Low-grade Central Osteosarcoma of the Metatarsal Bone: A Clinicopathological, Immunohistochemical, Cytogenetic and Molecular Cytogenetic Analysis

JUN NISHIO¹, HIROSHI IWASAKI², SATOSHI TAKAGI³, HAJIME SEO¹, MIKIKO AOKI², KAZUKI NABESHIMA² and MASATOSHI NAITO¹

Departments of ¹Orthopaedic Surgery, ²Pathology, and ³Plastic and Reconstructive Surgery, Faculty of Medicine, Fukuoka University, Fukuoka, Japan

Abstract. Low-grade central osteosarcoma (LGCOS) is a very rare low-grade malignant neoplasm that is often confused with a variety of benign fibro-osseous lesions. It rarely involves the small tubular bones of the feet. We present an unusual case of LGCOS arising in the third metatarsal bone of a 16-yearold boy. The radiographic appearance was suggestive of a benign lesion. An open biopsy was performed and the initial diagnosis was fibrous dysplasia. The patient underwent curettage of the lesion and packing of the bony defect with a synthetic bone substitute. Histologically, the curetted specimens consisted of spindle cells admixed with irregular bony trabeculae and osteoid. The spindle cells were fairly uniform with mild atypia, and cellularity varied from low to high. Immunohistochemistry showed that the tumor cells were focally-positive for cyclin-dependent kinase 4 and p53, but negative for murine double minute-2. The MIB-1 labeling index was 36.7% in the highest focus. Cytogenetic analysis exhibited the following clonal karvotypic abnormalities: 48,XY,del(6)(p11),add(8)(q24),add(12)(p11.2),+mar1,+mar-2. Spectral karyotyping demonstrated that marker chromosomes were composed mainly of chromosome 6. Metaphase-based comparative genomic hybridization analysis showed a highlevel amplification of 6p12-p21 and gains of 8q21-q24, 10p15, 12q13-q15, and 16q23-q24. Based on these findings, the final diagnosis was revised to LGCOS and the patient was treated with an additional wide excision, followed by reconstruction with a free-vascularized osteocutaneous scapular flap. At 18 months of follow-up, the patient is well with no evidence of

Correspondence to: Jun Nishio, MD, Department of Orthopaedic Surgery, Faculty of Medicine, Fukuoka University, 7-45-1 Nanakuma, Jonan-ku, Fukuoka 814-0180, Japan. Tel: +81 928011011, Fax: +81 928649055, e-mail: jnishio@cis.fukuokau.ac.jp

Key Words: CDK4, cytogenetics, low-grade central osteosarcoma, MDM2, metatarsal bone.

local recurrence or distant metastasis. Our case highlights the diagnostic difficulty of this tumor with limited tissue samples and the importance of immunohistochemical and molecular cytogenetic analyses in ambiguous cases.

Low-grade central osteosarcoma (LGCOS) is a rare but distinct variant of osteosarcoma, accounting for only 1-2% of all osteosarcomas (1). It tends to occur in the metaphysis of long bones, particularly the distal femur and the proximal tibia, and has a peak incidence in the second and third decades of life with no gender predilection. Patients present with intermittent pain, with or without swelling, usually of long duration. The treatment of choice for LGCOS is wide excision, and there is no role for chemotherapy. Unlike conventional osteosarcoma, LGCOS has a relatively good prognosis when treated appropriately.

Histologically, LGCOS consists of spindle cells arranged in an interlacing pattern. These cells exhibit subtle cytological atypia, and rarely carry mitotic figures. The matrix is produced as well-formed bony trabeculae. In some cases, the bone has a classic 'Chinese character' appearance of fibrous dysplasia. De-differentiation may be found either at the time of initial presentation (2-4) or at recurrence (5) and is associated with a poor prognosis (5). Immunohistochemically, the tumor cells are frequently positive for murine double minute-2 (MDM2) and/or cyclindependent kinase-4 (CDK4) (6, 7).

