

Immunohistochemical Analysis of WT1 Antigen Expression in Various Solid Cancer Cells

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Abstract. For a peptide-pulsed dendritic cell (DC) vaccine to work effectively in cancer treatment, it is significant that the target protein is expressed in cancer cells. Wilms' tumor 1 (WT1) has been identified as a molecular target for immune cell therapy of cancer. We evaluated the protein expression levels of WT1 in various solid tumors, as well as mucin 1 (MUC1) or major histocompatibility complex (MHC) class I molecules. Seven hundred and thirty-eight patients whose tissue samples were examined by immunohistochemical analysis agreed to undergo DC vaccine therapy. The positive staining of WT1 in tumor cells was observed in 25.3% of patients, with only 8.5% of them showing moderate to strong expression; moreover, WT1 tended to localize in the nucleus and cytoplasm. A positive staining of tumor cells by an anti-MHC class I monoclonal antibody was observed in 98.6% and by an anti-MUC1 monoclonal antibody in 76.8% of the patients. In relation to the application of cancer-specific immunotherapy, these findings provide useful information for determining the efficacy of MUC1- and WT1-targeted therapy.

Dendritic cell (DC)-based vaccines pulsed with various tumor-specific antigens (TSAs) have been developed for cancer immunotherapy (1). Our previous data suggest that immunotherapy with tumor antigen-pulsed immature DCs with zoledronate leads to activation of V γ 9 γ δ T cells and induction of CD40L on V γ 9 γ δ T cells (2). The activated V γ 9 γ δ T cells secrete T helper (Th)1-cytokines, such as interferon (IFN)- γ , and enhance the expansion of tumor antigen-specific CD8⁺ cells by tumor antigen-pulsed immature DCs. We utilized zoledronate-pulsed DCs as

cancer vaccines with various cancer antigens in treatment of solid tumors (3-6). For DC-based cancer vaccines, some reports have described insufficient clinical responses despite the good immunoresponses indicating delayed-type hypersensitivity (DTH) (7). Immune check-point inhibitors, such as antibodies against programmed cell death protein 1 (PD-1) and cytotoxic T-lymphocyte-associated protein 4 (CTLA4), are clinically used for patients with advanced or recurrent melanoma and non-small cell lung cancer to reverse immune suppression and activate effector T cells (8, 9). Furthermore, the efficacy of immune check-point inhibitors was reported to correlate with disorders related to TSAs, oncogenic viral proteins or DNA repair pathway mutations (10).

Tumor antigens can be categorized as TSAs, cancer/testis (CT) antigens or tumor-associated antigens (TAAs) (11, 12). TSAs are abnormal proteins that arise from non-synonymous somatic mutations in tumor cells; however, such antigens are not expressed in normal cells. CT antigens can be potential targets as cancer vaccines because their expression is normally restricted to the germ cells of the testis or ovary or certain tumor cells (11, 13). TAAs are overexpressed normal proteins, such as Wilms' tumor 1 (WT1) (14, 15) or mucin 1 (MUC1) that regulate growth-promoting functions (16). The antigenicity of TAAs was reported to depend on the levels of abnormal expressions (17) because of the lower affinity of the T cell receptor (TCR) against TAAs than of TCR against TSAs (18).

For DC vaccines loaded with various TAAs peptides, their phase I/IIa clinical trial for immunotherapy was carried out in elderly patients with acute myeloid leukemia (AML), using pulsed DCs with a modified WT1 peptide and zoledronate (19). In that trial, three human leukocyte antigen (HLA)-A2402-positive elderly patients with AML were enrolled. The induction of immunoresponses to the WT1 peptide detected as DTH was indicated in two of the three patients, with a transient decrease in the number of leukemic cells being observed in these two patients. Unfortunately, a rapid expansion of leukemic cells was observed in the patient showing no immunoresponses to the WT1-specific peptide

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after the third vaccination. Recently, we studied a DC vaccine pulsed with zoledronate and an overlapping pool of peptides derived from the full length of WT1 for patients with WT1-expressing solid tumors as both CD8⁺ cytotoxic T lymphocytes (CTLs) and CD4⁺ T-helper cells against WT1 can be potentially induced without HLA restriction. However, in order to be recognized by CTLs with low-affinity TCRs, it is essential that a sufficient amount of TAAs, such as WT1 or MUC1, should be presented by major histocompatibility complex (MHC) class I on tumor cells (20).

In the present study, we examined the protein expression levels of WT1 in various solid tumors, as well as MUC1 or MHC class I molecules by immunohistochemical analysis. We also analyzed the organ and histopathological profiles of WT1 protein expression in various malignancies classified on the basis of ICD10 and ICD-O-3.

Materials and Methods

Patients' background. Between October 2007 and September 2014, tumor samples were obtained intraoperatively or by biopsy from 738 patients who provided their informed consent to four facilities of the Seta Clinic Group. The patients agreed to undergo DC vaccine therapy and their tissue samples were examined by immunohistochemical analysis for the expression levels of WT1, MUC1 and MHC class I molecules. The mean age of the 738 patients was 58.6 years (range=6-87) and the male-to-female ratio was 1:1.10 (352:386). In this study, 55 cancer types (ICD-10) and 24 histological types (ICD-O-3M) were included (Table I).

Specimens. A total of 738 specimens were examined, comprising of 72 stomach cancers, 68 colon cancers, 47 rectum cancers, 63 pancreas cancers, 120 lung cancers, 48 breast cancers and 63 ovarian cancers. The remaining tumor types are summarized in Table I. The prepared paraffin-embedded or formalin-fixed tissues were examined at Tokyo Central Pathology Laboratory Company. The histological diagnosis of each tumor was confirmed on the basis of hematoxylin and eosin staining results and the pathological reports provided by each medical facility.

