Simultaneous Multianalyte Immunoassay Measurement of Five Serum Tumor Markers in the Detection of Colorectal Cancer*

FRANCO LUMACHI¹, FILIPPO MARINO², ROCCO ORLANDO³, GIORDANO B. CHIARA⁴ and STEFANO M.M. BASSO⁴

Departments of ¹Surgical, Oncological and Gastroenterological Sciences (DiSCOG),

²Pathology and ³Medical and Surgical Sciences, University of Padua, School of Medicine, Padova, Italy;

⁴Surgery 1, S. Maria degli Angeli Hospital, Pordenone, Italy

Abstract. Several serum tumor markers (STMs) have been proposed for the diagnosis of colorectal cancer (CRC), but their detection should be combined to increase accuracy. The measurement of a serum biomarker panel may improve the diagnostic value of single STM and a multianalyte immunoassay approach can shorten assay time and lower sample consumption. The aim of this study was to determine whether the simultaneous multianalyte immunoassay is useful for early detection of CRC. We measured a panel of five STMs namely, carcinoembryonic antigen (CEA), cancer antigen (CA) 19-9 and 72-4, cytokeratin fragment (CYFRA) 21-1, and osteopontin, in a selected homogeneous population of 102 consecutive patients (median age 66 years, range 42-75 years) with Dukes B, G1-2, colorectal adenocarcinoma (cases) and in a group of 99 age- and sex-matched patients suffering from confirmed benign colorectal diseases (controls). Overall, 141 (70.1%) men and 60 (29.9%) women were studied. The highest sensitivity was 45.1% (osteopontin), while the highest specificity was 90.9% (CEA). The accuracy was lower, ranging from 24.9% (CA 19-9) to 67.2% (CEA). CYFRA 21-1 and CA 72-4 had similar sensitivity (35.3% and 31.4%, respectively), but a significantly different specificity (37.4% vs. 89.9%). A combination of the five markers achieved 74.1% sensitivity

*Presented at the European Society of Medical Oncology, 13th World Congress on Gastrointestinal Cancer, Barcelona, Spain, 22-25 June, 2011.

Correspondence to: Professor Franco Lumachi, University of Padua, School of Medicine, Department of Surgical, Oncological and Gastroenterological Sciences (DiSCOG), via Giustiniani 2, 35128 Padova, Italy. Tel: +39 0498211812, Fax: +39 0498214394, e-mail: flumachi@unipd.it

Key Words: Colorectal cancer, serum tumor marker, CEA, CA 19-9, CA 72-4, CYFRA 21-1, osteopontin.

and 94.3% specificity. In conclusion, in patients with CRC all single STMs show low sensitivity and specificity, while the simultaneous measurement of a panel of STMs may increase the diagnostic accuracy. When the sample volume is limited, the multianalyte immunoassay can be a reliable tool for studying patients undergoing laboratory screening for CRC.

Colorectal cancer (CRC) still remains the second and third leading cause of cancer death in the USA and UK, respectively (1). However, both the mortality and incidence rate for CRC have decreased over the last two decades, likely because of advances in early detection and treatments (2, 3).

Several serum tumor markers (STMs) have been proposed for the diagnosis of CRC, but their detection should be combined to increase accuracy. It has been suggested that serum biomarker-based CRC screening should be combined with fecal protein markers, including immunologic fecal occult blood test and the tissue inhibitor of metalloproteinase-1 (TIMP-1) and S100A12, to achieve better results (4). However, measurement of a serum biomarker panel may improve the diagnostic value of single STM and a multianalyte immunoassay approach is useful to shorten assay time, lower sample consumption and reduce overall cost (5, 6).

The aim of this study was to determine whether a simultaneous multianalyte immunoassay technology can be useful for early detection of CRC.

Patients and Methods

We measured a panel of five STMs namely, carcinoembryonic antigen (CEA), cancer antigen (CA) 19-9 and 72-4, cytokeratin fragment (CYFRA) 21-1, and osteopontin in a selected homogeneous population of 102 consecutive patients (median age 66 years, range 42-75 years) with confirmed Dukes B, G1-2, colorectal adenocarcinoma (cases), and in a group of 99 age- and sex-matched patients suffering from confirmed benign colorectal diseases (controls). Overall, 141 (70.1%) men and 60 (29.9%) women were studied.

0250-7005/2012 \$2.00+.40

Table I. Sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV) and accuracy of the single serum tumor markers, odds ratio (OR) estimates, associated 95% confidence interval (CI) and relative p-value obtained using the Pearson's χ^2 test.

