

Review

The Role of Regulatory T Cells in Malignant Glioma

ADAM M. SONABEND, CLEO E. ROLLE and MACIEJ S. LESNIAK

The Brain Tumor Center, Section of Neurosurgery, The University of Chicago Pritzker School of Medicine, 5841 South Maryland Ave, MC 3026, Chicago, IL 60637, U.S.A.

Abstract. *The aggressive nature of gliomas is closely related to their capacity to evade the anti-tumoral immune response. The mechanisms implicated in this phenomenon are only partially understood. A subset of T cells, termed CD4⁺ CD25⁺ regulatory T cells (Treg), have been shown to inhibit the actions of effector lymphocytes. These Tregs are increased in the blood and tumors of glioma patients and animals with experimental brain tumors. Moreover, tumor infiltration by Tregs correlates with tumor grade and in animal models, depletion of Tregs is associated with prolonged survival. This review focuses on the role of Tregs in the immune suppression exhibited by malignant gliomas. The biology of these cells is briefly described in this context and finally, potential therapeutic strategies related to Treg ablation are explored.*

High grade gliomas represent the most common primary malignant tumor of the adult central nervous system (CNS) and unfortunately, the one with the worst prognosis. The aggressive nature of this neoplasia is closely related to its complex pathophysiology. In particular, evasion of the immune system by gliomas limits an effective anti-tumoral response.

The immunodeficient status associated with gliomas was described more than twenty years ago. Initially, it was noted that the cellular immune response of patients and animals with gliomas is impaired (1-3). The first reports described a lack of proliferation of the peripheral T cells of these patients after exposure to phytohemagglutinin (1, 2). More recently, the immunodeficiency induced by these tumors has been

Correspondence to: Maciej S. Lesniak, MD, Section of Neurosurgery, The University of Chicago Pritzker School of Medicine, 5841 South Maryland Ave, MC 3026, Chicago, IL 60637, U.S.A. Tel: +773 834 4757, Fax: +773 834 2608, e-mail: mlesniak@surgery.bs.d.uchicago.edu

Key Words: Glioma, regulatory T cells (Treg), brain tumor, CD4⁺ CD25⁺ cells, glioblastoma multiforme, immune function, review.

partially explained by the secretion of TGF-beta (4-8) and prostaglandins by tumor cells (9, 10). Moreover, tumor-infiltrating microglia express immunosuppressive cytokines such as interleukin 10 (IL-10) (11). In addition, the decreased level of major histocompatibility complex (MHC) class I expression by gliomas (12, 13) and the expression of human leukocyte antigen (HLA)-G, a non-typical class I MHC molecule (14) appear to play a role in this immunosuppressant status (15). To date, multiple factors contributing to the evasion of immune response by gliomas have been described, albeit, the precise relation between these mechanisms and their relative roles remain unknown.

CD4⁺ CD25⁺ regulatory T cells (Tregs) have been recently shown to infiltrate gliomas and their fraction is increased in the blood of patients bearing these tumors. Current evidence suggests a major role in the evasion of immune rejection by these cells (16-24). In this review, the role of Tregs in immune evasion by glioma is explored. In addition, the biology and the mechanism of action of these cells are described.

Biology of Regulatory T Cells

Tregs are lymphocytes that have a physiological role in the modulation of the immune response. Specifically, these cells prevent autoimmunity by inhibiting autoreactive effector T lymphocytes. Systemic depletion of Tregs has been associated with a wide variety of autoimmune diseases in murine models as well as in humans (25-31). In the context of cancer, the presence of Tregs is believed to maintain a lack of immune rejection of neoplastic cells in many malignancies including colorectal (32, 33), esophageal (34), gastric (34), pancreatic (35), breast (36-39), lung (39-42) and ovarian (42) tumors. Therefore, the precise understanding of the modus operandi of Tregs has potential therapeutic implications that should be explored.

Essentially, Tregs are distinguished from other lymphocytes by several characteristics. First, instead of being induced *de novo* from naïve T cells upon antigen exposure, Treg development takes place in the thymus. These cells leave the

thymus as a mature and distinct population with a defined functional phenotype (43). Moreover, these cells express IL-2 receptor α chain (IL-2R α chain or CD25) and are dependent on stimulation with IL-2 for their expansion and function (25, 44, 45). Consistent with this fact, mice that are deficient in IL-2, IL-2R, or signal transducer and activator of transcription 5 (STAT5) have a marked reduction in Tregs and subsequently develop autoimmunity (44, 46-50). The majority of the IL-2 needed for the activation of these cells is secreted by other T cells. Thereby, the presence of effector T cells and the resultant IL-2 secretion activate Tregs that subsequently modulate the activity of reactive cells. This feedback mechanism has been proposed to be constantly limiting the autoimmune response (reviewed by Sakaguchi in (51)). Tregs specifically express forkhead box P3 (FoxP3), a transcription factor that plays a key role in the definition of their phenotype (52-54). FoxP3 appears to be distinctive for Tregs as it is expressed in CD4⁺ CD25⁺ T cells and CD4⁺ CD25⁺ CD8⁻ thymocytes whereas it is not found in other thymic cells, T cells, B cells, natural killer or natural killer T cells (52, 53). FoxP3-deficient mice fail to develop Tregs and die from inflammatory diseases that can be abrogated by the transfer of these cells from naïve mice. Similarly, in the case of humans, a syndrome characterized by immune dysregulation, diabetes mellitus type I, thyroiditis, inflammatory bowel disease, and allergies is associated with mutations on the FoxP3 gene (55-57). Finally, Tregs are capable of suppressing the proliferation and action of other T cells. FoxP3 and CD25 are reliable and constant markers that have served to isolate and characterize Tregs.