Immunohistochemistry. A mouse monoclonal anti-human WT1 protein antibody (immunogen:human WT1 protein consisting of N-terminal amino acids 1-181, clone 6F-H2; Dako, Glostrup, Denmark), an anti-MUC-1 glycoprotein mouse monoclonal antibody (Novacastra Laboratories Ltd., Newcastle, UK) and an anti-HLA class I-A, B, C mouse monoclonal antibody (Hokudo Co., Ltd., Hokkaido, Japan) were used for the detection of WT1, MUC1 and MHC class I antigens, respectively.

WT1 immunohistochemical staining method. The mouse monoclonal anti-WT1 antibody (clone 6F-H2; Dako), an enzyme antibody LSAB method (labeled streptavidin-biotin) (21), and a Ventana Benchmark XT (Ventana Medical Systems, Inc., Arizona, USA) device were used for the immunohistochemical staining. Tissue sections were prepared as follows: enzyme and heat treatment for 8 and 60 min, respectively, to activate the antigen, reaction with the primary antibody for 32 min, reaction with a secondary antibody for 8 min and counterstaining with hematoxylin and eosin for 8 min.

Evaluation method. The relative ratio (proportion) and positive reaction strength (*i.e.*, staining intensity) were determined to analyze antigen expression. The level and distribution of expression were subjectively estimated and positive reaction strength was described as -, +, ++ and +++ (Table II).

Results

Immunohistochemical findings of WT1, MUC1 and MHC class I protein expressions in tumor cells. We studied WT1 protein expression in various solid tumors by immunohistochemical staining using monoclonal antibodies against WT1, MUC1 and MHC class I molecules. The results of the immunohistochemical staining of WT1 expressed in various solid tumors are shown in Figure 1. For malignant pleural mesothelioma, a strong expression of the WT1 protein was observed in the nucleus of tumor cells (Figure 1A). A weak expression of WT1 protein was observed only in the cytoplasm of cancer cells and in both the nucleus and cytoplasm of breast cancer and malignant pleural mesothelioma cells, respectively (Figure 1B and C). It is shown in Figure 1D that no WT1 protein expression was found in one patient with adenocarcinoma of the pancreas. Immunohistochemical staining showed MUC1 expression at the luminal and/or apical site of cancer cells (Figure 2A and B). MHC class I molecules were expressed in most of the solid tumors; however, loss or down-regulation of the expression of MHC class I molecules was observed in few tumors (Figure 2C and D).

Analysis of WT1, MUC1 and MHC class I protein expression in various tumors classified on the basis of ICD10. We categorized the WT1 expression patterns in tumors classified on the basis of ICD10 and ICD-O-3. Additionally, we also studied the expression patterns of MUC1, as one of the TAAs, and MHC class I molecules, as antigen presentation-associated molecules in various tumors. The expression levels of WT1, MUC1 and MHC class I in various tumors classified on the basis of ICD10 are shown in Table III. The expression of WT1 substantially differed depending on the tumor site, classified on the basis of ICD10. On the other hand, MUC1 expression was observed in most solid tumors. For malignant mesothelioma, WT1 expression was found in all tumors. Additionally, a high frequency of WT1 expression was shown (39.3%; 46/117) for the malignancies of female genital organs (ICD10; C52-C59), including cancers of the ovary (52.4%; 33/63). The frequency of WT1 expression was also relatively high in cancers of the bile duct (C23-24, 41.2%; 7/17), lung (C34, 35.0%; 42/120), breast (C50, 25.0%; 12/48) and prostate (C61, 28.6%; 4/14).

Regarding MUC1 protein expression, it was found in most solid tumors in the lip, oral cavity and pharynx (C02-13), respiratory and intrathoracic organs (C30-C38), skin

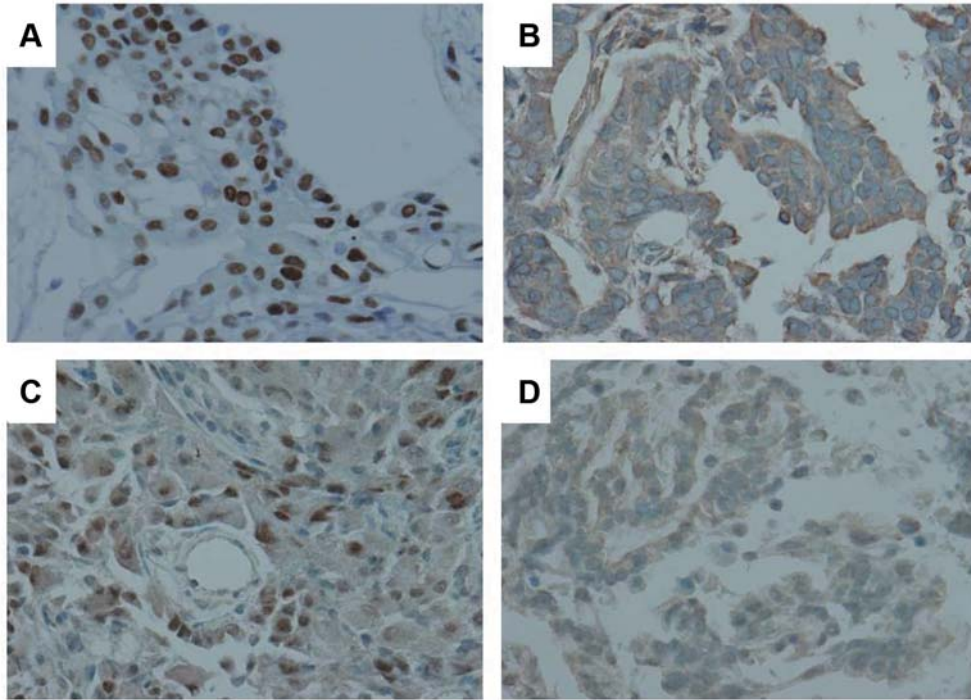


Figure 1. Immunohistochemical staining of Wilms' tumor 1 (WT1) protein by monoclonal anti-WT1 antibody. Pleural mesothelioma showing 90% nuclear staining (A), breast cancer showing 90% cytoplasmic staining (B), pleural mesothelioma showing 80% cytoplasmic and nuclear staining (C), adenocarcinoma of the pancreas showing negative staining (D). Original magnification, $\times 400$.

