproteins (such as LDLs). Albumin has the ability to non-covalently bind molecules, not only to internal cavities, but also to the outer surface (119, 120) and thus albumin has physiological significance in the modulation of the activity of bounded

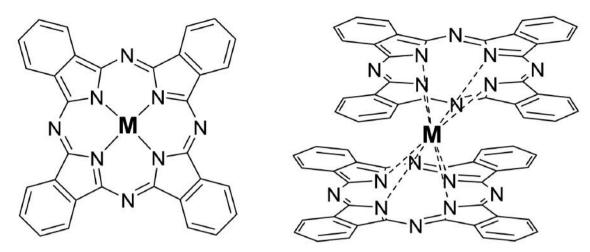


Figure 4. Structures of metallophthalocyanines (MPCs).

molecules and transport. Albumin delivers PCs to target tissue where they are absorbed using EPR effect (107). The high number of LDL receptors on tumour cell membranes enhance delivery efficacy to tumor tissue followed by the EPR effect (59, 82-84). Various MPCs non-covalently bound to albumin (121, 122) and LDL (122, 123) have been prepared. When one of these is administrated, redistribution of MPCs between albumin and LDL is expected. Covalent binding to albumin was described as well (124).

Typical peripheral substitutions leading to products with increased solubility and reduced tendency to aggregate are sulfonation, phosphonation (103, 125), glycosylation (68), carboxylation, or addition of other soluble substituents such as glucose, quarternary amino, hydroxyl or nitro groups to the periphery of macrocycle (1, 126). Sulfonation using fuming sulfuric acid leads to a mixture of many differently sulfonated PCs. The degree of sulfonation is inversely proportional to the degree of hydrophobicity and photodynamic activity of closed shell diamagnetic PCs (110, 127-131). Differently sulfonated aluminum complexes of PCs exhibit different subcellular localization (mono- and di- in cytoplasm while triand tetra-sulfonated in lysosomes) (132).

Axial substitution of central ion generally decrease tendency to aggregate due to sterical hindrance and also the nature of substitution modulates solubility and other properties. Suitable ions for this substitution are Al3+ with one or Si4+ and Ge4+, with two coordination sites. Note that opposite way unsubstituted MPCs with central ions such as silicon or iron tend to bind through oxo-bridges. Typical organic substituents are soluble polymers such as poly(ethylene glycol) or poly(vinyl alcohol). Shorter polymers tend to decrease the plasma half-life, but polymers with longer chains significantly prolong plasma half-life in comparison to unsubstituted metalophthalocyanine (133, 134).

Another option to increase solubility and bioavailability of hydrophobic PCs and their derivatives is their formulation using various Drug delivery systems. DDS can in principle provide enhanced efficacy and/or reduced toxicity of PS agents, deliver them to targeted tissues and allow their direct injection into the bloodstream. The most common and one of the simplest formulations is Cremophor oil emulsion and microemulsion. Cremophor formulations have been used for unsubstituted MPCs and MPCs with peripheral (fluorinated, hydroxylated) or axial substitutions (99).

More sofisticated colloidal carriers are micelles and liposomes. By chemical composition, liposomes are microscopic bilayer phospholipid vesicles similar to natural cell membranes separating an aqueous internal compartment from the bulk aqueous phase. When phospholipids dispersed in aqueous media unilamellar or multilamellar liposomes are spontaneously formed. Aqueous environment is an ideal medium for the existence of liposomes, rather than for microemulsions. Long circulating macromolecular carriers such as liposomes can exploit the EPR effect for preferential extravasation from tumor vessels (135). Their simple archetypal structures, facile preparation, controllable sizes and appropriate retention of PS incorporated into membranes explains why they are still popular as drug carriers. Examples of their utilization for MPCs include dipalmitoylphosphaditylcholine (DPPC), dipalmitoylphosphaditylglycerole (DPPG), 1-palmitoyl-2-oleoylphosphatidylcholine (POPC), 1,2-dioleoylphosphatidylserine (OOPS), DPPC/DPPG, DPPC/ cholesterol and egg-yolk lecithin liposomes (136-143).

Basic liposomal DDS provide passive targeting only and suffer from undesirable interactions with plasma proteins and cell membranes. Thus, liposome constructs featuring direct molecular targeting of cancer cells *via* antibody-mediated or the ligand-mediated interactions have been developed. This

represents an integration of biological components capable of tumor recognition with delivery technologies (135, 144, 145). The immunoliposome approach, in which monoclonal antibody (mAb) fragments are conjugated to liposomes, offers greater capacity of liposomes to load thousands of PSs in contrast to PS immunoconjugates (limited number of PSs linked to mAb) (135). A similar strategy for molecularly targeted drug delivery uses tumor-targeting peptides. Though direct conjugation of MPCs to tumor-targeting peptides has been described (62, 146), incorporation into liposomes is widely used (142, 143, 147-150), and the immunoliposome approach has been used in the formulation of many other drugs, not many PCs or MPCs have been formulated this way yet (151). Another approach used for topical applications to improve delivery of liposomes into viable dermis and epidermis are stratum corneum lipid liposomes (SCLLs). SCLLs composed of stratum corneum lipids use similar mechanisms with human cell to enhance interaction with skin (62, 152).

Micelles are closed monolayered vesicles with a hydrophobic core and a polar surface, typically formed by lipids with a fatty acid core or by polymers. An advantage of polymer micelles is their tunability allowing them to covalently bind targeting moieties, and to prepare thermoresponsive and/or hydrolytically degradable micelles to form advanced DDS (153, 154).

Polymer nanoparticles (PNPs), compared to liposomes, have a higher stability and their adjustable size and uniformity prevents recognition by macrophages and extends their circulation time in the bloodstream. While PSs are protected against enzymatic degradation, PNPs are able to penetrate into cells and allow controlled release of PSs (62, 155-158). Their properties could be tuned by the composition of polymer and by coating. Mostly used biocompatible biodegradable polymers are polylactide (PLA), polyglycolide (PGA), their copolymer poly(D,L-lactide-co-glycolide) (PLGA), polycaprolactone (PCL), poly(ethylene glycol) (PEG), chitosan, gelatin, β-glucan, as well as crosslinked polymers such as dioctyl sulfosuccinate with alginate (159) or alginate with cholesteryl residues (160). Mostly used nonbiodegradable PNPs are made of polyacrylamide (PAA), N-(2-hydroxypropyl)methacrylamide (HPMA) or polyalkylcyanoacrylates (PAC) polymers and copolymers (62, 161, 162). The advantages of PAA and HPMA PNPs include ease of synthesis and functionalization, and robustness of structure integrity (158, 163). PEG is often used for coating of particle surface to reduce uptake by the reticuloendothelial system (RES) after intravenous administration (a major drawback of PNPs) and thus prolong the plasma residence time. PSs can be physically entrapped in the core of PNPs with a hydrophilic periphery for efficient delivery to tumor cells in an aqueous environment. After successful delivery of PNP to tumor cells, the entrapped PS can be released (typical mechanism for biodegradable particles) or the porous structure of PNP may allow permeation of ROS and

interaction with the captured PS and thus maintaining their PDT performance in cells (158, 164). In case of drugs conjugated to polymer backbone, they can be cleaved enzymatically, hydrolytically or as a response of pH-sensitive bonds on change of pH (165). If targeting moieties such as oligopeptides are bound to the polymer, selective tumor uptake can be achieved (166). Examples of MPCs are PLA and PEG-coated-PLA PNPs containing perfluorinated MPCs (167) or poly(methyl vinyl ether-co-maleic anhydride) NPs which increased the singlet oxygen generation capacity of MPc by 10-fold compared to its free form (62, 168).

Other nanoparticle approaches include solid lipid nanoparticles (SLN), modified silica nanoparticles (SiNPs) (162) or metallic nanoparticles (MNPs). MNPs are functionalized mainly by gold (AuNPs) with covalently or non-covalently conjugated PSs (62, 169). These approaches have been described for MPCs, however, they are not widely used (170, 171). One specific type of extensively studied solid nanoparticles is upconversion nanoparticles (UCNPs).

UCNPs exhibit photon upconversion, which means that two or more incident photons of lower energy (typically in the infrared region) are absorbed and converted into one emitted photon with higher energy (typically in the visible or ultraviolet regions of the electromagnetic spectrum). Mostly used are fluorides of rare earths (group of elements consisting of Sc, Y, and the lanthanides) which are well known for their non-linear optical properties as downconversion or upconversion (172-175). Basically, the most of the rare earths have f-orbital partially filled with electrons and thus crystal structures of mixed rare earth fluorides (it means containing particular trivalent ions in various amounts in single crystal lattice) feature a wealth of electronic transitions within the 4f electron shells and transitions between closely matched intermediate-excited states of particular ions. Typically, compounds of general formula AREF4 (A=alkali, RE=rear earth), e.g. NaYF4 doped with metals like Er or Yb, are used (176-182). These mixed rare earths nanofluorides can be prepared by the broad variety of methods (183-189). Low systemic toxicity and cytotoxicity and negligible solubility in aqueous media make these materials suitable for medical applications. Their use for advanced drug delivery systems in anticancer treatment (190, 191) and advanced photodynamic therapy is very promising (178, 192). To achieve biocompatibility and water dispersibility, they are commonly subjected to surface modification with silica, porous silica, biocompatible polymers or hydrophilic functional groups (193). The main advantages of these approaches are possibility to connect tumor-targeting biomolecules and covalently bind or encapsulate molecules of PS such as various PCs and NPCs (194, 195). After the delivery of nanoparticles into the cancer tissue, light with deep tissue penetration (such as infrared lasers) is applied. Upconverted light excites PS followed by ROS production and surrounding tissue damage.

