# MicroRNA Expressions in HPV-induced Cervical Dysplasia and Cancer

KATALIN GOCZE<sup>1</sup>, KATALIN GOMBOS<sup>1</sup>, KRISZTINA KOVACS<sup>2</sup>, KRISZTINA JUHASZ<sup>1</sup>, PETER GOCZE<sup>3</sup> and ISTVAN KISS<sup>1</sup>

Institutes of <sup>1</sup>Public Health and <sup>2</sup>Pathology, University of Pecs, Pecs, Hungary; <sup>3</sup>Department of Obstetrics and Gynecology, University of Pecs, Pecs, Hungary

**Abstract.** Background/Aim: The role of oncogenic or highrisk human papillomavirus (HPV) in cervical carcinogenesis is inevitable, yet not fully understood. Detailed analysis of microRNA (miRNA) alterations occurring during high-risk HPV transformation will increase our current understanding over cervical carcinogenesis. The two main aims of the study were: (i) finding association between HPV infection characteristics and socio-demographic variables, (ii) finding an predictors of clinical outcome. Materials and Methods: The expression levels of different microRNAs (miR-21, miR-27a, miR-34a, miR-155, miR-196a, miR-203) were determined in formalin-fixed paraffin-embedded (FFPE) human HPV-positive cervical dysplastic and tumorous tissue samples using quantitative real-time PCR (qPCR). Sociodemographic and life-style factors were also analyzed. Results: The expression of miR-27a was significantly higher in cervical intraepithelial neoplasia (CIN)2-3 compared to CIN1 (p=0.023) and in squamous cell carcinoma (SCC) compared to CIN2-3 (p=0.033). Moreover, significantly lower levels of miR-34a were detected in CIN2-3 than in CIN1 (p=0.041) and in SCC than in CIN2-3 (p=0.021). Furthermore, we found significant differences in subjects with multiple HPV in miR-27a (p=0.015) and miR-203 (p=0.025) in CIN2-3 compared to CIN1 and miR-21 (p=0.002), mir-27a (p=0.001) and miR-34a (p=0.001) in SCC/CIN2-3. Expression of miR-27a, showing up-regulation in CIN2-3 compared to CIN1 (p=0.028) and miR-34a (downregulated), correlated with HPV 16 positivity (CIN2-3/CIN1: p=0.027 and SCC/CIN2-3: p=0.036). MiR-34a expression was also significantly altered in connection to smoking status and presence of HPV 16. Conclusion: The demand for

Correspondence to: Katalin Gocze, Institute of Public Health, University of Pecs, Szigeti ut 12, Pecs, H-7624, Hungary. Tel: +36 72 536394, Fax: +36 72 536395, e-mail: katag@hotmail.com

Key Words: Cervical intraepithelial neoplasia, cervical cancer, microRNA expression, single and multiple HPV infection.

additional, alternative molecular biomarkers with prognostic potential is strong. Evaluation of miRNA expression might be helpful to distinguish different cervical lesions and might be able to help in the prediction of HPV infection outcome.

Based on the latest worldwide cancer statistics, cervical cancer is the third most commonly diagnosed cancer in females with approximately 530,000 new cases annually and with 275,100 deaths in 2008 (1). The initial hypothesis concerning the link of human papilloma virus (HPV) infection with the development of cervical cancer was proposed by zur Hausen nearly 40 years ago (2). HPV 16 and 18 are known to be highly associated with cervical cancer (3). HPV 16 is invariably the most common HPV type, regardless of study design and geographical area. HPV 18 is also common but its prevalence and incidence is more variable across populations (4). Other common high-risk (HR) HPV types include: 31, 33, 35, 45, 52 and 58. Together with 16 and 18, these HR HPVs show a combined worldwide relative contribution of above 90% (5). Through the course of this virus-mediated carcinogenesis, the initial, persistent oncogenic or high-risk HPV infection can progress to well-defined precursor lesions of different grade, known as cervical intraepithelial neoplasia (CIN). More than 80% of cervical cancer (CC) cases are histologically squamous cell carcinomas (SCC) preceded by high-grade dysplasia (CIN2-3).

The Pap smear (cytology) was a revolutionary technique with a major public health impact - that has changed dramatically the incidence of cervical cancer since its widespread introduction in screening programs in developed countries. One of its limitations is the lack of sufficient information regarding infection and disease outcome (6) besides the frequently mentioned inter-observer variation (7). On the other hand, with extensive HPV-testing a high rate of insignificant HPV infections are revealed. Even though cotesting (HPV testing combined with cytology) is a reasonable alternative, the need for alternative molecular biomarkers with prognostic value, especially regarding HPV infection outcome, is overwhelming.