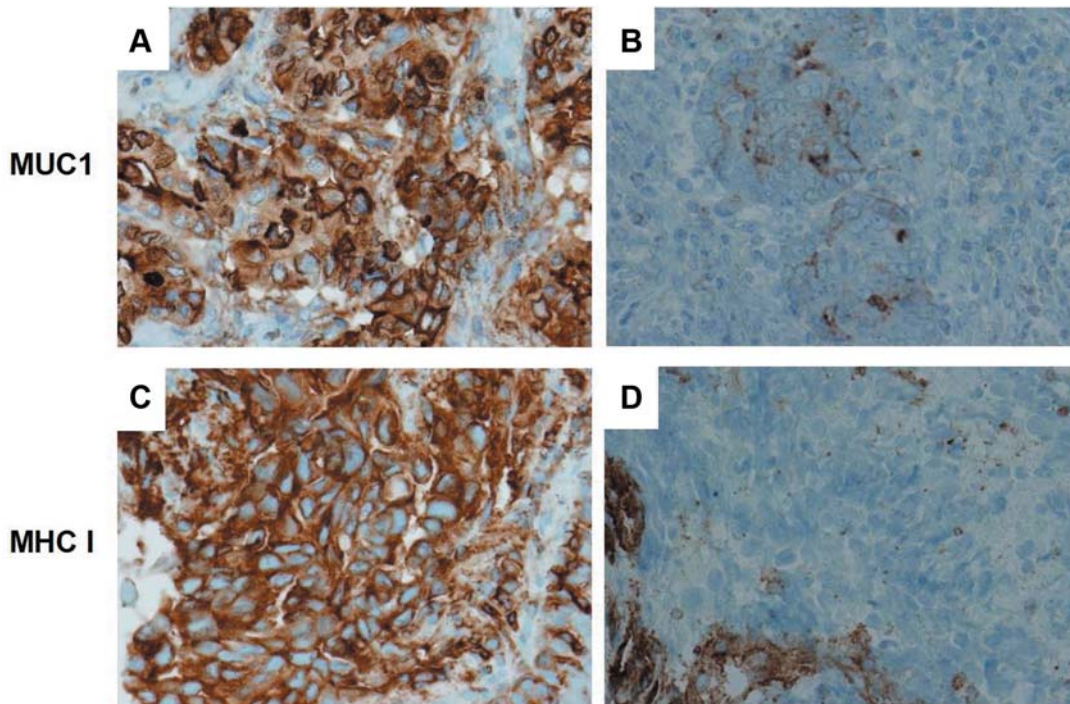


Figure 2. Immunohistochemical staining of mucin 1 (MUC1) protein (A), (B) and major histocompatibility complex (MHC) class I molecules (C), (D). Pancreatic cancer showing cell membrane and nuclear staining (A) and negative staining (B). Adenocarcinoma of the lung showing cell membrane staining (C), small cell carcinoma of the lung showing negative staining (D). Original magnification, $\times 400$.

Table I. Age and gender for each tumor type of malignancy.

ICD-10 Tumor type	Number of patients	Median age (range)	Gender (M:F)
Malignant neoplasms of lip, oral cavity and pharynx			
C02 Tongue cancer	6	53.5 (23-77)	1:5
C03 Gum cancer	1	81 -	1:0
C04 Floor of mouth cancer	1	66 -	1:0
C06 Mouth cancer	1	63 -	1:0
C07 Parotid gland cancer	1	67 -	1:0
C08 Salivary gland cancer	1	58 -	1:0
C10 Oropharyngeal cancer	3	53 (39-70)	2:1
C11 Nasopharyngeal cancer	2	34 (20-48)	1:1
C13 Hypopharyngeal cancer	2	67.5 (63-72)	2:0
Malignant neoplasms of digestive organs			
C15 Esopharyngeal cancer	26	64 (45-85)	20:6
C16 Stomach cancer	72	62 (15-85)	47:25
C17 Small intestine cancer	11	60 (37-84)	9:2
C18 Colon cancer	68	65 (23-85)	34:34
C19 Rectosigmoid junction cancer	3	62 (60-65)	2:1
C20 Rectal cancer	47	60 (29-87)	27:20
C22 Liver cancer	22	65 (25-83)	12:10
C23 Gallbladder cancer	5	55 (40-78)	5:0
C24 Bile duct cancer	12	61.5 (42-82)	9:3
C25 Pancreatic cancer	63	60 (32-86)	34:29
Malignant neoplasms of respiratory and intrathoracic organs			
C30 Nasal cavity and middle ear cancers	2	68 (65-71)	0:2
C31 Accessory sinus cancer	1	44 -	1:0
C32 Laryngeal cancer	1	59 -	1:0
C33 Tracheal cancer	2	43 (27-59)	2:0
C34 Lung cancer	120	64 (33-84)	74:46
C37 Thymus cancer	3	36 (36-58)	1:2
C38 Mediastinal cancer	1	72 -	0:1
Malignant neoplasms of bone and articular cartilage			
C40 Osteosarcoma	2	57.5 (50-65)	2:0
C41 Chondrosarcoma	1	16 -	1:0
Melanoma and other malignant neoplasms of skin			
C43 Malignant melanoma	8	60.5 (22-77)	3:5
C44 Skin cancer	3	80 (37-80)	2:1
Malignant neoplasms of mesothelial and soft tissues			
C45 Malignant mesothelioma	5	62 (36-74)	5:0
C48 Peritoneal cancer	5	68 (39-70)	0:5
C49 Sarcoma	4	30.5 (27-50)	3:1
Malignant neoplasm of breast			
C50 Breast cancer	48	50 (29-85)	-
Malignant neoplasms of female genital organs			
C52 Vaginal cancer	1	76 -	-
C53 Cervical cancer	14	50 (33-75)	-
C54 Corpus uteri cancer	26	61 (38-74)	-
C55 Uterine cancer	12	45.5 (31-65)	-
C56 Ovarian cancer	63	51 (19-82)	-
C57 Fallopian tube cancer	1	60 -	-
Malignant neoplasms of male genital organs			
C61 Prostate cancer	14	65 (55-77)	-
C63 Testicular cancer	1	79 -	-
Malignant neoplasms of urinary tract			
C64 Kidney cancer	9	56 (12-68)	8:1
C65 Renal pelvis cancer	3	63 (54-70)	2:1
C66 Ureter cancer	4	74 (70-82)	2:2
C67 Bladder cancer	10	64.5 (46-78)	8:2
C68 Urethral cancer	1	38 -	0:1