Recent studies have described the existence of other populations of T regulatory lymphocytes which unlike classic CD4⁺ CD25⁺ Tregs, are induced in the periphery *via* T cell receptor (TCR)-MHC/peptide stimulation. At least three populations of peripherally induced CD4⁺ regulatory T cells have been described, Tr1 cells, Th3 cells, and iTregs, which differ in their genesis, their suppressive mechanisms, and their respective FoxP3 expression. Tr1 cells are induced in the periphery in a process that is dependent on IL-10 (58) and interactions with immature dendritic cells (DC) (which lack the expression of co-stimulatory molecules). Tr1 cells are characterized by the expression of CD4⁺ CD25^{int/high} and mediate suppression by secreting large amounts of IL-10. In contrast to Tregs, FoxP3 is not constitutively expressed in Tr1 cells (59). Th3 cells are induced in the periphery through a TGF- β dependent process, and these cells require IL-10 for expansion. Th3 cells suppress *via* the secretion of transforming growth factor beta (TGF- β) (60). Both Tr1 and Th3 cells are implicated in oral tolerance (61). With respect to their role in neoplasia, these cells constitute part of the tumor infiltrating lymphocytes from gastric cancer patients (34, 62). Lastly, iTreg are induced from CD4⁺ CD25⁻ T cells upon the exposure to the suppressive cytokine milieu at the tumor site. These cells express high levels of FoxP3 and can suppress *via*

both cell-cell contact and soluble factors. iTreg can also be induced *via* interactions with Tregs (63).

While there are multiple populations of regulatory T cells with distinct suppressive mechanisms, this review will focus on Tregs, characterized by co-expression of CD4, CD25 and FoxP3. The precise means by which Tregs inhibit effector T cells has not been fully elucidated. Some evidence has suggested the implication of cell-cell contact, with a significant contribution of cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) (64-67) and membrane-bound TGF- β (68, 69). In addition, heme oxygenase-1 (HO-1), a rate-limiting enzyme in heme catabolism, seems to play a role in the immune suppressive phenotype of Tregs. HO-1 is constitutively expressed in human CD4⁺ CD25⁺ Tregs (70) and its expression is induced by FoxP3 (71). HO-1 suppresses T cell proliferation through the production of carbon monoxide (72, 73).

Regulatory T Cells in Glioma

Our group has found tumor infiltration of Tregs in patients with glioblastoma multiforme (GBM) (17). The expression of CD4⁺ CD25⁺ FoxP3⁺ T cells was significantly higher in patients with GBM than in controls, with a mean of 24.7% of Tregs among the glioma-infiltrating lymphocytes, whereas these cells were absent from control brain specimens ($p < 0.01$). Moreover, higher levels of FoxP3 expression in the CD4⁺ CD25⁺ cells were observed in regulatory T cells isolated from the tumor tissue (55.1%) in comparison to autologous patient blood (33.4%) and blood from control individuals (15.6%) ($p < 0.01$). In an *in vitro* suppression assay Tregs inhibited T cell proliferation in a dose-dependent manner. Among various markers analyzed, the expression of CD62L and CTLA-4 was elevated in the glioma Tregs in comparison to that of the controls.

At the same time, Fecci *et al.*, (20) found that whereas the absolute counts of both CD4⁺ T cells and CD4⁺ CD25⁺ FoxP3⁺ CD45RO⁺ Tregs were greatly diminished in the peripheral pool of patients with malignant glioma, the Treg fraction was increased in the remaining CD4 compartment in 5 out of the 8 patients evaluated (Figure 1). The proportion of Tregs in the peripheral blood of patients with GBM was 2.63 times higher than that found in the blood of normal volunteers ($p = 0.004$). Interestingly, their experiments suggested that despite the reduction in their total number, the increased Treg fraction ($p = 0.0003$ *versus* controls) was sufficient to elicit the characteristic impairment of T-cell responsiveness *in vitro* (Figure 2). The patients with an elevated Treg fraction showed significant CD4⁺ T cell proliferative dysfunction ($p < 0.0001$), whereas the patients without Treg elevation possessed CD4⁺ T cells that proliferated at normal levels. After Treg depletion *in vitro*, T cells from the patients bearing malignant gliomas regained their function and proliferated to levels equivalent to those of normal controls (20).

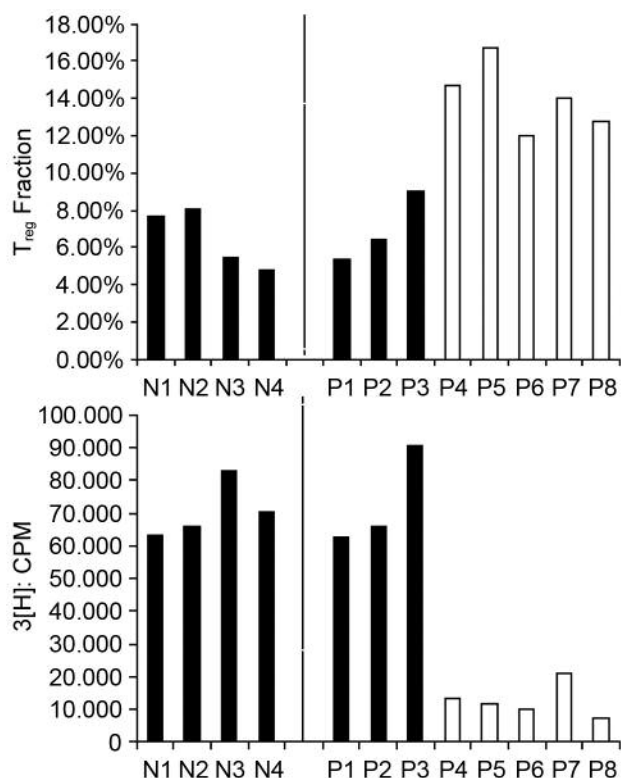


Figure 1. The fraction of Tregs (top) and the proliferation levels of CD4⁺ cells (bottom) in eight patients with GBM (P1 to P8) and four healthy individuals (N1 to N4) who underwent leukapheresis. Of note, the 5 patients with an increased Treg fraction had a poor CD4⁺ lymphocyte proliferation (white columns). 3[H]: CPM (counts per minute). Reprinted with permission from (20).