Marker	TP	FN	TN	FP	Sensitivity	Specificity	PPV	NPV	Accuracy	OR	95% CI	<i>p</i> -value
CEA	45	57	90	9	44.1%	90.9%	83.3%	61.2%	67.2%	7.89	3.59-17.37	<0.0001
CYFRA 21-1	36	66	37	62	35.3%	37.4%	36.7%	35.9%	36.3%	0.33	1.18-0.58	< 0.0001
Osteopontin	46	56	31	68	45.1%	31.3%	40.4%	35.6%	38.3%	0.37	0.21-0.67	0.007
CA 72-4	32	70	89	10	31.4%	89.9%	76.2%	56.0%	60.1%	4.07	1.87-8.84	0.002
CA 19-9	23	79	27	72	22.5%	27.3%	24.2%	25.5%	24.9%	0.11	0.06-0.21	<0.0001

CEA, Carcinoembryonic antigen; CYFRA 21-1, cytokeratin fragment 21-1; CA 72-4, cancer antigen 72-4; CA 19-9, cancer antigen 19-9; TP, true-positive; FN, false-negative; TN, true-negative; FP, false-positive.

According to the American Joint Committee on Cancer, Dukes B tumors invade the *muscolaris propria* into the subserosa (T3) or other organs or structures (T4), with no regional lymph node metastasis (N0) or distant metastasis (M0), while histologic grade (G) 1 was considered well-differentiated and G2 moderately differentiated (7). Written informed consent was obtained from all the participants. Blood samples were obtained from all participants following overnight fasting, were assayed in duplicates and the average was compared with the manufacturers' standard curves.

A multiplexed sandwich enzyme-linked immunosorbent assay (ELISA) array technology was used and the five microELISA assays were developed using commercially available antibodies. The antigens CEA, CA 19-9, CA 72-4, CYFRA and human osteopontin were analyzed simultaneously (8). The same CEA and osteopontin antibodies used in the microELISA format were also tested in a conventional ELISA format, based on two-sites monoclonal antibody against CEA and rabbit polyclonal antibodies against peptides 288-304 and 211-228 of human osteopontin, respectively (9, 10). ADVIA Centaur® Immunoassay system (Bayer Healthcare, Tarrytown, NY, USA) was used to determine if there was any difference in results between the microELISA.

The obtained cut-off limit values (at 95% specificity) were the following: 4.9 ng/ml CEA; 32.6 U/ml CA 19-9; 8.5 U/ml CA72-4, 2.7 ng/ml CYFRA 21-1; and 811.9 pmol/ml osteopontin.

Sensitivity was defined as true-positives (TP)/TP + falsenegatives (FN); specificity as true-negatives (TN)/TN + falsepositives (FP); positive predictive value (PPV) as TP/(TP+FP); negative predictive value (NPV) as TN/(TN+FN), and accuracy as (TN+TP)/overall patients. Odds ratio (OR) estimates and the associated 95% confidence interval (CI) were obtained. Testing ranges were determined by regression statistics obtained from a comparison of study methods used to calculate the reference limits of a new method (11). The coefficient of variation (R) of test samples at different dilutions calculation was used to determine the interassay precision. The Pearson's chi-square (χ^2) test and the relative *p*-value were also calculated.

Results

Regression analysis indicated that the two micro-ELISA assays for CEA and osteopontin were not statistically different from the conventional assays (R=0.96 and 0.98, respectively; p<0.0001). The sensitivity, specificity, positive predictive value, negative predictive value and accuracy of single STMs are reported in Table I.

None of the tested STMs was sufficiently sensitive for use as a screening marker at 95% specificity. The highest sensitivity was 45.1% for osteopontin and 44.1% (CEA), while the highest specificity was 90.9% for CEA. The accuracy was lower, ranging from 24.9% (CA 19-9) to 67.2% (CEA). Thus, in the diagnosis of CRC, CEA and CA 72-4 were the most useful single STMs.

CYFRA 21-1 and CA 72-4 had similar sensitivity (35.3% and 31.4%, respectively), but a significantly different specificity (37.4% vs. 89.9%). A combination of the five markers achieved 74.1% sensitivity and 94.3% specificity.

Discussion

STMs are chemical substances generated by the reactions of the human body to certain tumors, or are expressed and synthesized by genes in tumor cells and include proteins (glycoproteins), enzymes (isoenzymes), or peptide hormones, which may reveal the presence of cancer (12, 13). Increased serum levels of STMs are significantly associated with certain tumor types (14). Thus, the elevation of STM levels in human serum can be useful for early diagnosis of cancer or recurrence and for monitoring the curative effect of chemotherapy (15).

CEA is one of the most widely used tumor markers, especially for patients with CRC, and the most useful application of CEA is in the detection of liver metastasis. Several studies suggested the usefulness of a combination of CEA plus CA 72-4 and CA 19-9 (16, 17).

