

Review

Cancer Nanotargeted Radiopharmaceuticals for Tumor Imaging and Therapy

GANN TING¹, CHIH-HSIEN CHANG² and HSIN-ELL WANG³

¹Center of Nanomedicine and National Institute of Cancer Research, National Health Research Institutes, Miaoli;

²Isotope Application Division, Institute of Nuclear Energy Research, Taoyuan;

³Biomedical Imaging and Radiological Sciences, National Yang-Ming University, Taipei, Taiwan, R.O.C.

Abstract. Cancer is the second leading cause of death in the world. Radiolabeled nanocarriers or nanoparticles can be designed and used for cancer diagnostic and therapeutic purposes when tagged with appropriate radionuclides. Current progress in nanotechnology and nanomedicine has exploited the possibility of designing tumor-targeted nanocarriers able to deliver radionuclide payloads in a selective manner to improve the efficacy and safety of cancer imaging and therapy. The major nanocarriers include liposomes, dendrimers, quantum dots, iron oxide and carbon nanotubes. In addition, the combining of tumor specific multifunctional and multimodality nanocarriers will hopefully achieve earlier tumor detection and better tumor treatment. Several radiolabeled multifunctional and multimodality nanoparticles have been effectively demonstrated in detecting and treating cancer in animal models. However, further preclinical and clinical efficacy and toxicity studies are required to translate these advanced technologies to the health care of cancer patients. The aim of this article is to provide a brief overview of current status of applications, advantages and up-to-date research and development of nanotargeted radiopharmaceuticals in cancer imaging and therapy.

More than 12 million cancer cases and 7.6 million cancer deaths were reported in 2007 worldwide (1). The incidence and mortality of cancer were about 5.6 million and 2.7 million, respectively in 2007 in Asia. By 2050, the global

new cancer incidence is expected to grow to 27 million, and the cancer mortality will be 17.5 million (1). To conquer cancer is really a challenge of health care for mankind. Cancer exhibits up-regulated cell growth, with an ability for tumor cells to invade and metastasize. A century ago, Paul Ehrlich hypothesized that a 'magic bullet' could be developed to selectively target cancer disease (2). Over the past few decades, the progress in molecular biology and the understanding of malignant transformation and tumorigenesis have revealed the two major classes of anti-tumor therapeutics: i) Application of molecularly targeted therapeutics to block hallmarks of cancer, and ii) Employing drug delivery systems through tumor-targeted nanomedicines to improve the pharmacokinetics and bioavailability of vehicle-carried drugs. From 1980 to 2005, a total of 205 monoclonal antibodies (mAb) were studied in clinical trials (3-5). The US Food and Drug Administration (FDA) approved the first anti-CD20 mAb (Rituximab) for the treatment of non-Hodgkin's lymphoma in 1997. To date, 12 of these anticancer molecular-targeted mAbs have been approved worldwide (4, 5).

Conventional anticancer drugs exhibit a lack of specificity, poor solubility and distribution, unfavorable pharmacokinetics and high tissue damage or toxicity. Targeted drug delivery systems such as passive and active targeting nanocarriers, with diameters ranging from 10-100 nm have been developed to improve the biodistribution, pharmacological, therapeutic and toxicity properties of agents used in cancer diagnostics and therapeutics (6-9). The status of the development of targeting delivery systems, including targeting strategies, potential applications and the prospects of tumor-targeted nanocarriers were reviewed and discussed recently (6-9). Cancer nanotechnology is expected to transform current treatment systems by providing more efficient cancer diagnostics and therapeutics. Today, nanocarriers are used in detecting cancer at an early stage, delivering anticancer drugs specifically to malignant cells, and determining if these drugs

Correspondence to: Dr. Gann Ting, Center of Nanomedicine and National Institute of Cancer Research, National Health Research Institutes, 4F, 123-2, Section 1, Wan-Mei Street, Wen-Shan Districts, Taipei, 116, Taiwan, R.O.C. Fax: +886 2 22302573, e-mail: gann.ting@msa.hinet.net

Key Words: Cancer therapy, nanotargeted radiopharmaceuticals, nanocarriers, tumor imaging, review.

are killing malignant cells (8, 9). Two therapeutic nanocarrier-liposomes and albumin nanoparticles have been approved by US FDA for clinical practices (7, 10). As nanocarriers are evaluated for safety and efficacy, nanotechnology will bring with it significant advances in molecular imaging and specific targeting of tumor therapeutic agents, elevating therapeutic efficacy, and finally achieving the goal of early detection and control of cancer. Customized nanoscale constructs can serve as targeted drug delivery vehicles capable of delivering large doses of radionuclide or chemotherapeutic agents into malignant cells while sparing normal tissues, greatly reducing the side-effects that usually accompany many current cancer therapies (7-10).

Monoclonal antibody-guided radiation therapy, or radioimmunotherapy, demonstrated promise in preclinical and clinical anticancer applications (11-15). Two radiolabeled anti-CD20 monoclonal antibodies ^{90}Y -ibritumomab (Zevalin[®]) and ^{131}I -tositumomab (Bexxar[®]) were approved by the US FDA in 2002 and 2003, respectively, for treatment of B-cell non-Hodgkin's lymphoma (NHL), which indicates the potential benefit of antibody-guided systemic radionuclide-targeted therapy (11-15). Emerging new methods improve the specific uptake of radionuclides in tumor cells while sparing the normal tissues. Several advanced strategies for radionuclides delivery have been studied extensively, including the combination of chemotherapy agents with particle-emitting radionuclides and the development of novel multimodality and multifunctional therapeutics. Optimization of treatment protocols has significantly improved the therapeutic efficacy and reduced the toxicity to normal tissues. Nanoparticles delivering radionuclides for improving pharmacokinetics and therapeutic efficacy of cancer have been presented elsewhere (16, 17). The recent research progress and applications of advanced nanocarrier radiopharmaceuticals for *in vivo* cancer imaging and therapeutic applications will be briefly discussed and summarized in this review article.

Radionuclides for Tumor Imaging and Therapeutics

The research into tumor-targeted diagnostic and therapeutic radiopharmaceuticals is one of the potential areas of cancer drug development. Radiopharmaceuticals consist of two components, a targeting carrier and a trace amount of radionuclide with a specific radiation. The tumor therapeutic efficacy and diagnostic quality are determined by the selectivity or specificity of delivery systems and radionuclide radiation characteristics (11, 15-19).

The selection of potential radionuclides for tumor imaging (Table I) and radionuclide radiotherapy (Table II) involves the physical half-life, decay mode and the emission

Table I. Characteristics of potential radionuclides for nanotargeted tumor imaging (16, 17, 20, 21).

Radionuclide	Emission type	Half-life	E _{max} (γ), (keV)
^{131}I	γ (81.2%), β	8.0 days	284, 364, 637
^{67}Ga	γ	78.3 h	93, 184, 300, 393
^{111}In	Auger, γ	67.2 h	171, 245
^{123}I	Auger, γ	13.2 h	159
$^{99\text{m}}\text{Tc}$	γ	6.0 h	140
^{18}F	Positron	1.83 h	511
^{64}Cu	Positron	12.7 h	511

properties of the radionuclides. Gamma emitters with energy in the 150 keV range can be used for gamma imaging or single photon-emission tomography (SPECT), and high energy positron-emitters with energy at 511 keV energy can be applied for positron-emission tomography (PET) (16, 17, 20, 21). For targeted radionuclide radiotherapy applications, high and low energy β-emitters are ideal radioisotopes for the treatment of small to large clusters of tumor cells. The tissue penetration range (1-10 mm) (11, 18), and cross fire effect of β-particles can kill tumor cells in close proximity to neovasculature (11, 15, 18). Alpha-emitters hold great promise as therapeutics for small cancer lesions and micrometastatic cancers due to the high linear energy transfer (LET, 80 keV/μm) and short range energy depositions with tissue penetration range of 50-100 μm. Monoclonal antibody labeled with α-emitters has been demonstrated to have high specific killing effects and minimal normal-tissue damage in a tumor-bearing animal model (17). Auger electrons have an energy of <30 keV and subcellular pathlength of 2-12 μm. Thus, auger electron emitters can exert their radiotoxic effects on cells only when they are internalized into the cytoplasm (19, 22, 23).

