

The Role of FOXP3 in Human Cancers

ŁUKASZ SZYLBERG¹, DOMINIKA KARBOWNIK¹ and ANDRZEJ MARSZAŁEK^{1,2}

¹Department of Clinical Pathomorphology, Collegium Medicum in Bydgoszcz,
Nicolaus Copernicus University in Torun, Torun, Poland;

²Department of Oncologic Pathology, Poznan University of Medical Sciences
and Greater Poland Cancer Center, Poznan, Poland

Abstract. *FOXP3 transcription factor can be observed as the main component of the immune system expressed in regulatory T (Treg) cells that regulate hemostasis and self-tolerance. Moreover, the altered expression of FOXP3 was found in autoimmune diseases, benign tumors and carcinomas. Latest reports indicate that the FOXP3 gene mutation can contribute to carcinogenesis, which can be associated with immune response abnormalities. Infiltration of the Treg cells into tumor cells can be associated with prognosis. Understanding the biology of the FOXP3 gene may be crucial in developing new immunotherapeutics.*

Transcription factor forkhead box P3 (FOXP3) is a member of the forkhead family (1). This factor can be observed as the main component of the immune system expressed in regulatory T (Treg) cells (CD4+/CD25+ or CD4+/CD25-) both in cytoplasm and nucleus (2, 3). Aforementioned cells are the immunosuppressive cells (4) that regulate hemostasis and self-tolerance (5). Treg cells are divided into two types: natural Treg (nTreg) and induced Treg (iTreg) (4, 6). The function and development of the Treg cells is regulated by FOXP3 (lymphocyte FOXP3) (1, 4, 5). According to Müller *et al.* and Grimmig *et al.*, FOXP3 is the most specific biomarker of Treg cells (4, 7). but it could be also found in other cells, *e.g.* in B lymphocytes and thymocytes (Table I) (8). Moreover, it was revealed that FOXP3 may be expressed by normal tissues, such as lung, thymus, prostate and breast (2, 9, 10).

The FOXP3 gene includes 11 coding and 3 non-coding exons. (9) The coding exons encode a 47 kDa protein composed of 431 amino acid (8, 10). FOXP3 protein consists of three α helices (H1, H2, H3) and three β strands (S1, S2, S3). It consists of a N-terminal region, responsible for activation and repression, a central zinc finger and a leucine zipper domain (participate in dimerization) (8), as well as a C-terminal region that includes a forkhead domain (FKH, participation in DNA binding) (Figure 1) (5, 10). According to Chen *et al.* and Tavakoli *et al.*, FKH is a critical part of FOXP3 because, as they proved, most of the missense mutations in IPEX syndrome (immune dysregulation, polyendocrinopathy, enteropathy, X- linked) are located in this domain (5, 8).

Human cells synthesize three different isoforms of FOXP3 (Table II) (9, 10). FOXP3FL is a full-length isoform similar to the murine FoxP3. Its molecular weight is 58 kDa. Only this isoform includes exon 2. Exon 2 encodes a domain that inhibits retinoic acid receptor-related orphan receptors (ROR) α and can regulate Th17 differentiation (9, 10). *FOXP3 Δ E2* (54kDa) is devoid of exon 2. The consequence of this phenomenon is higher interleukin (IL)-2 secretion and also proliferation because of T-cell stimulation. *FOXP3 Δ E2 Δ 7* is devoid of the exon 2 and the exon 7. The exon 7 encodes a leucine zipper domain. Lack of this domain eliminates the suppression function of Treg cells (9, 10).

Mutation of FOXP3 Gene

In many studies, authors described the altered expression of FOXP3 that was found in cytoplasm and nucleus (1), in autoimmune diseases, benign tumors and carcinomas (Table III). This altered expression was associated with *FOXP3* gene mutations (1, 10, 13).

FOXP3 gene is positioned at the Xp11.23 (2). It is a critical locus because males have only one chromosome X (genotype XY), while females, despite genotype XX, have only one active allele. In consequence, only one hit to the genome could

This article is freely accessible online.