0250-7005/2015 \$2.00+.40 523

MicroRNAs (miRNAs) are small, non-protein-coding RNAs representing a new class of regulatory mechanisms with important functions in a wide range of biological processes. Aberrant or altered miRNA expression has been reported in a variety of human cancers with a wide range of clinical potential. miRNAs are not only tissue-, organ- or cell-specific. Their de-regulation, caused by genetic and/or epigenetic mechanisms, plays a role in malignant transformation (8, 9) as both mechanisms were shown to contribute to the alterations in miRNA expression in cervical cancer. A more detailed analysis of miRNA alterations occurring during high-risk HPV transformation will increase our current understanding over cervical carcinogenesis. In addition, miRNAs are considered to be a better choice for expression studies since they are more accurate for distinguishing disease states than mRNA expression analysis (9). Archival collection of formalin-fixed, paraffin-embedded (FFPE) tissues is widely used for biomarker research due to the link to rich clinical databases and utility. Previous studies have already demonstrated that miRNAs are minimally affected and display reliable expression levels regardless of fixation time or age of tissue blocks (10). Their small size and a possible protein protection by the RNA-induced silencing (RISC) complex are thought to contribute to their stability during fixation and processing (11). Furthermore, a high correlation has been shown by Goswani et al. between miRNA expression profiles in FFPE and fresh-frozen samples (12).

## Materials and Methods

Samples. A total of 98 female patients' (admitted between 2009-2014 to the Department of Obstetrics and Gynaecology, University of Pécs) cervical FFPE tissue samples (obtained either by conisation or hysterectomy), known to be high-risk HPV-positive (pre-selection criteria based on previous HPV genotyping) with definitive histological diagnosis of cervical lesions (CIN1, CIN2-3 and invasive cervical cancer), were used in this study. Histological diagnoses were produced by the same gynaecological pathologist. Tissue blocks were retrieved from the Department of Pathology, University of Pécs, according to approved protocols (Ethics Committee of Pécs University: 3440.316-8331/KK41/2009) and were microdissected restraining to selected morphologically distinctive areas. A total of 4 pieces of 10 µm thick FFPE sections were collected and used for further analysis.

Clinical specimens were grouped according to histological diagnosis: 1, mildly dysplastic lesions (CIN1: n=30), which typically spontaneously regress; 2, precancerous lesions (CIN2: n=10 and CIN3: n=20), about 20% would progress to invasive cervical cancer; and 3, squamous cell cervical carcinoma (SCC: n=38). Single (n=65) and multiple (n=33) HR HPV-positive samples were found in every category. Low-risk HPV types (6 and 11) were identified in one case simultaneously with HR HPV positivity.

RNA extraction and microRNA real-time polymerase chain reaction (qPCR) expression analysis. Steps include deparaffinization (xylol,

absolute alcohol) prior to total RNA isolation using the Trizol protocol. Reverse transcription and qPCR reactions, along with detection settings, were carried out as described previously (13). The miRNAs investigated in this study (miR-21, miR-27a, miR-34a, miR-155, miR-196a, miR-203) were systematically chosen based on our previous results and published literature (cell lines, clinical specimen) (13-18). Overall, the selected miRNAs were reported to be involved in oncogenesis and progression in cervical and other neoplasms (over- or under-expression). All runs, including no template controls (nuclease-free water), were carried out in triplicate. Inter-run calibrators were used to calculate correction factors to remove the run-to-run differences. Data normalization was performed using 5SrRNA and U6snRNA as endogenous references. Relative miRNA expression levels were compared via the  $2^{-\Delta\Delta CCT}$  method.

Statistical methods. Statistical analyses were performed using the IBM SPSS Version 21 (Armonk, New York, USA). Mann-Whitney *U*-test, unpaired *t*-test and one-way analysis of variance (ANOVA) were used to compare continuous and categorical variables based on data distribution. Logistic regression analyses were conducted to identify variables (miRNA expressions, sociodemographic and lifestyle factors) associated with worse pathological outcomes. The level of significance was set at *p*<0.05.

### Results

As far as for the distribution of HR HPV, HPV 16 was the most prevalent (57.53%) considering both single and multiple infections; types 18 and 31 were the second most frequent (10.96%) followed by 56 (9.59%) and types 51, 52 and 58 (8.22%). These findings correspond to the results of the first Hungarian HPV-Center in Budapest from 2007-2011 (19). Galamb *et al.* detected a relatively low representation (compared to data from developed countries) of HPV 18 along with high prevalence of other, not so common, HPV types. At the same time, the prevalence of types 51 and 31 among patients with cytological abnormality exceeded the expectations based on international data reflecting dominance of types 16, 18 and 45 (3-5). The prevalence of multiple HPV infection was 33.67%, much lower than expected based on previous data (19).

As described previously, the total of 98 samples were grouped according to histological diagnosis (CIN1: 30.61%, CIN2-3: 30.61% and SCC: 38.78%) and the presence of single or multiple HPV positivity. The mean age of the studied population was 36.17 (range: 18-65). Mean age was significantly lower among multi-HPV-positive patients than among mono-HPV-positive individuals ( $33.30\pm7.994$  compared to  $37.98\pm10.516$ , p=0.039). The mean age for patients infected with multiple HPV types was lower in every disease stage than the mean age for patients with single HPV infection, with the only exception being the CIN1 group where the mean age and the age range were the same or quite similar for both subcategories (single:  $33.2\pm8.804$  and multiple:  $33.2\pm10.207$ ).