More recently, our group has found a correlation between glioma Treg infiltration and tumor grade (Figure 3) (74). In this study, the correlation between FoxP3 and HO-1 was investigated. The highest level of FoxP3 expression was found in patients with grade IV tumors (11.54%) *versus* grade III (6.74%) or grade II (2.53%) ($p < 0.05$). Moreover, in grade IV tumors, the frequency of HO-1 mRNA expression in CD4⁺ CD25⁺ cells was $11.8 \pm 2.45\%$ *vs.* $7.42 \pm 0.31\%$ in grade III and $2.33 \pm 0.12\%$ in grade II. HO-1 has been shown to accumulate during glioma progression, and apparently, it plays a role in FoxP3 mediated immune suppression. Tumor infiltrating Tregs stained positively with anti-HO-1 antibody and the expression of HO-1 correlated with CD4⁺ CD25⁺ FoxP3⁺ infiltration ($r = 0.966$). These results suggested that the induction of HO-1 mRNA expression is linked to the expression of FoxP3 in CD4⁺ CD25⁺ glioma infiltrating Tregs. Collectively, this data supports the notion that HO-1 is a key suppressive factor for regulatory T-cells during the growth of malignant brain tumors (74).

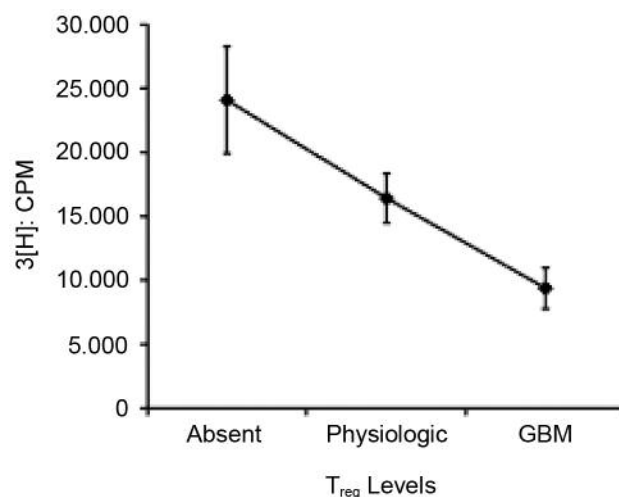


Figure 2. The increased Treg fraction in 5 out of the 8 patients with GBM is associated with a proliferative impairment of CD4⁺ lymphocytes from healthy human donors. For this experiment Tregs from GBM bearing patients and CD4⁺ cells from healthy controls were isolated and mixed to a final fraction of an absence of Tregs (0%), physiological fraction of Tregs (8%), or the Treg fraction observed in patients with GBM (16%). The cells were stimulated with anti-CD3. Increases in the Treg fraction to those levels found in patients with GBM were sufficient to produce significant deficits in the ability of T cells to proliferate. Reprinted with permission from (20).

Chemokines: Mediators of Tumor Infiltration by Tregs

Chemokines are a series of soluble peptides that have been implicated in various processes including angiogenesis and CNS development. Of utmost interest, chemokines play a central role within the immune system, as the secretion of these molecules leads to “chemotactic” migration of leucocytes (75-77). The vast variety of known chemokines are classified according to their cystin motifs (C), and accordingly, different families for these receptors such CXC, CC, C and CX3C have been described (78). Binding of specific chemokines to their cognate receptors, which are coupled to G proteins, promotes distinct chemotactic effects depending on the leucocyte population and chemokine receptor expression patterns. Chemokines appear to play a significant role in various human diseases including cancer (79-82). Since it is known that chronic inflammation can predispose to cancer formation and progression, it is suspected that the expression of chemokines could contribute to this process (reviewed by Rollins in (79)). On the other hand, chemokines might elicit an intrinsic effect on tumor cells. For instance, multiple human cancers including leukemias, lymphomas, gliomas and various epithelial carcinomas express CXC receptor 4 (CXCR4) and respond to its

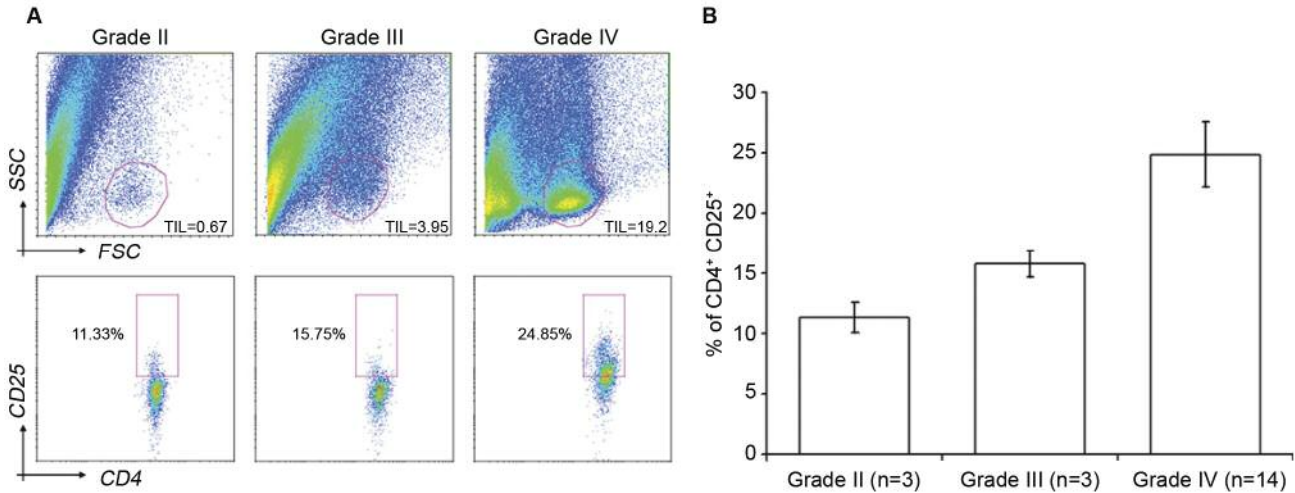


Figure 3. Frequency of tumor infiltrating lymphocytes in patients with astrocytoma (grade II-IV). (A) Tregs infiltration in human astrocytoma was examined by flow cytometry analysis of CD4 and CD25 expression of electronically gated lymphocytes. (B) shows the frequency of Tregs with grade of disease. The mean frequency of regulatory T-cells was $11.33 \pm 1.52\%$ (range: 10.46–13.5%) in grade II, $15.75 \pm 1.5\%$ (range: 14.88–17.5%) in grade III, and $24.85 \pm 2.67\%$ (range: 20–28.4%) in grade IV glioma tumors ($p < 0.02$) in total CD3⁺ T-cells. TIL (tumor infiltrating lymphocytes). Reprinted with permission from (74). SSC (Site scatter), FSC (Forward scatter).

ligand CXCL12 (CXCL12). This ligand-receptor interaction promotes the migration and metastatic establishment of tumor cells (82).