There have been very few cytogenetic studies of LGCOS (8, 9), and no recurrent chromosomal abnormalities have been detected. Metaphase and array-based comparative genomic hybridization (CGH) analyses have demonstrated gain or amplification of 12q13-q15 containing *MDM2* and *CDK4* (7, 10). Amplification of *tetraspanin-31* (formerly known as *SAS*), located on chromosome 12q13, has also been identified in 15% of LGCOS by quantitative polymerase chain reaction (11).

Only a very small number of LGCOS involving the metatarsal bone have been reported in the literature (12-14).

Herein we report an unusual example of LGCOS arising in the third metatarsal bone of an adolescent male with chromosome 6p alterations. We also review the cytogenetic and molecular cytogenetic characteristics of LGCOS, as well as its clinicopathological and immunohistochemical features.

Case Report

A previously good in general health, 16-year-old boy presented with an 18-month history of intermittent pain and swelling in the dorsum of the left foot. There was no history of antecedent trauma. Radiographs demonstrated an expansive lytic lesion in the third metatarsal bone without evidence of cortical destruction or periosteal new bone formation (Figure 1). No cortical destruction or soft tissue extension was seen on computed tomography nor on magnetic resonance imaging. The lesion exhibited intermediate signal intensity on T1-weighted sequences (Figure 2A) and moderately high signal intensity on T2weighted sequences (Figure 2B). T1-weighted contrastenhanced fat-suppressed sequences showed marked enhancement of the lesion (Figure 2C). A bone scintigraphy showed markedly increased uptake in the third metatarsal bone corresponding to the lytic lesion. Based on the clinical and radiological features, a benign condition, such as giantcell reparative granuloma, was strongly suggested.

The patient underwent an open biopsy, and the initial pathological diagnosis was fibrous dysplasia (Figure 3). Intralesional curettage and packing of the bony defect with a synthetic bone substitute were performed. Grossly, the tumor well-demarcated, firm, and was gravish-white. Microscopically, the tumor consisted of spindle cells admixed with irregular bony trabeculae and osteoid (Figure 4A). Hypercellularity, mild cellular atypia, and scattered mitotic figures observed (Figure were 4B). Immunohistochemically, the tumor cells were focally positive for CDK4 (Figure 4C) and p53, but negative for MDM2. The MIB-1 labeling index was 36.7% in the highest focus (Figure 4D). The pathological diagnosis of the curetted specimens was revised to LGCOS.

Cytogenetic analysis revealed a complex karyotypeincluding marker chromosomes (Figure 5). The karyotype was as follows: 48,XY,del(6)(p11),add(8)(q24), add(12)(p11.2), +mar1,+mar2[3]/48,idem,add(15)(q24),add (20)(p11.2)[2]/46,XY[15]. Spectral karyotyping (SKY) analysis showed that marker chromosomes were mainly composed of chromosome 6 (Figure 6). Metaphase-based CGH analysis showed a high-level amplification of 6p12-p21 and gains of 8q21-q24, 10p15, 12q13-q15, and 16q23-q24 (Figure 7). No significant loss of DNA sequences was found.

Two months after curettage, the patient underwent an additional wide excision, followed by reconstruction with a free vascularized osteocutaneous scapular flap. No



Figure 1. Plain radiographs of the left foot showing an expansive lytic lesion in the third metatarsal bone, without evidence of cortical destruction.

chemotherapy was administered. At 18 months of follow-up, the patient is well with no evidence of local recurrence or distant metastasis.

Discussion

LGCOS of the metatarsal bone is extremely rare; only four cases have been to date reported in literature (12-14). As in our case, a preoperative diagnosis of benign conditions, such as enchondroma and fibrous dysplasia, was made in two of those cases (12, 14). Moreover, even LGCOS involving the long bones was initially diagnosed as a benign condition in 40% of cases (15). These findings suggest that it may be difficult or impossible to clinico-radiologically distinguish LGCOS from benign fibro-osseous lesions without the presence of some degree of cortical destruction and/or soft tissue extension.

The differential diagnosis of LGCOS includes fibrous dysplasia and desmoplastic fibroma. The absence of cytological atypia and the presence of a permeative growth pattern may help to distinguish LGCOS from similar benign lesions histologically. In the present case, the open biopsy specimen showed a proliferation of plump spindle cells admixed with branching and anastomosing trabeculae of woven bone. There was neither cytological atypia nor mitotic activity. These features resulted in the initial mis-diagnosis of fibrous dysplasia. In fact, there are several reported cases of LGCOS mimicking fibrous dysplasia, where a limited tissue sample is used (15-17).