Hydrogels are polymeric materials with hydrophilic structures capable of holding large amounts of water in their three-dimensional mesh structures with physical properties similar to those of living tissues (196, 197). These are widely used for the controlled release of hydrophilic drugs including water soluble PCs and MPCs (198-200). They can combine chemo-photodynamic therapy (201) or can be used for *in vivo* imaging (202).

Dendrimers are typical examples of bottom-up approach in preparation of vehicles for drug delivery as they are synthesized stepwise. They are repetitively branched polymers with a precisely defined diameter in the order of nanometers and typically symmetric around the core with a spherical three-dimensional morphology. Ability to control its size, number of functional groups available for modifications and predictability of the amount of incorporated drug makes them ideal DDS with reproducible pharmacokinetics (62, 203). PS can be trapped in the voids of a dendrimer, covalently bound to the dendrimer (204) or used as a scaffold to form a dendrimer (205). They can be conjugated with tumor-targeting peptides as well (204). PCs and MPCs can play the role of core of dendrimer with significantly improved ability to photosensitize singlet oxygen (206).

Natural cyclodextrins (CDs) are macrocyclic oligosaccharides shaped like the truncated cone with slightly hydrophobic central cavity and hydrophilic outer surface obtained by action of cyclodextrin- α -glycosyl transferase enzyme. They have six, seven, or eight α -1,4 linked α -D-(+)-glucopyranose units named α -, β -, and γ -CD, respectively,. Their shape and hydrophobicity of cavity allow formation of an inclusion complex with various lipophilic drugs. CDs can be modified to improve their properties, such as complexation of particular drug moiety (62, 207-210). In the literature, there have been described covalently bonded MPCs to natural CDs (211) or biodegradable nanoassemblies of modified CDs with MPC (212), both to overcome poor aqueous solubility of phthalocyanines.

Conclusion

PDT is generally known as an alternative approach for cancer treatment. However, its future lies in multimodal treatment, dermatology and aesthetic medicine. In cancer treatment, the major advantages compared to surgery, chemo- or radiotherapy include limited and more tolerable side-effects because ROS are generated only in areas with accumulated PS that are illuminated by sufficient doses of light of specific wavelength. The main disadvantage of the localized damage is recurrence due to metastases, as well as insufficient accumulation of the PS in the tumor area far from the vasculature. However optimal integration with other therapies leads to improved effectiveness (*e.g.* photosterilization of the tumor bed after surgical resection) and decreased side effects

(lower doses of chemotherapeutics). The difficulties with administration and skin permeation and insufficient accumulation in targeted tissue can be overcome using appropriate drug delivery system with passive or active targeting. Limited tissue penetration of light commonly used to activate PSs to generate ROS can be overcome using photon upconverting nanoparticles with appropriate PSs, so far only phthalocyanines (PCs). The variability in metal cations coordinated to the centre and the peripheral substituents of the macrocycle results in many derivatives with tunable photophysical and photochemical properties. Their innovative formulations make them

Very powerful with immense promise in PDT.

Conflicts of Interest

There are no conflicts of interest regarding this study.

Authors' Contributions

JR, DV - literature search, defining the articles to be included in the systematic review, writing, drawing figures. JB, PP – review of the drafted manuscript.

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References

- Tedesco AC, Rotta JC and Lunardi CN: Synthesis, photophysical and photochemical aspects of phthalocyanines for photodynamic therapy. Curr Org Chem 7: 187-196, 2003. DOI: 10.2174/ 1385272033373076
- Allen CM, Sharman WM and Van Lier JE: Current status of phthalocyanines in the photodynamic therapy of cancer. J Porphyrins Phthalocyanines 05: 161-169, 2001. PMID: 7528127. DOI: 10.1002/jpp.324
- Moreira LM, Dos Santos FV, Lyon JP, Maftoum-Costa M, Pacheco-Soares C and Soares Da Silva N: Photodynamic therapy: Porphyrins and phthalocyanines as photosensitizers. Aust J Chem 61: 741-754, 2008. DOI: 10.1071/CH08145
- 4 Babilas P, Schreml S, Landthaler M and Szeimies RM: Photodynamic therapy in dermatology: State-of-the-art. Photodermatol Photoimmunol Photomed 26: 118-132, 2010. PMID: 20584250. DOI: 10.1111/j.1600-0781.2010.00507.x
- 5 Baldea I and Filip AG: Photodynamic therapy in melanoma An update. J Physiol Pharmacol 63: 109-118, 2012. PMID: 18400022. DOI: 10.1111/j.1600-0781.2005.00147.x
- 6 Ratour-Bigot C, Chemidling M, Montlahuc C, Abirached G, Madjlessi N, Bullier C, Battistella M, Bagot M, Lebbe C and Basset-Seguin N: Squamous cell carcinoma following photodynamic therapy for cutaneous bowen's disease in a series of 105 patients. Acta Derm Venereol 96: 658-663, 2016. PMID: 26714426. DOI: 10.2340/00015555-2330

- 7 Farhadi M, Kamrava SK, Behzadi AH, Rafiezadeh P, Asghari A, Rezvan F and Maleki S: The efficacy of photodynamic therapy in treatment of recurrent squamous cell and basal cell carcinoma. J Drugs Dermatol 9: 122-126, 2010. PMID: 20214173.
- 8 Thissen MRTM, Schroeter CA and Neumann HAM: Photodynamic therapy with delta-aminolaevulinic acid for nodular basal cell carcinomas using a prior debulking technique. Br J Dermatol 142: 338-339, 2000. PMID: 10730770. DOI: 10.1046/j.1365-2133.2000.03404.x
- 9 Christensen E, Mørk C and Skogvoll E: High and sustained efficacy after two sessions of topical 5-aminolaevulinic acid photodynamic therapy for basal cell carcinoma: a prospective, clinical and histological 10-year follow-up study. Br J Dermatol 166: 1342-1348, 2012. PMID: 22309486. DOI: 10.1111/j.1365-2133.2012.10878.x
- 10 Cosgarea R, Susan M, Crisan M and Senila S: Photodynamic therapy using topical 5-aminolaevulinic acid vs. surgery for basal cell carcinoma. J Eur Acad Dermatology Venereol 27: 980-984, 2013. PMID: 22738399. DOI: 10.1111/j.1468-3083.2012.04619.x
- 11 Gupta AK, Paquet M, Villanueva E and Brintnell W: Interventions for actinic keratoses. Cochrane Database Syst Rev 12: CD004415, 2012. PMID: 23235610. DOI: 10.1002/14651858.CD004415.pub2
- 12 Gupta AK and Paquet M: Network meta-analysis of the outcome 'participant complete clearance' in nonimmunosuppressed participants of eight interventions for actinic keratosis: A followup on a Cochrane review. Br J Dermatol 169: 250-259, 2013. PMID: 23550994. DOI: 10.1111/bjd.12343
- Truchuelo M, Fernández-Guarino M, Fleta B, Alcántara J and Jaén P: Effectiveness of photodynamic therapy in Bowen's disease: an observational and descriptive study in 51 lesions. J Eur Acad Dermatol Venereol 26: 868-874, 2012. PMID: 21740466. DOI: 10.1111/j.1468-3083.2011.04175.x
- 14 Zaar O, Fougelberg J, Hermansson A, Gillstedt M, Wennberg-Larkö A-M and Paoli J: Effectiveness of photodynamic therapy in Bowen's disease: a retrospective observational study in 423 lesions. J Eur Acad Dermatology Venereol 31: 1289-1294, 2017. PMID: 21740466. DOI: 10.1111/jdv.14164
- 15 Ge X, Liu J, Shi Z, Jing L, Yu N, Zhang X, Jiao Y, Wang Y and Li PA: Inhibition of MAPK signaling pathways enhances cell death induced by 5-Aminolevulinic acid-photodynamic therapy in skin squamous carcinoma cells. Eur J Dermatol 26: 164-172, 2016. PMID: 27032574. DOI: 10.1684/ejd.2015.2725
- 16 Coors EA and Von Den Driesch P: Topical photodynamic therapy for patients with therapy-resistant lesions of cutaneous T-cell lymphoma. J Am Acad Dermatol 50: 363-367, 2004. PMID: 14988676. DOI: 10.1016/S0190-9622(03)00868-5
- 17 Debu A, Girard C, Kluger N, Guillot B and Dereure O: Topical methyl aminolaevulinate-photodynamic therapy in erosive facial mycosis fungoides. Br J Dermatol 163: 884-885, 2010. PMID: 20854405. DOI: 10.1111/j.1365-2133.2010.09898.x
- 18 Markham T, Sheahan K and Collins P: Topical 5-aminolaevulinic acid photodynamic therapy for tumour-stage mycosis fungoides. Br J Dermatol 144: 1262-1263, 2001. PMID: 11422054. DOI: 10.1046/j.1365-2133.2001.04245.x
- 19 Díez Recio E, Zambrano B, Alonso ML, de Eusebio E, Martín M, Cuevas J and Jaén P: Topical 5-aminolevulinic acid photodynamic therapy for the treatment of unilesional mycosis fungoides: a report of two cases and review of the literature. Int J Dermatol 47: 410-413, 2008. PMID: 18377612. DOI: 10.1111/j.1365-4632.2008.03177.x.