Table I. Differential expression of miRNAs in dysplastic tissues of different grade. The expression values were calculated according to the 2nd derivative maximum method and normalized to 5S rRNA. The Mann-Whitney U-test was used to evaluate statistical significance.

	Cervical intraepithelial neoplasia (CIN1) Median (range)	Cervical intraepithelial neoplasia (CIN2-3) Median (range)	Fold change	p-Value
miR-21	4.68×10 <sup>-2</sup>	6.87×10 <sup>-2</sup>	1.468	0.051
	$(2.22 \times 10^{-3}$	$(7.33 \times 10^{-3}$		
	3.14)	9.97×101)		
miR-27a	$7.66 \times 10^{-4}$	$4.67 \times 10^{-3}$	6.097	0.023
	$(8.13 \times 10^{-5}$	$(3.60 \times 10^{-5}$ -		
	$1.20 \times 10^{-2}$	$6.27 \times 10^{-2}$		
miR-34a	$1.29 \times 10^{-2}$	$2.91 \times 10^{-3}$	0.226	0.041
	$(2.05 \times 10^{-4}$ -	$(2.16 \times 10^{-5}$ -		
	8.00)	$4.48 \times 10^{-2}$		
miR-155	$8.08 \times 10^{-4}$	$8.47 \times 10^{-4}$	1.048	0.023
	$(5.2 \times 10^{-5}$ -	$(1.06 \times 10^{-5}$ -		
	$1.69 \times 10^{-2}$	$1.23 \times 10^{-2}$		
miR-196a	$3.71 \times 10^{-4}$	$1.2 \times 10^{-3}$	3.234	0.105
	$(8.83 \times 10^{-6}$ -	$(4.78 \times 10^{-6}$ -		
	$7.48 \times 10^{-3}$ )	$2.93 \times 10^{-2}$		
miR-203	$1.36 \times 10^{-3}$	$7.74 \times 10^{-4}$	0.569	0.198
	$(1.48 \times 10^{-5}$	$(1.08 \times 10^{-5}$ -		
	5.00)	$8.76 \times 10^{-3}$ )		

Table II. Differential expression of miRNAs in dysplastic and tumour tissues. The expression values were calculated according to the 2nd derivative maximum method and normalized to 5S rRNA. The Mann-Whitney U-test was used to evaluate statistical significance.

	Cervical intraepithelial neoplasia (CIN2-3) Median (range)	Squamous cellular cancer (SCC) Median (range)	Fold change	<i>p</i> -Value
miR-21	6.87×10 <sup>-2</sup>	2.22	32.314	0.552
	$(7.33 \times 10^{-3}$	$(8.75 \times 10^{-3}$		
	$9.97 \times 10^{1}$	$1.22 \times 10^3$ )		
miR-27a	$4.67 \times 10^{-3}$	$3.18 \times 10^{-3}$	0.681	0.033
	$(3.60 \times 10^{-5}$ -	$(8.02 \times 10^{-6})$		
	$6.27 \times 10^{-2}$	-8.00)		
miR-34a	$2.91 \times 10^{-3}$	$5.46 \times 10^{-4}$	0.188	0.025
	$(2.16 \times 10^{-5}$ -	$(4.16 \times 10^{-5}$ -		
	$4.48 \times 10^{-2}$	$1.18 \times 10^{-2}$		
miR-155	8.47×10 <sup>-4</sup>	$1.27 \times 10^{-3}$	1.499	0.132
	$(1.06 \times 10^{-5}$ -	$(3.23 \times 10^{-6}$ -		
	$1.23 \times 10^{-2}$	5.00)		
miR-196a	$1.2 \times 10^{-3}$	$5.85 \times 10^{-4}$	0.488	0.016
	$(4.78 \times 10^{-6}$ -	$(7.25 \times 10^{-6}$		
	$2.93 \times 10^{-2}$	5.00)		
miR-203	$7.74 \times 10^{-4}$	4.23×10 <sup>-4</sup>	0.547	0.032
	$(1.08 \times 10^{-5}$ -	$(1.86 \times 10^{-7}$ -		
	$8.76 \times 10^{-3}$ )	5.82×10 <sup>-3</sup> )		

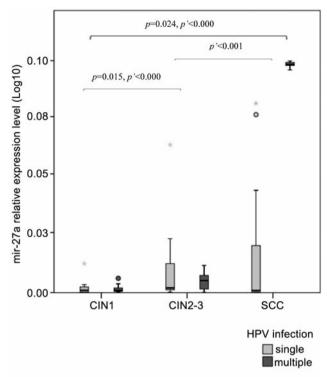


Figure 1. MiRNA-27a levels by patient group (low-grade dysplasia: cervical intraepithelial neoplasia (CIN)1, high-grade dysplasia: CIN2-3, squamous cellular cancer: SCC). Boxes represent the inter-quartile range and the line indicates the median value. Whiskers indicate 95th and 5th percentiles. 5s rRNA was used for normalization. Logistic regression was used to evaluate statistical significance (p' refers to multiple HPV infection).

