# Quantitative Structure–Activity Relationship (QSAR) Analysis of Tumor-specificity of 1,2,3,4-Tetrahydroisoquinoline Derivatives

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**Abstract.** Background: We have previously reported on the relative cytotoxicity of a total of 38 1,2,3,4-tetrahydroisoquinoline derivatives against human oral squamous cell carcinoma cell lines and human normal oral cells, and the correlation between the cytotoxicity and 17 chemical descriptors. However, the correlation between the tumorspecificity of these compounds and the chemical descriptors has never been investigated so far. Using these previous data, we investigated various parameters for their applicability in predicting tumor specificity. Materials and Methods: Original data of 50% cytotoxic concentration (CC<sub>50</sub>) values exceeding the maximum concentration in experimental conditions were corrected by the introduction of a harmonic mean, reducing the number of compounds analyzed to 30. The mean pCC<sub>50</sub> (=-log $CC_{50}$ ) values for normal and tumor cells were defined as N and T, respectively. Tumor specificity was defined as the ratio of the difference of these values to their sum: (T-N)/(T+N). The chemical descriptors were obtained by quantum chemical calculations using semi-empirical (AM1, PM3, and PM6) and density functional theory methods. The relationship between the chemical descriptors and tumor specificity was analyzed by linear regression and artificial neural networks. Results: Out of 17 chemical descriptors, water-accessible surface area showed the highest correlation coefficient with tumor specificity, regardless of the method of calculation. Furthermore, neural network analysis demonstrated the importance of quantum

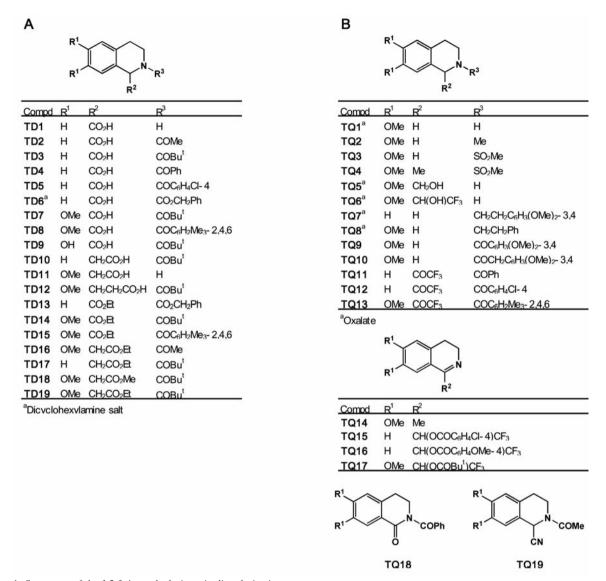
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chemical calculations in predicting the specificity of tetrahydroisoquinoline derivatives. Conclusion: The present study suggests the applicability of quantum chemical descriptor in the estimation of tumor specificity of related compounds.

The incorporation of the 1-methyl-1,2,3,4-tetrahydroisoquinoline (TIO) moiety is an important synthetic strategy in drug discovery (1), and in fact, TIQ is the only endogenous Parkinson-preventing agent discovered to date (2). The high therapeutic properties of the related drugs have encouraged medicinal chemists to synthesize a large number of novel chemotherapeutic agents. Pharmaceutical properties include antineoplastic (3, 4), nitric oxide (NO) inhibition (5), histamine  $H_3$  antagonism and serotonin reuptake inhibition (6),  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptor antagonism (7), bradycardic (8), orexin-2 receptor-selective antagonism (9), multidrug-resistance (MDR) reversion (10-12), γ-secretase inhibition (13), kinase insert domain containing receptor (KDR) inhibition (14), and antidiabetic activities (15). All are unique characteristics of newly synthesized TIQ derivatives. The TIQ family of alkaloids also includes potent cytotoxic agents that display a range of biological properties, such as antitumor and antimicrobial activities (16).

TIQ derivatives were shown to induce neurotoxicity in various animals *via* the decline of ATP levels due to mitochondrial inhibition of complex 1, and DNA damage (2), and inactivation of Cu,Zn-superoxide dismutase, induced by free radical formation (17). TIQ derivatives possessing bulky alkyl group substituents such as 1-cyclobutyl-, 1-cyclohexyl-, 1-phenyl-, or 1-benzyl-, at the C-1 position showed significant cytotoxicity against rat PC12 cells (18). This was confirmed by our recent finding that among 38 newly synthesized TIQ derivatives, TD1-to-19 (Figure 1A) and TQ1-to-19 (Figure 1B), (6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)(3,4-dimethoxyphenyl)methanone (TQ9), with has bulky



 $Figure\ 1.\ Structures\ of\ the\ 1,2,3,4-tetrahydroisoquinoline\ derivatives.$ 

substituents (such as a 3,4-dimethoxybenzoyl group), and ethyl 2-benzyloxycarbonyl-1,2,3,4-tetrahydroisoquinoline-1-carboxylate (TD13), which has an ethoxycarbonyl group and a benzyloxycarbonyl group, showed the highest tumor specific cytotoxicity (TS=12.5 and 5.3, respectively) towards human oral squamous cell carcinoma (OSCC) (19).