- 20 Mori M, Campolmi P, Mavilia L, Rossi R, Cappugi P and Pimpinelli N: Topical photodynamic therapy for primary cutaneous B-cell lymphoma: A pilot study. J Am Acad Dermatol 54: 524-526, 2006. PMID: 16488310. DOI: 10.1016/ j.jaad.2005.10.016
- 21 Abels C: Targeting of the vascular system of solid tumours by photodynamic therapy (PDT). Photochem Photobiol Sci *3*: 765-771, 2004. PMID: 15295633. DOI: 10.1039/b314241h
- 22 Ud-Din S, Thomas G, Morris J and Bayat A: Photodynamic therapy: an innovative approach to the treatment of keloid disease evaluated using subjective and objective non-invasive tools. Arch Dermatol Res 305: 205-214, 2013. PMID: 23117184. DOI: 10.1007/s00403-012-1295-4
- 23 Sakamoto FH, Izikson L, Tannous Z, Zurakowski D and Anderson RR: Surgical scar remodelling after photodynamic therapy using aminolaevulinic acid or its methylester: A retrospective, blinded study of patients with field cancerization. Br J Dermatol 166: 413-416, 2012. PMID: 21848691. DOI: 10.1111/j.1365-2133.2011.10576.x
- 24 Wan MT and Lin JY: Current evidence and applications of photodynamic therapy in dermatology. Clin Cosmet Investig Dermatol 7: 145-163, 2014. PMID: 24899818. DOI: 10.2147/ CCID.S35334
- 25 Hassani J and Feldman SR: Phototherapy in scleroderma. Dermatol Ther (Heidelb) 6: 519-553, 2016. DOI: 10.1007/ s13555-016-0136-3
- 26 Karrer S, Abels C, Landthaler M and Szeimies RM: Topical photodynamic therapy for localized scleroderma. Acta Derm Venereol 80: 26-27, 2000. PMID: 10721828. DOI: 10.1080/ 000155500750012469
- 27 Sakamoto FH, Torezan L and Anderson RR: Photodynamic therapy for acne vulgaris: A critical review from basics to clinical practice: Part II. Understanding parameters for acne treatment with photodynamic therapy. J Am Acad Dermatol 63: 195-211, 2010. PMID: 20633797. DOI: 10.1016/j.jaad.2009.09.057
- 28 Sakamoto FH, Lopes JD and Anderson RR: Photodynamic therapy for acne vulgaris: A critical review from basics to clinical practice - Part I. Acne vulgaris: When and why consider photodynamic therapy? J Am Acad Dermatol 63: 183-193, 2010. PMID: 20633796. DOI: 10.1016/j.jaad.2009.09.056
- 29 Hongcharu W, Taylor CR, Aghassi D, Suthamjariya K, Anderson RR and Chang Y: Topical ALA-photodynamic therapy for the treatment of acne vulgaris. J Invest Dermatol 115: 183-192, 2000. PMID: 10951234. DOI: 10.1046/j.1523-1747.2000.00046.x
- 30 Bryld LE and Jemec GBE: Photodynamic therapy in a series of rosacea patients. J Eur Acad Dermatol Venereol 21: 1199-1202, 2007. PMID: 17894705. DOI: 10.1111/j.1468-3083.2007.02219.x
- 31 Togsverd-Bo K, Wiegell S, Wulf H and Haedersdal M: Short and limited effect of long-pulsed dye laser alone and in combination with photodynamic therapy for inflammatory rosacea. J Eur Acad Dermatology Venereol 23: 200-201, 2009. PMID: 18452529. DOI: 10.1111/j.1468-3083.2008.02781.x
- 32 Gong Y, Labh S, Jin Y, Diao HY, Li XL, Liu ZY and Shi YL: Needle-free injection of 5-aminolevulinic acid in photodynamic therapy for the treatment of non-melanoma skin cancer. Dermatol Ther 29: 255-262, 2016. PMID: 23935753. DOI: 10.1111/dth.12335
- 33 Eibenschutz L, Silipo V, De Simone P, Buccini PL, Ferrari A, Carbone A and Catricalà C: A 9-month, randomized, assessorblinded, parallel-group study to evaluate clinical effects of film-

- forming medical devices containing photolyase and sun filters in the treatment of field cancerization compared with sunscreen in patients after successful p. Br J Dermatol *175*: 1391-1393, 2016. PMID: 27167413. DOI: 10.1111/bjd.14721
- 34 Jerjes W, Hamdoon Z, Abdulkareem AA and Hopper C: Photodynamic therapy in the management of actinic keratosis: Retrospective evaluation of outcome. Photodiagnosis Photodyn Ther 17: 200-204, 2017. PMID: 27174210. DOI: 10.1016/j.pdpdt.2016.04.017
- 35 Luderschmidt C and Plewig G: Circumscribed sebaceous gland hyperplasia: Autoradiographic and histoplanimetric studies. J Invest Dermatol 70: 207-209, 1978. PMID: 641372. DOI: 10.1111/1523-1747.ep12541329
- 36 Plewig G and Kligman AM: Proliferative activity of the sebaceous glands of the aged. J Invest Dermatol, 1978. PMID: 649977. DOI: 10.1111/1523-1747.ep12543478
- 37 Schroeter CA, Pleunis J, van Nispen Tot Pannerden C, Reineke T and Neumann HAM: Photodynamic therapy: New treatment for therapy-resistant plantar warts. Dermatologic Surg 31: 71-75, 2005. PMID: 15720099. DOI: 10.1097/00042728-200501000-00013
- 38 Stender IM, Lock-Andersen J and Christian Wulf H: Recalcitrant hand and foot warts successfully treated with photodynamic therapy with topical 5-aminolaevulinic acid: a pilot study. Clin Exp Dermatol 24: 154-159, 1999. PMID: 10354167. DOI: 10.1046/j.1365-2230.1999.00441.x
- 39 Gold MH and Pope A: Fractional resurfacing aiding photodynamic therapy of a recalcitrant plantar verruca: a case report and review of the literature. J Clin Aesthet Dermatol 1: 30-33, 2008. PMID: 21103307.
- 40 Gold MH, Boring MM, Bridges TM and Bradshaw VL: The successful use of ALA-PDT in the treatment of recalcitrant molluscum contagiosum. J Drugs Dermatol 3: 187-190, 2004. PMID: 15098976.
- 41 Sotiriou E, Panagiotidou D and Ioannides D: 5-Aminolevulininic acid photodynamic therapy treatment for tinea cruris caused by Trichophyton rubrum: report of 10 cases. J Eur Acad Dermatology Venereol 23: 341-342, 2009. PMID: 18637974. DOI: 10.1111/j.1468-3083.2008.02880.x
- 42 Sotiriou E, Koussidou T, Patsatsi A, Apalla Z and Ioannides D: 5-Aminolevulinic acid-photodynamic treatment for dermatophytic tinea pedis of interdigital type: A small clinical study. J Eur Acad Dermatology Venereol 23: 203-204, 2009. PMID: 18435726. DOI: 10.1111/j.1468-3083.2008.02783.x
- 43 Calzavara-Pinton PG, Venturini M, Capezzera R, Sala R and Zane C: Photodynamic therapy of interdigital mycoses of the feet with topical application of 5-aminolevulinic acid. Photodermatol Photoimmunol Photomed 20: 144-147, 2004. PMID: 15144392. DOI: 10.1111/j.1600-0781.2004.00095.x
- 44 Sohl S, Kauer F, Paasch U and Simon JC: Photodynamic treatment of cutaneous leishmaniasis. J Dtsch Dermatol Ges 5: 128-130, 2007. PMID: 17274779. DOI: 10.1111/j.1610-0387.2007.06177.x
- 45 Asilian A and Davami M: Comparison between the efficacy of photodynamic therapy and topical paromomycin in the treatment of Old World cutaneous leishmaniasis: A placebo-controlled, randomized clinical trial. Clin Exp Dermatol 31: 634-637, 2006. PMID: 16780497. DOI: 10.1111/j.1365-2230.2006.02182.x
- 46 Gardlo K, Horska Z, Enk CD, Rauch L, Megahed M, Ruzicka T and Fritsch C: Treatment of cutaneous leishmaniasis by photodynamic therapy. J Am Acad Dermatol 48: 893-896, 2003. PMID: 12789181. DOI: 10.1067/mjd.2003.218

- 47 da Silva NS, Ribeiro C de M, Machado AHA and Pacheco-Soares C: Ultrastructural changes in Tritrichomonas foetus after treatments with AlPcS4 and photodynamic therapy. Vet Parasitol 146: 175-181, 2007. PMID: 17399904. DOI: 10.1016/j.vetpar.2007.02.006
- 48 Otsuji T, Sho K, Tsumura A, Koike N, Nishimura T and Takahashi K: Three-year results of a modified photodynamic therapy procedure (Ironing PDT) for age-related macular degeneration patients with large lesions. Clin Ophthalmol 10: 431-436, 2016. PMID: 27041985. DOI: 10.2147/OPTH.S102171
- 49 Battaglia Parodi M, La Spina C, Berchicci L, Petruzzi G and Bandello F: Photosensitizers and photodynamic therapy: verteporfin. Dev Ophthalmol 55: 330-336, 2015. PMID: 26501167. DOI: 10.1159/000434704
- 50 Crescenzi E, Chiaviello A, Canti G, Reddi E, Veneziani BM and Palumbo G: Low doses of cisplatin or gemcitabine plus Photofrin/photodynamic therapy: Disjointed cell cycle phaserelated activity accounts for synergistic outcome in metastatic nonsmall cell lung cancer cells (H1299). Mol Cancer Ther 5: 776-785, 2006. PMID: 16546993. DOI: 10.1158/1535-7163.MCT-05-0425
- 51 Crescenzi E, Varriale L, Iovino M, Chiaviello A, Veneziani BM and Palumbo G: Photodynamic therapy with indocyanine green complements and enhances low-dose cisplatin cytotoxicity in MCF-7 breast cancer cells. Mol Cancer Ther 3: 537-544, 2004. PMID: 15141011. DOI: 10.1039/b108586g
- 52 van Straten D, Mashayekhi V, de Bruijn H, Oliveira S and Robinson D: Oncologic photodynamic therapy: Basic principles, current clinical status and future directions. Cancers (Basel) 9: 19, 2017. PMID: 28218708. DOI: 10.3390/cancers9020019
- 53 Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, Hahn SM, Hamblin MR, Juzeniene A, Kessel D, Korbelik M, Moan J, Mroz P, Nowis D, Piette J, Wilson BC and Golab J: Photodynamic therapy of cancer: An update. CA Cancer J Clin 61: 250-281, 2011. PMID: 21617154. DOI: 10.3322/caac 20114
- 54 Konan YN, Gurny R and Allémann E: State of the art in the delivery of photosensitizers for photodynamic therapy. J Photochem Photobiol B Biol 66: 89-106, 2002. PMID: 11897509. DOI: 10.1016/S1011-1344(01)00267-6
- 55 Dougherty TJ, Gomer CJ, Henderson BW, Jori G, Kessel D, Korbelik M, Moan J and Peng Q: Photodynamic therapy. J Natl Cancer Inst 90: 889-905, 1998. PMID: 9637138.
- 56 Kalka K, Merk H and Mukhtar H: Photodynamic therapy in dermatology. J Am Acad Dermatol 42: 389-416, 2000. PMID: 10688709. DOI: 10.1016/S0190-9622(00)90209-3
- 57 Isaacs NS: Physical organic chemistry. 2nd ed. London, Longman Scientific & Technical, 1998.
- 58 Kalyanasundaram K: Photochemistry of polypyridine and porphyrin complexes. London; San Diego; New York, Academic Press, 1992.
- 59 Ochsner M: Photophysical and photobiological processes in the photodynamic therapy of tumours. J Photochem Photobiol B Biol 39: 1-18, 1997. PMID: 9210318. DOI: 10.1016/S1011-1344(96) 07428-3
- 60 MacDonald IJ and Dougherty TJ: Basic principles of photodynamic therapy. J Porphyrins Phthalocyanines 05: 105-129, 2001. DOI: 10.1002/jpp.328
- 61 Dolmans D, Fukumura D and Jain R: Photodynamic therapy for cancer. Nat Rev Cancer 3: 380-387, 2003. PMID: 12724736. DOI: 10.1038/nrc1071