Significant associations were found as the expression of miR-27a increased with progression: CIN2-3 related to CIN1 (p=0.023) and SCC to CIN2-3 (p=0.033). MiR-155 also showed an increase in the transition from CIN1 to SCC with statistically significant correlation in CIN2-3 over CIN1 (p=0.023). MiR-196a expression showed an increase from CIN1 to CIN2-3 but was significantly lower in SCC than in CIN2-3 (p=0.016). These associations were unaffected by differences in the type or number of HPV. Even though we registered elevations in miR-21 expression through every disease stage from the progression of CIN1 to CC, the differences were not significant. On the other hand, significantly lower levels of miR-34a were detected in CIN2-3 than in CIN1 (p=0.041) and in SCC than in CIN2-3 (p=0.021) (Tables I and II).

Furthermore, we found significant differences in subjects with multiple HPV for miR-27a (p<0.000), miR-34a (p<0.001) and miR-203 (p=0.025) in CIN2-3 compared to CIN1 and for miR-21 (p=0.002) and mir-27a (p<0.001) in SCC/CIN2-3 with miR-21 (p<0.000), miR-27a (p<0.000)

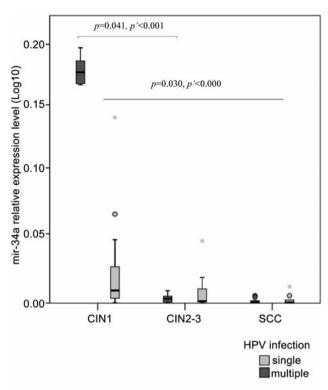


Figure 2. MiRNA-34a levels by patient group (low-grade dysplasia: cervical intraepithelial neoplasia (CIN)1, high-grade dysplasia: CIN2-3, squamous cellular cancer: SCC). Boxes represent the inter-quartile range and the line indicates the median value. Whiskers indicate 95th and 5th percentiles. 5S rRNA was used for normalization. Logistic regression was used to evaluate statistical significance (p refers to multiple HPV infection).

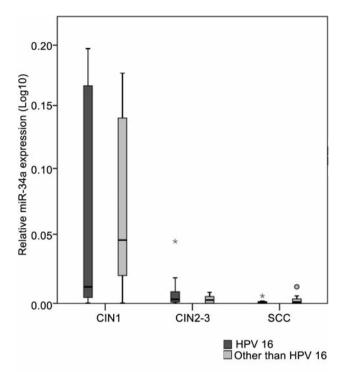


Figure 3. MiRNA-34a levels by patient group (low-grade dysplasia: cervical intraepithelial neoplasia (CIN)1, high-grade dysplasia: CIN2-3, squamous cellular cancer: SCC). Boxes represent the inter-quartile range and the line indicates the median value. Whiskers indicate 95th and 5th percentiles. 5S rRNA was used for normalization. Logistic regression was used to evaluate statistical significance. Transition from CIN1 to CIN2-3: miR-34a correlated with HPV 16 positivity (p=0.027), CIN2-3 to SCC: p=0.036.

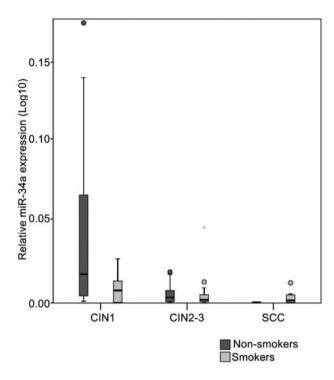
and miR-34a (p<0.000) in CIN1/SCC. The analysis regarding the correlation between miRNA expression and the role of single or multiple HPV showed significant association with disease outcome in the case of miR-27a (p=0.029) and miR-34a (p=0.045) (Figures 1 and 2, respectively).

Regarding other factors, we focused on cigarette smoking and HR HPV type. The presence of HPV 16 has a significant effect on disease outcome (p=0.004) (CIN1, 40%; CIN2-3, 63.33%; SCC, 65.22%). During the transition from CIN1 to CIN2-3, miR-27a and miR-34a correlated with HPV 16 positivity (p=0.028 and p=0.027, respectively). As for the progression of CIN2-3 to SCC, in correlation with HPV 16, miR-34a reached significance (p=0.036) (Figure 3). Our study population had a rate of cigarette smoking at 65% among women with CIN1, 46.67% among women with CIN2-3 and 53.33% among CC subjects. Alteration in miRNA expressions between smokers and non-smokers appeared statistically significant in the case of miR-27a and miR-34a among CIN1 subjects (p=0.036 and p=0.029, respectively) (Figure 4). When, based on smoking history, the

expressions of miR-155 were compared, opposite trends with relative increase among non-smokers and a decrease among smokers in the transition from pre-malignant dysplasia to cancer were shown (Figure 5).