Figure 2. Coronal magnetic resonance images of low-grade central osteosarcoma involving the third metatarsal bone. The lesion exhibits intermediate signal intensity on T1-weighted sequence (A) and moderately-high signal intensity on T2-weighted sequence (B). T1-weighted contrast-enhanced fat-suppressed sequence reveals marked enhancement of the lesion (C).

Recent immunohistochemical studies demonstrated that MDM2 and/or CDK4 were expressed in LGCOS, but not in benign fibrous and fibro-osseous lesions (6, 7). The combination of these two markers can therefore serve as a valuable adjunct in the diagnosis of LGCOS. In the present case, we observed that the tumor cells were positive for CDK4. but negative for MDM2. Previous immunohistochemical studies also showed that LGCOS was more commonly positive for CDK4 than MDM2 (6, 11). These observations may be explained by the possibility that CDK4 staining is relatively resistant to de-calcification by acid-based products. On the other hand, Okada et al. (18) showed that proliferative cell activity evaluated by MIB-1 staining was significantly higher in LGCOS than in fibrous dysplasia, offering an advantage in differential diagnosis. Park et al. (19) also reported that the MIB-1 labeling index was lower in LGCOS than in conventional osteosarcoma. These results indicate that the overall mean MIB-1 labeling index is approximately 10% in LGCOS. In contrast, the present case had a high MIB-1 labeling index of 36.7%.

CGH analyses of LGCOS have revealed the gain or amplification of 12q13-q15 involving *MDM2*, *CDK4*, and *TSPAN31* genes (7, 10). It is interesting to note that the 12q13q15 amplification is more common in low-grade osteosarcoma than in high-grade osteosarcoma (20, 21). Moreover, the gain of 12q13-q15 sequences has been shown to correlate with the presence of ring chromosomes in parosteal osteosarcoma (22). The amplification of *MDM2* and *CDK4* can lead to the deregulation of the cell cycle and may therefore play an important role in the progression of LGCOS.

Cytogenetic analysis of LGCOS revealed an inv(6)(p23q15) as the sole anomaly (9). In addition, a previous CGH study

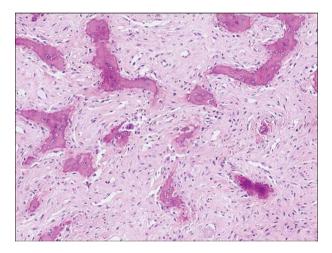


Figure 3. The open biopsy specimen shows a proliferation of spindle cells admixed with bracing trabeculae of woven bone lacking osteoblastic rimming. There is no evidence of cytological atypia. Based on these features, we first made a diagnosis of fibrous dysplasia.

showed a gain of 6p21 in 33% of LGCOS (10). We also identified a rearrangement and an amplification involving chromosome 6p by SKY and CGH. These findings suggest that chromosome 6p alterations seem to recur in LGCOS. On the other hand, amplification of 6p12-p21 has frequently been observed in conventional osteosarcoma (23-27) and appears to be an early event in its pathogenesis. Interestingly, the gain and amplification of murine chromosome regions homologous to human chromosome 6p have also been reported in conditional mouse models of osteosarcoma (28). Several genes with

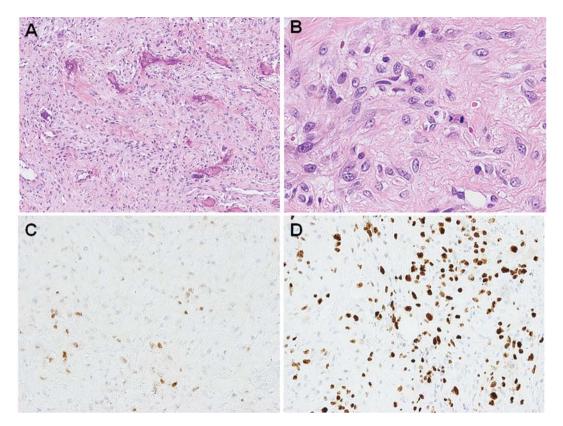


Figure 4. Histological and immunohistochemical findings of low-grade central osteosarcoma. The tumor is composed of spindle cells admixed with irregular bony trabeculae and osteoid (A). Mild cellular atypia and mitotic activity are found (B). The tumor cells are focally positive for cyclindependent kinase-4 (C). The MIB-1 labeling index is 36.7% (D).