- 62 Calixto GMF, Bernegossi J, De Freitas LM, Fontana CR, Chorilli M and Grumezescu AM: Nanotechnology-based drug delivery systems for photodynamic therapy of cancer: A review. Molecules 21: 1-18, 2016. PMID: 26978341. DOI: 10.3390/molecules21030342
- 63 De Rosa FS and Bentley M V: Photodynamic therapy of skin cancers: sensitizers, clinical studies and future directives. Pharm Res 17: 1447-1455, 2000. PMID: 11303952. DOI: 10.1023/A:1007612905378
- 64 Kharkwal GB, Sharma SK, Huang Y-Y, Dai T and Hamblin MR: Photodynamic therapy for infections: clinical applications. Lasers Surg Med 43: 755-767, 2011. PMID: 22057503. DOI: 10.1002 /lsm.21080
- 65 Peng Q, Berg K, Moan J, Kongshaug M and Nesland JM: 5-aminolevulinic acid-based photodynamic therapy: Principles and experimental research. Photochem Photobiol 65: 235-251, 1997. PMID: 9066303. DOI: 10.1111/j.1751-1097.1997.tb08549.x.
- 66 Gannon MJ and Brown SB: Photodynamic therapy and its applications ingynaecology. BJOG An Int J Obstet Gynaecol 106: 1246-1254, 1999. PMID: 10609717. DOI: 10.1111/j.1471-0528.1999.tb08177.x
- 67 Moan J, Streckyte G, Bagdonas S, Bech Ø and Berg K: Photobleaching of protoporphyrin IX in cells incubated with 5aminolevulinic acid. Int J Cancer 70: 90-97, 1997. PMID: 8985096. DOI: 10.1002/(SICI)1097-0215(19970106)70:1<90:: AID-IJC14>3.0.CO;2-H
- 68 Leznoff CC and Lever ABP: Phthalocyanines: Properties and applications. Wiley, 1989.
- 69 Tokumaru K: Photochemical and photophysical behaviour of porphyrins and phthalocyanines irradiated with violet or ultraviolet light. J Porphyr Phthalocyanines 05: 77-86, 2001. DOI: 10.1002/1099-1409(200101)5:1<77::AID-JPP302>3.0.CO; 2-X
- 70 Ahmad N and Mukhtar H: Mechanism of photodynamic therapyinduced cell death. *In*: Singlet Oxygen, UV-A, and Ozone. Academic Press, pp. 342-358, 2000.
- 71 Allison RR and Moghissi K: Photodynamic therapy (PDT): PDT mechanisms. Clin Endosc 46: 24-29, 2013. PMID: 23422955. DOI: 10.5946/ce.2013.46.1.24
- 72 Moan J: On the diffusion length of singlet oxygen in cells and tissues. J Photochem Photobiol B Biol 6: 343-344, 1990. PMID: 49212696. DOI: 10.1016/1011-1344(90)85104-5
- 73 Andrzejak M, Price M and Kessel DH: Apoptotic and autophagic responses to photodynamic therapy in 1c1c7 murine hepatoma cells. Autophagy 7: 979-984, 2011. PMID: 21555918. DOI: 10.4161/auto.7.9.15865
- 74 Chiaviello A, Postiglione I and Palumbo G: Targets and mechanisms of photodynamic therapy in lung cancer cells: A brief overview. Cancers (Basel) 3: 1014-1041, 2011. PMID: 24212652. DOI: 10.3390/cancers3011014.
- 75 Kim J, Lim W, Kim S, Jeon S, Hui Z, ni K, Kim C, Im Y, Choi H and Kim O: Photodynamic therapy (PDT) resistance by PARP1 regulation on PDT-induced apoptosis with autophagy in head and neck cancer cells. J Oral Pathol Med 43: 675-684, 2014. DOI: 10.1111/jop.12195
- 76 Buytaert E, Dewaele M and Agostinis P: Molecular effectors of multiple cell death pathways initiated by photodynamic therapy. Biochim Biophys Acta - Rev Cancer 1776: 86-107, 2007. PMID: 17693025. DOI: 10.1016/j.bbcan.2007.07.001

- 77 Piette J: Signalling pathway activation by photodynamic therapy: NF-κB at the crossroad between oncology and immunology. Photochem Photobiol Sci 14: 1510-1517, 2015. DOI: 10.1039/ C4PP00465E
- 78 Santus R, Morliere P and Kohen E: The photobiology of the living cell as studied by microspectrometric techniques. Photochem Photobiol 54: 1071-1077, 1991.
- 79 Moan J and Berg K: Photochemotherapy of cancer: Experimental research. Photochem Photobiol 55: 931-948, 1992. PMID: 1409894. DOI: 10.1111/j.1751-1097.1992.tb08541.x
- 80 de Castro Pazos M, Pacheco-Soares C, Soares da Silva N, DaMatta RA and Pacheco MTT: Ultrastructural effects of two phthalocyanines in CHO-K1 and HeLa cells after laser irradiation. Biocell 27: 301-309, 2003. PMID: 15002747.
- 81 Cavalcante AKD, Martinez GR, Di Mascio P, Menck CFM and Agnez-Lima LF: Cytotoxicity and mutagenesis induced by singlet oxygen in wild type and DNA repair deficient Escherichia coli strains. DNA Repair (Amst) 1: 1051-1056, 2002. PMID: 12531014. DOI: 10.1016/S1568-7864(02)00164-7
- 82 Donnelly R, McCarron P and Woolfson D: Drug delivery systems for photodynamic therapy. Recent Pat Drug Deliv Formul 3: 1-7, 2009. PMID: 19149725. DOI: 10.2174/187221 109787158319
- 83 Jori G, Beltramini M, Reddi E, Salvato B, Pagnan A, Ziron L, Tomio L and Tsanov T: Evidence for a major role of plasma lipoproteins as hematoporphyrin carriers in vivo. Cancer Lett 24: 291-297, 1984. PMID: 6498807. DOI: 10.1016/0304-3835(84)90025-9
- 84 Pottier R and Kennedy JC: New trends in photobiology. The possible role of ionic species in selective biodistribution of photochemotherapeutic agents toward neoplastic tissue. J Photochem Photobiol B Biol, 1990. PMID: 2127428. DOI: 10.1016/1011-1344(90)85183-W
- 85 Harada M, Woodhams J, MacRobert AJ, Feneley MR, Kato H and Bown SG: The vascular response to photodynamic therapy with ATX-S10Na(II) in the normal rat colon. J Photochem Photobiol B Biol 79: 223-230, 2005. PMID: 15896649. DOI: 10.1016/j.jphotobiol.2004.08.011
- 86 Oleinick NL, Morris RL and Belichenko I: The role of apoptosis in response to photodynamic therapy: What, where, why, and how. Photochem Photobiol Sci 1: 1-21, 2002. PMID: 12659143. DOI: 10.1039/b108586g
- 87 Nyokong T and Gledhill I: The use of phthalocyanines in cancer therapy. AIP Conf Proc 1517: 49-52, 2013. DOI: 10.1063/ 1.4794220
- 88 Banfi S, Caruso E, Buccafurni L, Ravizza R, Gariboldi M and Monti E: Zinc phthalocyanines-mediated photodynamic therapy induces cell death in adenocarcinoma cells. J Organomet Chem 692: 1269-1276, 2007. DOI: 10.1016/j.jorganchem.2006.11.028
- Edelson RL: Light-Activated Drugs. Sci Am 259: 68-75, 1988.
 PMID: 3064295. DOI: 10.1038/scientificamerican0888-68
- 90 Allison RR, Mota HC and Sibata CH: Clinical PD/PDT in North America: An historical review. Photodiagnosis Photodyn Ther 1: 263-277, 2004. PMID: 25048431. DOI: 10.1016/S1572-1000(04)00084-5
- 91 Ball DJ, Wood SR, Vernon DI, Griffiths J, Dubbelman TM and Brown SB: The characterisation of three substituted zinc phthalocyanines of differing charge for use in photodynamic therapy. A comparative study of their aggregation and photosensitising ability in relation to mTHPC and polyhaemato-