We previously applied a quantitative structure–activity relationship (QASR) analysis to delineate the relationship between the cytotoxicity (evaluated by 50% cytotoxic concentration,  $CC_{50}$ ) of these TIQ derivatives, their molecular weight and 17 chemical parameters (descriptors), using a semi-empirical molecular-orbital method (CAChe 4.9, PM5). There was a good correlation between the  $CC_{50}$  of TQ1-to-19 and their hydrophobicity (logP) and the

descriptors for the molecular size such as surface area, volume and width (20). The cytotoxicity of TD1-to-19 depended on hydrophobicity and the distance between C–R2 in the 3-dimensional configuration (21).

However, the relationship between tumor specificity and these chemical descriptors has not yet been reported. We therefore investigated the correlation between tumor specificity of TIQ derivatives and various chemical descriptors.

#### Materials and Methods

Preparation of 1,2,3,4-tetrahydroisoquinoline (TIQ) derivatives. Thirty-eight TIQ derivatives (Figure 1) were prepared as described elsewhere (19).

Table I. Estimated 50% cytotoxic concentration ( $CC_{50}$ ), average log  $CC_{50}^{-1}$  ( $pCC_{50}$ ), and tumor-specificity index values of 38 1,2,3,4-tetrahydroisoquinoline derivatives. N1, N2, and N3 represent human normal cells, gingival fibroblast HGF, pulp cell HPC, and periodontal ligament fibroblast HPLF, respectively. T1, T2, T3, and T4 represent human OSCC cell lines (HSC-2, HSC-3, HSC-4) and human promyelocytic leukemic HL-60 cells, respectively.

Comp. no.			Estin	Average	$pCC_{50}$	Tumor-specificity index				
	N1	N2	N3	T1	T2	Т3	T4	N	Т	(T-N)/(T+N)
TD1	0.800	0.800	0.800	0.800	0.800	0.800	0.800	0.097	0.097	0.000
TD2	0.800	0.800	0.800	0.800	0.800	0.800	0.800	0.097	0.097	0.000
TD3	0.800	0.800	0.800	0.800	0.800	0.800	0.800	0.097	0.097	0.000
TD4	0.800	0.800	0.800	0.800	0.800	0.800	0.788	0.097	0.099	
TD5	0.800	0.800	0.800	0.800	0.800	0.800	0.342	0.097	0.189	0.323
TD6	0.800	0.800	0.800	0.800	0.800	0.800	0.199	0.097	0.248	0.438
TD7	0.800	0.800	0.800	0.800	0.800	0.800	0.800	0.097	0.097	0.000
TD8	0.309	0.330	0.690	0.730	0.800	0.772	0.131	0.384	0.307	
TD9	0.800	0.800	0.800	0.728	0.752	0.784	0.146	0.097	0.301	0.513
TD10	0.800	0.800	0.800	0.800	0.800	0.800	0.720	0.097	0.108	
TD11	0.800	0.800	0.800	0.115	0.275	0.072	0.108	0.097	0.902	
TD12	0.800	0.800	0.800	0.800	0.800	0.800	0.792	0.097	0.098	0.006
TD13	0.301	0.272	0.226	0.061	0.063	0.072	0.010	0.578	1.389	
TD14	0.748	0.764	0.792	0.708	0.800	0.800	0.088	0.115	0.350	
TD15	0.120	0.134	0.108	0.068	0.064	0.083	0.016	0.920	1.310	
TD16	0.800	0.800	0.800	0.800	0.800	0.800	0.201	0.097	0.247	
TD17	0.325	0.344	0.324	0.258	0.307	0.300	0.038	0.480	0.761	
TD18	0.336	0.714	0.710	0.706	0.800	0.780	0.112	0.256	0.327	
TD19	0.314	0.317	0.315	0.706	0.782	0.746	0.084	0.501	0.365	
TQ1	0.728	0.764	0.742	0.167	0.310	0.343	0.315	0.128	0.563	
TQ2	0.772	0.800	0.774	0.800	0.800	0.756	0.790	0.107	0.104	
TQ3	0.800	0.800	0.800	0.710	0.800	0.800	0.336	0.097	0.204	
TQ4	0.720	0.780	0.794	0.790	0.800	0.800	0.293	0.117	0.207	
TQ5	0.708	0.800	0.748	0.176	0.248	0.175	0.193	0.124	0.708	
TQ6	0.800	0.800	0.800	0.740	0.800	0.800	0.326	0.097	0.203	
TQ7	0.307	0.313	0.299	0.276	0.298	0.250	0.071	0.514	0.709	
TQ8	0.298	0.318	0.289	0.255	0.238	0.222	0.087	0.521	0.733	
TQ9	0.334	0.346	0.282	0.020	0.042	0.030	0.010	0.496	1.650	
TQ10	0.279	0.314	0.303	0.296	0.794	0.358	0.098	0.525	0.521	
TQ11	0.272	0.282	0.249	0.116	0.083	0.092	0.032	0.573	1.137	
TQ12	0.348	0.742	0.650	0.161	0.166	0.148	0.032	0.258	0.956	
TQ13	0.274	0.232	0.030	0.131	0.100	0.146	0.037	0.654	1.101	
TQ14	0.800	0.800	0.768	0.329	0.363	0.282	0.746	0.103	0.400	
TQ15	0.387	0.256	0.700	0.209	0.155	0.262	0.043	0.557	0.400	
TQ15	0.264	0.209	0.184	0.209	0.133	0.080	0.043	0.664	1.167	
TQ17	0.269	0.261	0.226	0.152	0.141	0.182	0.032	0.600	0.983	
TQ17	0.287	0.201	0.220	0.132	0.141	0.182	0.030	0.540	0.718	
TQ19	0.287	0.800	0.273	0.200	0.240	0.248	0.085	0.119	0.718	