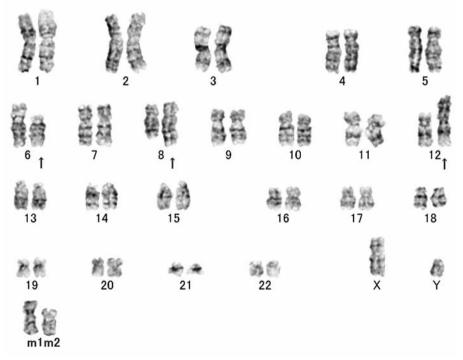


Figure 5. A representative G-banded karyotype of low-grade central osteosarcoma, including two marker chromosomes. Arrows indicate the structural chromosomal aberrations.



Figure 6. Spectral karyotyping of low-grade central osteosarcoma illustrating the origins of two marker chromosomes (arrows). A classified image is displayed alongside the reverse 4',6-diamidino-2-phenylindole image.

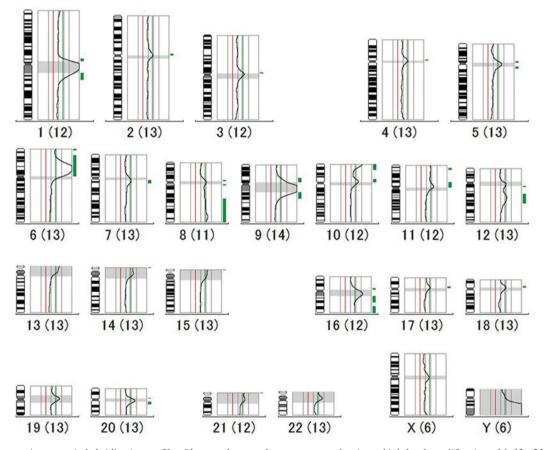


Figure 7. Comparative genomic hybridization profile of low-grade central osteosarcoma showing a high-level amplification of 6p12-p21 and gains of 8q21-q24, 10p15, 12q13-q15, and 16q23-q24. The line in the middle (gray) is the baseline ratio (1.0); the left (red) and right (green) lines indicate ratio values of 0.8 and 1.2, respectively. Bars on the left (red) and right (green) of each frame indicate losses and gains, respectively. The terminology "1 (12)" represents 12 aberrations detected on chromosome 1; the same applies to other chromosomes shown in the profile.

oncogenic potential lie within this chromosome region, such as *CDC5 cell division cycle 5-like (CDC5L)*, *runt-related transcription factor 2 (RUNX2)*, *Cyclin D3 (CCND3)*, *vascular endothelial growth factor A (VEGFA)*, and *pim-1 oncogene (PIM1)*. *CDC5L* is a cell cycle regulator important for the G_2/M transition, and its overexpression may promote mitotic

entry and shorten the G_2 phase (29). Lu *et al.* (26) suggested that overexpression of *CDC5L* through genomic amplification is likely to lead to aberrant cell-cycle control and may contribute to the malignant phenotype of osteosarcoma. *RUNX2* is a member of the RUNX family of transcription factors and encodes a nuclear protein with a Runt DNA-

binding domain. RUNX2 regulates osteoblast lineage determination and expansion, osteoblast maturation, and terminal differentiation via a complex variety of pathways (30). Recently, Sadikovic et al. (31) reported that RUNX2 overexpression was correlated with a poor response to chemotherapy in osteosarcoma. CCND3 is a member of the cyclin-D family and encodes a protein involved in the regulation of the G_1/S transition (32). The amplification of CCND3 has been reported in other types of cancer (33-35). VEGFA is a member of the platelet-derived growth factor/VEGF growth factor family and encodes a protein that is often found as a disulfide-linked homodimer. Its amplification and overexpression have been shown to be poor prognostic factors for survival in osteosarcoma (36, 37). PIM1 is a protooncogene and encodes a serine/threonine protein kinase that is involved in cell proliferation, survival, differentiation, apoptosis, and tumourigenesis (38). PIM1 overexpression has been found in several cancer types (39). However, the precise roles of these amplifications in the pathogenesis and progression of LGCOS remain to be elucidated.