- porphyrin. J Photochem Photobiol B 45: 28-35, 1998. PMID: 9819897. DOI: 10.1016/S1011-1344(98)00156-0
- 92 Calzavara-Pinton PG, Venturini M and Sala R: Photodynamic therapy: update 2006. Part 1: Photochemistry and photobiology. J Eur Acad Dermatol Venereol 21: 293-302, 2006. PMID: 17309449. DOI: 10.1111/j.1468-3083.2006.01902.x
- 93 Rak J, Král V and Šmucler R: Photosensitive polymer film based on 5-aminolevulinic acid and its derivatives and its utilisation. EP3085359A1, 2016.
- 94 Juzeniene A and Moan J: The history of PDT in Norway. Part one: Identification of basic mechanisms of general PDT. Photodiagnosis Photodyn Ther 4: 3-11, 2007. PMID: 25047184. DOI: 10.1016/j.pdpdt.2006.11.002
- 95 Ajioka RS, Phillips JD and Kushner JP: Biosynthesis of heme in mammals. Biochim Biophys Acta 1763: 723-736 2006. PMID: 16839620. DOI: 10.1016/j.bbamcr.2006.05.005
- 96 Bung N, Pradhan M, Srinivasan H and Bulusu G: Structural Insights into E. coli Porphobilinogen Deaminase during Synthesis and Exit of 1-Hydroxymethylbilane. PLoS Comput Biol 10: e1003484, 2014. PMID: 24603363. DOI: 10.1371/ journal.pcbi.1003484
- 97 Tewari KM and Eggleston IM: Chemical approaches for the enhancement of 5-aminolevulinic acid-based photodynamic therapy and photodiagnosis. Photochem Photobiol Sci 17: 1553-1572, 2018. PMID: 30328459. DOI: 10.1039/C8PP00362A
- 98 Wachowska M, Muchowicz A, Firczuk M, Gabrysiak M, Winiarska M, Wańczyk M, Bojarczuk K and Golab J: Aminolevulinic acid (ALA) as a prodrug in photodynamic therapy of cancer. Molecules 16: 4140-4164, 2011. DOI: 10.3390/molecules16054140
- 99 Brasseur N: Sensitizers for PDT: phthalocyanines. *In*: Photodynamic Therapy. Cambridge, Royal Society of Chemistry, pp. 105-118, 2003
- 100 Ogura S, Tabata K, Fukushima K, Kamachi T and Okura I: Development of phthalocyanines for photodynamic therapy. J Porphyr Phthalocyanines 10: 1116-1124, 2006. DOI: 10.1142/ S1088424606000466.
- 101 Mantareva V, Kussovski V, Angelov I, Borisova E, Avramov L, Schnurpfeil G and Wöhrle D: Photodynamic activity of watersoluble phthalocyanine zinc(II) complexes against pathogenic microorganisms. Bioorganic Med Chem 15: 4829-4835, 2007. PMID: 17517508. DOI: 10.1109/ROBIO.2010.5723479
- 102 Reddi E, Lo Castro G, Biolo R and Jori G: Pharmacokinetic studies with zinc(II)-phthalocyanine in tumour-bearing mice. Br J Cancer 56: 597-600, 1987. PMID: 3426922. DOI: 10.1038/ bjc.1987.247
- 103 Ali H and van Lier JE: Metal complexes as photo- and radiosensitizers. Chem Rev 99: 2379-2450, 1999. PMID: 11749485. DOI: 10.1021/cr980439y
- 104 Nyman ES and Hynninen PH: Research advances in the use of tetrapyrrolic photosensitizers for photodynamic therapy. J Photochem Photobiol B Biol 73: 1-28, 2004. PMID: 14732247. DOI: 10.1016/j.jphotobiol.2003.10.002
- 105 Ball DJ, Wood SR, Vernon DI, Griffiths J, Dubbelman TMAR and Brown SB: The characterisation of three substituted zinc phthalocyanines of differing charge for use in photodynamic therapy. A comparative study of their aggregation and photosensitising ability in relation to mTHPC and polyhaematoporphyrin. J Photochem Photobiol B Biol 45: 28-35, 1998. PMID: 9819897. DOI: 10.1016/S1011-1344(98)00156-0

- 106 Ben-Hur E and Rosenthal I: Photosensitization of Chinese hamster cells by water-soluble phthalocyanines. Photochem Photobiol 43: 615-619, 1986. PMID: 3749287. DOI: 10.1111/ j.1751-1097.1986.tb05636.x
- 107 Spikes JD: Phthalocyanines as photosensitizers in biological systems and for the photodynamic therapy of tumors. Photochem Photobiol 43: 691-699, 1986. PMID: 3092251. DOI: 10.1111/ j.1751-1097.1986.tb05648.x
- 108 Zaidi SI, Agarwal R, Eichler G, Rihter BD, Kenney ME and Mukhtar H: Photodynamic effects of new silicon phthalocyanines: in vitro studies utilizing rat hepatic microsomes and human erythrocyte ghosts as model membrane sources. Photochem Photobiol 58: 204-210, 1993. PMID: 8415911. DOI: 10.1111/j.1751-1097.1993.tb09550.x
- 109 Agarwal R, Athar M, Elmets CA, Bickers DR and Mukhtar H: Photodynamic therapy of chemically- and ultraviolet B radiationinduced murine skin papillomas by chloroaluminum phthalocyanine tetrasulfonate. Photochem Photobiol 56: 43-50, 1992. PMID: 1508981. DOI: 10.1111/j.1751-1097.1992.tb0 9600.x
- 110 Brasseur N, Ali H, Autenrieth D, Langlois R and Liert JE van: Biological activities of phthalocyanines-III. Photoinactivation of V-79 chinese hamster cells by tetrasulfophthalocyanines. Photochem Photobiol 42: 515-521, 1985. PMID: 4089034. DOI: 10.1111/j.1751-1097.1985.tb01603.x
- 111 Gregory P: Industrial applications of phthalocyanines. J Porphyr Phthalocyanines 4: 432-437, 2000. PMID: 19404370. DOI: 10.1002/(SICI)1099-1409(200006/07)4:4<432::AID-JPP254>3.3.CO;2-E
- 112 Enokida T, Hirohashi R and Nakamura T: Polymorphism of oxotitanium phthalocyanines and applications for electrophotographic receptors. Denshi Shashin Gakkaishi (Electrophotography) 29: 373-383, 1990. DOI: 10.11370/isjepj.29.373
- 113 Gregory P: High-Technology Applications of Organic Colorants. Boston, MA, Springer US, 1991.
- 114 Cook MJ, Chambrier I, Cracknell SJ, Mayes DA and Russell DA: Octa-alkyl zinc phthalocyanines: Potential photosensitizers for use in the photodynamic therapy of cancer. Photochem Photobiol 62: 542-545, 1995. PMID: 8570709. DOI: 10.1111/j.1751-1097.1995.tb02381.x
- 115 Stilts CE, Nelen MI, Hilmey DG, Davies SR, Gollnick SO, Oseroff AR, Gibson SL, Hilf R and Detty MR: Water-soluble, core-modified porphyrins as novel, longer-wavelength- absorbing sensitizers for photodynamic therapy. J Med Chem 45: 449-461, 2000. PMID: 10882367. DOI: 10.1021/jm000044i
- 116 Hilmey DG, Abe M, Nelen MI, Stilts CE, Baker GA, Baker SN, Bright F V., Davies SR, Gollnick SO, Oseroff AR, Gibson SL, Hilf R and Detty MR: Water-soluble, core-modified porphyrins as novel, longer-wavelength-absorbing sensitizers for photodynamic therapy. II. Effects of core heteroatoms and mesosubstituents on biological activity. J Med Chem 45: 449-461, 2002. PMID: 11784149. DOI: 10.1021/jm0103662
- 117 Yates NC, Moan J and Western A: Water-soluble metal naphthalocyanines-near-IR photosensitizers: Cellular uptake, toxicity and photosensitizing properties in nhik 3025 human cancer cells. J Photochem Photobiol B Biol 4: 379-390, 1990. PMID: 2111384. DOI: 10.1016/1011-1344(90)85017-Q
- 118 Cruse-Sawyer JE, Griffiths J, Dixon B and Brown SB: The photodynamic response of two rodent tumour models to four zinc (II)-substituted phthalocyanines. Br J Cancer 77: 965-972, 1998. PMID: 9528842. DOI: 10.1038/bjc.1998.159