Nanocarriers for the Targeted Delivery of Radionuclide

Targeted radionuclide therapy is often limited by insufficient delivery of radionuclide to tumor sites using the currently available targeting strategies, such as monoclonal antibodies and peptides, due to relatively low and heterogeneous expression of receptor on tumor cells, as well as dose-limiting toxicities to normal tissues. To maximize the therapeutic index and to minimize the outcome of toxicity, it is very important to deliver the radionuclides to the right site at the right concentration and at the right time. The rapidly advancing field of cancer nanotechnology has generated several innovative drug delivery systems, such as liposomes, dendrimers, quantum dots, iron oxide and carbon nanotubes, to improve and enhance targeted transport of cytotoxic drugs and radionuclides to tumor

Table II. Characteristics of potential radionuclides for tumor radiotherapy (11, 15-19).

Radionuclide	Emission type	Half-life	E _{max} (MeV)	R _{max} (mean) ¹	Size of tumor cells ²
¹⁸⁶ Re	β, γ (9.4%)	89.2 h	1.07	5 mm (0.9 mm)	Intermediate clusters
¹⁸⁸ Re	β, γ (15.1%)	17 h	2.12	11 mm (2.4 mm)	Large clusters
¹⁷⁷ Lu	β	161 h	0.49	1.6 mm (0.67 mm)	Small clusters
¹³¹ I	γ (81.2%), β	8 d	0.28, 0.36, 0.64	2.0 mm	Small clusters
⁹⁰ Y	β	64.1 h	2.28	12 mm (2.8 mm)	Large clusters
⁶⁷ Cu	β	2.6 d	0.19	0.7 mm	Small clusters
²²⁵ Ac	α	10 d	5.83, 5.79, 5.79, 5.73	40-80 μm	Single cells and small clusters
²¹¹ At	α	7.2 h	5.87	60-80 μm	Single cells and small clusters
¹¹¹ In	Auger, γ	67 h	0.42	2-500 nm	Single cells

¹Radiation tumor tissue penetration maximum and mean range. ²Small, intermediate and large clusters correspond approximately to the intervals 10⁴-10⁶, 10⁶-10⁸, and 10⁸-10¹⁰ tumor cells per cluster, respectively (18).

Table III. Characteristics of nanotargeted nuclear imaging modalities (16, 17).

Modality	Image probe (amount of probe)	Type of radiation	Sensitivity (mole/l)	Spatial resolution	Depth
SPECT	^{99m} Tc, ¹¹¹ In <i>etc.</i> loaded or labeled nanocarriers (ng)	γ-ray	10 ⁻¹⁰ -10 ⁻¹¹ (pM)	0.5-1 mm	No limit
PET	¹⁸ F, ⁶⁴ Cu <i>etc.</i> loaded or labeled nanocarriers (ng)	Positron high energy γ-ray	10 ⁻¹¹ -10 ⁻¹² (pM)	1-2 mm	No limit

SPECT: Single photon-emission computed tomography; PET: positron-emission tomography.

lesions. It is estimated that approximately 240 nano-enabled products entered pharmaceutical research pipelines in 2006. These nanocarrier systems could provide the delivery platforms needed for improving the delivery of radionuclide to tumor sites. Nanocarrier delivery systems have also revealed enhanced imaging and therapeutic efficacy by targeted delivery of drugs to the tumor site and by reducing their toxic side-effects (6-10). Major advantages of nanocarriers are that they can be prepared in sizes <100 nm, and selectively increase the localization of drugs and radionuclides in the tumor through passive targeting or active targeting, while sparing non-targeted tissue, ensuring minimal drug or radionuclide leakage during circulation, and facilitating intracellular drug or radionuclide delivery and uptake for active targeting (16, 17). Figure 1 shows the schematic concept of passive (A, B) and active (C, D) targeting nanoliposome encapsulated with radionuclides (A, C) and co-delivery of radiochemotherapeutics (B, D).

There are three generations of nanocarriers: i) The first generation of nanocarriers (passive targeting) which are rapidly trapped in the reticuloendothelial system (RES) organs (*e.g.* liver and/or spleen); ii) The second generation of pegylated nanocarriers (passive targeting), which can evade the RES of the liver and spleen, enjoys a prolonged circulation in the blood and allows for passive targeting through the enhanced permeability and retention (EPR) effect in leaky tumor tissues; iii) The third generation of nanocarriers (active targeting) has a bioconjugated surface modification using specific antibodies or peptides to actively

targeted specific tumor or tissues. The pharmacokinetics and bioavailability of drugs and radionuclides delivered by the third generation of nanocarrier were much improved. One of the major challenges is to design a nanocarrier with less immunotoxic effect and to avoid higher biological barriers in the body such as to reduce delivered diagnostic and therapeutic agent uptake in the reticuloendothelial system (RES) (10).

There are five approaches generally used for labeling or encapsulating radionuclides on nanocarriers: i) Labeling nanocarriers by encapsulation during preparation; ii) nanocarrier surface labeling after preparation; iii) nanocarrier surface labeling of bioconjugates after preparation; iv) incorporation into the lipid bilayer after preparation; and v) after-loading of the aqueous phase of the nanocarriers after preparation. The after-loading method has provided higher labeling efficiencies (>90%) and the greatest *in vivo* stability for ^{99m}Tc, ¹¹¹In, and ⁶⁷Ga radionuclides for nuclear imaging (16, 17, 20, 21).

Nanotargeted Radiopharmaceuticals for Tumor Imaging

The major characteristics of nanotargeted nuclear imaging modalities such as SPECT and PET are listed in Table III (16, 17, 20, 21). Liposomes are spherical bilayers of small phospholipid vesicles which spontaneously form when water is added to a dried lipid mixture. Significant progress has