In summary, we have reported a rare case of LGCOS of the metatarsal bone with complex chromosomal rearrangements. Our case highlights the diagnostic difficulty of this tumor with limited tissue samples and the importance of immunohistochemical and molecular cytogenetic analyses in ambiguous cases. Moreover, our results indicate that chromosome 6p alterations may play a critical role in the pathogenesis of LGCOS.

Acknowledgements

This study was supported in part by Fukuoka Cancer Society, Ogata Foundation, and Foundation for the Promotion of Medical Science.

References

- Inwards CY and Knuutila S: Low grade central osteosarcoma. *In*: World Health Organization Classification of Tumours: Pathology and Genetics of Tumours of Soft Tissue and Bone. Fletcher CDM, Unni KK and Mertens F (eds.). Lyon, IARC Press, pp. 275-276, 2002.
- 2 Iemoto Y, Ushigome S, Fukunaga M, Nikaido T and Asanuma K: Case report 679: Central low-grade osteosarcoma with foci of dedifferentiation. Skeletal Radiol 20: 379-382, 1991.
- 3 Ogose A, Hotta T, Emura I, Imaizumi S, Takeda M and Yamamura S: Repeated de-differentiation of low-grade intraosseous osteosarcoma. Hum Pathol 31: 615-618, 2000.
- 4 Kenan S, Ginat DT and Steiner GC: De-differentiated high-grade osteosarcoma originating from low-grade central osteosarcoma of the fibula. Skeletal Radiol 36: 347-351, 2007.
- 5 Kurt AM, Unni KK, McLeod RA and Pritchard DJ: Low-grade intraosseous osteosarcoma. Cancer 65: 1418-1428, 1990.
- 6 Yoshida A, Ushiku T, Motoi T, Shibata T, Beppu Y, Fukayama M and Tsuda H: Immunohistochemical analysis of MDM2 and CDK4 distinguishes low-grade osteosarcoma from benign mimics. Mod Pathol 23: 1279-1288, 2010.