With regards to the migration of Tregs into tumors, there is data suggesting that cancers express a series of chemokines that promote the infiltration by these regulatory lymphocytes. For instance, chemokine CCL22 promotes the migration of Tregs into prostate and ovarian carcinomas (83, 84). Human gliomas express chemokines CCL2 and CCL22, and secrete CCL2. This has been investigated in the human glioma cell lines D-54, U-87, U-251, and LN-229 as well as in tumor cells from eight patients with GBM. Interestingly, the Tregs from these brain tumor patients had significantly higher expression of the CCL2 receptor CCR4 than the Tregs from healthy controls. Migration experiments have suggested that Treg migration is mediated by CCL2 and CCL22. Moreover, this migration was blocked by antibodies to the chemokine receptors CCR2 and CCR4 (85).

Future Perspectives: Therapeutic Ablation of Tregs

Ideally, the ablation of Tregs could lead to an effective immune response against gliomas. This rationale is supported by two facts. Firstly, effective anti-tumoral responses documented in anti-glioma immunotherapies suggest an important role for T cells. Secondly, the main targets of Tregs's suppressive features are also T cells.

Anti-CD25 antibodies. Different alternatives to neutralize Tregs are being explored. The use of an anti-CD25 antibody

is an illustrative example of this principle (16, 21, 22). Consistent with findings in human patients with gliomas (86), in addition to CD4 lymphopenia, the Treg fraction is increased in glioma-bearing mice (16, 87). To evaluate the role of Tregs in tumor development, our group has tested an anti-CD25 antibody in a murine model for glioma, where tumors were established by intracranial implantation of the cell line GL261. The tumor-infiltrating lymphocytes isolated from mice with GL261 tumors were found to have a significant increase in Tregs compared with the control animals ($p < 0.05$). The animals injected with anti-CD25 antibody exhibited a decrease in Tregs (CD4⁺ CD25⁺) and lived significantly longer than the untreated tumor-bearing control animals (Figure 4) ($p < 0.05$).

Fecci *et al.*, have also found that the anti-CD25 antibody is beneficial for the treatment of experimental brain tumors (21). Consistent with their findings in human patients with gliomas, in addition to CD4 lymphopenia, the Treg fraction was increased in glioma-bearing mice, but systemic anti-CD25 administration failed to completely eliminate Tregs, reducing their number only moderately. Nonetheless, the suppressive function of the Tregs decreased leading to enhanced lymphocyte proliferative and interferon gamma (IFN- γ) responses and up to 80% specific lysis of glioma cell targets *in vitro* (21).

Targeting by TLR ligands. Toll-like receptors (TLR) are interesting molecules in the context of tumor immunity. Specifically, stimulation of TLR9 by DNA containing CpG sequences has been shown to elicit an effective anti-tumor

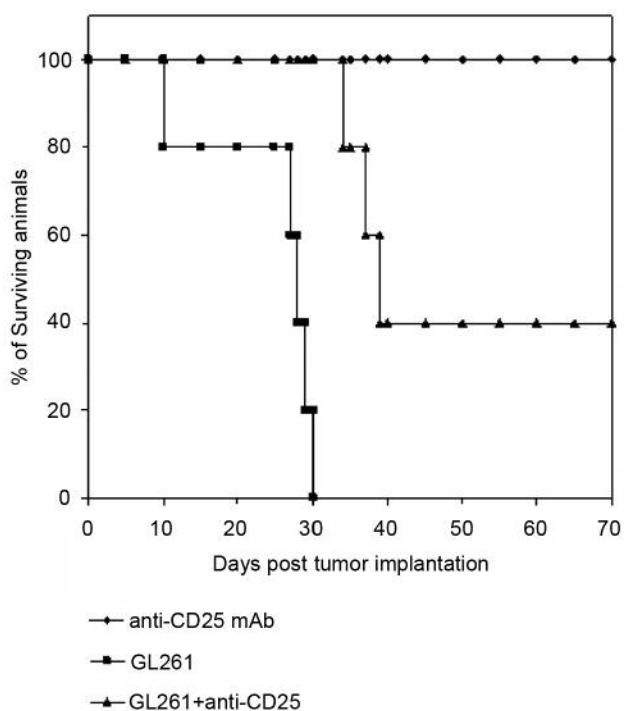


Figure 4. Prolongation of survival of mice with experimental GL261 gliomas after treatment with anti CD25 antibody is represented on the Kaplan-Meier plot. Median survival was increased from 27 to 40 days. 40% of the mice treated with the anti CD25 antibody remained long-term survivors (>70 days) ($p < 0.005$). Reprinted with permission from (16).

response for various neoplasias including experimental brain tumors (19, 88-92). With regard to Tregs, some evidence suggests the possibility of neutralizing their effects by stimulating TLRs. For instance, one study has described the reversal of Treg suppressive function by stimulation with synthetic or natural ligands for human TLR8 (93). Interestingly, in this case, the effect was independent of DC, but required functional TLR8, MyD88 and IRAK4 (molecules implicated in the intracellular signaling pathway for various TLRs) signaling in the Treg cells. Most importantly, the transfer of TLR8 ligand-stimulated Treg cells into tumor-bearing mice led to the enhancement of anti-tumor immunity (93).

Systemic treatment with TLR9 ligands has a deleterious effect on Tregs. We have described apoptosis induction in experimental brain tumors and the prolongation of survival following stimulation of TLR9 with CpGs. Interestingly, in addition to apoptosis, CpG stimulation of murine gliomas enhanced the antigen presenting capacity of microglia, shifted the immune response toward CD8⁺ T cells, and decreased the number of infiltrating Tregs (19). The mechanism for this phenomenon is not clear, and in this study the extent to which Treg decrease contributed to the therapeutic benefit of CpG was not explored. Nevertheless,

this finding supports the possibility of antagonizing Tregs by stimulation of TLRs, an illustrative example of modulation of lymphocyte function by the innate immunity.