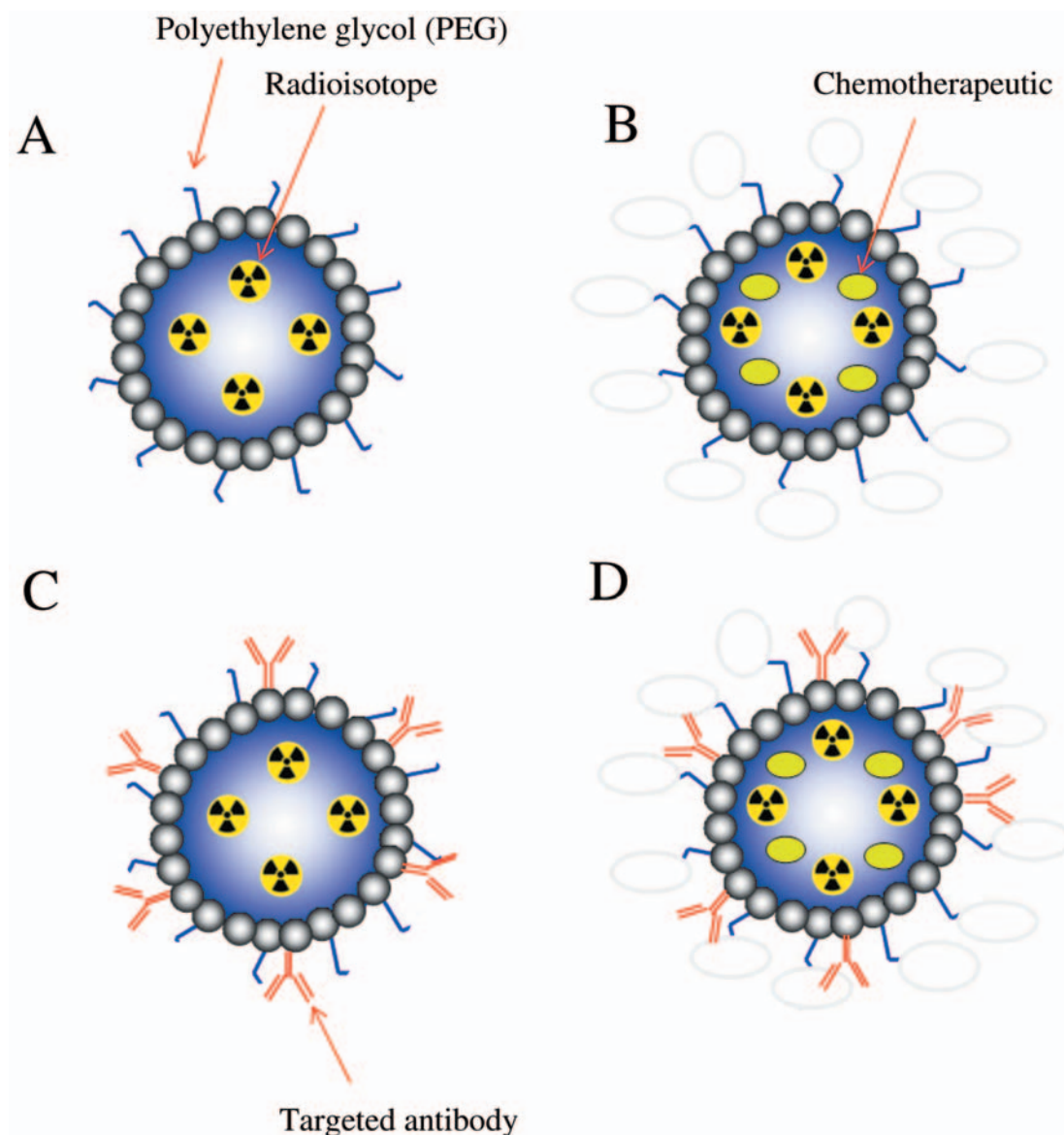


Figure 1. Schematic representation of passive (A, B) and active (C, D) targeting nanoliposome encapsulated with radionuclides (A, C) and co-delivery of radiochemotherapeutics (B, D).

been made in the use of liposome as nanocarriers for the delivery of imaging radionuclides. The ability to modify the surface of nanocarriers permits improvement in the pharmacokinetics, bioavailability, toxicity and customization of nanocarrier formulations for particular tumor imaging agents (24). Selected current cancer nanotargeted nuclear imaging agents are summarized in Table IV.

Delivery of ^{99m}Tc , ^{111}In and ^{67}Ga radionuclides by liposomes for gamma-imaging and monitoring drug treatment have been reviewed and reported (20, 21, 25). The biodistribution, pharmacokinetics and nuclear imaging of ^{111}In -DTPA-labeled pegylated liposome were studied in

patients with advanced local cancer (26). Effective targeting of solid tumors of breast (5.3 ± 2.6 % ID/kg for a tumor volume of 234.7 ± 101.4 cm³), head and neck (highest uptake of 33.0 ± 15.8 % ID/kg for a tumor volume of 36.2 ± 18.0 cm³), lung (18.3 ± 5.7 % ID/kg for a tumor volume of 114.5 ± 42.0 cm³), brain and cervix were also observed with gamma camera and SPECT imaging (26). Liposome encapsulating the positron-emitter ^{18}F -FDG was applicable for diagnostic imaging and real-time liposomal tracking *in vivo* (27). Wang *et al.* demonstrated an intravenous administration of ^{111}In -liposome by conjugating ^{111}In -oxine to DTPA/PEG-liposome followed by whole-body

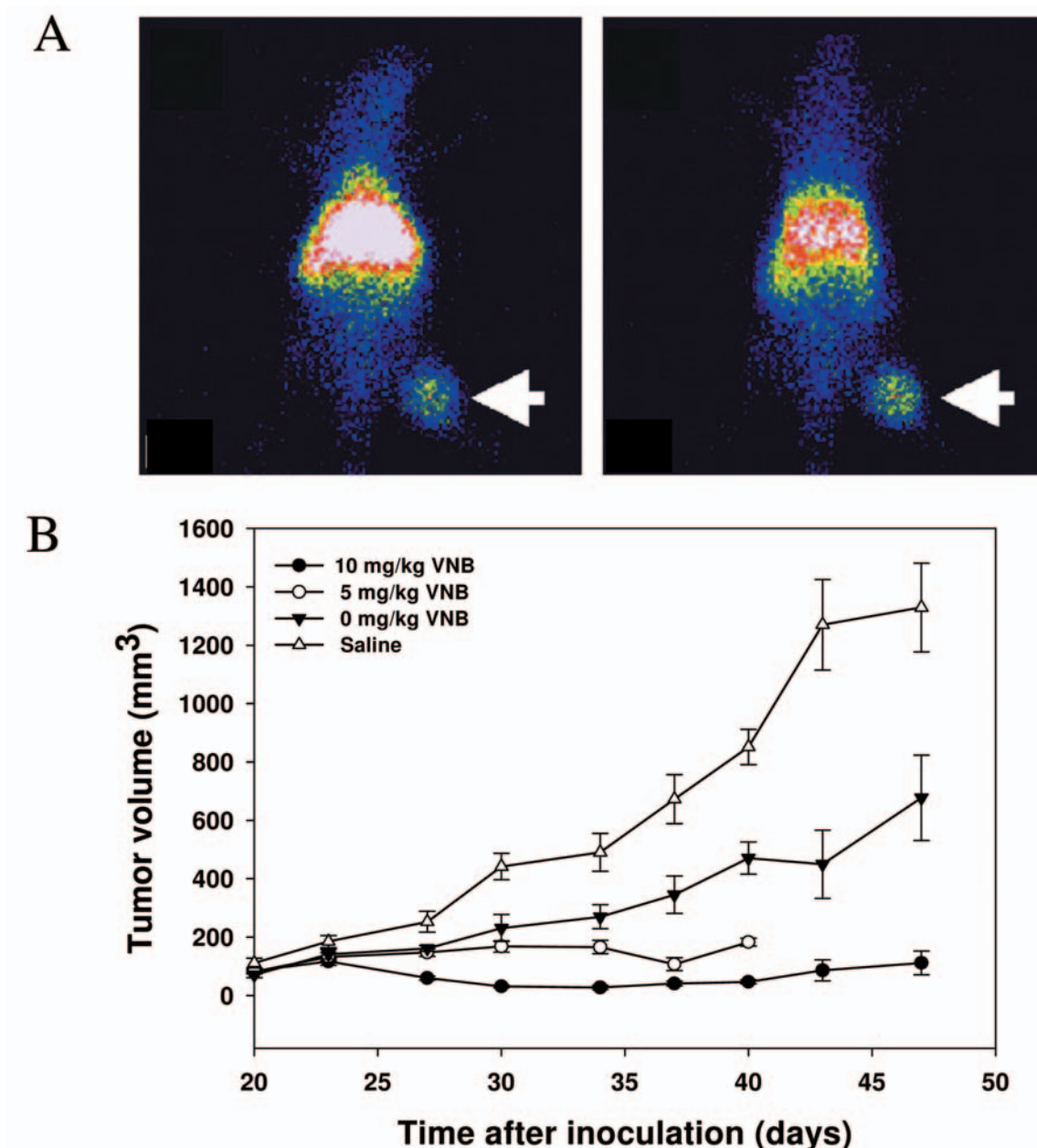


Figure 2. A, Imaging of passive nanotargeted ^{111}In -liposome in CT-26 carcinoma-bearing mice (28). B, Therapeutic efficacy of ^{111}In -VNB-liposomes in HT-29/luc tumor-bearing mice. From (28) and (29) with permission.

scintigraphy. Images revealed that the tumor clearly accumulated ^{111}In -liposome up to 48 h post injection (*p.i.*) (28) (Figure 2A). In addition to the diagnostic imaging of ^{111}In -liposome, Lee *et al.* demonstrated the bifunctional imaging and bimodality therapeutic efficacy of ^{111}In -VNB-liposomes in HT-29/luc mouse xenografts (29) (Figure 2B).