- 7 Dujardin F, Binh MB, Bouvier C, Gomez-Brouchet A, Larousserie F, Muret A, Louis-Brennetot C, Aurias A, Coindre JM, Guillou L, Pedeutour F, Duval H, Collin C and de Pinieux G: MDM2 and CDK4 immunohistochemistry is a valuable tool in the differential diagnosis of low-grade osteosarcomas and other primary fibroosseous lesions of the bone. Mod Pathol 24: 624-637, 2011.
- 8 Bridge JA, Nelson M, McComb E, McGuire MH, Rosenthal H, Vergara G, Maale GE, Spanier S and Neff JR: Cytogenetic findings in 73 osteosarcoma specimens and a review of the literature. Cancer Genet Cytogenet 95: 74-87, 1997.
- 9 Grubb G, Hoctor V, Venter D and Choong P: Inversion (6)(p23q15) as the sole anomaly in a low-grade intraosseous osteosarcoma. Cancer Genet Cytogenet *109*: 70-71, 1999.
- 10 Tarkkanen M, Böhling T, Gamberi G, Ragazzini P, Benassi MS, Kivioja A, Kallio P, Elomaa I, Picci P and Knuutila S: Comparative genomic hybridization of low-grade central osteosarcoma. Mod Pathol 11: 421-426, 1998.
- 11 Ragazzini P, Gamberi G, Benassi MS, Orlando C, Sestini R, Ferrari C, Molendini L, Sollazzo MR, Merli M, Magagnoli G, Bertoni F, Bohling T, Pazzagli M and Picci P: Analysis of *SAS* gene and CDK4 and MDM2 proteins in low-grade osteosarcoma. Cancer Detect Prev 23: 129-136, 1999.
- 12 Lee EY, Seeger LL, Nelson SD and Eckardt JJ: Primary osteosarcoma of a metatarsal bone. Skeletal Radiol 29: 474-476, 2000.
- 13 Andresen KJ, Sundaram M, Unni KK and Sim FH: Imaging features of low-grade central osteosarcoma of the long bones and pelvis. Skeletal Radiol 33: 373-379, 2004.
- 14 Bugnone AN, Temple HT and Pitcher JD: Low-grade central osteosarcoma of the foot and ankle: Radiographic and pathologic features in two patients: Case report and literature review. Foot Ankle Int 26: 494-500, 2005.
- 15 Choong PF, Pritchard DJ, Rock MG, Sim FH, McLeod RA and Unni KK: Low-grade central osteogenic sarcoma. A long-term followup of 20 patients. Clin Orthop Relat Res 322: 198-206, 1996.
- 16 Franceschina MJ, Hankin RC and Irwin RB: Low-grade central osteosarcoma resembling fibrous dysplasia. A report of two cases. Am J Orthop 27: 432-440, 1997.
- 17 Muramatsu K, Hashimoto T, Seto S, Gondo T, Ihara K and Taguchi T: Low-grade central osteosarcoma mimicking fibrous dysplasia: A report of two cases. Arch Orthop Trauma Surg *128*: 11-15, 2008.
- 18 Okada K, Nishida J, Morita T, Kakizaki H, Ishikawa A and Hotta T: Low-grade intraosseous osteosarcoma in northern Japan: advantage of AgNOR and MIB-1 staining in differential diagnosis. Hum Pathol 31: 633-639, 2000.
- 19 Park HR, Jung WW, Bertoni F, Bacchini P, Park JH, Kim YW and Park YK: Molecular analysis of *p53*, *MDM2* and *H-RAS* genes in low-grade central osteosarcoma. Pathol Res Pract 200: 439-445, 2004.
- 20 Wunder JS, Eppert K, Burrow SR, Gokgoz N, Bell RS and Andrulis IL: Co-amplification and overexpression of *CDK4*, *SAS* and *MDM2* occurs frequently in human parosteal osteosarcomas. Oncogene *18*: 783-788, 1999.
- 21 Mejia-Guerrero S, Quejada M, Gokgoz N, Gill M, Parkes RK, Wunder JS and Andrulis IL: Characterization of the 12q15 *MDM2* and 12q13-14 *CDK4* amplicons and clinical correlations in osteosarcoma. Genes Chromosomes Cancer 49: 518-525, 2010.

- 22 Szymanska J, Mandahl N, Mertens F, Tarkkanen M, Karaharju E and Knuutila S: Ring chromosomes in parosteal osteosarcoma contain sequences from 12q13-15: A combined cytogenetic and comparative genomic hybridization study. Genes Chromosomes Cancer 16: 31-34, 1996.
- 23 Forus A, Weghuis DO, Smeets D, Fodstad O, Myklebost O and Geurts van Kessel A: Comparative genomic hybridization analysis of human sarcomas: II. Identification of novel amplicons at 6p and 17p in osteosarcomas. Genes Chromosomes Cancer 14: 15-21, 1995.
- 24 Lau CC, Harris CP, Lu XY, Perlaky L, Gogineni S, Chintagumpala M, Hicks J, Johnson ME, Davino NA, Huvos AG, Meyers PA, Healy JH, Gorlick R and Rao PH: Frequent amplification and rearrangement of chromosomal bands 6p12p21 and 17p11.2 in osteosarcoma. Genes Chromosomes Cancer 39: 11-21, 2004.
- 25 Man TK, Lu XY, Jaeweon K, Perlaky L, Harris CP, Shah S, Ladanyi M, Gorlick R, Lau CC and Rao PH: Genome-wide array comparative genomic hybridization analysis reveals distinct amplifications in osteosarcoma. BMC Cancer 4: 45, 2004.
- 26 Lu XY, Lu Y, Zhao YJ, Jaeweon K, Kang J, Xiao-Nan L, Ge G, Meyer R, Perlaky L, Hicks J, Chintagumpala M, Cai WW, Ladanyi M, Gorlick R, Lau CC, Pati D, Sheldon M and Rao PH: Cell cycle regulator gene CDC5L, a potential target for 6p12p21 amplicon in osteosarcoma. Mol Cancer Res 6: 937-946, 2008.
- 27 Selvarajah S, Yoshimoto M, Ludkovski O, Park PC, Bayani J, Thorner P, Maire G, Squire JA and Zielenska M: Genomic signatures of chromosomal instability and osteosarcoma progression detected by high resolution array CGH and interphase FISH. Cytogenet Genome Res 128: 199-213, 2010.
- 28 Walkley CR, Qudsi R, Sankaran VG, Perry JA, Gostissa M, Roth SI, Rodda SJ, Snay E, Dunning P, Fahey FH, Alt FW, McMahon AP and Orkin SH: Conditional mouse osteosarcoma, dependent on p53 loss and potentiated by loss of Rb, mimics the human disease. Genes Dev 22: 1662-1676, 2008.
- 29 Bernstein HS and Coughlin SR: A mammalian homolog of fission yeast cdc5 regulates G 2 progression and mitotic entry. J Biol Chem 273: 4666-4671, 1998.
- 30 Martin JW, Zielenska M, Stein GS, van Wijnen AJ and Squire JA: The role of RUNX2 in osteosarcoma oncogenesis. Sarcoma 2011: 282745, 2011.