Table I. Continued

Table I. *Continued*

ICD-10 Tumor type	Number of patients	Median age (range)	Gender (M:F)
Malignant neoplasms of eye, brain and other parts of central nervous system			
C70 Malignant meningioma	1	14	-
C71 Malignant brain tumor	9	44	(6-80)
Malignant neoplasms of thyroid and other endocrine glands			
C73 Thyroid cancer	3	59	(53-73)
C74 Adrenal gland cancer	1	55	-
C75 Parathyroid cancer	1	63	-
Malignant neoplasms of ill-defined, secondary and unspecified sites			
C80 Cancer of unknown primary site	5	47	(39-64)
Malignant neoplasms, stated or presumed to be primary, of lymphoid, hematopoietic and related tissues			
C84 Peripheral T-cell lymphoma	2	74.5	(69-80)
C85 Malignant lymphoma	4	71	(63-85)
Total	738	61	(6-87)
			352:386

(C44), mesothelial tissues (C45), breast (C50), female genital organs (C52-57) and urinary tract (C64.65). For cancers of the digestive organs (C15-25), the MUC1 protein was expressed in more than 80% of the tumors in the esophagus, small intestine, bile duct or pancreas; however, the MUC1 positivity rates were 50-60% for cancers of the stomach, colon, rectum, liver and gallbladder. In this study, MHC class I expression was also investigated in various solid tumors and MHC class I molecules were found to be expressed in all cancer cells. We found nine malignant tumors with loss of MHC class I expression among 645 solid tumors, including four lung cancers, two corpus uteri cancers, one esophageal cancer, one ovarian cancer and one thyroid cancer.

Analysis of WT1 protein expression in various tumors histologically classified on the basis of ICD-O-3. We studied the expression patterns of WT1 in various solid tumors classified on the basis of ICD-O-3 (Table IV). For the histological types of cancer, the frequencies of WT1 expression were 44.2% (19/43) in cystic, mucinous and serous tumors, such as serous adenocarcinoma, mucinous cancer and signet ring cell carcinoma (M844-849); 62.5% (5/8) in complex mixed and stromal tumors, such as carcinosarcoma and adenocarcinoma (M893-899); and 100% (5/5) in malignant mesothelioma (M905). Additionally, these three types of solid tumors demonstrated moderate to strong expression of WT1 (++ and +++) in cancer cells. A high expression positivity rate of WT1 was observed in myosarcoma (80.0%; 4/5), although a strong expression of WT1 was found only in one patient. For other histological types of cancer, the positivity rates of WT1 expression ranged from 6.3 to 50.0%; however, these tumor cells

Table II. *Evaluation method for immunohistochemical analysis.*

Relative ratio (proportion)	
(-)	A positive cell is not recognized as a neoplastic cell (0%)
(+)	About 1/4 of the neoplastic cells are positive cells (less than 1/2)
(++)	About 1/2 of the neoplastic cells are positive cells (less than 3/4)
(+++)	About 3/4 of the neoplastic cells are positive cells with high reaction strength (intensity) (with the positive cells mostly neoplastic cells)
Positive reaction strength (intensity)	
(-)	Negative
(+)	Weak reaction: observable but judged to be relatively weak under high magnification
(++)	Moderate reaction: observable but judged to be slightly weak under low magnification
(+++)	Strong reaction: easily observable under low magnification

showed a weak expression of WT1. For the localization analysis of WT1 in cancer cells, the expression frequencies were 33.7, 59.4 and 4.3% in the nucleus, cytoplasm and both nucleus and cytoplasm of tumor cells, respectively (Table V). In most tumors with a strong expression of WT1, such as malignant mesothelioma, cystic, mucinous, serous tumors and complex mixed and stromal tumors, the WT1 protein localized in the nucleus of cells. However, for tumor cells with a weak (+) WT1 expression, the WT1 protein distributed uniformly in the cytoplasm of cancer cells. There were few tumors with the WT1 protein distributed in both the nucleus and cytoplasm.

Table III. Results of immunohistochemical analysis of Wilms' tumor 1 (WT1), major histocompatibility complex (MHC) class I and mucin 1 (MUC1).