Enhanced tumor accumulation and visualization by γ -scintigraphy with ^{111}In -labeled nucleosome-specific monoclonal antibody 2C5 bioconjugated immunoliposome has

been studied, and the results indicated better and faster imaging in various tumor-bearing mice (30, 31). $\alpha_v\beta_3$ -Integrin-targeted ^{111}In perfluorocarbon nanoparticles have been developed and studied for the detection of rabbit Vx-2 tumor angiogenesis. The circulatory half-life was estimated to be 5 h. The mean tumor uptake was 4-fold higher than in the nontargeted control. The specificity activity ($^{111}\text{In}/\text{NP}$) of ^{111}In to nanoparticle (NP) may affect the tumor-to-muscle uptake ratio in patients. The tumor-to-muscle ratio for the

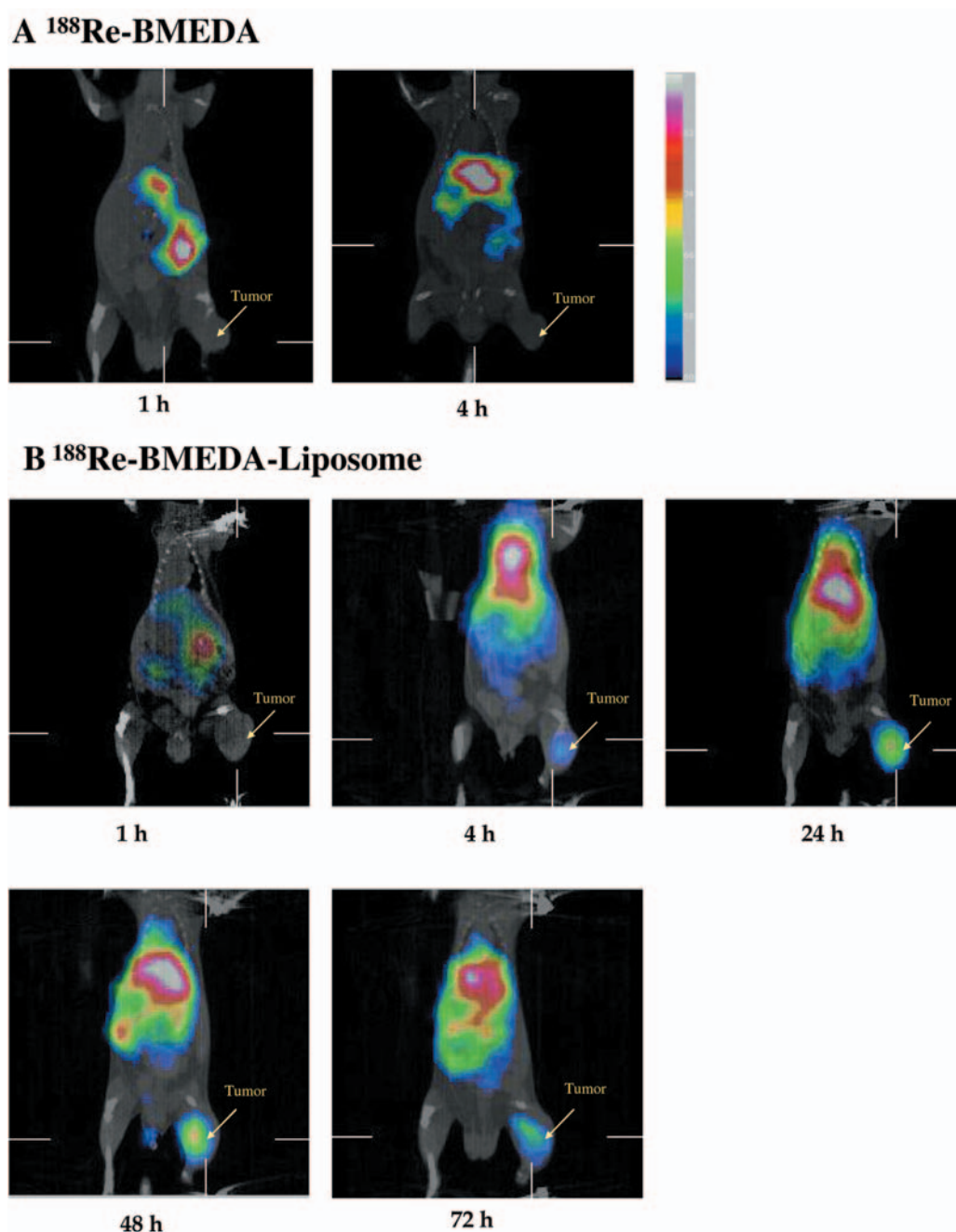


Figure 3. Comparisons of microSPECT/CT images of ^{188}Re -BMEDA and ^{188}Re -liposomes passive targeting tumors in C26 colorectal carcinoma-bearing mice. From (46) with permission.

nanotargeted $^{111}\text{In}/\text{NP}=10$ to $^{111}\text{In}/\text{NP}=1$ were 6.3 ± 0.2 to 5.1 ± 0.1 , respectively. The data suggest that $\alpha_v\beta_3$ -targeted ^{111}In perfluorocarbon nanoparticles may provide a clinically useful tool for detecting angiogenesis in nascent tumors (32). ^{111}In radiolabeled soluble functionalized multifunctional drug delivery platforms of active targeting with rituximab monoclonal antibody bioconjugated on single-wall carbon

nanotubes have been developed, and the selectivity of targeting to disseminated human lymphoma were evaluated *in vitro* and *in vivo* (33). The results of the ability to specifically target tumor with prototype-radiolabeled or fluorescent-labeled, antibody-appended carbon nanotube constructs was encouraging and suggested further investigation of carbon nanotubes as a novel radionuclide delivery platform (33).