- 31 Sadikovic B, Thorner P, Chilton-Macneill S, Martin JW, Cervigne NK, Squire J and Zielenska M: Expression analysis of genes associated with human osteosarcoma tumors shows correlation of *RUNX2* overexpression with poor response to chemotherapy. BMC Cancer *10*: 202, 2010.
- 32 Motokura T, Keyomarsi K, Kronenberg HM and Arnold A: Cloning and characterization of human cyclin D3, a cDNA closely related in sequence to the *PRAD1*/cyclin D1 protooncogene. J Biol Chem 267: 20412-20415, 1992.
- 33 Büschges R, Weber RG, Actor B, Lichter P, Collins VP and Reifenberger G: Amplification and expression of cyclin D genes (*CCND1*, *CCND2* and *CCND3*) in human malignant gliomas. Brain Pathol 9: 435-442, 1999.
- 34 Kasugai Y, Tagawa H, Kameoka Y, Morishima Y, Nakamura S and Seto M: Identification of *CCND3* and *BYSL* as candidate targets for the 6p21 amplification in diffuse large B-cell lymphoma. Clin Cancer Res *11*: 8265-8272, 2005.
- 35 Lopez-Beltran A, Ordóñez JL, Otero AP, Blanca A, Sevillano V, Sanchez-Carbayo M, Muñoz E, Cheng L, Montironi R and de Alava E: Cyclin D₃ gene amplification in bladder carcinoma *in situ*. Virchows Arch 457: 555-561, 2010.
- 36 Yang J, Yang D, Sun Y, Sun B, Wang G, Trent JC, Araujo DM, Chen K and Zhang W: Genetic amplification of the vascular endothelial growth factor (VEGF) pathway genes, including VEGFA, in human osteosarcoma. Cancer 117: 4925-4938, 2011.
- 37 Ługowska I, Woźniak W, Klepacka T, Michalak E and Szamotulska K: A prognostic evaluation of vascular endothelial growth factor in children and young adults with osteosarcoma. Pediatr Blood Cancer *57*: 63-68, 2011.
- 38 Wang Z, Bhattacharya N, Weaver M, Petersen K, Meyer M, Gapter L and Magnuson NS: Pim-1: A serine/threonine kinase with a role in cell survival, proliferation, differentiation and tumorigenesis. J Vet Sci 2: 167-179, 2001.
- 39 Shah N, Pang B, Yeoh KG, Thorn S, Chen CS, Lilly MB and Salto-Tellez M: Potential roles for the PIM1 kinase in human cancer-a molecular and therapeutic appraisal. Eur J Cancer 44: 2144-2151, 2008.

Received October 2, 2012 Revised October 27, 2012 Accepted October 30, 2012