ICD-10	Tumor type	WT1		MUC1		MHC-I	
		No. of positive cases	Positive ratio (%)	No. of positive cases	Positive ratio (%)	No. of positive cases	Positive ratio (%)
Malignant neoplasms of lip, oral cavity, and pharynx							
C02	Tongue cancer	1/6	16.7	6/6	100.0	6/6	100.0
C03	Gum cancer	0/1	0.0	0/1	0.0	1/1	100.0
C04	Floor of mouth cancer	1/1	100.0	1/1	100.0	1/1	100.0
C06	Mouth cancer	0/1	0.0	1/1	100.0	1/1	100.0
C07	Parotid gland cancer	1/1	100.0	1/1	100.0	1/1	100.0
C08	Salivary gland cancer	0/1	0.0	1/1	100.0	1/1	100.0
C10	Oropharyngeal cancer	0/3	0.0	3/3	100.0	3/3	100.0
C11	Nasopharyngeal cancer	0/2	0.0	1/2	50.0	2/2	100.0
C13	Hypopharyngeal cancer	0/2	0.0	2/2	100.0	2/2	100.0
Malignant neoplasms of digestive organs							
C15	Esopharyngeal cancer	5/26	19.2	20/21	95.2	20/21	95.2
C16	Stomach cancer	13/72	18.1	37/63	58.7	61/61	100.0
C17	Small intestine cancer	2/11	18.2	8/10	80.0	10/10	100.0
C18	Colon cancer	6/68	8.8	28/56	50.0	57/57	100.0
C19	Rectosigmoid junction cancer	0/3	0.0	0/2	0.0	2/2	100.0
C20	Rectal cancer	5/47	10.6	24/42	57.1	41/41	100.0
C22	Liver cancer	2/22	9.1	9/17	52.9	17/17	100.0
C23	Gallbladder cancer	2/5	40.0	3/5	60.0	5/5	100.0
C24	Bile duct cancer	5/12	41.7	8/10	80.0	10/10	100.0
C25	Pancreatic cancer	19/63	30.2	56/62	90.3	60/60	100.0
Malignant neoplasms of respiratory and intrathoracic organs							
C30	Nasal cavity and middle ear cancers	2/2	100.0	2/2	100.0	2/2	100.0
C31	Accessory sinus cancer	0/1	0.0	1/1	100.0	1/1	100.0
C32	Laryngeal cancer	0/1	0.0	0/1	0.0	1/1	100.0
C33	Tracheal cancer	1/2	50.0	2/2	100.0	2/2	100.0
C34	Lung cancer	42/120	35.0	97/109	89.0	101/105	96.2
C37	Thymus cancer	1/3	33.3	2/3	66.7	3/3	100.0
C38	Mediastinal cancer	0/1	0.0	0/0	0/0		
Malignant neoplasms of bone and articular cartilage							
C40	Osteosarcoma	0/2	0.0	1/2	50.0	2/2	100.0
C41	Chondrosarcoma	0/1	0.0	0/1	0.0	1/1	100.0
Melanoma and other malignant neoplasms of skin							
C43	Malignant melanoma	3/8	37.5	1/4	25.0	7/7	100.0
C44	Skin cancer	0/3	0.0	3/3	100.0	2/2	100.0
Malignant neoplasms of mesothelial and soft tissues							
C45	Malignant mesothelioma	5/5	100.0	3/3	100.0	3/3	100.0
C48	Peritoneal cancer	4/5	80.0	4/5	80.0	5/5	100.0
C49	Sarcoma	1/4	25.0	0/4	0.0	4/4	100.0
Malignant neoplasm of breast							
C50	Breast cancer	12/48	25.0	45/45	100.0	43/43	100.0
Malignant neoplasms of female genital organs							
C52	Vaginal cancer	1/1	100.0	0/0	1/1	100.0	
C53	Cervical cancer	3/14	21.4	11/13	84.6	11/11	100.0
C54	Corpus uteri cancer	6/26	23.1	17/22	77.3	20/22	90.9
C55	Uterine cancer	2/12	16.7	9/11	81.8	10/10	100.0
C56	Ovarian cancer	33/63	52.4	51/54	94.4	55/56	98.2
C57	Fallopian tube cancer	1/1	100.0	1/1	100.0	1/1	100.0
Malignant neoplasms of male genital organs							
C61	Prostate cancer	4/14	28.6	5/11	45.5	12/12	100.0
C63	Testicular cancer	0/1	0.0	0/0	0/0		
Malignant neoplasms of urinary tract							
C64	Kidney cancer	1/9	11.1	8/9	88.9	9/9	100.0
C65	Renal pelvis cancer	1/3	33.3	3/3	100.0	3/3	100.0

Table III. Continued

Table III. *Continued*

ICD-10	Tumor type	WT1		MUC1		MHC-I	
		No. of positive cases	Positive ratio (%)	No. of positive cases	Positive ratio (%)	No. of positive cases	Positive ratio (%)
C66	Ureter cancer	0/4	0.0	4/4	100.0	4/4	100.0
C67	Bladder cancer	0/10	0.0	9/9	100.0	9/9	100.0
C68	Urethral cancer	0/1	0.0	1/1	100.0	1/1	100.0
Malignant neoplasms of eye, brain and other parts of central nervous system							
C70	Malignant meningioma	0/1	0.0	1/1	100.0	1/1	100.0
C71	Malignant brain tumor	1/9	11.1	1/5	20.0	7/7	100.0
Malignant neoplasms of thyroid and other endocrine glands							
C73	Thyroid cancer	0/3	0.0	2/3	66.7	2/3	66.7
C74	Adrenal gland cancer	0/1	0.0	0/0	0/0		
C75	Parathyroid cancer	0/1	0.0	0/1	0.0	1/1	100.0
Malignant neoplasms of ill-defined, secondary and unspecified sites							
C80	Cancer of unknown primary site	1/5	20.0	2/5	40.0	5/5	100.0
Malignant neoplasms, stated or presumed to be primary, of lymphoid, hematopoietic and related tissues							
C84	Peripheral T-cell lymphoma	0/2	0.0	1/1	100.0	2/2	100.0
C85	Malignant lymphoma	0/4	0.0	1/2	50.0	4/4	100.0
Total	187/738	25.3	497/647	76.8	636/645	98.6	