N1, N2, and N3 mean human normal cells, gingival fibroblast HGF, pulp cell HPC, and periodontal ligament fibroblast HPLF, respectively. T1, T2, T3, and T4 mean human OSCC cell lines (HSC-2, HSC-3, HSC-4) and human promyelocytic leukemic HL-60 cells, respectively.

Determination of  $CC_{50}$ . Three human OSCC cell lines (HSC-2, HSC-3, HSC-4) (purchased from Riken Cell Bank, Tsukuba, Japan), three human normal cells (gingival fibroblast HGF, pulp cell HPC, periodontal ligament fibroblast HPLF) (established as described previously (19)), and human promyelocytic leukemic HL-60 cells (purchased from Riken Cell Bank) were treated for 48 h with different concentrations of test compounds. The 50% cytotoxic concentration ( $CC_{50}$ ) was determined from the dose–response curve (19). The original data of  $CC_{50}$  values (mM) are listed in Table I where names of tumor cells (HSC-2, HSC-3, HSC-4, and HL-60)

are indicated by T1, T2, T3, and T4, respectively, and normal cells (HGF, HPC, and HPLF) by N1, N2, and N3, respectively.

Estimation of  $CC_{50}$  values. Original data contain the sign of inequality such as '>'. For the convenience of analysis, these values were changed into forms suitable for the arithmetic calculation. Since '>400' is equal to '400 $\sim$ ', we calculated the harmonic mean as follows:  $1/(average(1/400,1/\infty))=800$ 

As a result of the estimation by harmonic mean, the value became two-fold (Table I). Eight compounds, TD1, TD2, TD3, TD4,

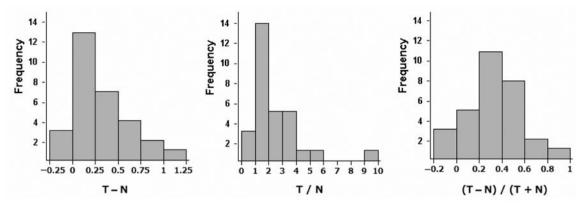


Figure 2. Comparison of normality produced by three calculation approaches: T-N, T/N, and (T-N)/(T+N), where N and T are the estimated 50% cytotoxic concentrations of agent against normal and tumor cells, respectively.

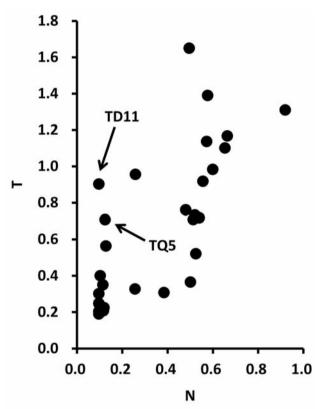


Figure 3. Correlation between N and T, where N and T are the estimated 50% cytotoxic concentrations of agent against normal and tumor cells, respectively.