#### Discussion

De-regulation of microRNAs, in connection with malignant transformation, is well known. The etiological relevance of high-risk HPV types in the development of cervical cancer has been demonstrated by a series of epidemiological and biological studies during the past few decades. Nevertheless, a high-risk HPV infection does not inevitably progress to invasive cervical cancer. Multiple HPV genotypes have been found to be a risk factor for persistent infection in healthy young women and co-infections involving HPV 16 and 58 seemed particularly prone to increased risk (4). The phenomenon that we observed in the case of miR-27a and miR-34a, in correlation with HPV 16 positivity during the transition from low-grade to high-grade dysplasia, seems to



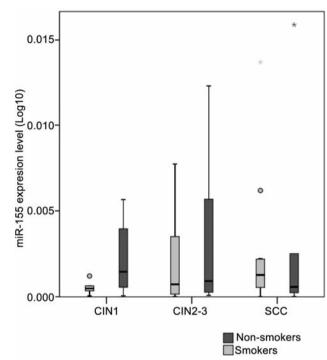


Figure 4. MiRNA-34a levels by patient group (low-grade dysplasia: cervical intraepithelial neoplasia (CIN)1, high-grade dysplasia: CIN2-3, squamous cellular cancer: SCC). Boxes represent the inter-quartile range and the line indicates the median value. Whiskers indicate 95th and 5th percentiles. 5S rRNA was used for normalization. The Mann-Whitney Utest was used to evaluate statistical significance. MiR-34a expression was significantly lower in smokers than non-smokers, p=0.029.

Figure 5. MiRNA-155 levels by patient group (low-grade dysplasia: cervical intraepithelial neoplasia (CIN)1, high-grade dysplasia: CIN2-3, squamous cellular cancer: SCC). Boxes represent the inter-quartile range and the line indicates the median value. Whiskers indicate 95th and 5th percentiles. 5s rRNA was used for normalization.

reinforce this previous observation. Alterations in the expressions of both oncogenic and tumor suppressor miRNAs (miR-27a (Figure 1) and miR-34a (Figure 2)) also verify the opinion that multiple HPV types appear to act synergistically in cervical carcinogenesis (20, 21).

Previous studies have used different types of samples (cultured cells, cancer-derived cell-lines, human tissue samples - fresh-frozen or FFPE, serum) for the analysis of miRNA expression profiles related to cervical cancer (13-18) and, despite the overlapping results obtained, there is a considerable amount of variability (16, 17).

In our study, we wanted to distinguish characteristic alterations in miR-21, miR-27a, miR-34a, miR-155, miR-196a, miR-203 expression in different stages of cervical dysplasia and SCC. MiR-27a is an oncogenic miRNA modulated by p53, E2F and c-Myc. Both HR HPV E6 and E7 interact with c-Myc and augment c-Myc transactivation activities (18). We found that in HR HPV positive tissue samples miR-27a expression correlated with the progression of high grade cervical lesions to cancer making it a credible

biomarker for HR HPV-associated cervical carcinogenesis. A similar phenomenon was observed with higher level of miR-27a in CC compared with CIN3, CIN1+2 or normal cervical tissues in oncogenic HPV-infected raft tissues by Wang *et al.* (22). Our results are unique regarding associations with single and multiple HPV positivity.

MiR-34a is directly regulated by p53 and acts as a tumour suppressor, while HR HPV E6 induces the inhibition of miR-34a through the p53-pathway. It is a downstream target of the p53-network with key regulatory functions in cellular apoptosis, G1-arrest, DNA repair and senescence, which are essential for the maintenance of cellular stability and stress response. Li *et al.* have demonstrated that the expression of miR-34a is significantly lower in HR HPV-infected tissues (23). Our observations clearly confirm that miR-34a acts as a tumour suppressive miRNA in HPV-induced cervical transformation. We also note that alterations in miR-34a expression are associated with the presence of single or multiple HR HPV types supporting the theory of synergism on multiple HPV infection. Cigarette smoke-induced

dysregulation of miRNA expression has been demonstrated in both rats (24) and humans but mainly focusing on lung cells or the bronchial airway epithelium (25). One of the miRNAs significantly dysregulated by cigarette smoke (CS) and implicated in lung carcinogenesis is miR-34a, causing disruption of transcription. Smoke seems to leave a unique "molecular signature" in the p53 gene in smoke-exposed lung tumours (26). In the light of these observations, we can rightly hypothesize that CS might influence miRNA regulatory mechanisms in other organs, such as the cervix as well. Based on these findings, we can conclude that miR-34a could help to predict HPV infection outcome and assist in distinguishing different cervical lesions.

The fact that the expression of miR-155 correlated with the progression of low grade to high grade cervical lesions makes miR-155 eligible as a possible biomarker for HR HPV-associated cervical carcinogenesis. Even though we did not find a significant correlation, the difference in miR-155 expression linked to smoking history must be mentioned. Functionally miR-155 mediates immune response by shaping the transcriptome of lymphoid cells that control diverse biological functions vital in adaptive immunity (27). Its altered expression has also been implicated in viral infections and persistence (Epstein-Barr Virus) (28). Lao et al. registered up-regulated expression of miR-155 in cervical cancer tissues compared to adjacent non-cancer tissues. They found that over-expression of miR-155 promoted the proliferation of Hela and SiHa cells and the down-regulation of miR-155 inhibited the growth of cervical cancer cells and promoted apoptosis and induced cell cycle arrest. They also suggested that miRNA-155 promoted the proliferation of cervical cancer cells by regulating tumour suppressor liver kinase B1 (LKB1) expression (29). Functional studies revealed that miR-155 directly targets the tumour protein 53induced nuclear protein 1 (TP53INP1) and suppresses apoptosis in tumour cells (30, 31). MiR-155 over-expression has been observed in other types of tumours, including oral SCC (OSCC) (32) and head and neck squamous cell carcinomas (HNSCC) (33), also known to be associated with HPV infection.

