Review

The Role of KRAS in Endometrial Cancer: A Mini-Review

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Abstract. Endometrial cancer (EC) is the most common cancer of the female genital tract, resulting annually in 76,000 related deaths worldwide. EC originates either from oestrogen-related proliferative endometrium (type I, endometrioid), or from atrophic endometrium (type II, non-endometrioid). Each type of EC is characterized by different molecular profile alterations. The Kirsten rat sarcoma viral oncogene homolog (KRAS) gene encodes a signalling protein which moderates response to various extracellular signals via down-regulation of the mitogenactivated protein kinase (MAPK) or phosphoinositide-3-kinase/vakt murine thymoma viral oncogene (PI3K/AKT) pathways. This article reviews the role of KRAS in predicting transition from hyperplastic endometrium to early-stage well-differentiated EC, as well as further invasive proliferation of the tumour to advanced-stage disease. KRAS seems to be directly associated with type I EC, and most studies support its early involvement in carcinogenesis. Current evidence correlates KRAS mutations with increased cell proliferation and apoptosis, as well as upregulation of endometrial cell oestrogen receptors. Tumours positive for KRAS mutation can harbour hypermethylation-

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related changes in genome expression, and this can be the cause of concurrent loss of DNA repair proteins. Despite some evidence that KRAS mutation status affects cancer progression, a consensus is yet to be reached. Based on the available evidence, we suggest that screening for KRAS mutations in patients with hyperplastic endometrium or early-stage type I EC, may provide important information for prognosis stratification, and further provision of personalised treatment options.

Endometrial cancer (EC) is the most common cancer of the female genital tract in developed countries (1). Each year 319,500 women are diagnosed with EC resulting in 76,000 deaths worldwide (2). EC develops from the inner lining of the uterine corpus (3), and it is currently divided into two types as firstly described by Llobet et al. (4). Type I tumours tend to be low or intermediate tumour grade; they overlap considerably (80%) with oestrogen-related endometrioid carcinomas. Contrary to type I, type II EC results from a sequence of genetic alterations occurring in atrophic endometrium; this can occasionally reflect a progression from polyps or pre-cancerous lesions to EC. Type II EC is mostly considered as non-endometrioid serous carcinomas (4); it tends to be high grade, deeply invasive into the myometrium, and of more advanced stage at presentation (4, 5). The estimated 5-year overall survival for patients with any type of EC is 81.5% (any stage) (6).

Current Staging

EC staging consensus keeps with the 2009 International Federation of Gynaecology and Obstetrics (FIGO) revised classification. Revised FIGO staging defines four stages

(I-IV) following radical surgical resection (7). Stage I refers to a uterus-confined tumour, stage II to involvement of the cervix, stage III to adnexal or lymph node involvement, and stage IV to the presence of any metastatic deposits outside the pelvis (7).

Treatment Stratification

To-date, the stratification of treatment options relies on the disease stage; this includes certain histopathological features which are integrated into the FIGO staging. The cornerstone of EC treatment is to offer (radical) excision of the tumour; this includes total hysterectomy with/without bilateral salpingo-oophorectomy, and if indicated, systematic pelvic/para-aortic lymphadenectomy. Besides its role in treatment, radical surgery is also the basis for staging and stratifying patients for further adjuvant treatment options (8-10). Gupta et al. suggest an EC risk group classification; this is primarily based on disease staging after primary resection and stratifies the need for further adjuvant treatment depending on the potential for disease recurrence (11). Adjuvant treatment options include chemo-radiotherapy, pelvic external beam radiation therapy or vaginal cuff brachytherapy. Based on this model, surgery is the only treatment in early, low-risk EC, whereas intermediate high-risk EC would require additional adjuvant treatment (11).

Molecular Staging

Although FIGO remains the gold standard in EC staging, there is an increasing need to identify novel molecular biomarkers in order to achieve individualised treatment options. Several efforts have been described in the literature, however, consensus is yet to be reached. All efforts aim to provide a more accurate framework which can predict both prognosis as well as response to treatment and the need for additional adjuvant therapy schemes.