TD7, TD10, TD12, and TQ2, which had estimated  $CC_{50}$  values >400  $\mu$ M for all 7 cell lines were omitted, since the validity of their tumor specificity value calculated is very low. Therefore, the number of compounds analyzed was reduced from 38 to 30.

Inverse logarithmic ratio. In the case of inhibition constant (Ki), which shows a logarithmic normal distribution, the use of pKi (=-log Ki) instead of Ki facilitates the analysis. Since the CC<sub>50</sub>

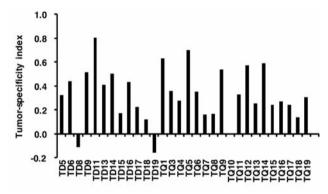


Figure 4. The tumor-specificity index for each compound as given by (T-N)/(T+N), where N and T are the estimated 50% cytotoxic concentrations of agent against normal and tumor cells, respectively.

values had a distribution pattern close to the logarithmic normal distribution, we used the pCC<sub>50</sub> (=-log CC<sub>50</sub>) (Table I).

Mean values. The mean pCC $_{50}$  values for normal cells and tumor cell lines were defined as N and T, respectively (Table I).

Calculation of the representative value for tumor specificity. Parameters that indicate tumor specificity were determined for each compound. It is conceivable that this parameter is defined by the balance between pCC $_{50}$  values for normal cells and that for tumor cell lines (N, T). The following three parameters are available: (i) difference (T–N), (ii) ratio (T/N) and (iii) ratio of difference to their sum: (T–N)/(T+N). Of these, (T–N)/(T+N) produced the highest normality (Figure 2) and correlation coefficient with chemical descriptors (data not shown). Furthermore, the ratio of T/N produced the same value, whenever both T and N had higher or low values. On the other hand, the ratio of (T–N)/(T+N) produced a higher correlation coefficient for compounds with lower cytotoxicity. Based on these considerations, (T–N)/(T+N) was used for the following analyses as a tumor-specificity index.

Calculation of chemical descriptors. Spartan10 for Windows (Wavefunction, Inc., Irvine, CA, USA) was used for the calculations.

Table II. Quantum chemical parameters calculated by PM3.