Discussion

We evaluated the WT1 expression in cancer tissues of 738 patients using the anti-WT1 monoclonal (6F-H2) antibody. The positive staining of WT1 in tumor cells was observed in 25.3% of patients, indicating that WT1 was not strongly expressed in many cancer cells. WT1 was expressed in malignant mesothelioma, peritoneal cancer and ovarian cancer (malignancy classification according to ICD-10). On the other hand, malignant mesothelioma, serous adenocarcinoma, mucinous cancer, signet ring cell carcinoma, carcinosarcoma and adenosarcoma (malignancy classification according to ICD-O-3) showed strong WT1 expressions. These findings may provide potentially meaningful information when considering WT1-targeted therapies, such as peptide vaccine and DC vaccine therapies. Antigenic expression in cancer tissues is one of the important factors for the aforementioned antigen-specific immunotherapies. Despite the clinical development of WT1-targeted therapies in recent years, the expression level of WT1 in cancer tissue remains controversial (22, 23).

Nakatsuka *et al.* focused on issues associated with the analysis of the expression of WT1 in the cytoplasm, as well as the nucleus of cells (23). Even though we confirmed the cytoplasmic, as well as nuclear, WT1 expression, a low WT1 positivity rate was observed. Such discrepancies may be explained by the current use of non-standardized

immunohistochemical techniques for measuring WT1 expression levels in cancer cells. Therefore, to effectively implement WT1-targeted therapies, the staining method should be standardized. Similarly, the same issue-solving must be considered when using the tissue expression of programmed death-ligand 1 (PDL-1) as a biomarker of PD-1/PD-L1 pathway blockade for WT1-targeted therapies (24).

From the results of this study, WT1 was found to have the tendency to localize in the nucleus and cytoplasm. In malignancies of the female genital organ and mesothelioma, WT1 was observed to mainly localize in the nucleus and cytoplasm in cancers of the digestive organs. Interestingly, among adenocarcinomas, WT1 localization in the nucleus was observed in 94.1% of patients with ovarian cancer, while only 6.8% of patients with other cancer types showed mostly cytoplasmic WT1 localization.

WT1 was expressed (+, ++ and +++) in 25.3% of the patients; however, only 8.5% of them showed moderate to strong expression (++ and +++). It is in our practice to apply the DC vaccine therapy to patients with more than moderate (++) WT1 expression. Therefore, immunohistochemical staining is essential for DC vaccine therapy.

Since TSAs are not encoded in the normal host genome, oncogenic viral proteins and abnormal proteins arising from mutated somatic cells have recently been receiving attention as a target for immunotherapy (25). TAAs, including WT1, have versatility but not affinity compared with TSAs; thus,

Table IV. Intensity of Wilms' tumor 1 (WT1) immunohistochemical staining.

ICD-O-3	Histological type	Case number (%)						
		-		1+	2+ - 3+	Total		
M801-M804	Large cell carcinoma	20	(71.4)	7	(25.0)	1	(3.6)	28
	Undifferentiated carcinoma							
	Small cell carcinoma							
M805-M808	Squamous cell carcinoma	64	(75.3)	18	(21.2)	3	(3.5)	85
M809-M811	Basal cell carcinoma	1	(100.0)	0		0		1
M812-M813	Transitional cell carcinoma	15	(93.8)	1	(6.3)	0		16
M814-M838	Adenocarcinoma	363	(77.2)	79	(16.8)	28	(6.0)	470
M843	Mucoepidermoid carcinoma	1	(50.0)	1	(50.0)	0		2
M844-M849	Serous adenocarcinoma	24	(55.8)	3	(7.0)	16	(37.2)	43
	Mucinous adenocarcinoma							
	Signet ring cell carcinoma							
M850-M854	Infiltrating duct adenocarcinoma	24	(77.4)	4	(12.9)	3	(9.7)	31
	Paget disease							
M855	Acinar cell carcinoma	1	(100.0)	0		0		1
M856-M857	Adenosquamous carcinoma	3	(75.0)	1	(25.0)	0		4
M858	Malignant thymoma	1	(50.0)	1	(50.0)	0		2
M872-879	Malignant melanoma	5	(62.5)	3	(37.5)	0		8
M881-M883	Fibromyxosarcoma	2	(100.0)	0		0		2
M885-M888	Liposarcoma	1	(100.0)	0		0		1
M889-M892	Myosarcoma	1	(20.0)	3	(60.0)	1	(20.0)	5
M893-M899	Adenosarcoma	3	(37.5)	0		5	(62.5)	8
	Carcinosarcoma							
M904	Synovial sarcoma	2	(100.0)	0		0		2
M905	Malignant mesothelioma	0	(0.0)	0		5	(100.0)	5
M906-M909	Germ cell tumor	1	(100.0)	0	0	1		2
M912-M916	Hemangiosarcoma	0	(0.0)	1	(100.0)	0		1
M918-M924	Chondrosarcoma	2	(100.0)	0		0		2
M938-M948	Malignant glioma	7	(87.5)	1	(12.5)	0		8
M949-M952	Neuroepithelioma	1	(100.0)	0		0		1
M953	Malignant meningioma	1	(100.0)	0		0		1
M959-M972	Malignant lymphoma	6	(100.0)	0		0		6
Unknown	Unknown	2	(50.0)	1	(25.0)	1	(25.0)	4
Total	551		(74.7)	124	(16.8)	63	(8.5)	738

TAAAs may not be capable of eliciting effective antitumor immunoresponses. Therefore, an effective extensive induction of the immunoresponses of T lymphocytes, along with DCs, CD4+ T cells and CD8+ T cells in the body, using long peptides, is a way to realize the beneficial effects of DC vaccine therapy *via* TAAAs (26).