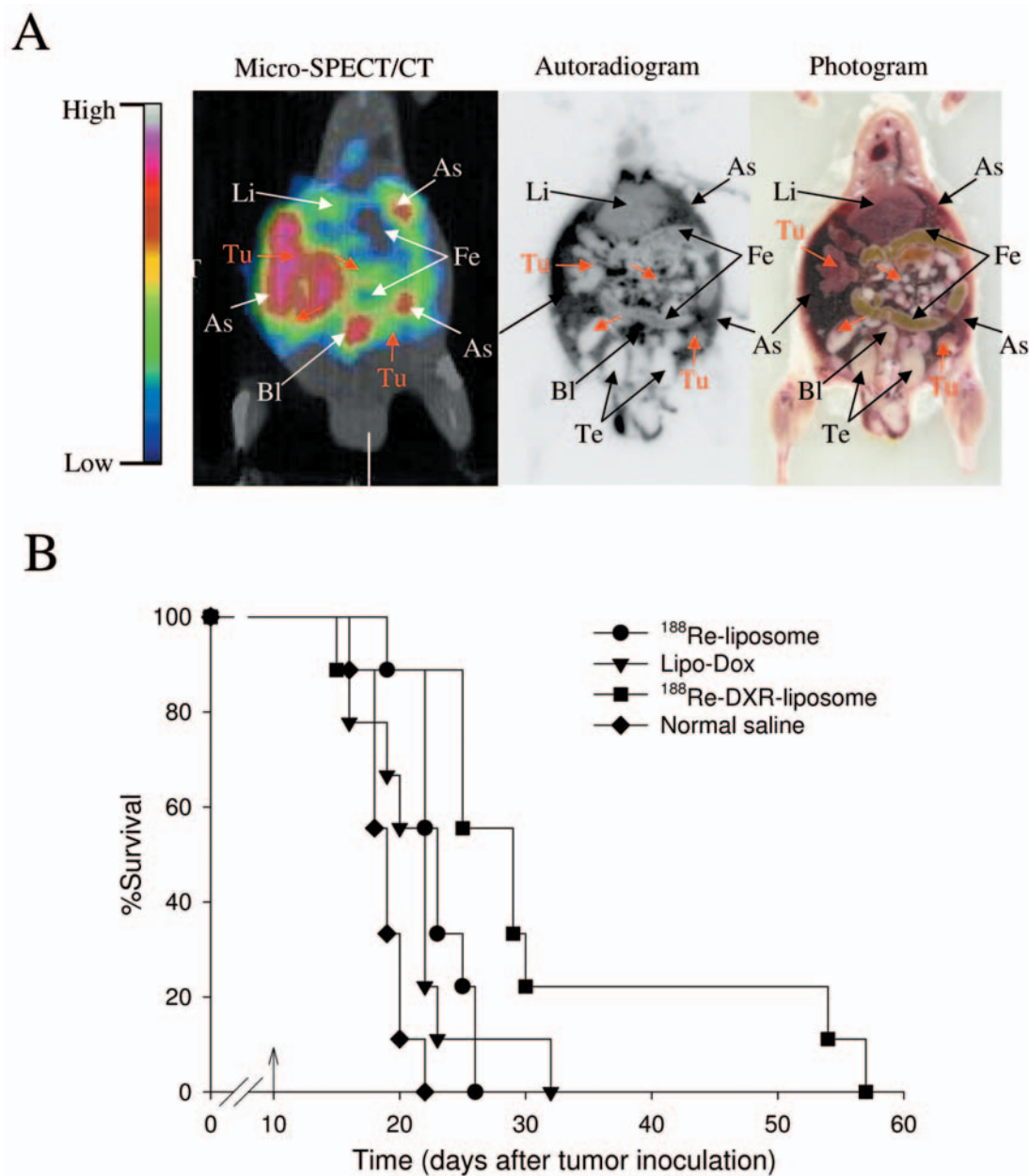


Figure 4. A, Coronal microSPECT/CT image of co-delivery of nanotargeted radiochemotherapeutics of ^{188}Re -DXR-liposome correlated with whole-body autoradiography in C26 colon carcinoma ascites mice. B, Therapeutic efficacy studies of survival curves for mice bearing peritoneal C26 tumor and ascites animal model of passive nanotargeted ^{188}Re -liposome, Lipo-Dox, ^{188}Re -DXR-liposome, and normal saline. From (48) with permission.

The development of a dual-function PET/near-infrared fluorescence (NIRF) molecular probe for the accurate assessment of pharmacokinetics and tumor-targeting efficacy of U87MG human glioblastoma tumor-bearing mice has been reported (11, 34). The amine-functionalized surface of quantum dot (QD) bioconjugated with arginine-glycine-aspartic acid (RGD) peptides and 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA)

chelators for ^{64}Cu radiolabeled ^{64}Cu -DOTA-QD-RGD nanoconstructs with 90 RGD per QD to target angiogenesis of integrin- $\alpha_v\beta_3$ PET/NIRF imaging was also illustrated (34). This dual-function nuclear/optical *in vivo* molecular probe revealed a quantitative targeting ability in deep tumor lesions (34). Dual modality optical and PET imaging of vascular endothelial growth factor receptor (VEGFR) on tumor vasculature using QDs of ^{64}Cu radiolabeled ^{64}Cu -

Table IV. *Selected cancer nanotargeted nuclear imaging agents.*

Nanocarriers	Radionuclides	Passive or active targeting	Applications	References
Liposome	^{99m}Tc , ^{111}In , ^{67}Ga	Passive, gamma imaging	Multitude diagnostics of tumor, infection, Inflammation, and lymphoscintigraphy	(20, 21, 25)
Liposome	^{111}In	Passive, gamma/SPECT imaging	Clinical biodistribution, PK and imaging studies of breast, head and neck, glioma and lung cancer patients	(26)
Immunoliposome	^{111}In	Active, gamma imaging	^{111}In -liposome-2C5 (mAb) nucleosome-specific monoclonal 2C5 targeting delivery vehicles for tumor visualization of murine lewis lung carcinoma and human HT-29 tumors	(30, 31)
Perfluorocarbon nanoparticles	^{111}In	Active, gamma imaging	Imaging of targeted tumor angiogenesis of $\alpha_v\beta_3$ -integrin in Vx-2 rabbit tumors	(32)
Carbon nanotubes	^{111}In	Active, gamma or SPECT imaging	Multifunctional targeted delivery and imaging with functionalized and bioconjugated ^{111}In -DOTA-CNT-Rituximab nanoconstructs.	(33)
Quantum dots	^{64}Cu	Active, bifunctional PET/NIRF imaging	Dual-functional targeted delivery with amine functionalized ^{64}Cu -DOTA-QD-RGD for tumor angiogenesis PET/NIRF imaging	(34)
Quantum dots	^{64}Cu	Active, bifunctional PET/NIRF imaging	Dual-functional targeted delivery with amine functionalized ^{64}Cu -DOTA-QD-VEGF for tumor angiogenesis PET/NIRF imaging	(35)
Iron oxide	^{64}Cu	Active, bifunctional PET/MRI imaging	PET/MRI dual-modality tumor angiogenesis imaging with ^{64}Cu -DOTA-IO-RGD nanoconstructs	(36)

mAb: Monoclonal antibody; CNT: carbon nanotube; QD: quantum dots; IO: iron oxide; PET: positron-emission tomography; NIRF: near-infrared fluorescence.

DOTA-QD-VEGF was also investigated (35). The U87MG tumor uptake of active nanotargeted ^{64}Cu -DOTA-QD-VEGF (1.52 ± 0.6 % ID/g, 2.81 ± 0.3 % ID/g, 3.84 ± 0.4 % ID/g, and 4.16 ± 0.5 % ID/g at 1, 4, 16 and 24 h, respectively, post injection) was one percentage injected dose per gram (% ID/g) higher than that passive targeted ^{64}Cu -DOTA-QD (35). Development of a bifunctional polyaspartic acid-coated nanotargeted iron oxide (IO) molecular probe for PET and magnetic resonance imaging (MRI) of tumor integrin- $\alpha_v\beta_3$ expression was reported; this bifunctional ^{64}Cu -DOTA-IO-RGD nanotargeted molecular imaging approach may allow for earlier tumor detection and may provide insight into the molecular mechanisms of cancer (36).

Nanotargeted Radiopharmaceuticals for Tumor Therapeutics

Typically, nanotargeted radiopharmaceuticals have a two-component architecture for passive targeting therapeutics, *e.g.* a pegylated nanoliposome loaded with radionuclide payloads, and three-component architecture for active targeting therapeutics, such as pegylated nanoliposome bioconjugated with targeting antibody or peptide and encapsulated radionuclide payloads (16, 17). Selected cancer nanotargeted therapeutics are summarized in Table V. An

analytical dosimetry study for the use of ^{131}I , ^{90}Y , ^{188}Re , and ^{67}Cu radionuclide labeled liposome for internal radiotherapy has been reported, and the analysis suggested that the optimal liposome system for radiotherapy differs from that for chemotherapy delivery (37). The results of the effective targeting of solid tumors in patients with advanced local cancer by radiolabeled pegylated liposomes support the possibility of the delivery of β -emitting radionuclide loaded pegylated liposome for the treatment of solid tumors, particularly those liposomes in head and neck patients with highest levels of tumor uptake 33.0 ± 15.8 % ID/kg for a tumor volume of 36.2 ± 18.0 cm³ (26).

Bao *et al.* have developed a method of labeling liposomes with radionuclides using *N, N*-bis (2-mercaptoethyl)-*N', N'*-diethylethylenediamine (BMEDA) to post-load ^{99m}Tc or ^{186}Re into liposomes (38, 39). In addition to therapy *via* intravenous administration, the intratumoral and intraoperative therapy were also investigated for the potential use of ^{186}Re -liposomes (40-42). High-resolution SPECT/CT images revealed the intratumoral distribution of therapeutic liposomes and indicated the potential use of ^{186}Re -liposomes for intratumoral therapy (40). Intraoperative passive nanotargeted ^{186}Re -liposome therapy showed excellent tumor suppression and minimal side-effect profile in a head and neck squamous cell carcinoma xenograft positive surgical margin model (41).