Summary

Tregs contribute to the evasion of the immune response required for the development of malignant gliomas. These cells are found infiltrating tumors, and in the blood of patients and animals with gliomas. Compelling evidence suggests that tumoral infiltration by Tregs leads to the suppression of effector T cells, which would otherwise be capable of mounting an immune response against gliomas.

The causative relation of Tregs tumor infiltration and the progression of malignant gliomas is not clearly defined. Nevertheless, the possibility of Treg suppression of the anti-tumor immunity contributing to the development of gliomas is suggested by the correlation of Treg tumor infiltration and tumor grade, and by the fact that these cells are capable of suppressing tumor immunity.

In contrast to the development of other lymphocyte populations, naturally occurring Tregs seem to mature in the thymus rather than in the periphery. An interesting question that remains unanswered is the means by which gliomas can promote the generation of Tregs in the thymus or in the tumor site.

The role of Tregs in the tumor biology of gliomas might be of interest for its potential therapeutic implications. Indeed, animal models have shown evidence of antitumor effects derived from the ablation of Tregs and some preliminary studies suggest that these cells can be targeted by various means. Neutralization with anti-CD25 antibodies or DNA oligonucleotides that stimulate TLR ligands are two examples of such a principle. Further research is needed to investigate the best way to limit the activity of these regulatory cells. The modulation of anti-tumor immunity is a rapidly evolving field within brain tumor biology. This area of research warrants close attention by the professionals who treat patients with such a devastating disease, since a thorough understanding of this process might lead to interesting therapeutic implications in the near future.

Acknowledgements

This work was supported by the National Institute of Neurological Disorders and Stroke (K08-NS046430), The Alliance for Cancer Gene Therapy Young Investigator Award, the American Cancer Society (RSG-07-276-01-MGO), and the Elsa U Pardee Foundation.

References

- 1 Wood GW and Morantz RA: *In vitro* reversal of depressed T-lymphocyte function in the peripheral blood of brain tumor patients. *J Natl Cancer Inst* 68(1): 27-33, 1982.