A classic example is The Cancer Genome Atlas (TCGA) classification. TCGA provides a molecular taxonomy for EC based on an integrated multi-platform incorporating genomic, transcriptomic and proteomic profiling (1, 12). TCGA classifies EC into four groups, each of which is based on different histopathology or molecular sub-type, as well as prognostic potential. Group 1 includes tumours with a hypermutant profile and mainly DNA polymerase epsilon, catalytic subunit (POLE) exonuclease inactivation mutations, which have a favourable overall prognosis. Group 2 refers to EC which is associated with microsatellite instability (MSI), and specifically more hypermethylation of the promoter region of the mutL homolog 1 (MLH1) gene; the latter has been found to be the primary MSI-associated mechanism of carcinogenesis in

sporadic colorectal cancer (CRC) (13). Group 3 includes tumours with low somatic copy number alterations (SCNA); the latter refers to various segmental aneuploidies, focal events, and whole-chromosome aneuploidies (14). SCNA are strongly associated with chromosomal instability; these mechanisms explain why cancer cells can potentially deviate from a diploid karyotype and can also be the fundamental cause for a degree of general heterogeneity within an individual tumour. Groups 2 and 3 have similar prognosis. Group 4 represents a high SCNA group, which mostly incorporates *TP53* mutation, and includes serous-like EC, indicating a poor overall prognosis.

Further to stratifying prognosis, the incorporation of molecular features into the classification of EC aims to optimise personalised treatment options and to predict potential responses to (neo)adjuvant treatment (12). Recent studies have shown that in the case of Kirsten rat sarcoma viral oncogene homolog (*KRAS*)-mutant EC, a combination therapy of mitogen-activated extracellular kinase (MEK) inhibitors plus anti-oestrogen agents may alter oestrogen signalling and thus improve the response rate (15).

KRAS: A Marker in Cancer Molecular Biology

KRAS is a proto-oncogene (Gene ID: 3845) located at chromosome 12 (12p12.1) and is primarily involved in the cellular response to extracellular signals. It is strongly associated with down-regulation of mitogen-activated protein kinase (MAPK) and phosphoinositide-3-kinase/v-akt murine thymoma viral oncogene (PI3K/AKT) pathways (16-18). KRAS encodes a 21-kDa signalling protein which connects activated membrane receptors to MAPK and PI3K/AKT pathways (5). Mutant KRAS promotes down-regulation of MAPK or PI3K/AKT, which further results in excessive cell proliferation and subsequently carcinogenesis (5). KRAS mutations refer to a frequent alteration of guanine to adenine (G>A), a mutation most frequently found in codon 12. Codons 12 and 13 in exon 2 constitute 90% of all KRAS mutations (19). In total, there have been 85 different mutations reported, many of which are pathway-specific (20).

KRAS is tethered to several cell membrane receptors and acts as a signalling transducer molecule. A classic example of such receptors are the surface tyrosine kinase receptors, including the epidermal growth factor receptor (EGFR) across the cell membrane of colonic and rectal epithelial cells. There is a large family of anti-EGFR chemotherapy agents, for example cetuximab and panitumumab, that target these receptors. *KRAS* mutations can cause resistance to EGFR inhibitors (20). Hence, especially in the case of CRC, *KRAS*-mutant status is directly associated with chemotherapy resistance. Therefore, *KRAS* is currently being used in clinical practice as a predictive biomarker for response to anti-EGFR chemotherapy agents (21-25).

KRAS Mutations as Biomarkers of Early-stage Type I EC

Type I and II EC are thought to be associated with different distinct mutations (4, 5). *KRAS* mutations have been mostly associated with type I oestrogen-related EC and their frequency is estimated at around 10-30% (4, 5).

In their review of mechanisms of EC development, Banno et al. supported the assumption that KRAS mutations occur at the early stages of the EC pathway (5). Further to this, KRAS mutations are present in 6-16% of endometrial hyperplasia specimens (26). Similar notions have been discussed in the case of CRC, where in the classic adenoma-carcinoma pathway, KRAS mutations seem to appear early in the neoplastic route. Several studies concluded that KRAS may play a significant role in early stage CRC (27-29). In 1988, Vogelstein et al. stated that early mutations of adenomatous polyposis coli gene result in deregulation of the wingless-related integration site (WNT) pathway (30); KRAS mutations follow deregulation of the WNT pathway and certainly take place prior to TP53 gene inactivation. Similarly, in the case of EC, KRAS mutations appear to be a stage ahead of TP53 involvement, and TP53 signifies the transition from low-grade to highgrade type I EC (5).

Further to this, Tsuda *et al.* stress the role of *KRAS* in predicting invasive proliferation of well-differentiated (grade I) tumours (31). Therefore, the role of KRAS in both an early checkpoint of transition from hyperplasia to EC, as well as a marker of the invasive potential in the case of grade I tumours, is clear.