Table V. *Selected cancer nanotargeted therapeutics.*

Nanocarrier	Radionuclide	Passive or active targeting	Applications	References
Liposomes	¹³¹ I, ⁹⁰ Y, ¹⁸⁸ Re, ⁶⁷ Cu	Passive, radiotherapeutics	An analytical dosimetry study for the use of radionuclide-liposome conjugates in internal radiotherapy	(37)
Liposomes	¹⁸⁶ Re	Passive, radiotherapeutics	Intraoperative ¹⁸⁶ Re-liposome radionuclide therapy in a head and neck squamous cell carcinoma xenograft positive surgical margin model	(41)
Liposomes	¹¹¹ In, ¹⁸⁸ Re	Passive, radiotherapeutics	Imaging, biodistribution, pharmacokinetics, and therapeutic efficacy studies of ¹¹¹ In/ ¹⁸⁸ Re-liposome on C26 and HT-29 tumor-bearing animal models	(28, 46, 47)
Liposomes	¹¹¹ In, ¹⁸⁸ Re	Passive, radiochemotherapeutics	Imaging, biodistribution, pharmacokinetics, therapeutic efficacy, and dosimetry studies of ¹¹¹ In/ ¹⁸⁸ Re-VNB/DXR-liposome on C26 and HT-29 tumor/ascites-bearing animal models	(29, 48-50)
Liposomes/ immunoliposomes	²²⁵ Ac	Passive and active, radiotherapeutics	Targeted α -particles emitters of ²²⁵ Ac-generators encapsulated in liposomes as therapeutic agents for micrometastases cancer	(53-55)
Immunoliposomes	⁹⁰ Y	Active, radiotherapeutics	Targeted antiangiogenesis of $\alpha_v\beta_3$ -integrin or VEGFR2 anti-FLK-1 therapy with nanotargeted therapeutics of ⁹⁰ Y-DTPA-liposome-IA (integrin antagonist) or ⁹⁰ Y-DTPA-liposome-mAb	(56)
Liposomes and dendrimers	¹⁰ B	Passive or active, therapeutics	¹⁰ B-liposomes and ¹⁰ B-PAMAM dendrimers-anti-folate nanotargeted therapeutics for boron neutron capture therapy (BNCT)	(57, 58)

Concomitant chemotherapy and radiotherapy has been illustrated to improve treatment outcome in a range of solid tumors. Pegylated liposome-encapsulation of doxorubicin and cisplatin has been shown to target drugs to tumors, increase therapeutic efficacy and reduce toxicity (43). Trimodal cancer therapy combining antiangiogenesis, chemotherapy and radiotherapy has beneficial effects and is emerging as a clinical antitumor strategy (44). Image-guided and passive nanocarrier-based polymeric nanomedicine for radiotherapy holds significant potential for improving the treatment of advanced solid tumors (45). Biodistribution, pharmacokinetics, and nuclear imaging of passive nanotargeted radio-therapeutics of ¹¹¹In/¹⁸⁸Re-liposome on C26 and HT-29 colon carcinoma-bearing animal models have been studied by our group (28, 46, 47). ¹¹¹In has γ -rays with 171 keV energy for nuclear imaging and auger electrons with 0.42 MeV energy in the nm tissue penetration range, with specific single tumor cell or small tumor cluster killing effect (as shown in Table II). ¹⁸⁸Re has γ -rays with 155 keV energy for nuclear imaging and is a high energy beta emitter with 2.12 MeV energy for nonspecific large tumor cluster killing effect. Both radionuclides can be used in bifunctional nuclear imaging and internal radio-therapeutic applications. Long-circulating pegylated liposomes radiolabeled with ¹⁸⁸Re (¹⁸⁸Re-liposomes) showed a higher uptake in the tumor as compared with ¹⁸⁸Re-BMEDA alone. Passive nanotargeted ¹⁸⁸Re-liposomes were found to have a 7.1-fold higher tumor-to-muscle uptake ratio as compared to

intravenously administered unencapsulated ¹⁸⁸Re-BMEDA in an animal model of C26 murine colon carcinoma solid tumor (46). Biodistribution, pharmacokinetics, nuclear imaging and therapeutic efficacies were investigated for nanotargeted bifunctional radiochemotherapeutics of ¹¹¹In/¹⁸⁸Re-(vinorelbine/doxorubicin, VNB/DXR)-liposomes on colorectal carcinoma of HT-29 and C26 tumor, and ascites-bearing animal models (29, 48-50). The accumulation and localization of passive nanotargeted ¹⁸⁸Re-liposomes have been studied in C26 solid-mouse model (Figure 3). The images revealed that ¹⁸⁸Re-liposomes remained in the tumor for up to 72 h *p.i.*, while the corresponding images for free ¹⁸⁸Re-BMEDA revealed that it could not efficiently accumulate in tumor at 4 h *p.i.* (46) (Figure 3).

In addition to the diagnostic imaging of ¹¹¹In/¹⁸⁸Re-liposome, additive therapeutic efficacy was observed for the comparative co-delivery radiochemotherapeutics of specific-killing auger electron emitters of ¹¹¹In-(VNB)-liposomes on HT-29/luc mouse xenografts (29, 49) (Figure 2B), and nonspecific-killing high energy β -emitters of ¹⁸⁸Re-DXR-liposomes on C26 ascites animal models (48) (Figure 4). Coronal microSPECT/CT image of ¹⁸⁸Re-DXR-liposomes correlated with its whole-body autoradiography image in C26 colon carcinoma ascites in mice is illustrated in Figure 4A. Furthermore, ¹⁸⁸Re-DXR-liposomes could provide a beneficial and promising strategy for the delivery of passive nanotargeted bimodality radiochemotherapeutics in the

treatment of tumor (48) (Figure 4B). Previous theoretical dosimetry studies have addressed the potential use of radiotherapeutic liposomes for the treatment of tumors *via* intravenous injection (37, 51, 52). The comparative dosimetric evaluation of nanotargeted ^{188}Re -(DXR)-liposome derived from the biodistribution indicated that the delivery radiation doses were safe and feasible for further clinical translation research from bench to bedside (50). The results for major organ doses for ^{188}Re -(DXR)-liposome revealed that similar doses were received by spleen and liver, but a lower dose was given to kidney, compared to that of ^{111}In -DTPA-octreotide therapy. Lower doses were also received by total body and liver, compared to ^{111}In -DTPA-hEGF radiotherapeutics (0.19 and 0.76 mGy/MBq, respectively). The absorbed doses for spleen, liver, kidney and red marrow in these studies are much lower than those from ^{90}Y -DOTATOC therapy (50).

Targeted α -particle emitters are promising therapeutics for micrometastatic tumors. Enhanced loading of ^{225}Ac and retention of three α -particle-emitting daughters of ^{225}Ac by passively targeted liposomes and actively targeted immunoliposomes have already been established (53-55). Targeted angiogenesis of $\alpha_v\beta_3$ and VEGFR2 with three-component actively nanotargeted radiopharmaceuticals of ^{90}Y -liposome-IA (integrin antagonist) and ^{90}Y -liposome-anti-Flk-1 (mAb) have been reported in the murine melanoma K1735-M2 and colon cancer CT26 animal models (56). The results demonstrated that ^{90}Y -liposome-anti-Flk-1 (mAb) was significantly more efficacious than conventional radioimmunotherapy in the mouse melanoma model (14). Boron neutron capture therapy (BNCT) is a binary approach to cancer therapy based on the nuclear reaction that occurs when ^{10}B is irradiated with thermal neutrons to yield high LET of α -particles and lithium nuclei. These particles have a short range ($<10\text{ }\mu\text{m}$) and deposit their energy within single cells. The efficacy and successful treatment of tumors by BNCT depend on the selective delivery of relatively high amounts of ^{10}B to tumors. Application of passive stealth liposomes and active folate receptor-targeted pamam-dendrimer-entrapped ^{10}B delivery systems have been studied for BNCT in animal models (57, 58). The results of the study of ^{10}B -PEG-liposome through intravenous injection suggested that passively targeted delivery of ^{10}B SH can increase the retention of ^{10}B by tumor cells, causing the suppression of tumor growth *in vivo* by BNCT (58).

Conclusion and Future Prospects

This review article briefly summarizes the recent research on tumor nanotargeted diagnostic and therapeutic radiopharmaceuticals, which is a potential field of novel cancer drug development and application. As compared to conventional targeted radionuclide therapy or radioimmunotherapy, the use

of nanocarriers can allow for the specific multivalent attachment of targeted molecules of antibodies, peptides or ligands to the surface of nanocarriers which can deliver a high payload of radionuclides, chemotherapeutics, and/or imaging agents to achieve multifunctional and multimodality targeting of tumor cells, and can enhance the efficacy and safety of targeted therapy. The simultaneous attainment of preferential location, reduction of immunotoxic effect and avoidance of the sequential biological barriers is a major challenge in passively and actively nanotargeted drug delivery systems.