With respect to variations of miR-196a expression during HPV-mediated cervical carcinogenesis, we confirmed previously reported alterations in tissue samples (16). We found that these alterations were uninfluenced by the dimension of HPV infection (single-multiple, HPV 16 or other). Villegas-Ruiz *et al.* have validated *HOXC8*, involved in embryonic development and cellular remodelling, as a target of miR-196a in CC (34). Hou *et al.* suggested that upregulation of miR-196a enhanced G1/S-phase transition and the proliferative ability of cervical cancer cells through the regulation of the phosphoinositide 3-kinase (PI3K)/Akt signalling pathway (35). It is important to note that miR-196a also plays a distinct role in the regulation of cell proliferation

in HNSCC (36). Wu *et al.* have demonstrated that miR-196a might be involved in tumour invasion and in the interactions of tumour microenvironment in patients with squamous cell lung carcinoma (37). In the case of OSCC, high expression of miR-196a was associated with tumour recurrence, nodal metastasis, mortality and poor survival (38).

We have proven, in conjunction with Pereira *et al.*, that miR-203 shows a relative reduction in expression during the transition from normal tissue to atypical dysplasia and cancer (16). This observation is due to the assumption that high levels of miR-203 (targeting, amongst others, the p63 family of transcription factors) are inhibitory to HPV amplification and that HPV proteins (E7 in particular) act to suppress expression of this microRNA to allow productive replication in differentiating cells (39). It has also been indicated that miR-203 down-regulation in cervical cancer is caused by promoter hypermethylation (40). Investigations regarding invasion and metastasis showed that low expression of miR-203 correlated with lymph node metastasis (41).

# Acknowledgements

We thank Zsuzsanna Brunner and Monika Herczeg for their excellent (precise, coherent) technical assistance. This research was supported by the European Union and the State of Hungary, cofinanced by the European Social Fund in the framework of TÁMOP 4.2.4.A/2-11-1-2012-0001 'National Excellence Program'.

# **Conflicts of Interest**

The Authors have declared that no competing interests exist.

## References

- 1 Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D: Global cancer statistics. CA Cancer J Clin 61(2): 69-90, 2011.
- 2 zur Hausen H: Condylomata acuminata and human genital cancer. Cancer Res 36(2 pt 2): 794, 1976.
- 3 Muñoz N, Bosch FX, de Sanjosé S, Herrero R, Castellsagué X, Shah KV, Snijders PJ and Meijer CJ; International Agency for Research on Cancer Multicenter Cervical Cancer Study Group: Epidemiologic classification of human papilloma virus types associated with cervical cancer. N Engl J Med 348(6): 518-527, 2003.
- 4 Trottier H, Mahmud S, Costa MC, Sobrinho JP, Duarte-Franco E, Rohan TE, Ferenczy A, Villa LL and Franco EL: Human papilloma virus infections with multiple types and risk of cervical neoplasia. Cancer Epidemiol Biomarkers Prev 15(7): 1274-1280, 2006.
- 5 de Sanjose S, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Lloveras B, Tous S, Felix A, Bravo LE, Shin HR, Vallejos CS, de Ruiz PA, Lima MA, Guimera N, Clavero O, Alejo M, Llombart-Bosch A, Cheng-Yang C, Tatti SA, Kasamatsu E, Iljazovic E, Odida M, Prado R, Seoud M, Grce M, Usubutun A, Jain A, Suarez GA, Lombardi LE, Banjo A, Menéndez C, Domingo EJ, Velasco J, Nessa A, Chichareon SC, Qiao YL, Lerma E, Garland SM, Sasagawa T, Ferrera A, Hammouda D,