CYFRA 21-1 is a marker of advanced urothelial carcinoma of the bladder, as well of stage I-II non-small cell lung carcinoma (18, 19). However, in combination with other STMs, it has also been tested in the detection of CRC and the performance of the marker combination was comparable with fecal immunochemical testing (20, 21).

Osteopontin is a phosphoprotein associated with tumor progression in several types of solid tumors, including CRC (22). Its expression is strongly elevated in patients with metastatic disease and inversely correlates with the interval between diagnosis and resection of colorectal liver metastases

(23). Moreover, osteopontin contributes to CRC development and progression, regulating several tumor functions (24).

Various approaches have been proposed to perform multianalyte immunoassays, including label-free immunoassays and labeled probe methods, which are difficult to develop because of the lack of a detectable protein signal or a signal too weak to quantify the trace amount of analytes (25). Multilabel and spatially resolved assays can offer amplified detection signals for multianalyte immunoassays (26). Unfortunately, they need several labels, such as radioisotopes, fluorescent dyes, enzymes, metal ions, or quantum dots, which limits their application (24, 27, 28). The electrochemical immunoassay has the advantages of small analyte volume required, low detection limit, simple instrumentation, and minimal manipulation, due to a miniaturized assay system (29).

Conclusion

In patients with CRC all single STMs show low sensitivity and specificity, while the simultaneous measurement of a panel of STMs may increase the diagnostic accuracy (14). When the sample volume is limited, the multianalyte immunoassay can be a reliable tool for studying patients undergoing laboratory screening for CRC.

References

- Poston GJ, Tait D, O'Connell S, Bennett A and Berendse S: Diagnosis and management of colorectal cancer: summary of NICE guidance. BMJ 343: d6751, 2011.
- 2 Hao Y, Jemal A, Zhang X and Ward EM: Trends in colorectal cancer incidence rates by age, race/ethnicity, and indices of access to medical care, 1995-2004 (United States). Cancer Causes Control 20: 1855-1863, 2009.
- 3 Van Cutsem E, Dicato M, Arber N, Berlin J, Cervantes A, Ciardiello F, De Gramont A, Diaz-Rubio E, Ducreux M, Geva R, Glimelius B, Glynne Jones R, Grothey A, Gruenberger T, Haller D, Haustermans K, Labianca R, Lenz HJ, Minsky B, Nordlinger B, Ohtsu A, Pavlidis N, Rougier P, Schmiegel W, Van de Velde C, Schmoll HJ, Sobrero A and Tabernero J: Molecular markers and biological targeted therapies in metastatic colorectal cancer: expert opinion and recommendations derived from the 11th ESMO/World Congress on Gastrointestinal Cancer, Barcelona, 2009. Ann Oncol 21(Suppl 6): vi1-vi10, 2010.
- 4 Karl J, Wild N, Tacke M, Andres H, Garczarek U, Rollinger W and Zolg W: Improved diagnosis of colorectal cancer using a combination of fecal occult blood and novel fecal protein markers. Clin Gastroenterol Hepatol 6: 1122-1128, 2008.
- 5 Wilson MS and Nie W: Multiplex measurement of seven tumor markers using an electrochemical protein chip. Anal Chem 78: 6476-6483, 2006.
- 6 Careri M, Elviri L and Mangia A: Element-tagged immunoassay with inductively coupled plasma mass spectrometry for multianalyte detection. Anal Bioanal Chem 393: 57-61, 2009.