Several passively and actively nanotargeted radiolabeled nanocarriers have been successfully applied to image and treat tumor models both preclinically and clinically. Future studies should be designed to optimize these novel approaches and extend them to combine potent radionuclides, imaging agents, chemotherapeutics and/or radiosensitizing agents. We have demonstrated that co-delivery of radiochemotherapeutics and simultaneous multifunctional imaging is an advantageous characteristic of nanotargeted radiopharmaceuticals for cancer imaging and therapy. An integrated 'bench to clinic' translational approach with good multidiscipline and multiinstitute collaboration between academia, research institutes and industry would accelerate the progression of research into nanotargeted radiopharmaceuticals toward clinical applications for the healthcare of cancer patients.

References

- Garcia M, Jemal A, Ward EM, Center MM, Hao Y, Siegel RL and Thun MJ: Global Cancer Facts and Figures. American Cancer Society 2008.
- Strebhardt K and Ullrich A: Paul Ehrlich's magic bullet concept: 100 years of progress. *Nat Rev Cancer* 8: 473-480, 2008.
- Adams GP and Weiner LM: Monoclonal antibody therapy of cancer. *Nat Biotechnol* 23: 1147-1157, 2005.
- Reichert JM and Valge-Archer VE: Development trends for monoclonal antibody cancer therapeutics. *Nat Rev Drug Discov* 6: 349-356, 2007.
- Carter PJ: Potent antibody therapeutics by design. *Nat Rev Immunol* 6: 343-357, 2006.
- Allen TM and Cullis PR: Drug delivery systems: entering the mainstream. *Science* 303: 1818-1822, 2004.
- Davis ME, Chen ZG and Shin DM: Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nat Rev Drug Discov* 7: 771-782, 2008.
- Ferrari M: Cancer nanotechnology: opportunities and challenges. *Nat Rev Cancer* 5: 161-171, 2005.
- Lammers T, Hennink WE and Storm G: Tumour-targeted nanomedicines: principles and practice. *Br J Cancer* 99: 392-397, 2008.
- Sanhai WR, Sakamoto JH, Canady R and Ferrari M: Seven challenges for nanomedicine. *Nat Nanotechnol* 3: 242-244, 2008.
- Milenic DE, Brady ED and Brechbiel MW: Antibody-targeted radiation cancer therapy. *Nat Rev Drug Discov* 3: 488-499, 2004.

- 12 Davies AJ: Radioimmunotherapy for B-cell lymphoma: ^{90}Y -ibritumomab tiuxetan and ^{131}I -tositumomab. *Oncogene* 26: 3614-3628, 2007.
- 13 Macklis RM and Pohlman B: Radioimmunotherapy for non-Hodgkin's lymphoma: a review for radiation oncologists. *Int J Radiat Oncol Biol Phys* 66: 833-841, 2006.
- 14 Jacene HA, Filice R, Kasecamp W and Wahl RL: Comparison of ^{90}Y -ibritumomab tiuxetan and ^{131}I -tositumomab in clinical practice. *J Nucl Med* 48: 1767-1776, 2007.
- 15 Wong JY: Systemic targeted radionuclide therapy: potential new areas. *Int J Radiat Oncol Biol Phys* 66: S74-82, 2006.
- 16 Mitra A, Nan A, Line BR and Ghandehari H: Nanocarriers for nuclear imaging and radiotherapy of cancer. *Curr Pharm Des* 12: 4729-4749, 2006.
- 17 Hamoudeh M, Kamleh MA, Diab R and Fessi H: Radionuclides delivery systems for nuclear imaging and radiotherapy of cancer. *Adv Drug Deliv Rev* 60: 1329-1346, 2008.
- 18 Carlsson J, Forssell Aronsson E, Hietala SO, Stigbrand T and Tennvall J: Tumour therapy with radionuclides: assessment of progress and problems. *Radiother Oncol* 66: 107-117, 2003.
- 19 Reilly RM, Kiarash R, Cameron RG, Porlier N, Sandhu J, Hill RP, Vallis K, Hendler A and Gariepy J: ^{111}In -labeled EGF is selectively radiotoxic to human breast cancer cells overexpressing EGFR. *J Nucl Med* 41: 429-438, 2000.
- 20 Phillips WT: Delivery of gamma-imaging agents by liposomes. *Adv Drug Deliv Rev* 37: 13-32, 1999.
- 21 Boerman OC, Laverman P, Oyen WJ, Corstens FH and Storm G: Radiolabeled liposomes for scintigraphic imaging. *Prog Lipid Res* 39: 461-475, 2000.
- 22 Chen P, Cameron R, Wang J, Vallis KA and Reilly RM: Antitumor effects and normal tissue toxicity of ^{111}In -labeled epidermal growth factor administered to athymic mice bearing epidermal growth factor receptor-positive human breast cancer xenografts. *J Nucl Med* 44: 1469-1478, 2003.
- 23 Reilly RM, Chen P, Wang J, Scollard D, Cameron R and Vallis KA: Preclinical pharmacokinetic, biodistribution, toxicology, and dosimetry studies of ^{111}In -DTPA-human epidermal growth factor: an auger electron-emitting radiotherapeutic agent for epidermal growth factor receptor-positive breast cancer. *J Nucl Med* 47: 1023-1031, 2006.
- 24 Huwyler J, Drewe J and Krahenbuhl S: Tumor targeting using liposomal antineoplastic drugs. *Int J Nanomedicine* 3: 21-29, 2008.
- 25 Kleiter MM, Yu D, Mohammadian LA, Niehaus N, Spasojevic I, Sanders L, Viglianti BL, Yarmolenko PS, Hauck M, Petry NA, Wong TZ, Dewhirst MW and Thrall DE: A tracer dose of technetium-99m-labeled liposomes can estimate the effect of hyperthermia on intratumoral doxil extravasation. *Clin Cancer Res* 12: 6800-6807, 2006.
- 26 Harrington KJ, Mohammadtaghi S, Uster PS, Glass D, Peters AM, Vile RG and Stewart JS: Effective targeting of solid tumors in patients with locally advanced cancers by radiolabeled pegylated liposomes. *Clin Cancer Res* 7: 243-254, 2001.
- 27 Oku N: Delivery of contrast agents for positron-emission tomography imaging by liposomes. *Adv Drug Deliv Rev* 37: 53-61, 1999.
- 28 Wang HE, Yu HM, Lu YC, Hsieh NN, Tseng YL, Huang KL, Chung KT, Chen CH, Hwang JJ, Lin WJ, Wang SJ, Ting G, Whang-Peng J and Deng WP: Internal radiotherapy and dosimetric study for $^{111}\text{In}/^{177}\text{Lu}$ -pegylated liposomes conjugates in tumor-bearing mice. *Nucl Instrum Meth A* 569: 533-537, 2006.
- 29 Lee WC, Hwang JJ, Tseng YL, Wang HE, Chang YF, Lu YC, Ting G, Whang-Peng J and Wang SJ: Therapeutic efficacy evaluation of ^{111}In -VNB-liposome on human colorectal adenocarcinoma HT-29/luc mouse xenografts. *Nucl Instrum Meth A* 569: 497-504, 2006.
- 30 Elbayoumi TA and Torchilin VP: Enhanced accumulation of long-circulating liposomes modified with the nucleosome-specific monoclonal antibody 2C5 in various tumours in mice: gamma-imaging studies. *Eur J Nucl Med Mol Imaging* 33: 1196-1205, 2006.
- 31 Erdogan S, Roby A and Torchilin VP: Enhanced tumor visualization by gamma-scintigraphy with ^{111}In -labeled polychelating-polymer-containing immunoliposomes. *Mol Pharm* 3: 525-530, 2006.
- 32 Hu G, Lijowski M, Zhang H, Partlow KC, Caruthers SD, Kiefer G, Gulyas G, Athey P, Scott MJ, Wickline SA and Lanza GM: Imaging of Vx-2 rabbit tumors with alpha(nu)beta3-integrin-targeted ^{111}In nanoparticles. *Int J Cancer* 120: 1951-1957, 2007.
- 33 McDevitt MR, Chattopadhyay D, Kappel BJ, Jaggi JS, Schiffman SR, Antczak C, Njardarson JT, Brentjens R and Scheinberg DA: Tumor targeting with antibody-functionalized, radiolabeled carbon nanotubes. *J Nucl Med* 48: 1180-1189, 2007.
- 34 Cai W, Chen K, Li ZB, Gambhir SS and Chen X: Dual-function probe for PET and near-infrared fluorescence imaging of tumor vasculature. *J Nucl Med* 48: 1862-1870, 2007.
- 35 Chen K, Li ZB, Wang H, Cai W and Chen X: Dual-modality optical and positron emission tomography imaging of vascular endothelial growth factor receptor on tumor vasculature using quantum dots. *Eur J Nucl Med Mol Imaging* 35: 2235-2244, 2008.
- 36 Lee HY, Li Z, Chen K, Hsu AR, Xu C, Xie J, Sun S and Chen X: PET/MRI dual-modality tumor imaging using arginine-glycine-aspartic (RGD)-conjugated radiolabeled iron oxide nanoparticles. *J Nucl Med* 49: 1371-1379, 2008.
- 37 Emfietzoglou D, Kostarelos K and Sgouros G: An analytic dosimetry study for the use of radionuclide-liposome conjugates in internal radiotherapy. *J Nucl Med* 42: 499-504, 2001.
- 38 Bao A, Goins B, Klipper R, Negrete G and Phillips WT: ^{186}Re -liposome labeling using ^{186}Re -SNS/S complexes: *in vitro* stability, imaging, and biodistribution in rats. *J Nucl Med* 44: 1992-1999, 2003.
- 39 Bao A, Goins B, Klipper R, Negrete G and Phillips WT: Direct $^{99\text{m}}\text{Tc}$ labeling of pegylated liposomal doxorubicin (Doxil) for pharmacokinetic and non-invasive imaging studies. *J Pharmacol Exp Ther* 308: 419-425, 2004.
- 40 Phillips WT, Goins B and Bao A: Radioactive liposomes. *Wiley Interdiscipl Rev Nanomed Nanobiotechnol* 1: 69-83, 2009.
- 41 Wang SX, Bao A, Herrera SJ, Phillips WT, Goins B, Santoyo C, Miller FR and Otto RA: Intraoperative ^{186}Re -liposome radionuclide therapy in a head and neck squamous cell carcinoma xenograft positive surgical margin model. *Clin Cancer Res* 14: 3975-3983, 2008.
- 42 Phillips WT, Klipper R and Goins B: Novel method of greatly enhanced delivery of liposomes to lymph nodes. *J Pharmacol Exp Ther* 295: 309-313, 2000.
- 43 Harrington KJ, Rowlinson-Busza G, Syrigos KN, Vile RG, Uster PS, Peters AM and Stewart JS: Pegylated liposome-encapsulated doxorubicin and cisplatin enhance the effect of radiotherapy in a tumor xenograft model. *Clin Cancer Res* 6: 4939-4949, 2000.