- Mariani L, Pelayo A, Steiner I, Oliva E, Meijer CJ, Al-Jassar WF, Cruz E, Wright TC, Puras A, Llave CL, Tzardi M, Agorastos T, Garcia-Barriola V, Clavel C, Ordi J, Andújar M, Castellsagué X, Sánchez GI, Nowakowski AM, Bornstein J, Muñoz N and Bosch FX; Retrospective International Survey and HPV Time TrendsStudy Group: Human papilloma virus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study. Lancet Oncol 11(11): 1048-1056, 2011.
- 6 Franco EL: A new generation of studies in human papillomavirus DNA testing in cervical cancer screening. J Natl Cancer Inst 101(23): 1600-1601, 2009.
- 7 Pinto AP, Degen M, Villa LL and Cibas ES: Immunomarkers in gynecologic cytology: the search for the ideal 'biomolecular Papanicolaou test'. Acta Cytol 56(2): 109-121, 2012.
- 8 Calin GA and Croce CM: MicroRNA-cancer connection: the beginning of a new tale. Cancer Res *66(15)*: 7390-7394, 2006.
- 9 Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA, Downing JR, Jacks T, Horvitz HR and Golub TR: MicroRNA expression profiles classify human cancers. Nature 435(7043): 834-838, 2005.
- 10 Liu A, Tetzlaff MT, Vanbelle P, Elder D, Feldman M, Tobias JW, Sepulveda AR and Xu X: MicroRNA expression profiling outperforms mRNA expression profiling in formalin-fixed paraffin-embedded tissues. Int J Clin Exp Pathol 2: 519-27, 2009.
- 11 Li J, Smyth P, Flavin R, Cahill S, Denning K Aherne S, Guenther SM, O'Leary JJ and Sheils O: Comparison of miRNA expression patterns using total RNA extracted from matched samples of formalin-fixed paraffin-embedded (FFPE) cells and snap frozen cells. BMC Biotechnol 7: 36, 2007.
- 12 Goswami RS, Waldron L, Machado J, Cervigne NK, Xu W, Reis PP,Bailey DJ, Jurisica I, Crump MR and Kamel-Reid S: Optimization and analysis of a quantitative real-time PCR-based technique to determine microRNA expression in formalin-fixed paraffin-embedded samples. BMC Biotechnol 10: 47, 2010.
- 13 Gocze K, Gombos K, Juhasz K, Kovacs K, Kajtar B, Benczik M, Gocze P, Patczai B, Arany I and Ember I: Unique microRNA expression profiles in cervical cancer. Anticancer Res 33(6): 2561-2567, 2013.
- 14 Banno K, Iida M, Yanokura M, Kisu I, Iwata T, Tominaga E, Tanaka K and Aoki D: MicroRNA in cervical cancer: OncomiRs and tumorsuppressor miRs in diagnosis and treatment. Scientific World Journal 2014: 178075, 2014.
- 15 Kaczkowski B, Morevati M, Rossing M, Cilius F and Norrild B: A Decade of Global mRNA and miRNA Profiling of HPV-Positive Cell Lines and Clinical Specimens. Open Virol J 6: 216-231, 2012.
- 16 Pereira PM, Marques JP, Soares AR, Carreto L and Santos MA: MicroRNA expression variability in human cervical tissues. PLoS One 5(7): e11780, 2010.
- 17 Gómez-Gómez Y, Organista-Nava J and Gariglio P: Deregulation of the miRNAs expression in cervical cancer: human papilloma virus implications. Biomed Res Int *2013*: 407052, 2013.
- 18 Wang X, Wang HK, Li Y, Hafner M, Banerjee NS, Tang S, Briskin D, Meyers C, Chow LT, Xie X, Tuschl T and Zheng ZM: microRNAs are biomarkers of oncogenic human papilloma virus infections. Proc Natl Acad Sci USA 111(11): 4262-4267, 2014.
- 19 Galamb A, Pajor A, Langmár Z and Sobel G: Results of the first human papillomavirus center in Hungary (2007-2011). Orv Hetil 152(45): 1804-1807, 2011.