- 2 Wood GW and Morantz RA: Depressed T lymphocyte function in brain tumor patients: monocytes as suppressor cells. *J Neurooncol* *1*(2): 87-94, 1983.
- 3 Yamasaki T, Handa H, Yamashita J, Namba Y and Hanaoka M: Characteristic immunological responses to an experimental mouse brain tumor. *Cancer Res* *43*(10): 4610-4617, 1983.
- 4 Grauer O, Poschl P, Lohmeier A, Adema GJ and Bogdahn U: Toll-like receptor triggered dendritic cell maturation and IL-12 secretion are necessary to overcome T-cell inhibition by glioma-associated TGF-beta2. *J Neurooncol* *82*(2): 151-161, 2007.
- 5 Platten M, Wick W and Weller M: Malignant glioma biology: role for TGF-beta in growth, motility, angiogenesis, and immune escape. *Microsc Res Tech* *52*(4): 401-410, 2001.
- 6 Weller M and Fontana A: The failure of current immunotherapy for malignant glioma. Tumor-derived TGF-beta, T-cell apoptosis, and the immune privilege of the brain. *Brain Res Brain Res Rev* *21*(2): 128-151, 1995.
- 7 Olofsson A, Miyazono K, Kanzaki T, Colosetti P, Engstrom U and Heldin CH: Transforming growth factor-beta 1, -beta 2, and -beta 3 secreted by a human glioblastoma cell line. Identification of small and different forms of large latent complexes. *J Biol Chem* *267*(27): 19482-19488, 1992.
- 8 Kuppner MC, Hamou MF, Sawamura Y, Bodmer S and de Tribolet N: Inhibition of lymphocyte function by glioblastoma-derived transforming growth factor beta 2. *J Neurosurg* *71*(2): 211-217, 1989.
- 9 Lauro GM, Di Lorenzo N, Grossi M, Maleci A and Guidetti B: Prostaglandin E2 as an immunomodulating factor released *in vitro* by human glioma cells. *Acta Neuropathol (Berl)* *69*(3-4): 278-282, 1986.
- 10 Nakano Y, Kuroda E, Kito T, Yokota A and Yamashita U: Induction of macrophagic prostaglandin E2 synthesis by glioma cells. *J Neurosurg* *104*(4): 574-582, 2006.
- 11 Wagner S, Czub S, Greif M *et al*: Microglial/macrophage expression of interleukin 10 in human glioblastomas. *Int J Cancer* *82*(1): 12-16, 1999.
- 12 Ito A, Shinkai M, Honda H, Wakabayashi T, Yoshida J and Kobayashi T: Augmentation of MHC class I antigen presentation *via* heat shock protein expression by hyperthermia. *Cancer Immunol Immunother* *50*(10): 515-522, 2001.
- 13 Prins RM and Liau LM: Cellular immunity and immunotherapy of brain tumors. *Front Biosci* *9*: 3124-3136, 2004.
- 14 Wiendl H, Mitsdoerffer M, Hofmeister V *et al*: A functional role of HLA-G expression in human gliomas: an alternative strategy of immune escape. *J Immunol* *168*(9): 4772-4780, 2002.
- 15 Wiendl H, Mitsdoerffer M and Weller M: Hide-and-seek in the brain: a role for HLA-G mediating immune privilege for glioma cells. *Semin Cancer Biol* *13*(5): 343-351, 2003.
- 16 El Andaloussi A, Han Y and Lesniak MS: Prolongation of survival following depletion of CD4+ CD25+ regulatory T cells in mice with experimental brain tumors. *J Neurosurg* *105*(3): 430-437, 2006.
- 17 El Andaloussi A and Lesniak MS: An increase in CD4+ CD25+ FOXP3+ regulatory T cells in tumor-infiltrating lymphocytes of human glioblastoma multiforme. *Neuro-oncol* *8*(3): 234-243, 2006.
- 18 El Andaloussi A and Lesniak MS: CD4+ CD25+ FoxP3+ T-cell infiltration and heme oxygenase-1 expression correlate with tumor grade in human gliomas. *J Neurooncol* *83*(2): 145-152, 2007.
- 19 El Andaloussi A, Sonabend AM, Han Y and Lesniak MS: Stimulation of TLR9 with CpG ODN enhances apoptosis of glioma and prolongs the survival of mice with experimental brain tumors. *Glia* *54*(6): 526-535, 2006.
- 20 Fecci PE, Mitchell DA, Whitesides JF *et al*: Increased regulatory T-cell fraction amidst a diminished CD4 compartment explains cellular immune defects in patients with malignant glioma. *Cancer Res* *66*(6): 3294-3302, 2006.
- 21 Fecci PE, Sweeney AE, Grossi PM *et al*: Systemic anti-CD25 monoclonal antibody administration safely enhances immunity in murine glioma without eliminating regulatory T cells. *Clin Cancer Res* *12*(14 Pt 1): 4294-4305, 2006.
- 22 Grauer OM, Nierkens S, Binnink E *et al*: CD4+ FoxP3+ regulatory T cells gradually accumulate in gliomas during tumor growth and efficiently suppress anti-glioma immune responses *in vivo*. *Int J Cancer* *121*(1): 95-105, 2007.
- 23 Hussain SF, Yang D, Suki D, Aldape K, Grimm E and Heimberger AB: The role of human glioma-infiltrating microglia/macrophages in mediating antitumor immune responses. *Neuro-oncol* *8*(3): 261-279, 2006.
- 24 Learn CA, Fecci PE, Schmittling RJ *et al*: Profiling of CD4+, CD8+, and CD4+ CD25+ CD45RO+ FoxP3+ T cells in patients with malignant glioma reveals differential expression of the immunologic transcriptome compared with T cells from healthy volunteers. *Clin Cancer Res* *12*(24): 7306-7315, 2006.
- 25 Sakaguchi S, Sakaguchi N, Asano M, Itoh M and Toda M: Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol* *155*(3): 1151-1164, 1995.
- 26 Balandina A, Lecart S, Darteville P, Saoudi A and Berrih-Aknin S: Functional defect of regulatory CD4(+)CD25+ T cells in the thymus of patients with autoimmune myasthenia gravis. *Blood* *105*(2): 735-741, 2005.
- 27 Kriegl MA, Lohmann T, Gabler C, Blank N, Kalden JR and Lorenz HM: Defective suppressor function of human CD4+ CD25+ regulatory T cells in autoimmune polyglandular syndrome type II. *J Exp Med* *199*(9): 1285-1291, 2004.
- 28 Longhi MS, Hussain MJ, Mitry RR *et al*: Functional study of CD4+ CD25+ regulatory T cells in health and autoimmune hepatitis. *J Immunol* *176*(7): 4484-4491, 2006.
- 29 Longhi MS, Ma Y, Bogdanos DP, Cheeseman P, Mieli-Vergani G and Vergani D: Impairment of CD4(+)CD25(+) regulatory T-cells in autoimmune liver disease. *J Hepatol* *41*(1): 31-37, 2004.
- 30 Longhi MS, Ma Y, Mitry RR *et al*: Effect of CD4+ CD25+ regulatory T-cells on CD8 T-cell function in patients with autoimmune hepatitis. *J Autoimmun* *25*(1): 63-71, 2005.
- 31 Ryan KR, Lawson CA, Lorenzi AR, Arkwright PD, Isaacs JD and Lilic D: CD4+ CD25+ T-regulatory cells are decreased in patients with autoimmune polyendocrinopathy candidiasis ectodermal dystrophy. *J Allergy Clin Immunol* *116*(5): 1158-1159, 2005.
- 32 Ling KL, Pratap SE, Bates GJ *et al*: Increased frequency of regulatory T cells in peripheral blood and tumour infiltrating lymphocytes in colorectal cancer patients. *Cancer Immun* *7*: 7-14, 2007.
- 33 Clarke SL, Betts GJ, Plant A *et al*: CD4+ CD25+ FOXP3+ regulatory T cells suppress anti-tumor immune responses in patients with colorectal cancer. *PLoS ONE* *1*(e129), 2006.