Comp.	Energy (au)	E <sub>HOMO</sub> (au)	E <sub>LUMO</sub> (au)	Dipole moment (debye)	MSA	MV (Å <sup>2</sup> )	PSA (Å <sup>3</sup> )	WSA (Å <sup>2</sup> )	χ (Å <sup>2</sup> )	η (au)	ω (au)	EPmin (au)	EPmax (au)	Ovality (au)	Log	HBD P	HBA
TD5	-0.130	-0.356	-0.0170	4.33	314.1	300.3	49.1	168.5	0.186	0.169	0.103	-0.105	0.0485	1.448	0.498	1	3
TD6	-0.128	-0.353	-0.0005	4.69	316.4	305.2	48.7	173.3	0.177	0.176	0.089	-0.108	0.0469	1.443	0.578	1	3
TD8	-0.276	-0.334	-0.0054	3.53	415.7	395.4	62.8	219.8	0.170	0.164	0.088	-0.110	0.0483	1.596	-0.791	1	5
TD9	-0.337	-0.330	-0.0005	2.93	303.9	288.9	85.8	156.9	0.165	0.165	0.083	-0.107	0.0528	1.438	-0.704	. 3	5
TD11	-0.232	-0.343	-0.0009	2.59	278.5	254.7	61.6	150.1	0.172	0.171	0.086	-0.100	0.0471	1.433	-1.946	1	4
TD13	-0.125	-0.350	0.0000	4.62	354.8	343.9	34.6	189.2	0.175	0.175	0.088	-0.107	0.0344	1.495	1.179	0	3
TD14	-0.305	-0.332	-0.0021	3.45	390.0	368.5	46.8	209.0	0.167	0.165	0.084	-0.108	0.0296	1.569	0.112	0	5
TD15	-0.272	-0.346	-0.0072	2.26	460.5	435.0	49.7	245.1	0.177	0.170	0.092	-0.112	0.0235	1.658	-0.189	0	5
TD16	-0.296	-0.348	-0.0063	2.85	362.2	334.3	50.6	197.4	0.177	0.171	0.092	-0.109	0.0363	1.555	-1.697	0	5
TD17	-0.201	-0.351	0.0048	2.17	347.0	332.0	35.0	192.1	0.173	0.178	0.084	-0.105	0.0327	1.497	2.182	0	3
TD18	-0.307	-0.329	0.0024	2.92	389.0	368.3	48.7	213.0	0.163	0.166	0.081	-0.107	0.0326	1.565	-0.108	0	5
TD19	-0.315	-0.329	0.0027	2.88	409.5	386.8	48.5	224.2	0.163	0.166	0.080	-0.109	0.0323	1.595	0.230	0	5
TQ1	-0.086	-0.335	0.0036	1.50	230.8	208.7	27.3	141.6	0.166	0.169	0.081	-0.098	0.0336	1.356	-1.485	0	3
TQ3	-0.196	-0.343	-0.0166	4.60	291.9	263.0	51.0	158.7	0.180	0.163	0.099	-0.140	0.0731	1.471	-2.001	0	6
TQ4	-0.206	-0.352	-0.0148	4.48	310.1	281.3	50.8	165.4	0.183	0.169	0.100	-0.140	0.0695	1.494	-1.683	0	6
TQ5	-0.157	-0.337	0.0018	2.28	257.7	234.3	46.8	146.8	0.168	0.169	0.083	-0.094	0.0530	1.402	-2.022	. 1	4
TQ6	-0.406	-0.353	-0.0122	2.43	290.4	266.8	44.9	138.0	0.183	0.170	0.098	-0.080	0.0473	1.449	-1.063	1	4
TQ7	-0.054	-0.334	0.0015	0.86	352.2	330.6	17.1	213.3	0.166	0.168	0.082	-0.100	0.0201	1.523	0.106	0	3
TQ8	-0.054	-0.333	0.0036	1.39	352.5	330.6	17.1	210.4	0.165	0.169	0.081	-0.096	0.0209	1.524	0.106	0	3
TQ9	-0.216	-0.339	-0.0089	2.70	394.5	369.3	44.9	222.9	0.174	0.165	0.092	-0.109	0.0332	1.585	-2.690	0	6
TQ10	-0.227	-0.347	-0.0055	1.75	413.8	387.8	45.9	209.6	0.176	0.171	0.091	-0.105	0.0354	1.609	-2.746	0	6
TQ11	-0.274	-0.356	-0.0162	5.48	327.1	313.3	29.7	161.8	0.186	0.170	0.102	-0.105	0.0351	1.466	2.286	0	3
TQ12	-0.284	-0.358	-0.0203	5.05	341.9	326.4	29.7	163.4	0.189	0.169	0.106	-0.104	0.0363	1.491	2.151	0	3
TQ13	-0.429	-0.354	-0.0212	5.45	444.7	421.8	44.0	212.9	0.187	0.166	0.106	-0.108	0.0376	1.635	0.862	0	5
TQ14	-0.072	-0.340	-0.0184	1.52	243.4	221.9	21.7	153.7	0.179	0.161	0.100	-0.109	0.0235	1.373	-0.527	0	3
TQ15	-0.280	-0.352	-0.0303	3.20	349.0	327.2	26.1	192.4	0.191	0.161	0.114	-0.104	0.0336	1.520	3.050	0	2
TQ16	-0.332	-0.350	-0.0217	2.99	364.0	341.3	33.1	205.4	0.186	0.164	0.105	-0.105	0.0325	1.541	2.210	0	3
TQ17	-0.463	-0.347	-0.0277	2.53	391.2	357.7	41.1	196.2	0.187	0.160	0.110	-0.101	0.0365	1.605	2.062	0	4
TQ18	-0.038	-0.362	-0.0210	3.33	275.0	262.6	28.9	170.4	0.191	0.170	0.107	-0.104	0.0291	1.387	1.091	0	3
TQ19	0.055	-0.326	-0.0239	4.53	224.5	209.5	30.2	142.3	0.175	0.151	0.101	-0.109	0.0436	1.316	0.314	0	3

 $E_{\text{HOMO}}$ : Highest occupied molecular orbital energy;  $E_{\text{LUMO}}$ : lowest unoccupied molecular orbital energy; MSA: surface area of the molecule; MV: volume of the molecule; PSA: polar surface area; WSA: water-accessible surface area;  $\chi$ : negativity;  $\eta$ : absolute hardness;  $\omega$ : reactivity index; EPmin: minimal electrostatic potential; EPmax: maximal electrostatic potential; HBA: hydrogen-bond acceptor; HBD: hydrogen-bond donor.