Positive staining of tumor cells by the anti-MUC1 monoclonal antibody was observed in 76.8% of samples. Many samples of cancer tumor cells showed MUC1 expression, whereas a weak expression was observed in some types of cancer of the stomach, colon, rectum, liver or gallbladder. These findings provide information useful for determining the efficacy of MUC1-targeted therapy similarly to WT1-targeted therapy. MUC1, a TAA, is expressed in various types of somatic cells; consequently, there is an undeniable possibility that lymphocytes are anergic (27) towards MUC1 and may not also be activated by a vaccine.

A positive staining of tumor cells by the anti-MHC class I monoclonal antibody was observed in 98.6% of samples. For application of specific immunotherapy, this finding indicates that the presence of MHC class I is not a limiting factor. It is the selection of the appropriate cancer antigens that is of importance. Therefore, CD8+ T lymphocyte-based therapies, in relation to the cancer immunity cycle, can form the basis for immune-cell therapy. Inhibitory NK cell receptors recognize self-MHC class I molecules, that prevent NK cell activation (28), on the basis of which the 'missing-self' hypothesis has been proposed. In the practice of NK cell therapy, this finding shows the importance of confirming the presence of rare MHC class I abnormalities by prior examinations.

In conclusion, relating to the application of cancer-specific immunotherapy, these findings provide information useful for determining the efficacy of MUC1- and WT1-targeted therapy.

Table V. Distribution of Wilms' tumor 1 (WT1) protein in cells determined by immunohistochemical staining.

ICD-O-3	Histological type	Case number (%)							
		Nucleus		Cytoplasm		Nucleus & cytoplasm		Unknown	Total
M801-M804	Large cell carcinoma	2	(25.0)	6	(75.0)	0		0	8
	Undifferentiated carcinoma								
	Small cell carcinoma								
M805-M808	Squamous cell carcinoma	4	(19.0)	16	(76.2)	1	(4.8)	0	21
M812-M813	Transitional cell carcinoma	0		1	(100.0)	0		0	1
M814-M838	Adenocarcinoma	22	(20.6)	79	(73.8)	3	(2.8)	3	107
M843	Mucoepidermoid carcinoma	0		1	(100.0)	0		0	1
M844-M849	Serous adenocarcinoma	18	(94.7)	1	(5.3)	0		0	19
	Mucinous adenocarcinoma								
	Signet ring cell carcinoma								
M850-M854	Infiltrating duct adenocarcinoma	2	(28.6)	5	(71.4)	0		0	7
	Paget disease								
M856-M857	Adenosquamous carcinoma	0		1	(100.0)	0		0	1
M858	Malignant thymoma	0		0		1	(100.0)	0	1
M872-879	Malignant melanoma	2	(66.7)	0		1	(33.3)	0	3
M889-M892	Myosarcoma	3	(75.0)	0		1	(25.0)	0	4
M893-M899	Adenosarcoma	5	(100.0)	0		0		0	5
	Carcinosarcoma								
M905	Malignant mesothelioma	4	(80.0)	0		1	(20.0)	0	5
M912-M916	Hemangiosarcoma	0		1	(100.0)	0		0	1
M938-M948	Malignant glioma	0		0		0		1	(100.0)
Unknown	Unknown	1	(50.0)	0		0		1	(50.0)
Total		63	(33.7)	111	(59.4)	8	(4.3)	5	(2.7)

References

- Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, Ly A, Lie WR, Hildebrand WH, Mardis ER and Linette GP: Cancer immunotherapy. A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. *Science* 348: 803-808, 2015.
- Takahara M, Miyai M, Tomiyama M, Mutou M, Nicol AJ and Nieda M: Copulsing tumor antigen-pulsed dendritic cells with zoledronate efficiently enhance the expansion of tumor antigen-specific CD8⁺ T cells *via* Vgamma9gamma delta T cell activation. *J Leukoc Biol* 83: 742-754, 2008.
- Kamigaki T, Takahara M, Maekawa R and Goto S: Zoledronate-pulsed dendritic cell-based anticancer vaccines. *Oncoimmunology* 2: e25636, 2013.
- Kamigaki T, Kaneko T, Naitoh K, Takahara M, Kondo T, Ibe H, Matsuda E, Maekawa R and Goto S: Immunotherapy of autologous tumor lysate-loaded dendritic cell vaccines by a closed-flow electroporation system for solid tumors. *Anticancer Res* 33: 2971-2976, 2013.
- Kamigaki T, Matsuda E, Okada S, Naitoh K, Kondo T, Ibe H, Maekawa R and Goto S: Prospective evaluation of safety of immune-cell therapy for patients with various types of advanced cancer. *Anticancer Res* 34: 4601-4607, 2014.
- Kamigaki T, Ibe H, Okada S, Matsuda E, Tanaka M, Oguma E, Kinoshita Y, Ogasawara S, Ono A, Makita K, Naitoh K and Goto S: Improvement of impaired immunological status of patients with various types of advanced cancers by autologous immune cell therapy. *Anticancer Res* 35: 4535-4543, 2015.
- Anguille S, Smits EL, Lion E, van Tendeloo VF and Berneman ZN: Clinical use of dendritic cells for cancer therapy. *Lancet Oncol* 15: 257-267, 2014.
- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A and Urba WJ: Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 363: 711-723, 2010.
- Brahmer J, Reckamp KL, Baas P, Crinò L, Eberhardt WE, Poddubskaya E, Antonia S, Pluzanski A, Vokes EE, Holgado E, Waterhouse D, Ready N, Gainor J, Arén Frontera O, Havel L, Steins M, Garassino MC, Aerts JG, Domine M, Paz-Ares L, Reck M, Baudelet C, Harbison CT, Lestini B and Spigel DR: Nivolumab versus Docetaxel in advanced squamous-cell non-small-cell lung Cancer. *N Engl J Med* 373: 123-135, 2015.
- Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, Skora AD, Lubner BS, Azad NS, Laheru D, Biedrzycki B, Donehower RC, Zaheer A, Fisher GA, Crocenzi TS, Lee JJ, Duffy SM, Goldberg RM, de la Chapelle A, Koshiji M, Bhajee F, Huebner T, Hruban RH, Wood LD, Cuka N, Pardoll DM, Papadopoulos N, Kinzler KW, Zhou S, Cornish TC, Taube JM, Anders RA, Eshleman JR, Vogelstein B and Diaz LA Jr.: PD-1 Blockade in tumors with mismatch-repair deficiency. *New Eng J Med* 372: 2509-2520, 2015.