Another interesting feature is the association of KRAS mutations with MSI-positive EC. Microsatellites are short repetitive DNA sequences which are involved in the DNA repair system. The vast majority of MSI involvement is either via direct base substitutions (point mutations), or via hypermethylation of promoters of involved genes (epigenetic changes). In the case of Lynch syndrome, the whole series of mismatch repair (MMR) genes, including MLH1, mutS homolog 2 (MSH2), mutS homolog 6 (MSH6) and postmeiotic segregation increased 2 (PMS2), is affected (13). Nevertheless, in sporadic CRC and type I EC, MMR defects are primarily a consequence of hypermethylation of gene promoters, and this primarily affects the MLH1 (4, 13). A classic example was shown by Muraki et al., who reported hypermethylation of MLH1 promoter in 40% of type I ECs (32). KRAS promoter can equally be affected by hypermethylation and this can explain its concurrent presence with defective expression of MMR genes (in MSIpositive EC). Both hypermethylation changes, as reflected by reduced presence of DNA repair proteins (MMR), and KRAS mutations, are generally thought to occur early in the EC pathway.

Translation of KRAS Mutation Status into Clinical Information for Type I EC

A narrative review of the literature was performed to summarize the current views on the prognostic and predictive value of *KRAS* mutations in EC. PubMed database was searched using any (AND, OR) combination of keywords "*KRAS*" and "Endometrial Cancer". Any original study which involved *KRAS* mutation in EC was identified and critically commented on.

Most studies focus on explaining the role of KRAS in type I oestrogen-related EC. van der Putten et al. supported the view that KRAS mutations are found adjacent to hyperplastic endometrial tissue (33). Based on this, KRAS status was used as a prognostic marker to describe a possible transition from hyperplastic tissue to malignancy; 27% of their type I EC specimens were positive for KRAS mutation, most of which were next to hyperplastic endometrium. In 5% of cases, hyperplastic (non-malignant) endometrial specimens were also positive for KRAS mutation. Further to this study, Zauber et al. supported the involvement of KRAS early in the carcinogenesis pathway, suggesting that biopsies confirming endometrial hyperplasia should be analysed for KRAS status, along with MSI status (34). Similarly, Berg et al. concluded that KRAS involvement happens early, and that molecular alterations related to KRAS mutations and inflammation are more common in obese patients (35). Similarly, Duggan et al. concluded that there is early-stage involvement of KRAS gene in type I EC, prior to clonal expansion (36).

Many studies correlated *KRAS* status with certain histopathological features; Xiong *et al.* supported the role of *KRAS* mutations in the formation of superficial epithelial changes in endometrioid EC, which has been further associated with focal mucinous differentiation (37). A similar association between *KRAS* and mucinous differentiation was reported by He *et al.* (38). Another interesting study by Steward *et al.* identified *KRAS* mutations in 12 out of 42 endometriosis-associated endometrioid adenocarcinomas (39).

As discussed previously, current literature concludes that *KRAS* mutations are primarily found in type I oestrogenrelated EC (5). An interesting question would be to explore the relationship between the *KRAS* gene and oestrogen receptors (ER), as there is extremely limited evidence for this. Tu *et al.* supported the assumption that the transcriptional activity of the ER is up-regulated by *KRAS* mutation (40). In other words, ER expression may be seen as a regulator of the RAS signalling pathway which directly affects directly the tumorigenesis of EC.

Several studies support the role of *KRAS* as a potential prognostic marker, both in terms of transition from premalignant to malignant cell status, as well as progression from early to more advanced invasive cancer. Ninomiya *et*

Table I. Summary of the role of Kirsten rat sarcoma viral oncogene homolog (KRAS) mutation in endometrial cancer (EC).

I	General information							
_	KRAS is a proto-oncogene primarily involved incellular response to several signals; Most authors describe KRAS as a signalling transducer protein molecule.	Mutations cause down-regulation of MAPK/PI3K/AKT pathway			Location: 12p12.1 Encodes: 21 kDa protein Mutation: G>A, most common at codon 12, 13 (90%); 85 different mutations in total		Involved in several carcinogenesis pathways (lung, CRC, endometrial etc.); most authors attribute KRAS mutations to being fairly early in the process	
II	What we know about KRAS							
	Associated with type I EC (endometrioid) and subsequently hyperplastic endometrium; supposed to be an early stage mutation.	ndometrioid) and subsequently hypermethylation of perplastic endometrium; mismatch repair general promoters (MLH1		Mutations present in 6-16% of endometrial hyperplasia specimens; and 10-30% of type I EC. Positive predictive value of 88% in diagnosing complex atypical hyperplasia		Potentially dual role: - Transition from hyperplastic endometrium to early stage malignancy - Progression from early to more advanced invasive cancer.		Relationship with ER: Transcriptional activity of the ER might be up-regulated by KRAS mutations.
III	What we do not know about KRAS							
	Prognostic and predictive (response to chemotherapy) value still equivocal for EC.	Limited use in clinical practice.			alue in type Iometrioid EC	Association of <i>KRAS</i> and non-malignant conditions <i>i.e.</i> endometriosis		Association of KRAS mutations with exposure to tamoxifen following breast cancer chemotherapy.
IV	Future Directions – Applicability in Clinical Practice							
	specimens for KRAS to flag high-risk cases for potential transition to malignancy? Could w treatmen type I Ed		RAS predict response to (neo) chemotherapy? e stratify need for adjuvant at options following early-stage C based on KRAS mutation k for aggressive EC pattern).		Should <i>KRAS</i> be included as a screening tool for polyps or other (pre-/non-) malignant conditions <i>i.e.</i> endometriosis in pre-menopausal women?			