Each structure was optimized with Merck Molecular Force Field (MMFFaq), and then checked by the semi-empirical method (AM1, PM3, PM6) and density functional theory (DFT-B3LYP/6-31+G\*). During each step of the calculation, quantum chemical, molecular shape, and molecular property parameters were obtained. The parameters used were: energy (au), highest occupied molecular orbital (HOMO) energy ( $E_{\text{HOMO}}$ ; au), lowest unoccupied molecular orbital (LUMO) energy ( $E_{\rm LUMQ}$ ; au), dipole moment (debye), surface area of the molecule (MSA; Å<sup>2</sup>), volume of the molecule (MV; Å<sup>3</sup>), polar surface area (PSA; Å<sup>2</sup>), hydrogen-bond acceptor (HBA), hydrogenbond donor (HBD), negativity [ $\chi = -(E_{LUMO} + E_{HOMO})/2$ ], absolute hardness  $[\eta = (E_{\text{UMO}} - E_{\text{HOMO}})/2]$ , reactivity index  $(\omega = \chi^2/2\eta)$ , ovality, hydrophobicity (logP), minimal and maximal electrostatic potentials (EPmin and EPmax; au), and water-accessible surface area (WSA; Å<sup>2</sup>). These had different values in each calculation method used for precise structural determination.

Statistical analysis. We determined the relationship between N and T (Figure 3) and the tumor-specificity index (T–N)/(T+N) for each chemical descriptor (Figure 4). We attempted to construct a tumor-

specificity estimation model using artificial neural networks. Thirty compounds were divided at random into two sets, one consisting of 24 compounds (training set for construction of the model: TD6, TD8, TD9, TD11, TD13, TD15, TD16, TD17, TD18, TD19, TQ1, TQ3, TQ6, TQ8, TQ9, TQ10, TQ11, TQ12, TQ14, TQ15, TQ16, TQ17, TQ18, and TQ19) and the other consisting of 6 compounds (test set for confirmation of the model: TD5, TD14, TQ4, TQ5, TQ7, and TQ13). Using major PM3 descriptors and the 'neuralnet' script in JMP ver.9 (SAS Institute Inc., Cary, NC, U.S.A.) for calculation, an estimation model was constructed from a nonlinear regression of multilayer perception neural networks employing back propagation-training algorithm. Descriptor selection was performed with the leave-one-out cross validation. The model constructed was validated by prediction of the test set compounds.

## Results and Discussion

Correlation between N and T. The structures of TD11 and TQ5 are desirable for chemotherapy (Figure 3) because of

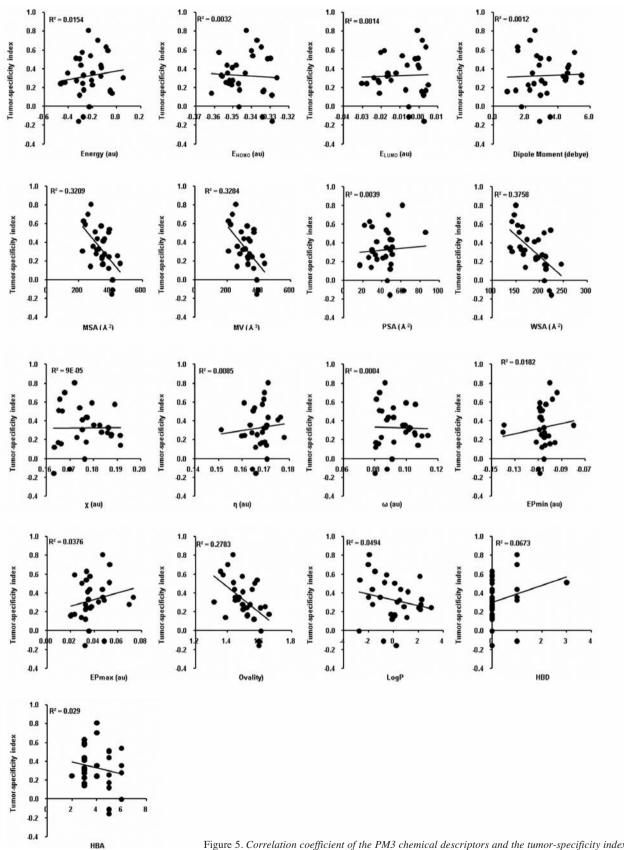


Figure 5. Correlation coefficient of the PM3 chemical descriptors and the tumor-specificity index.

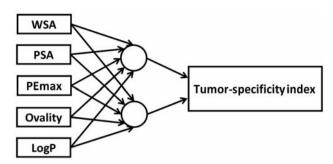


Figure 6. Input, hidden, and output layers in the artificial neural network model.

high T and the low N values. The tumor-specificity index of each compound is shown in Figure 4. Both TD11 and TQ5 had high values of a tentative cut-off point of 0.7 units. Both compounds include methoxy groups and hydrogen at R<sup>1</sup> and R<sup>3</sup> positions, respectively, of the tetrahydroisoquinoline skeleton (Figure 1). These structural properties may be responsible for the tumor-specificity.