- 11 Simpson AJ, Caballero OL, Jungbluth A, Chen YT and Old LJ: Cancer/testis antigens, gametogenesis and cancer. *Nat Rev Cancer* 5: 615-625, 2005.
- 12 Vigneron N, Stroobant V, Van den Eynde BJ and van der Bruggen P: Database of T cell-defined human tumor antigens: the 2013 update. *Cancer Immun* 13: 15. 2013.
- 13 Caballero OL and Chen YT: Cancer/testis (CT) antigens: potential targets for immunotherapy. *Cancer Sci* 100: 2014-2021, 2009.
- 14 Ohnishi H, Yasukawa M and Fujita S: HLA class I-restricted lysis of leukemia cells by a CD8(+) cytotoxic T-lymphocyte clone specific for WT1 peptide. *Blood* 95: 286-293, 2000.
- 15 Ohno S, Dohi S, Ohno Y, Kyo S, Sugiyama H, Suzuki N and Inoue M: Immunohistochemical detection of WT1 protein in endometrial cancer. *Anticancer Res* 29: 691-695, 2009.
- 16 Nakamori S, Ota DM, Cleavy KR, Shirota K and Irimura T: MUC1 mucin expression as a marker of progression and metastasis of human colorectal carcinoma. *Gastroenterology* 106: 353-361, 1994.
- 17 Pardoll D: Does the immune system see tumors as foreign or self? *Annu Rev Immunol* 21: 807-839, 2003.
- 18 Stone JD, Harris DT and Kranz DM: TCR affinity for p/MHC formed by tumor antigens that are self-proteins: impact on efficacy and toxicity. *Curr Opin Immunol* 33: 16-22, 2015.
- 19 Kitawaki T, Kadowaki N, Fukunaga K, Kasai Y, Maekawa T, Ohmori K, Kondo T, Maekawa R, Takahara M, Nieda M, Kuzushima K, Ishikawa T and Uchiyama T: A phase I/IIa clinical trial of immunotherapy for elderly patients with acute myeloid leukaemia using dendritic cells co-pulsed with WT1 peptide and zoledronate. *Br J Haematol* 153: 796-799, 2011.
- 20 Rivoltini L, Barracchini KC, Viggiano V, Kawakami Y, Smith A, Mixon A, Restifo NP, Topalian SL, Simonis TB, Rosenberg SA and Marincola FM: Quantitative correlation between HLA class I allele expression and recognition of melanoma cells by antigen-specific cytotoxic T lymphocytes. *Cancer Res* 55: 3149-3157, 1995.
- 21 Chilosi M, Iestani M, Pedron S, Montagna L, Benedelli A, Pizzolo V G and Menestrina F: A rapid immunostaining method for frozen sections. *Biotech Histochem* 69: 235, 1994.
- 22 Oji Y, Miyoshi S, Maeda H, Hayashi S, Tamaki H, Nakatsuka S, Yao M, Takahashi E, Nakano Y, Hirabayashi H, Shintani Y, Oka Y, Tsuboi A, Hosen N, Asada M, Fujioka T, Murakami M, Kanato K, Motomura M, Kim EH, Kawakami M, Ikegame K, Ogawa H, Aozasa K, Kawase I and Sugiyama H: Overexpression of the Wilms' tumor gene WT1 in de novo lung cancers. *Int J Cancer* 100: 297-303, 2002.
- 23 Nakatsuka S, Oji Y, Horiuchi T, Kanda T, Kitagawa M, Takeuchi T, Kawano K, Kuwae Y, Yamauchi A, Okumura M, Kitamura Y, Oka Y, Kawase I, Sugiyama H and Aozasa K: Immunohistochemical detection of WT1 protein in a variety of cancer cells. *Mod Pathol* 19: 804-814, 2006.
- 24 Teixidó C, Karachaliou N, González-Cao M, Morales-Espinosa D and Rosell R: Assays for predicting and monitoring responses to lung cancer immunotherapy. *Cancer Biol Med* 12: 87-95, 2015.
- 25 Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, Ivanova Y, Hundal J, Arthur CD, Krebber WJ, Mulder GE, Toebes M, Vesely MD, Lam SS, Korman AJ, Allison JP, Freeman GJ, Sharpe AH, Pearce EL, Schumacher TN, Aebbersold R, Rammensee HG, Melief CJ, Mardis ER, Gillanders WE, Artyomov MN and Schreiber RD: Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature* 515: 577-581, 2014.
- 26 Rosalia RA, Quakkelaar ED, Redeker A, Khan S, Camps M, Drijfhout JW, Silva AL, Jiskoot W, van Hall T, van Veelen PA, Janssen G, Franken K, Cruz LJ, Tromp A, Oostendorp J, van der Burg SH, Ossendorp F and Melief CJ: Dendritic cells process synthetic long peptides better than whole protein, improving antigen presentation and T-cell activation. *Eur J Immunol* 43: 2554-2565, 2013.
- 27 Agrawal B, Krantz MJ, Reddish MA and Longenecker BM: Cancer-associated MUC1 mucin inhibits human T-cell proliferation, which is reversible by IL-2. *Nat Med* 4: 43-49, 1998.
- 28 Kärre K, Ljunggren HG, Piontek G and Kiessling R: Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy. *Nature* 319: 675-678, 1986.

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