- 44 Huber PE, Bischof M, Jenne J, Heiland S, Peschke P, Saffrich R, Grone HJ, Debus J, Lipson KE and Abdollahi A: Trimodal cancer treatment: beneficial effects of combined antiangiogenesis, radiation, and chemotherapy. *Cancer Res* 65: 3643-3655, 2005.
- 45 Lammers T, Subr V, Peschke P, Kuhnlein R, Hennink WE, Ulbrich K, Kiessling F, Heilmann M, Debus J, Huber PE and Storm G: Image-guided and passively tumour-targeted polymeric nanomedicines for radiochemotherapy. *Br J Cancer* 99: 900-910, 2008.
- 46 Chang YJ, Chang CH, Chang TJ, Yu CY, Chen LC, Jan ML, Luo TY, Lee TW and Ting G: Biodistribution, pharmacokinetics and microSPECT/CT imaging of ^{188}Re -BMEDA-liposome in a C26 murine colon carcinoma solid tumor animal model. *Anticancer Res* 27: 2217-2225, 2007.
- 47 Chen LC, Chang CH, Yu CY, Chang YJ, Hsu WC, Ho CL, Yeh CH, Luo TY, Lee TW and Ting G: Biodistribution, pharmacokinetics and imaging of ^{188}Re -BMEDA-labeled pegylated liposomes after intraperitoneal injection in a C26 colon carcinoma ascites mouse model. *Nucl Med Biol* 34: 415-423, 2007.
- 48 Chen LC, Chang CH, Yu CY, Chang YJ, Wu YH, Lee WC, Yeh CH, Lee TW and Ting G: Pharmacokinetics, micro-SPECT/CT imaging and therapeutic efficacy of ^{188}Re -DXR-liposome in C26 colon carcinoma ascites mice model. *Nucl Med Biol* 35: 883-893, 2008.
- 49 Chow TH, Lin YY, Hwang JJ, Wang HE, Tseng YL, Pang VF, Wang SJ, Whang-Peng J and Ting G: Diagnostic and therapeutic evaluation of ^{111}In -vinorelbine-liposomes in a human colorectal carcinoma HT-29/luc-bearing animal model. *Nucl Med Biol* 35: 623-634, 2008.
- 50 Chang CH, Stabin MG, Chang YJ, Chen LC, Chen MH, Chang TJ, Lee TW and Ting G: Comparative dosimetric evaluation of nanotargeted ^{188}Re -(DXR)-liposome for internal radiotherapy. *Cancer Biother Radiopharm* 23: 749-758, 2008.
- 51 Kostarelos K and Emfietzoglou D: Tissue dosimetry of liposome-radionuclide complexes for internal radiotherapy: toward liposome-targeted therapeutic radiopharmaceuticals. *Anticancer Res* 20: 3339-3345, 2000.
- 52 Syme AM, McQuarrie SA, Middleton JW and Fallone BG: Dosimetric model for intraperitoneal targeted liposomal radioimmunotherapy of ovarian cancer micrometastases. *Phys Med Biol* 48: 1305-1320, 2003.
- 53 Sofou S, Thomas JL, Lin HY, McDevitt MR, Scheinberg DA and Sgouros G: Engineered liposomes for potential alpha-particle therapy of metastatic cancer. *J Nucl Med* 45: 253-260, 2004.
- 54 Chang MY, Seideman J and Sofou S: Enhanced loading efficiency and retention of ^{225}Ac in rigid liposomes for potential targeted therapy of micrometastases. *Bioconjug Chem* 19: 1274-1282, 2008.
- 55 Sofou S, Kappel BJ, Jaggi JS, McDevitt MR, Scheinberg DA and Sgouros G: Enhanced retention of the alpha-particle-emitting daughters of actinium-225 by liposome carriers. *Bioconjug Chem* 18: 2061-2067, 2007.
- 56 Li L, Wartchow CA, Danthi SN, Shen Z, Dechene N, Pease J, Choi HS, Doede T, Chu P, Ning S, Lee DY, Bednarski MD and Knox SJ: A novel antiangiogenesis therapy using an integrin antagonist or anti-Flk-1 antibody coated ^{90}Y -labeled nanoparticles. *Int J Radiat Oncol Biol Phys* 58: 1215-1227, 2004.
- 57 Shukla S, Wu G, Chatterjee M, Yang W, Sekido M, Diop LA, Muller R, Sudimack JJ, Lee RJ, Barth RF and Tjarks W: Synthesis and biological evaluation of folate receptor-targeted boronated PAMAM dendrimers as potential agents for neutron capture therapy. *Bioconjug Chem* 14: 158-167, 2003.
- 58 Yanagie H, Maruyama K, Takizawa T, Ishida O, Ogura K, Matsumoto T, Sakurai Y, Kobayashi T, Shinohara A, Rant J, Skvarc J, Ilic R, Kuhne G, Chiba M, Furuya Y, Sugiyama H, Hisa T, Ono K, Kobayashi H and Eriguchi M: Application of boron-entrapped stealth liposomes to inhibition of growth of tumour cells in the *in vivo* boron neutron-capture therapy model. *Biomed Pharmacother* 60: 43-50, 2006.

Received May 4, 2009

Revised July 15, 2009

Accepted July 20, 2009