- 119 Lee P and Wu X: Review: modifications of human serum albumin and their binding effect. Curr Pharm Des 21: 1862-1865, 2015. PMID: 25732553. DOI: 10.2174/13816128216 66150302115025
- 120 Rak J, Jakubek M, Kaplánek R, Matějíček P and Král V: Cobalt bis(dicarbollide) derivatives: Solubilization and self-assembly suppression. Eur J Med Chem 46: 1140-1146, 2011. PMID: 21315496. DOI: 10.1016/j.ejmech.2011.01.032
- 121 Larroque C, Pelegrin A and Van Lier JE: Serum albumin as a vehicle for zinc phthalocyanine: Photodynamic activities in solid tumour models. Br J Cancer 74: 1886-1890, 1996. PMID: 8980386. DOI: 10.1038/bjc.1996.649.
- 122 Reddi E, Zhou C, Biolo R, Menegaldo E and Jori G: Liposomeor LDL-administered Zn (II)-phthalocyanine as a photodynamic agent for tumours. I. Pharmacokinetic properties and phototherapeutic efficiency. Br J Cancer 61: 407-411, 1990. PMID: 2328207. DOI: 10.1007/BF02032589
- 123 Reddi E, Cernuschi S, Biolo R and Jori G: Liposome- or LDL-administered Zn(II)-phthalocyanine as a photodynamic agent for tumours III. Effect of cholesterol on pharmacokinetic and phototherapeutic properties. Lasers Med Sci 5: 339-343, 1990. DOI: 10.1007/BF02032589
- 124 Brasseur N, Langlois R, Madeleine C La, Ouellet R and Lier JE: Receptor-mediated targeting of phthalocyanines to macrophages via covalent coupling to native or maleylated bovine serum albumin. Photochem Photobiol 69: 345-352, 2008. PMID: 10089827. DOI: 10.1111/j.1751-1097.1999.tb03297.x
- 125 Sharman WM, Kudrevich SV and Van Lier JE: Novel watersoluble phthalocyanines substituted with phosphonate moieties on the benzo rings. Tetrahedron Lett *33*: 5831-5834, 1996. DOI: 10.1016/0040-4039(96)01243-9
- 126 Pandey RK: Recent advances in photodynamic therapy. J Porphyrins Phthalocyanines 4: 368-373, 2000. DOI: 10.1002/ (SICI)1099-1409(200006/07)4:4<368::AID-JPP244>3.0.CO;2-6
- 127 Dhami S, Cosa JJ, Bishop SM and Phillips D: Photophysical characterization of sulfonated aluminum phthalocyanines in a cationic reversed micellar system. Langmuir 12: 293-300, 1996. DOI: 10.1021/la950968m
- 128 Chan W-S, Marshall JF, Svensen R, Bedwell J and Hart IR: Effect of Sulfonation on the Cell and Tissue Distribution of the Photosensitizer Aluminum Phthalocyanine. Cancer Res *50*: 4533-4538, 1990. PMID: 2369730.
- 129 Margaron P, Grégoire M-J, Ŝĉasnár V, Ali H and Lier JE: Structure-photodynamic activity relationships of a series of 4substituted zinc phthalocyanines. Photochem Photobiol 63: 217-223, 1996. PMID: 8657735. DOI: 10.1111/j.1751-1097.1996. tb03017.x
- 130 Berg K, Bommer JC and Moan J: Evaluation of sulfonated aluminium phthalocyanines for use in photochemotherapy. A study on the relative efficiencies of photoinactivation. Photochem Photobiol 49: 587-594, 1989. PMID: 2755994. DOI: 10.1111/ j.1751-1097.1989.tb08428.x
- 131 Berg K, Bommer JC and Moan J: Evaluation of sulfonated aluminum phthalocyanines for use in photochemotherapy. Cellular uptake studies. Cancer Lett 44: 7-15, 1989. PMID: 2735971. DOI: https://doi.org/10.1016/0304-3835(89)90101-8
- 132 Paquette B, Ali H, Langlois R and Lier JE: Biological activities of phthalocyanines—VIII. Cellular distribution in V-79 Chinese hamster cells and phototoxicity of celectively sulfonated aluminium phthalocyanines. Photochem Photobiol 47: 215-220,

- 1988. PMID: 3344290. DOI: 10.1111/j.1751-1097.1988. tb02717.x
- 133 Yamaoka T, Tabata Y and Ikada Y: Distribution and tissue uptake of poly(ethylene glycol) with different molecular weights after intravenous administration to mice. J Pharm Sci 83: 601-606, 1994. PMID: 8046623. DOI: 10.1002/jps.2600830432
- 134 Brasseur N, Ouellet R, La Madeleine C and Van Lier JE: Water soluble aluminium phthalocyanine-polymer conjugates for PDT: Photodynamic activities and pharmacokinetics in tumour bearing mice. Br J Cancer 80: 1533-1541, 1999. PMID: 10408394. DOI: 10.1038/sj.bjc.6690557
- 135 Tiwari G, Tiwari R, Bannerjee S, Bhati L, Pandey S, Pandey P and Sriwastawa B: Drug delivery systems: An updated review. Int J Pharm Investig 2: 1-11, 2012. PMID: 23071954. DOI: 10.4103/2230-973X.96920
- 136 Reddi E, Zhou C, Biolo R, Menegaldo E and Jori G: Liposomeor LDL-administered Zn (II)-phthalocyanine as a photodynamic agent for tumours. I. Pharmacokinetic properties and phototherapeutic efficiency. Br J Cancer 61: 407-411, 1990. DOI: 10.1007/BF02032589
- 137 Shopova M, Mantareva V, Krastev K, Hadjiolov D, Milev A, Spirov K, Jori G and Ricchelli F: Comparative pharmacokinetic and photodynamic studies with zinc(II) phthalocyanine in hamsters bearing an induced or transplanted rhabdomyosarcoma. J Photochem Photobiol B Biol *16*: 83-89, 1992. PMID: 1469513. DOI: 10.1016/1011-1344(92)85155-N
- 138 Cuomo V, Jori G, Rihter B, Kenney ME and Rodgers MAJ: Tumour-localising and -photosensitizing properties of liposomedelivered ge(IV)-octabutoxy-phthalocyanine. Br J Cancer 64: 93-95, 1991. PMID: 19233766. DOI: 10.1038/bjc.1991.247
- 139 Daziano J-P, Steenken S, Chabannon C, Mannoni P, Chanon M and Julliard M: Photophysical and redox properties of a series of phthalocyanines: relation with their photodynamic activities on TF-1 and daudi leukemic cells. Photochem Photobiol 64: 712-719, 1996. PMID: 8863479. DOI: 10.1111/j.1751-1097.1996.tb03129.x
- 140 Decreau R, Richard MJ, Verrando P, Chanon M and Julliard M: Photodynamic activities of silicon phthalocyanines against achromic M6 melanoma cells and healthy human melanocytes and keratinocytes. J Photochem Photobiol B Biol 48: 48-56, 1999. PMID: 10205878. DOI: 10.1016/S1011-1344(99)00008-1
- 141 Soncin M, Polo L, Reddi E, Jori G, Kenney ME, Cheng G and Rodgers MA: Effect of axial ligation and delivery system on the tumour-localising and -photosensitising properties of Ge(IV)-octabutoxy-phthalocyanines. Br J Cancer 71: 727-732, 1995.
- 142 Sutoris K, Vetvicka D, Horak L, Benes J, Nekvasil M, Jezek P, Zadinova M and Pouckova P: Evaluation of topical photodynamic therapy of mammary carcinoma with an experimental gel containing liposomal hydroxyl-aluminium phthalocyanine. Anticancer Res 32: 3769-3774, 2012. PMID: 22993318. DOI: 32/9/3769 [pii]
- 143 Sutoris K, Rakusan J, Karaskova M, Mattova J, Benes J, Nekvasil M, Jezek P, Zadinova M, Pouckova P and Vetvicka D: Novel Topical photodynamic therapy of prostate carcinoma using hydroxy-aluminum phthalocyanine entrapped in liposomes. Anticancer Res 33: 1563-1568, 2013. PMID: 23564798.
- 144 Krishna R and Pandit JK: Carboxymethylcellulose-sodium based transdermal drug delivery system for propranolol. J Pharm Pharmacol 48: 367-370, 1996. PMID: 8794984. DOI: 10.1111/ j.2042-7158.1996.tb05934.x

- 145 Rao PR and Diwan PV: Permeability studies of cellulose acetate free films for transdermal use: Influence of plasticizers. Pharm Acta Helv 72: 47-51, 1997. PMID: 9063088. DOI: 10.1016/ S0031-6865(96)00060-X
- 146 Ranyuk E, Cauchon N, Klarskov K, Guérin B and Van Lier JE: Phthalocyanine-peptide conjugates: Receptor-targeting bifunctional agents for imaging and photodynamic therapy. J Med Chem 56: 1520-1534, 2013. PMID: 23356907. DOI: 10.1021/jm301311c
- 147 Broekgaarden M, Weijer R, Van Wijk AC, Cox RC, Egmond MR, Hoebe R, Van Gulik TM and Heger M: Photodynamic therapy with liposomal zinc phthalocyanine and tirapazamine increases tumor cell death via DNA damage. J Biomed Nanotechnol 13: 204-220, 2017. DOI: 10.1166/jbn.2017.2327
- 148 Longo JPF, Leal SC, Simioni AR, De Fátima Menezes Almeida-Santos M, Tedesco AC and Azevedo RB: Photodynamic therapy disinfection of carious tissue mediated by aluminum-chloride-phthalocyanine entrapped in cationic liposomes: An *in vitro* and clinical study. Lasers Med Sci 27: 575-584, 2012. PMID: 21809069. DOI: 10.1007/s10103-011-0962-6
- 149 Nombona N, Maduray K, Antunes E, Karsten A and Nyokong T: Synthesis of phthalocyanine conjugates with gold nanoparticles and liposomes for photodynamic therapy. J Photochem Photobiol B Biol 107: 35-44, 2012. PMID: 22209036. DOI: 10.1016 /j.jphotobiol.2011.11.007
- 150 Barbugli PA, Alves CP, Espreafico EM and Tedesco AC: Photodynamic therapy utilizing liposomal ClAlPc in human melanoma 3D cell cultures. Exp Dermatol 24: 970-972, 2015. PMID: 26194528. DOI: 10.1111/exd.12815
- 151 Derycke ASL, Kamuhabwa A, Gijsens A, Roskams T, De Vos D, Kasran A, Huwyler J, Missiaen L and de Witte PAM: Transferrin-conjugated liposome targeting of photosensitizer AlPcS4 to rat bladder carcinoma cells. JNCI J Natl Cancer Inst 96: 1620-1630, 2004. PMID: 15523091. DOI: 10.1093/jnci/djh314
- 152 Pierre MB, Tedesco AC, Marchetti JM and Bentley MV: Stratum corneum lipids liposomes for the topical delivery of 5-aminolevulinic acid in photodynamic therapy of skin cancer: preparation and *in vitro* permeation study. BMC Dermatol 1: 5, 2001. PMID: 11545679. DOI: 10.1186/1471-5945-1-5
- 153 Vetvicka D, Hruby M, Hovorka O, Etrych T, Vetrik M, Kovar L, Kovar M, Ulbrich K and Rihova B: Biological evaluation of polymeric micelles with covalently bound doxorubicin. Bioconjug Chem 20: 2090-2097, 2009. PMID: 19835372. DOI: 10.1021/bc900212k
- 154 Hruby M, Konak C, Kucka J, Vetrik M, Filippov SK, Vetvicka D, Mackova H, Karlsson G, Edwards K, Rihova B and Ulbrich K: Thermoresponsive, hydrolytically degradable polymer micelles intended for radionuclide delivery. Macromol Biosci 9: 1016-1027, 2009. DOI: 10.1002/mabi.200900083
- 155 Kataoka K, Harada A and Nagasaki Y: Block copolymer micelles for drug delivery: Design, characterization and biological significance. Adv Drug Deliv Rev 47: 113-131, 2001. DOI: 10.1016/S0169-409X(00)00124-1
- 156 Bugaj AM: Targeted photodynamic therapy A promising strategy of tumor treatment. Photochem Photobiol Sci 10: 1097-1109, 2011. PMID: 14706442. DOI: 10.1039/c0pp00147c
- 157 Van Nostrum CF: Polymeric micelles to deliver photosensitizers for photodynamic therapy. Adv Drug Deliv Rev 56: 9-16, 2004. PMID: 14706442. DOI: 10.1016/j.addr.2003.07.013