- 7 American Joint Committee on Cancer (AJCC): Colon and Rectum. *In*: Cancer Staging Handbook, Sixth edition. New York, Springer-Verlag, pp. 131-132, 2002.
- 8 Liew M, Groll MC, Thompson JE, Call SL, Moser JE, Hoopes JD, Voelkerding K, Wittwer C and Spendlove RS. Validating a custom multiplex ELISA against individual commercial immunoassays using clinical samples. Biotechniques 42: 327-333, 2007.
- 9 Lumachi F, Basso SM, Bonamini M, Marzano B, Milan E, Waclaw BU and Chiara GB: Relationship between preoperative serum markers CA 15-3 and CEA and relapse of the disease in elderly (>65 years) women with breast cancer. Anticancer Res 30: 2331-2334, 2010.
- 10 Wild N, Andres H, Rollinger W, Krause F, Dilba P, Tacke M and Karl J: A combination of serum markers for the early detection of colorectal cancer. Clin Cancer Res 16: 6111-6121, 2010.
- 11 Westgard JO. Internal quality control: planning and implementation strategies. Ann Clin Biochem 40: 593-611, 2003.
- 12 Lumachi F and Basso SMM: Serum tumor markers in breast cancer. In: Progress in Tumor Marker Research. Swenson LI (eds.) New York, Nova Science Publishers, pp. 83-100, 2007.
- 13 Zhang B, Zhang X, Yan H, Xub S, Tang D and Fua W: A novel multi-array immunoassay device for tumor markers based on insert-plug model of piezoelectric immunosensor. Biosens Bioelectron 23: 15-25, 2007.
- 14 Lumachi F and Basso SMM: Serum tumor markers in patients with breast cancer. Expert Rev Anticancer Ther 4: 921-931, 2004.
- 15 Carpelan-Holmström M, Louhimoa J, Stenmanb U-H, Alfthanb H, Järvinena H and Haglunda C: CEA, CA 242, CA 19-9, CA 72-4 and hCGß in the diagnosis of recurrent colorectal cancer. Tumor Biol 25: 228-234, 2004.
- 16 Duffy MJ: Carcinoembryonic antigen as a marker for colorectal cancer: is it clinically useful? Clin Chem 47: 624-630, 2001.
- 17 Carpelan-Holmström M, Louhimo J, Stenman UH, Alfthan H and Haglund C: CEA, CA 19-9 and CA 72-4 improve the diagnostic accuracy in gastrointestinal cancers. Anticancer Res 22: 2311-2316, 2002.
- 18 Washino S, Hirai M, Matsuzaki A and Kobayashi Y: Clinical usefulness of CEA, CA19-9, and CYFRA 21-1 as tumor markers for urothelial bladder carcinoma. Urol Int 87: 420-428, 2011.
- 19 Patel JL, Erickson JA, Roberts WL and Grenache DG: Performance characteristics of an automated assay for the quantitation of CYFRA 21-1 in human serum. Clin Biochem 43: 1449-1452, 2010.
- 20 Hanagiri T, Sugaya M, Takenaka M, Oka S, Baba T, Shigematsu Y, Nagata Y, Shimokawa H, Uramoto H, Takenoyama M, Yasumoto K and Tanaka F: Preoperative CYFRA 21-1 and CEA as prognostic factors in patients with stage I non-small cell lung cancer. Lung Cancer 74: 112-117, 2011.
- 21 Levin B, Lieberman DA, McFarland B, Smith RA, Brooks D, Andrews KS, Dash C, Giardiello FM, Glick S, Levin TR, Pickhardt P, Rex DK, Thorson A and Winawer SJ; American Cancer Society Colorectal Cancer Advisory Group; US Multi-Society Task Force; American College of Radiology Colon Cancer Committee: Screening and surveillance for the early detection of colorectal cancer and adenomatous polyps, 2008: a joint guideline from the American Cancer Society, the US Multi-Society Task Force on Colorectal Cancer, and the American College of Radiology. CA Cancer J Clin 58: 130-160, 2008.

- 22 Rohde F, Rimkus C, Friederichs J, Rosenberg R, Marthen C, Doll D, Holzmann B, Siewert JR and Janssen KP: Expression of osteopontin, a target gene of de-regulated Wnt signaling, predicts survival in colon cancer. Int J Cancer *121*: 1717-1723, 2007.
- 23 Mole DJ, O'Neill C, Hamilton P, Olabi B, Robinson V, Williams L, Diamond T, El-Tanani M and Campbell FC: Expression of osteopontin coregulators in primary colorectal cancer and associated liver metastases. Br J Cancer 104: 1007-1012, 2011.
- 24 Irby RB, McCarthy SM and Yeatman TJ: Osteopontin regulates multiple functions contributing to human colon cancer development and progression. Clin Exp Metastasis 21: 515-523, 2004.
- 25 Wu J, Yan F, Tang J, Zhai C and Ju H: A disposable multianalyte electrochemical immunosensor array for automated simultaneous determination of tumor markers. Clin Chem 53: 1495-1502, 2007.
- 26 Fu Z, Yang Z, Tang J, Liu H, Yan F and Ju F: Channel and substrate zone two-dimensional resolution for chemiluminescent multiplex immunoassay. Analyt Chem 79: 7376-7382, 2007.

- 27 Swartzman EE, Miraglia SJ, Mellentin-Michelotti J, Evangelista L and Yuan PM: A homogeneous and multiplexed immunoassay for high-throughput screening using fluorometric microvolume assay technology. Anal Biochem 271: 143-151, 1999.
- 28 Fu ZF, Liu H and Ju HX: Flow-through multianalyte chemiluminescent immunosensing system with designed substrate zone-resolved technique for sequential detection of tumor markers. Anal Chem 78: 6999-7005, 2006.
- 29 Thomas JH, Kim SK, Hesketh PJ, Halsall HB and Heineman WR: Bead-based electrochemical immunoassay for bacteriophage MS2. Analyt Chem 76: 2700-2707, 2004.

Received December 27, 2011 Revised January 31, 2012 Accepted February 2, 2012