- 34 Ichihara F, Kono K, Takahashi A, Kawaida H, Sugai H and Fujii H: Increased populations of regulatory T cells in peripheral blood and tumor-infiltrating lymphocytes in patients with gastric and esophageal cancers. *Clin Cancer Res* 9(12): 4404-4408, 2003.
- 35 Liyanage UK, Moore TT, Joo HG *et al*: Prevalence of regulatory T cells is increased in peripheral blood and tumor microenvironment of patients with pancreas or breast adenocarcinoma. *J Immunol* 169(5): 2756-2761, 2002.
- 36 Bates GJ, Fox SB, Han C *et al*: Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. *J Clin Oncol* 24(34): 5373-5380, 2006.
- 37 Slavina EG, Chertkova AI, Zabortina TN, Gan'shina IP and Lichinitser MR: Variations in the number of regulatory T cells (CD4⁺ CD25⁺) in patients with breast cancer during herceptin therapy. *Bull Exp Biol Med* 141(3): 361-363, 2006.
- 38 Hueman MT, Stojadinovic A, Storrer CE *et al*: Levels of circulating regulatory CD4⁺ CD25⁺ T cells are decreased in breast cancer patients after vaccination with a HER2/neu peptide (E75) and GM-CSF vaccine. *Breast Cancer Res Treat* 98(1): 17-29, 2006.
- 39 Okita R, Saeki T, Takashima S, Yamaguchi Y and Toge T: CD4⁺ CD25⁺ regulatory T cells in the peripheral blood of patients with breast cancer and non-small cell lung cancer. *Oncol Rep* 14(5): 1269-1273, 2005.
- 40 Meloni F, Morosini M, Solari N *et al*: Foxp3 expressing CD4⁺ CD25⁺ and CD8⁺ CD28⁻ T regulatory cells in the peripheral blood of patients with lung cancer and pleural mesothelioma. *Hum Immunol* 67(1-2): 1-12, 2006.
- 41 Woo EY, Yeh H, Chu CS *et al*: Cutting edge: Regulatory T cells from lung cancer patients directly inhibit autologous T cell proliferation. *J Immunol* 168(9): 4272-4276, 2002.
- 42 Woo EY, Chu CS, Goletz TJ *et al*: Regulatory CD4⁺ CD25⁺ T cells in tumors from patients with early-stage non-small cell lung cancer and late-stage ovarian cancer. *Cancer Res* 61(12): 4766-4772, 2001.
- 43 Itoh M, Takahashi T, Sakaguchi N *et al*: Thymus and autoimmunity: production of CD25⁺ CD4⁺ naturally anergic and suppressive T cells as a key function of the thymus in maintaining immunologic self-tolerance. *J Immunol* 162(9): 5317-5326, 1999.
- 44 Furtado GC, Curotto de Lafaille MA, Kutchukhidze N and Lafaille JJ: Interleukin 2 signaling is required for CD4⁺ regulatory T cell function. *J Exp Med* 196(6): 851-857, 2002.
- 45 Malek TR and Bayer AL: Tolerance, not immunity, crucially depends on IL-2. *Nat Rev Immunol* 4(9): 665-674, 2004.
- 46 Antov A, Yang L, Vig M, Baltimore D and Van Parijs L: Essential role for STAT5 signaling in CD25⁺ CD4⁺ regulatory T cell homeostasis and the maintenance of self-tolerance. *J Immunol* 171(7): 3435-3441, 2003.
- 47 Bayer AL, Yu A, Adeegbe D and Malek TR: Essential role for interleukin-2 for CD4⁺ CD25⁺ T regulatory cell development during the neonatal period. *J Exp Med* 201(5): 769-777, 2005.
- 48 Bayer AL, Yu A and Malek TR: Function of the IL-2R for thymic and peripheral CD4⁺ CD25⁺ Foxp3⁺ T regulatory cells. *J Immunol* 178(7): 4062-4071, 2007.
- 49 Burchill MA, Yang J, Vogtenhuber C, Blazar BR and Farrar MA: IL-2 receptor beta-dependent STAT5 activation is required for the development of Foxp3⁺ regulatory T cells. *J Immunol* 178(1): 280-290, 2007.
- 50 Yao Z, Kanno Y, Kerenyi M *et al*: Nonredundant roles for Stat5a/b in directly regulating Foxp3. *Blood* 109(10): 4368-4375, 2007.
- 51 Sakaguchi S: Naturally arising Foxp3-expressing CD25⁺ CD4⁺ regulatory T cells in immunological tolerance to self and non-self. *Nat Immunol* 6(4): 345-352, 2005.
- 52 Hori S, Nomura T and Sakaguchi S: Control of regulatory T cell development by the transcription factor Foxp3. *Science* 299(5609): 1057-1061, 2003.
- 53 Fontenot JD, Gavin MA and Rudensky AY: Foxp3 programs the development and function of CD4⁺ CD25⁺ regulatory T cells. *Nat Immunol* 4(4): 330-336, 2003.
- 54 Khattri R, Cox T, Yasayko SA and Ramsdell F: An essential role for Scurfin in CD4⁺ CD25⁺ T regulatory cells. *Nat Immunol* 4(4): 337-342, 2003.
- 55 Levy-Lahad E and Wildin RS: Neonatal diabetes mellitus, enteropathy, thrombocytopenia, and endocrinopathy: Further evidence for an X-linked lethal syndrome. *J Pediatr* 138(4): 577-580, 2001.
- 56 Wildin RS, Ramsdell F, Peake J *et al*: X-linked neonatal diabetes mellitus, enteropathy and endocrinopathy syndrome is the human equivalent of mouse scurfy. *Nat Genet* 27(1): 18-20, 2001.
- 57 Bennett CL, Christie J, Ramsdell F *et al*: The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet* 27(1): 20-21, 2001.
- 58 Levings MK, Gregori S, Tresoldi E, Cazzaniga S, Bonini C and Roncarolo MG: Differentiation of Tr1 cells by immature dendritic cells requires IL-10 but not CD25⁺ CD4⁺ Tr cells. *Blood* 105(3): 1162-1169, 2005.
- 59 Vieira PL, Christensen JR, Minaee S *et al*: IL-10-secreting regulatory T cells do not express Foxp3 but have comparable regulatory function to naturally occurring CD4⁺ CD25⁺ regulatory T cells. *J Immunol* 172(10): 5986-5993, 2004.
- 60 Miller A, Lider O, Roberts AB, Sporn MB and Weiner HL: Suppressor T cells generated by oral tolerization to myelin basic protein suppress both *in vitro* and *in vivo* immune responses by the release of transforming growth factor beta after antigen-specific triggering. *Proc Natl Acad Sci USA* 89(1): 421-425, 1992.
- 61 Zhang X, Izikson L, Liu L and Weiner HL: Activation of CD25⁺ CD4⁺ regulatory T cells by oral antigen administration. *J Immunol* 167(8): 4245-4253, 2001.
- 62 Kono K, Kawaida H, Takahashi A *et al*: CD4⁺ CD25⁺ high regulatory T cells increase with tumor stage in patients with gastric and esophageal cancers. *Cancer Immunol Immunother* 55(9): 1064-1071, 2006.
- 63 Qiao M, Thornton AM and Shevach EM: CD4⁺ CD25⁺ [corrected] regulatory T cells render naive CD4⁺ CD25⁻ T cells anergic and suppressive. *Immunology* 120(4): 447-455, 2007.
- 64 Manzotti CN, Tipping H, Perry LC *et al*: Inhibition of human T cell proliferation by CTLA-4 utilizes CD80 and requires CD25⁺ regulatory T cells. *Eur J Immunol* 32(10): 2888-2896, 2002.
- 65 Read S, Malmstrom V and Powrie F: Cytotoxic T lymphocyte-associated antigen 4 plays an essential role in the function of CD25⁺ CD4⁺ regulatory cells that control intestinal inflammation. *J Exp Med* 192(2): 295-302, 2000.
- 66 Takahashi T, Tagami T, Yamazaki S *et al*: Immunologic self-tolerance maintained by CD25⁺ CD4⁺ regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J Exp Med* 192(2): 303-310, 2000.