- 158 Lim CK, Heo J, Shin S, Jeong K, Seo YH, Jang WD, Park CR, Park SY, Kim S and Kwon IC: Nanophotosensitizers toward advanced photodynamic therapy of Cancer. Cancer Lett 334: 176-187, 2013. PMID: 23017942. DOI: https://doi.org/10.1016/j.canlet.2012.09.012
- 159 Khdair A, Gerard B, Handa H, Mao G, Shekhar MP V and Panyam J: Surfactant–polymer nanoparticles enhance the effectiveness of anticancer photodynamic therapy. Mol Pharm 5: 795-807, 2008. PMID: 17625926. DOI: 10.1021/mp800026t
- 160 Yu Z, Li H, Zhang LM, Zhu Z and Yang L: Enhancement of phototoxicity against human pancreatic cancer cells with photosensitizer-encapsulated amphiphilic sodium alginate derivative nanoparticles. Int J Pharm 473: 501-509, 2014. PMID: 25089506. DOI: 10.1016/j.ijpharm.2014.07.046
- 161 Manmode AS, Sakarkar DM and Mahajan NM: Nanoparticlestremendous therapeutic potential: A review. Int J PharmTech Res *I*: 1020-1027, 2009.
- 162 Ohulchanskyy TY, Roy I, Goswami LN, Chen Y, Bergey EJ, Pandey RK, Oseroff AR and Prasad PN: Organically modified silica nanoparticles with covalently incorporated photosensitizer for photodynamic therapy of cancer. Nano Lett 7: 2835-2842, 2007. PMID: 17718587. DOI: 10.1021/nl0714637
- 163 Chatterjee DK, Fong LS and Zhang Y: Nanoparticles in photodynamic therapy: An emerging paradigm. Adv Drug Deliv Rev 60: 1627-1637, 2008. DOI: 10.1016/j.addr.2008.08.003
- 164 Moffat BA, Ramachandra Ready G, McConville P, Hall DE, Chenevert TL, Kopelman RR, Philbert M, Weissleder R, Rehemtulla A and Ross BD: A Novel Polyacrylamide Magnetic Nanoparticle Contrast Agent for Molecular Imaging using MRI. Mol Imaging 2: 324-332, 2003. DOI: 10.1162/15353500332 2750664
- 165 Kovar L, Etrych T, Kabesova M, Subr V, Vetvicka D, Hovorka O, Strohalm J, Sklenar J, Chytil P, Ulbrich K and Rihova B: Doxorubicin attached to HPMA copolymer *via* amide bond modifies the glycosylation pattern of EL4 cells. Tumor Biol 31: 233-242, 2010. DOI: 10.1007/s13277-010-0019-7
- 166 Pola R, Studenovský M, Pechar M, Ulbrich K, Hovorka O, Větvička D and Říhová B: HPMA-copolymer conjugates targeted to tumor endothelium using synthetic oligopeptides. J Drug Target 17: 763-776, 2009. DOI: 10.3109/106118609 03115282.
- 167 Allemann E, Brasseur N, Benrezzak O, Rousseau J, Kudrevich S V., Boyle RW, Leroux J-C, Gurny R and Lier JE: PEG-coated poly(lactic acid) nanoparticles for the delivery of hexadecafluoro zinc phthalocyanine to EMT-6 mouse mammary tumours. J Pharm Pharmacol 47: 382-387, 1995. PMID: 7494187. DOI: 10.1111/j.2042-7158.1995.tb05815.x
- 168 Muehlmann LA, Ma BC, Longo JPF, Almeida Santos M de FM and Azevedo RB: Aluminum-phthalocyanine chloride associated to poly(methyl vinyl ether-co-maleic anhydride) nanoparticles as a new third-generation photosensitizer for anticancer photodynamic therapy. Int J Nanomedicine 9: 1199-1213, 2014. PMID: 24634582. DOI: 10.2147/IJN.S57420
- 169 Benergossi J, Calixto G, Fonseca-Santos B, Aida K, Negrini T, Duque C, Gremião M and Chorilli M: Highlights in peptide nanoparticle carriers intended to oral diseases. Curr Top Med Chem 15: 345-355, 2015. PMID: 25579347. DOI: 10.2174/1568026615666150108125040
- 170 Camerin M, Moreno M, Marín MJ, Schofield CL, Chambrier I, Cook MJ, Coppellotti O, Jori G and Russell DA: Delivery of a

- hydrophobic phthalocyanine photosensitizer using PEGylated gold nanoparticle conjugates for the: *in vivo* photodynamic therapy of amelanotic melanoma. Photochem Photobiol Sci *15*: 618-625, 2016. PMID: 27064601. DOI: 10.1039/c5pp00463b
- 171 Xin J, Wang S, Wang B, Wang JJ, Wang JJ, Zhang L, Xin B, Shen L, Zhang Z and Yao C: AlPcS4-PDT for gastric cancer therapy using gold nanorod, cationic liposome, and Pluronic[®] F127 nanomicellar drug carriers. Int J Nanomedicine *13*: 2017-2036, 2018. PMID: 29670347. DOI: 10.2147/IJN.S154054
- 172 Fedorov PP, Luginina AA, Kuznetsov SV and Osiko VV: Nanofluorides. J Fluor Chem *132*: 1012-1039, 2011. DOI: 10.1016/j.jfluchem.2011.06.025
- 173 Chen D, Zhou Y, Wan Z, Ji Z and Huang P: Tuning into single-band red upconversion luminescence in Yb3+/Ho3+ activated nano-glass-ceramics through Ce3+ doping. Dalt Trans 44: 5288-5293, 2015. DOI: 10.1039/C5DT00061K
- 174 Chen D, Zhou Y, Wan Z, Yu H, Lu H, Ji Z and Huang P: Tunable upconversion luminescence in self-crystallized Er3+:K (Y1-xYbx)3F10 nano-glass-ceramics. Phys Chem Chem Phys 17: 7100-7103, 2015. DOI: 10.1039/C4CP05706F
- 175 Chen D and Huang P: Highly intense upconversion luminescence in Yb/Er:NaGdF4@NaYF4 core–shell nanocrystals with complete shell enclosure of the core. Dalt Trans 43: 11299-11304, 2014. DOI: 10.1039/C4DT01237B
- 176 Wen S, Zhou J, Zheng K, Bednarkiewicz A, Liu X and Jin D: Advances in highly doped upconversion nanoparticles. Nat Commun 9: 2415, 2018. PMID: 29925838. DOI: 10.1038/s41467-018-04813-5
- 177 Mackenzie LE, Goode JA, Vakurov A, Nampi PP, Saha S, Jose G and Millner PA: The theoretical molecular weight of NaYF4:RE upconversion nanoparticles. Sci Rep 8: 1106, 2018. PMID: 29348590. DOI: 10.1038/s41598-018-19415-w
- 178 Chen G, Qiu H, Prasad PN and Chen X: Upconversion nanoparticles: Design, nanochemistry, and applications in theranostics. Chem Rev *114*: 5161-5214, 2014. PMID: 24605868. DOI: 10.1021/cr400425h
- 179 Tu L, Liu X, Wu F and Zhang H: Excitation energy migration dynamics in upconversion nanomaterials. Chem Soc Rev 44: 1331-1345, 2015. PMID: 25223635. DOI: 10.1039/C4CS00168K
- 180 Chen X, Peng D, Ju Q and Wang F: Photon upconversion in core–shell nanoparticles. Chem Soc Rev 44: 1318-1330, 2015. PMID: 25058157. DOI: 10.1039/C4CS00151F
- 181 Liu Y, Lu Y, Yang X, Zheng X, Wen S, Wang F, Vidal X, Zhao J, Liu D, Zhou Z, Ma C, Zhou J, Piper JA, Xi P and Jin D: Amplified stimulated emission in upconversion nanoparticles for super-resolution nanoscopy. Nature 543: 229-233, 2017. PMID: 28225761. DOI: 10.1038/nature21366
- 182 Zhou B, Shi B, Jin D and Liu X: Controlling upconversion nanocrystals for emerging applications. Nat Nanotechnol 10: 924-936, 2015. PMID: 26530022. DOI: 10.1038/nnano. 2015.251
- 183 Wang F, Han Y, Lim CS, Lu Y, Wang J, Xu J, Chen H, Zhang C, Hong M and Liu X: Simultaneous phase and size control of upconversion nanocrystals through lanthanide doping. Nature 463: 1061-1065, 2010. PMID: 20182508. DOI: 10.1038/ nature08777
- 184 Bartůněk V, Rak J, Pelánková B, Sofer Z, Ulbrich P, Kuchař M and Král V: Preparation and luminescent properties of cubic potassium-erbium fluoride nanoparticles. J Fluor Chem 156: 363-366, 2013. DOI: 10.1016/j.jfluchem.2013.07.020