ER: Oestrogen receptor; MAPK/PI3K/AKT; MLH1: mitogen-activated protein kinase/phosphoinositide-3-kinase/v-akt murine thymoma viral oncogene; MLH1: mutL homolog 1.

al. identified K- and NRAS-mediated signalling pathways as potential inducers of cell apoptosis (41). Birkeland et al. noted an increase in KRAS amplification and KRAS mRNA expression during transition from primary to metastatic disease (42). Alexander-Sefre et al. suggests a molecular assessment of the depth of myometrial invasion of EC based on KRAS (43). Mizuuchi et al. correlated the presence of KRAS mutation (codon 12 or 13) with poor prognosis (44). Ito et al. attributed KRAS mutations as being responsible for more aggressive clinical behaviour of EC in postmenopausal women (type II EC) (45). On the other hand, Varras et al. (46) and Trowbridge et al. (47) did not find correlation of KRAS status with any clinicopathological features. From the aforementioned evidence, it is apparent that a consensus on the exact way that KRAS overall affects EC prognosis is yet to be achieved.

Another interesting question is the association of *KRAS* status following tamoxifen exposure after breast cancer. Wallen *et al.* supported the existence of a link between tamoxifen use and *KRAS* codon 12 mutation (48). Nagy *et al.* noted a higher trend in *KRAS* mutation following exposure to tamoxifen (49).

Finally, although *KRAS* has an established predictive value in CRC, there are extremely limited data on this aspect in the case of EC. Byron *et al.* state that KRAS and fibroblast growth factor receptor 2 (*FGFR2*) mutations may alter the effectiveness of anti-FGR or anti-MEK biological therapies (50).

A promising study by Alomari *et al.* showed that *KRAS* mutation had a positive predictive value of 88% in diagnosing complex atypical hyperplasia (51). Based on the previous discussion, this could be an extremely important finding which may alter the current management of

endometrial hyperplasia. Current practice in the United Kingdom, as defined by the Green-top guideline (No. 67), suggests first line management of endometrial hyperplasia with atypia in premenopausal women who wish to maintain fertility, by offering the Levonorgestrel Intrauterine System (LNG-IUS) and second-line by administering oral progestogen supplements or combination of both for at least 6 months (52). Identifying novel biomarkers which can predict the course of lesions and their progression to EC would be useful to optimise care provision and reduce anxiety from both the patient's and clinician's point of view.

Limitations

We recognise a series of limitations in this narrative minireview. Although PubMed was searched systematically using certain key words, only the studies that were thought to be relevant and of high quality were considered in raising the discussion points. Secondly, most of the included cohorts were small with several limitations. Furthermore, existing evidence was conflicting. Lastly, a single biomarker was searched, which may provide a biased view as it may be optimistic to explain a complex carcinogenesis progression using a single gene.

Conclusion and Future Endeavours

Table I summarises the current knowledge on the role of KRAS in EC. Although evidence is limited and occasionally conflicting, there is a clear trend in the literature showing that KRAS plays a role early in EC progression, especially when the disease originates from hyperplastic endometrium. Given the lack of focused biomarkers, it would be interesting to conduct a prospective cohort study to delineate the role of KRAS in predicting response of hyperplasia to standard treatment or cancer progression after hyperplasia with or without atypia. In conclusion, findings of this review may allow for revision of the current management of premalignant endometrial lesions, especially hyperplasia with or without atypia. Screening of such cases for KRAS mutation would allow individualisation of treatment approach via flagging potentially high-risk pre-malignant cases for relapse of hyperplasia or future cancer progression.

Conflicts of Interest

None of the Authors has any conflict of interest to declare in regard to this article.

Authors' Contributions

MS conceived the methodology and reviewed structure, drafted the manuscript and is the guarantor for the accuracy of the data. EIE contributed to literature screening and editing parts of the article. ZA contributed to editing parts of the article. KS, VS, EE and JH contributed to literature screening. SP, SV, TH, JO, FW are equal contributors, senior authors of the article and clinicians with interest in Surgical Oncology (SP) and Advanced Gynecological Surgery and Oncology (SV, TH, JO, FW), and edited the article. All Authors have agreed to the final version of the article.

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