Correlation between each chemical descriptor and tumorspecificity index. Regardless of the calculation method used, the best correlation was shown between WSA and tumorspecificity index. That is, the determined coefficients (r<sup>2</sup>) for the index and WSA calculated by AM1, PM3, PM6, and DFT-B3LYP/6-31+G\* methods were 0.357, 0.376, 0.338, and 0.374, respectively, suggesting that WSA calculated by PM3, as well as DFT, is a useful chemical descriptor to evaluate the tumor-specificity. Scatter plots with correlation coefficients obtained in the linear regression analyses on PM3 chemical descriptors (Table II) and tumor-specificity index are shown in Figure 5.

Artificial neural network. Neural networks are powerful tools for the numerical formulation of nonlinear relationships, although it is difficult to determine how much each descriptor contributes to the constructed model. As a result of trials for variable selection, we constructed a model with 5 descriptors in an input layer (WSA, PSA, PEmax, ovality and logP) and 2 nodes in a hidden layer (Figure 6). Training and test sets enabled the successful construction of a model that can predict tumor specificity at a high probability (R<sup>2</sup><sub>training</sub>=0.909, Q<sup>2</sup><sub>leave-one-out</sub>=0.543, R<sup>2</sup><sub>test</sub>=0.923). This suggests that these quantum chemical descriptors have the most information on tumor specificity and can be used to estimate the tumor specificity of related compounds (Figure 7).

# Conclusion

The present study demonstrates for the first time that there is a significant correlation between the tumor-specificity of

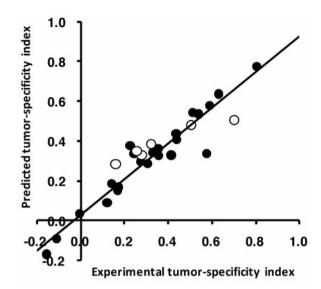


Figure 7. Scatter plots between the experimental tumor-specificity indices and those predicted by the artificial neural network model using the descriptors shown in Figure 6. Training set and test set compounds are indicated by closed and open symbols, respectively.

TIQ derivatives and WSA. Furthermore, we succeeded in constructing a predictive neural network model for tumor specificity with 5 parameters, including WSA and logP.

### References

- 1 Page D, Nguyen N, Bernard S, Coupal M, Gosselin M, Lepage J, Adam L and Brown W: New scaffolds in the development of mu opioid-receptor ligands. Bioorg Med Chem Lett 13: 1585-1589, 2003.
- 2 Abe K, Saitoh T, Horiguchi Y, Utsunomiya I and Taguchi K: Review. Synthesis and neurotoxicity of tetrahydroisoquinoline derivatives for studying Parkinson's disease. Biol Pharm Bull 28: 1355-1362, 2005.
- 3 Mohler ML, Kang G-S, Hong S-S, Patil R, Kirichenko OV, Li W, Rakov IM, Geisert EE and Miller DD: Discovery of antiglioma activity of biaryl 1,2,3,4-tetrahydroisoquinoline derivatives and conformationally flexible analogues. J Med Chem 49: 5845-5848, 2006.
- 4 Lin H-R, Safo MK and Abraham DJ: Identification of a series of teterhydroisoquinoline derivatives as potential therapeutic agents for breast cancer. Bioorg Med Chem Lett 17: 2581-2589, 2007.
- 5 Seo JW, Srisook E, Son HJ, Hwang O, Cha Y-N and Chi DY: Syntheses of terahydroisoquinoline derivatives that inhibit NO production in activated BV-2 microglial cells. Eur J Med Chem 43: 1160-1170, 2008.
- 6 Letavic MA, Keith JM, Jablonowski JA, Stocking EM, Gomez LA, Ly KS, Miller JM, Barbier AJ, Bonaventure P, Boggs JD, Wilson SJ, Miller KL, Lord B, McAllister HM, Tognarelli DJ, Wu J, Abad MC, Schubert C, Lovenberg TW and Carruther NI: Novel tetrahydroisoquinolines are histamine H3 antagonists and serotonin reuptake inhibitors. Bioorg Med Chem Lett 17: 1047-1051, 2007.