- 185 Bartůněk V, Rak J, Sofer Z and Král V: Nano-crystals of various lanthanide fluorides prepared using the ionic liquid bmimPF6. J Fluor Chem 149: 13-17, 2013. DOI: 10.1016/j.jfluchem. 2013.01.035
- 186 Bartůněk V, Rak J, Král V and Smrčková O: Simple one-step preparation of cerium trifluoride nanoparticles. J Fluor Chem 132: 298-301, 2011. DOI: 10.1016/j.jfluchem.2011.02.009
- 187 Bartůněk V, Jakeš V, Král V and Rak J: Lanthanum trifluoride nanoparticles prepared using ionic liquids. J Fluor Chem *135*: 358-361, 2012. DOI: 10.1016/j.jfluchem.2011.09.003
- 188 Bartůněk V, Rak J, Pelánková B, Junková J, Mezlíková M, Král V, Kuchař M, Engstová H, Ježek P and Šmucler R: Large scale preparation of up- converting YF3:YbEr nanocrystals with various sizes by solvothermal syntheses using ionic liquid bmimCl. J Fluor Chem 188: 14-17, 2016. DOI: 10.1016/j.jfluchem.2016.05.015
- 189 Bartůněk V, Pinc J, Ulbrich P, Rak J, Pelánková B, Král V, Kuchař M, Ježek P, Engstová H and Smolková K: Tunable rapid microwave synthesis of up-converting hexagonal NaYxGdy YbzEr(1-x-y-z)F4 nanocrystals in large quantity. J Fluor Chem *178*: 56-60, 2015. DOI: 10.1016/j.jfluchem. 2015.06.021
- 190 Chen D, Liu L, Huang P, Ding M, Zhong J and Ji Z: Nd3+-Sensitized Ho3+ Single-Band Red Upconversion Luminescence in Core–Shell Nanoarchitecture. J Phys Chem Lett 6: 2833-2840, 2015. PMID: 26266869. DOI: 10.1021/acs.jpclett.5b01180
- 191 Wu Y, Yang D, Kang X, Ma P, Huang S, Zhang Y, Li C and Lin J: Core–shell structured luminescent and mesoporous β-NaYF4:Ce3+/Tb3+@mSiO2-PEG nanospheres for anti-cancer drug delivery. Dalt Trans 42: 9852-9861, 2013. DOI: 10.1039/C3DT50658D
- 192 Chen X, Zhao Z, Jiang M, Que D, Shi S and Zheng N: Preparation and photodynamic therapy application of NaYF4:Yb, Tm-NaYF4:Yb, Er multifunctional upconverting nanoparticles. New J Chem 37: 1782-1788, 2013. DOI: 10.1039/C3NJ00065F
- 193 Niemeyer CM: Nanoparticles, Proteins, and Nucleic Acids: Biotechnology Meets Materials Science. Angew Chemie Int Ed 40: 4128-4158, 2001. PMID: 29712109. DOI: 10.1002/1521-3773(20011119)40:22<4128::AID-ANIE4128>3.0.CO;2-S
- 194 Mader HS, Kele P, Saleh SM and Wolfbeis OS: Upconverting luminescent nanoparticles for use in bioconjugation and bioimaging. Curr Opin Chem Biol 14: 582-596, 2010. DOI: 10.1016/j.cbpa.2010.08.014
- 195 Kostiv U, Patsula V, Noculak A, Podhorodecki A, Větvička D, Poučková P, Sedláková Z and Horák D: Phthalocyanine-conjugated upconversion NaYF4:Yb3+/Er3+@SiO2 Nanospheres for NIR-triggered photodynamic therapy in a tumor mouse model. Chem Med Chem 12: 2066-2073, 2017. PMID: 29105372. DOI: 10.1002/cmdc.201700508
- 196 Ahmed EM: Hydrogel: Preparation, characterization, and applications: A review. J Adv Res 6: 105-121, 2015. PMID: 25750745. DOI: 10.1016/j.jare.2013.07.006
- 197 Mohy Eldin MS, Kamoun EA, Sofan MA and Elbayomi SM: l-Arginine grafted alginate hydrogel beads: A novel pH-sensitive system for specific protein delivery. Arab J Chem 8: 355-365, 2015. PMID: 28723551. DOI: 10.1016/j.arabjc.2014.01.007
- 198 Wang Y, Han B, Shi R, Pan L, Zhang H, Shen Y, Li C, Huang F and Xie A: Preparation and characterization of a novel hybrid hydrogel shell for localized photodynamic therapy. J Mater Chem B *1*: 6411, 2013. DOI: 10.1039/c3tb20779j

- 199 Karimi AR, Khodadadi A and Hadizadeh M: A nanoporous photosensitizing hydrogel based on chitosan cross-linked by zinc phthalocyanine: an injectable and pH-stimuli responsive system for effective cancer therapy. RSC Adv 6: 91445-91452, 2016. DOI: 10.1039/C6RA17064A
- 200 Mermut O, Noiseux I, Bouchard J-P, Cormier J-F, Desroches P, Fortin M, Gallant P, Leclair S, Vernon ML, Diamond KR and Patterson MS: Effect of liposomal confinement on photothermal and photo-oximetric fluorescence lifetimes of photosensitizers with varying hydrophilicity. J Biomed Opt 13: 041314, 2008. PMID: 19021322. DOI: 10.1117/1.2952298
- 201 Xu L, Zhao M, Yang Y, Liang Y, Sun C, Gao W, Li S, He B and Pu Y: A reactive oxygen species (ROS)-responsive low molecular weight gel co-loaded with doxorubicin and Zn(ii) phthalocyanine tetrasulfonic acid for combined chemo-photodynamic therapy. J Mater Chem B 5: 9157-9164, 2017. DOI: 10.1039/C7TB02359F
- 202 Liang J, Dong X, Wei C, Kong D, Liu T and Lv F: Phthalocyanine incorporated alginate hydrogel with near infrared fluorescence for non-invasive imaging monitoring in vivo. RSC Adv 7: 6501-6510, 2017. DOI: 10.1039/C6RA27756J
- 203 Tomalia DA and Khanna SN: A systematic framework and nanoperiodic concept for unifying nanoscience: hard/soft nanoelements, superatoms, meta-atoms, new emerging properties, periodic property patterns, and predictive mendeleev-like nanoperiodic tables. Chem Rev 116: 2705-2774, 2016. PMID: 26821999. DOI: 10.1021/acs.chemrev.5b00367
- 204 Narsireddy A, Vijayashree K, Adimoolam MG, Manorama SV and Rao NM: Photosensitizer and peptide-conjugated PAMAM dendrimer for targeted *in vivo* photodynamic therapy. Int J Nanomedicine 10: 6865-6878, 2015. PMID: 26604753. DOI: 10.2147/JJN.S8947
- 205 Caminade A-M, Turrin C-O and Majoral J-P: Biological properties of water-soluble phosphorhydrazone dendrimers. Brazilian J Pharm Sci 49: 33-44, 2013. PMID: 369917130. DOI: 10.1590/S1984-82502013000700004.
- 206 Setaro F, Ruiz-González R, Nonell S, Hahn U and Torres T: Synthesis, photophysical studies and ¹O₂ generation of carboxylate-terminated zinc phthalocyanine dendrimers. J Inorg Biochem 136: 170-176, 2014. PMID: 24629699. DOI: 10.1016/j.jinorgbio.2014.02.007

- 207 Schneider H-J, Hacket F, Rüdiger V and Ikeda H: NMR studies of cyclodextrins and cyclodextrin complexes. Chem Rev 98: 1755-1786, 1998. DOI: 10.1021/cr970019t
- 208 Loftsson T, Jarho P, Másson M and Järvinen T: Cyclodextrins in drug delivery. Expert Opin Drug Deliv 2: 335-351, 2005. PMID: 16296758. DOI: 10.1517/17425247.2.1.335
- 209 Saenger W, Jacob J, Gessler K, Steiner T, Hoffmann D, Sanbe H, Koizumi K, Smith SM and Takaha T: Structures of the common cyclodextrins and their larger analoguesbeyond the doughnut. Chem Rev 98: 1787-1802, 1998. PMID: 11848949. DOI: 10.1021/cr9700181
- 210 Rak J, Kaplánek R and Král V: Solubilization and deaggregation of cobalt bis(dicarbollide) derivatives in water by biocompatible excipients. Bioorg Med Chem Lett 20: 1045-1048, 2010. PMID: 20036123. DOI: 10.1016/j.bmcl.2009.12.038
- 211 Lourenço LMO, Pereira PMR, Maciel E, Válega M, Domingues FMJ, Domingues MRM, Neves MGPMS, Cavaleiro JAS, Fernandes R and Tomé JPC: Amphiphilic phthalocyanine—cyclodextrin conjugates for cancer photodynamic therapy. Chem Commun 50: 8363-8366, 2014. PMID: 24943806. DOI: 10.1039/C4CC02226B
- 212 Conte C, Scala A, Siracusano G, Leone N, Patanè S, Ungaro F, Miro A, Sciortino MT, Quaglia F and Mazzaglia A: Nanoassembly of an amphiphilic cyclodextrin and Zn(ii)-phthalocyanine with the potential for photodynamic therapy of cancer. RSC Adv 4: 43903-43911, 2014. PMID: 27424090. DOI: 10.1039/C4RA07847K

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