- 67 Tang Q, Boden EK, Henriksen KJ, Bour-Jordan H, Bi M and Bluestone JA: Distinct roles of CTLA-4 and TGF-beta in CD4+ CD25+ regulatory T cell function. *Eur J Immunol* 34(11): 2996-3005, 2004.
- 68 Nakamura K, Kitani A and Strober W: Cell contact-dependent immunosuppression by CD4(+)CD25(+) regulatory T cells is mediated by cell surface-bound transforming growth factor beta. *J Exp Med* 194(5): 629-644, 2001.
- 69 Ostroukhova M, Qi Z, Oriss TB, Dixon-McCarthy B, Ray P and Ray A: Treg-mediated immunosuppression involves activation of the Notch-HES1 axis by membrane-bound TGF-beta. *J Clin Invest* 116(4): 996-1004, 2006.
- 70 Pae HO, Oh GS, Choi BM, Chae SC and Chung HT: Differential expressions of heme oxygenase-1 gene in CD25- and CD25+ subsets of human CD4+ T cells. *Biochem Biophys Res Commun* 306(3): 701-705, 2003.
- 71 Choi BM, Pae HO, Jeong YR, Kim YM and Chung HT: Critical role of heme oxygenase-1 in Foxp3-mediated immune suppression. *Biochem Biophys Res Commun* 327(4): 1066-1071, 2005.
- 72 Pae HO, Oh GS, Choi BM *et al*: Carbon monoxide produced by heme oxygenase-1 suppresses T cell proliferation *via* inhibition of IL-2 production. *J Immunol* 172(8): 4744-4751, 2004.
- 73 Song R, Mahidhara RS, Zhou Z *et al*: Carbon monoxide inhibits T lymphocyte proliferation *via* caspase-dependent pathway. *J Immunol* 172(2): 1220-1226, 2004.
- 74 El Andaloussi A and Lesniak MS: CD4(+)CD25 (+)FoxP3 (+) T-cell infiltration and heme oxygenase-1 expression correlate with tumor grade in human gliomas. *J Neurooncol* 83(2): 145-152, 2007.
- 75 Mantovani A: Chemokines in neoplastic progression. *Semin Cancer Biol* 14(3): 147-148, 2004.
- 76 Rollins BJ: Chemokines. *Blood* 90(3): 909-928, 1997.
- 77 Mantovani A: The chemokine system: redundancy for robust outputs. *Immunol Today* 20(6): 254-257, 1999.
- 78 Ward SG and Westwick J: Chemokines: understanding their role in T-lymphocyte biology. *Biochem J* 333(Pt 3): 457-470, 1998.
- 79 Rollins BJ: Inflammatory chemokines in cancer growth and progression. *Eur J Cancer* 42(6): 760-767, 2006.
- 80 Gerard C and Rollins BJ: Chemokines and disease. *Nat Immunol* 2(2): 108-115, 2001.
- 81 Robinson SC and Coussens LM: Soluble mediators of inflammation during tumor development. *Adv Cancer Res* 9: 159-187, 2005.
- 82 Balkwill F: The significance of cancer cell expression of the chemokine receptor CXCR4. *Semin Cancer Biol* 14(3): 171-179, 2004.
- 83 Curiel TJ, Coukos G, Zou L *et al*: Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 10(9): 942-949, 2004.
- 84 Miller AM, Lundberg K, Ozenci V *et al*: CD4+ CD25^{high} T cells are enriched in the tumor and peripheral blood of prostate cancer patients. *J Immunol* 177(10): 7398-7405, 2006.
- 85 Jordan JT, Sun W, Hussain SF, Deangulo G, Prabhu SS and Heimberger AB: Preferential migration of regulatory T cells mediated by glioma-secreted chemokines can be blocked with chemotherapy. *Cancer Immunol Immunother* 57(1): 123-131, 2008.
- 86 El Andaloussi A and Lesniak MS: An increase in CD4+ CD25+ FOXP3+ regulatory T cells in tumor-infiltrating lymphocytes of human glioblastoma multiforme. *Neuro Oncol* 8(3): 234-243, 2006.
- 87 Grauer OM, Nierkens S, Bennink E *et al*: CD4+ FoxP3+ regulatory T cells gradually accumulate in gliomas during tumor growth and efficiently suppress antiglioma immune responses *in vivo*. *Int J Cancer* 121(1): 95-105, 2007.
- 88 Du YC, Lin P, Zhang J, Lu YR, Ning QZ and Wang Q: Fusion of CpG-ODN-stimulating dendritic cells with Lewis lung cancer cells can enhance anti-tumor immune responses. *Tissue Antigens* 67(5): 368-376, 2006.
- 89 Friedberg JW, Kim H, McCauley M *et al*: Combination immunotherapy with a CpG oligonucleotide (1018 ISS) and rituximab in patients with non-Hodgkin lymphoma: increased interferon-alpha/beta-inducible gene expression, without significant toxicity. *Blood* 105(2): 489-495, 2005.
- 90 Link BK, Ballas ZK, Weisdorf D *et al*: Oligodeoxynucleotide CpG 7909 delivered as intravenous infusion demonstrates immunologic modulation in patients with previously treated non-Hodgkin lymphoma. *J Immunother* 29(5): 558-568, 2006.
- 91 Lubaroff DM, Karan D, Andrews MP *et al*: Decreased cytotoxic T cell activity generated by co-administration of PSA vaccine and CpG ODN is associated with increased tumor protection in a mouse model of prostate cancer. *Vaccine* 24(35-36): 6155-6162, 2006.
- 92 Meng Y, Carpentier AF, Chen L *et al*: Successful combination of local CpG-ODN and radiotherapy in malignant glioma. *Int J Cancer* 116(6): 992-997, 2005.
- 93 Peng G, Guo Z, Kiniwa Y *et al*: Toll-like receptor 8-mediated reversal of CD4+ regulatory T cell function. *Science* 309(5739): 1380-1384, 2005.

Received December 10, 2007

Revised January 23, 2008

Revised February 6, 2008