- 7 Gitto R, Ficarra R, Stancanelli R, Guardo M, De Luca L, Barreca ML, Pagano B, Rotondo A, Bruno G, Russo E, De Sarro G and Chimirri A: Synthesis, resolution, stereochemistry, and molecular modeling of (R)- and (S)-2-acetyl-1-(4'-chlorophenyl)-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline AMPAR antagonists. Bioorg Med Chem 15: 5417-5423, 2007.
- 8 Kubota H, Watanabe T, Kakefuda A, Masuda N, Wada K, Ishii N, Sakamoto S and Tsukamoto S: Synthesis and pharmacological evaluation of piperidinoalkanoyl-1,2,3,4-tetrahydroisoquinoline derivatives as novel specific bradycardic agents. Bioorg Med Chem Lett 14: 3049-3052, 2004.
- 9 Hirose M, Egashira S, Goto Y, Hashihayata T, Ohtake N, Iwaasa H, Hata M, Fukami T, Kanatani A and Yamada K: N-Acyl 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline: the first orexin-2 receptor selective non-peptidic antagonist. Bioorg Med Chem Lett 13: 4497-4499, 2003.
- 10 Fang W, Li Y, Cai Y, Kang K, Yan F, Liu G and Huang W: Substituted tetrahydroisoquinoline compounds B3 inhibited Pglycoprotein-mediated multidrug resitance in vitro and in vivo. J Pharm Pharmacol 59: 1649-1655, 2007.
- 11 Yan F, Jiang Y, Li Y-M, Zhen X, Cen J and Fang W-R: Reversal of P-glycoprotein and multidrug resistance-associated protein 1mediated multidrug resistance in cancer cells by HZ08 isomers, tetraisohydroquinoline derivatives. Biol Pharm Bull 31: 1258-1264, 2008.
- 12 Li Y, Zhang H-B, Huang W-L and Li Y-M: Design and synthesis of tetrahydroisoquinoline derivatives as potential multidrug resistance reversal agents in cancer. Bioorg Med Chem Lett 18: 3652-3655, 2008.
- 13 Hu M-K, Liao Y-F, Chen J-F, Wang B-J, Tung Y-T, Lin H-C and Lee K-P: New 1,2,3,4-tetrahydroisoquinoline derivatives as modulators of proteolytic cleavage of amyloid precursor proteins. Bioorg Med Chem 16: 1957-1965, 2008.
- 14 Choquette D, Teffera Y, Polverino A and Harmange J-C: Discovery of novel 1,2,3,4-tetrahydroisoquinolines and 3,4-dihydroisoquinoline-1(2*H*)-ones as potent and selective inhibitors of KDR: synthesis, SAR, and pharmacokinetic properties. Bioorg Med Chem Lett *18*: 4054-4058, 2008.

- 15 Azukizawa S, Kasai M, Takahashi K, Mike T, Kunishiro K, Kanda M, Mukai C and Shirahase H: Synthesis and biological evaluation of (S)-1,2,3,4-tetrahydroisoquinoline-3-carboxylic acids: a novel series of PPARγ agonists. Chem Pharm Bull 56: 335-345, 2008.
- 16 Scott JD and Williams RM: Chemistry and biology of the tetrahydroisoquinoline antitumor antibiotics. Chem Rev 102: 1669-1730, 2002.
- 17 Kang JH: Salsolinol, a tetrahydroisoquinoline catechol neurotoxin, induces human Cu,Zn-superoxide dismutase modification. J Biochem Mol Biol 40: 684-689, 2007.
- 18 Saitoh T, Abe K, Ishikawa M, Nakatani M, Shimazu S, Satoh N, Yoneda F, Taguchi K and Horiguchi Y: Synthesis and *in vitro* cytotoxicity of 1,2,3,4-tetrahydroisoquinoline derivatives. Eur J Med Chem 41: 241-253, 2006.
- 19 Hatano H, Takekawa H, Hashimoto K, Ishihara M, Kawase M, Chu Q, Wang QT and Sakagami H: Tumor-specific cytotoxic activity of 1,2,3,4-tetrahydroisoquinoline derivatives against human oral squamous cell carcinoma cell lines Anticancer Res 29: 3079-3086, 2009.
- 20 Ishihara M, Hatano H, Kawase M and Sakagami H: Estimation of relationship between the structure of 1,2,3,4-tetrahydroisoquinolin derivatives determined by a semi-empirical molecular-orbital method and their cytotoxicity. Anticancer Res 29: 2265-2272, 2009.
- 21 Ishihara M, Hatano H, Takekawa F, Kawase M and Sakagami H: Estimation of relationship between descriptors and cytotoxicity of newly synthesized 1,2,3,4-tetrahydroisoquinoline derivatives. Anticancer Res 29: 4077-4082, 2009.

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