- 20 Dickson EL, Vogel RI, Geller MA and Downs LS J: Cervical cytology and multiple type HPV infection: a study of 8182 women ages 31-65. Gynecol Oncol 133(3): 405-408, 2014.
- 21 Spinillo A, Dal Bello B, Gardella B, Roccio M, Dacco' MD and Silini EM: Multiple human papilloma virus infection and high grade cervical intraepithelial neoplasia among women with cytological diagnosis of atypical squamous cells of undetermined significance of low grade squamous intraepithelial lesions. Gynecol Oncol 113(1): 115-119, 2009.
- 22 Wang X, Tang S, Le SY, Lu R, Rader JS, Meyers C and Zheng ZM: Aberrant expression of oncogenic and tumour-suppressive microRNAs in cervical cancer is required for cancer cell growth. PLoS One 3(7): e2557, 2008.
- 23 Li B, Hu Y, Ye F, Li Y, Lv W and Xie X: Reduced miR-34a expression in normal cervical tissues and cervical lesions with high-risk human papilloma virus infection. Int J Gynecol Cancer 20(4): 597-604, 2010.
- 24 Izzotti A, Calin GA, Arrigo P, Steele VE, Croce CM and De Flora S: Down-regulation of microRNA expression in the lungs of rats exposed to cigarette smoke. FASEB J 23(3): 806-812, 2009.
- 25 Schembri F, Sridhar S, Perdomo C, Gustafson AM, Zhang X, Ergun A, Lu J, Liu G, Zhang X, Bowers J, Vaziri C, Ott K, Sensinger K, Collins JJ, Brody JS, Getts R, Lenburg ME and Spira A: MicroRNAs as modulators of smoking-induced gene expression changes in human airway epithelium. Proc Natl Acad Sci USA 106(7): 2319-2324, 2009.
- 26 Russ R and Slack FJ: Cigarette-Smoke-Induced Dysregulation of MicroRNA Expression and Its Role in Lung Carcinogenesis. Pulm Med 2012: 791234, 2012.
- 27 Seddiki N, Brezar V, Ruffin N, Lévy Y and Swaminathan S: Role of miR-155 in the regulation of lymphocyte immune function and disease. Immunology *142(1)*: 32-38, 2014.
- 28 Lu F, Weidmer A, Liu CG, Volinia S, Croce CM and Lieberman PM: Epstein-Barr virus-induced miR-155 attenuates NF-kappaB signaling and stabilizes latent virus persistence. J Virol 82: 10436-10443, 2008.
- 29 Lao G, Liu P, Wu Q, Zhang W, Liu Y, Yang L and Ma C: Mir-155 promotes cervical cancer cell proliferation through suppression of its target gene LKB1. Tumour Biol 2014. 2014 Aug 26. [Epub ahead of print]
- 30 Gironella M, Seux M, Xie MJ, Cano C, Tomasini R, Gommeaux J, Garcia S, Nowak J, Yeung ML, Jeang KT, Chaix A, Fazli L, Motoo Y, Wang Q, Rocchi P, Russo A, Gleave M, Dagorn JC, Iovanna JL, Carrier A, Pebusque MJ and Dusetti NJ: Tumor protein 53-induced nuclear protein 1 expression is repressed by miR-155, and its restoration inhibits pancreatic tumour development. Proc Natl Acad Sci USA 104(41): 16170-16175, 2007.
- 31 Ovcharenko D, Kelnar K, Johnson C, Leng N and Brown D: Genome-scale microRNA and small interfering RNA screens identify small RNA modulators of TRAIL-induced apoptosis pathway. Cancer Res *67*(22): 10782-10788, 2007.
- 32 Shi LJ, Zhang CY, Zhou ZT, Ma JY, Liu Y, Bao ZX and Jiang WW: MicroRNA-155 in oral squamous cell carcinoma: Overexpression, localization, and prognostic potential. Head Neck 2014. 2014 Apr 2. doi: 10.1002/hed.23700. [Epub ahead of print]
- 33 Chen D, Cabay RJ, Jin Y, WangA, Lu Y, Shah-Khan M and Zhou X: MicroRNA Deregulations in Head and Neck Squamous Cell Carcinomas. J Oral Maxillofac Res 4(1): e2, 2013.

- 34 Villegas-Ruiz V, Juárez-Méndez S, Pérez-González OA, Arreola H, Paniagua-García L, Parra-Melquiadez M, Peralta-Rodríguez R, López-Romero R, Monroy-GarcíaA, Mantilla-Morales A, Gómez-Gutiérrez G, Román-Bassaure E and Salcedo M: Heterogeneity of microRNAs expression in cervical cancer cells: over-expression of miR-196a. Int J Clin Exp Pathol 7(4): 1389-1401, 2014.
- 35 Hou T, Ou J, Zhao X, Huang X, Huang Y and Zhang Y: MicroRNA-196a promotes cervical cancer proliferation through the regulation of FOXO1 and p27Kip1. Br J Cancer 110(5): 1260-1268, 2014.
- 36 Severino P, Brüggemann H, Andreghetto FM, Camps C, Klingbeil M de F, de Pereira WO, Soares RM, Moyses R, Wünsch-Filho V, Mathor MB, Nunes FD, Ragoussis J and Tajara EH: MicroRNA expression profile in head and neck cancer: HOX-cluster embedded microRNA-196a and microRNA-10b dysregulation implicated in cell proliferation. BMC Cancer 13: 533, 2013.
- 37 Wu H, Haag D, Muley T, Warth A, Zapatka M, Toedt G, Pscherer A, Hahn M, Rieker RJ, Wachter DL, Meister M, Schnabel P, Müller-Decker K, Rogers MA, Hoffmann H and Lichter P: Tumor-microenvironment interactions studied by zonal transcriptional profiling of squamous cell lung carcinoma. Genes Chromosomes Cancer 52(3): 250-264, 2013.

- 38 Liu CJ, Tsai MM, Tu HF, Lui MT, Cheng HW and Lin SC: miR-196a overexpression and miR-196a2 gene polymorphism are prognostic predictors of oral carcinomas. Ann Surg Oncol 20(Suppl 3): S406-414, 2013.
- 39 Melar-New M and Laimins LA: Human Papillomaviruses Modulate Expression of MicroRNA 203 upon Epithelial Differentiation to Control Levels of p63 Proteins. J Virol 84(10): 5212-5221, 2010.
- 40 Zhu X, Er K, Mao C, Yan Q, Xu H, Zhang Y, Zhu J, Cui F, Zhao W and Shi H: miR-203 suppresses tumour growth and angiogenesis by targeting VEGFA in cervical cancer. Cell Physiol Biochem *32(1)*: 64-73, 2013.
- 41 Zhao S, Yao DS, Chen JY and Ding N: Aberrant expression of miR-20a and miR-203 in cervical cancer. Asian Pac J Cancer Prev 14(4): 2289-2293, 2013.

Received September 19, 2014 Revised October 15, 2014 Accepted October 21, 2014