Correspondence to: Łukasz Szyłberg, MD, Ph.D., Department of Clinical Pathomorphology, A. Jurasz University Hospital No. 1, Skłodowskiej-Curie 9, 85-094 Bydgoszcz, Poland. Tel: +48 525854200, Fax: +48 525854049, e-mail: l.szyłberg@cm.umk.pl

Key Words: FOXP3, cancer, tumor immunity, review.

Table I. The level of the *FOXP3* expression in lymphocytes.

| High level of FOXP3 | Low level of FOXP3 |
|--|--|
| CD4+/CD25+ T cells CD4+/CD25+/CD8– thymocytes | CD4+/CD25– T cells CD4–/CD8+ T cells B cells |

Table II. *FOXP3* isoforms.

| Isoform | Structure |
|-------------------|------------------------------|
| <i>FOXP3FL</i> | A full-length isoform |
| <i>FOXP3ΔE2</i> | Lacks exon 2 |
| <i>FOXP3ΔE2Δ7</i> | Lacks both exon 2 and exon 7 |

transform a normal cell into tumor cell (10, 13). It can be associated with single nucleotide polymorphism or microsatellite polymorphism in the *FOXP3* gene (9). The single nucleotide polymorphism is observed in a promoter, introns and coding region. However, microsatellite polymorphism is observed only in the promoter and in the introns' region (9). Two of the most common *FOXP3* genotype polymorphisms are rs3761549 (C>T) and rs3761548 (C>A) located in the promoter. Previous studies proved that changes in *FOXP3* promoter can be associated with carcinogenesis (2), mainly in a group of patients related to tobacco smoking, asbestosis, pollution and viral infection (14, 15).

Changes in the immune system could be associated with carcinogenesis (similarly as in autoimmune diseases) (Table III) (4, 6, 16). The stimulation of T cells by antigen-presenting cells induces FOXP3 expression and the “acquisition” of suppressor function (9). Such mechanism could play a role in the process when cancer cells escape from the antitumor response (11). The expression of FOXP3 in Treg cells is more intensified in the peripheral blood in patients with cancer comparing to control group (10). Moreover, infiltration of Treg cells into tumor cells can be associated with prognosis. In some neoplasms, such as colorectal cancer (7), melanoma (9), non-small and small lung carcinoma (14), high level of the FOXP3 is associated with poor prognosis. However, in breast (9, 10), prostate (9, 17) and gastric cancer (18, 19), the high level of FOXP3 is associated with good prognosis (7, 10). The differences in prognosis of different tumor types are probably related to different function activation of FOXP3 in given cases. *FOXP3* regulates transcription of target genes (up- and down-regulation) (5). If *FOXP3* binds to the promoter and the 5' regulatory region of CTLA4 (encoding CTLA-4 receptor) or IL2RA (encoding CD25 and IL-2 receptor-α) induces up-regulation of these genes. Such

Table III. Examples of diseases caused by *FOXP3* mutations (1, 2, 8-22).

| Cancers | Other diseases |
|----------------------------|--|
| Breast cancer | Allergic rhinitis |
| Cervix carcinoma | Alopecia areata |
| Colorectal carcinoma | Crohn's disease |
| Gastric cancer | Graves' disease |
| Head and neck carcinoma | Hashimoto's disease |
| Hepatocellular carcinoma | Inflammatory primary biliary cirrhosis |
| Lymphoma | IPEX syndrome (immune dysregulation, polyendocrinopathy, enteropathy, X- linked) |
| Non-small cell lung cancer | Myasthenia gravis |
| Ovary carcinoma | Psoriasis |
| Pancreatic carcinoma | Type I diabetes |
| Prostate cancer | Vitiligo |
| Small cell lung cancer | |
| Thyroid carcinoma | |
| Tongue cancer | |

Table IV. The mechanism of thyroid carcinogenesis.

| High level of FOXP3 | Mechanism | Effect |
|---------------------|--|--------------------------------------|
| | Inhibition of PPARγ | Proliferation and migration of cells |
| | Inhibition of caspase-3 | Blockade of apoptosis |
| | Induced expression of NF-κB p65 and cyclin D | |

interaction causes histone acetylation and transcription of CTLA-4 and CD25 by T cells. On the other hand, if it binds to the promoter of IL-2, IL-7RA or interferon (IFN)γ, this process inhibits acetylation of histones and chromatin remodeling. In consequence, the expression of IL-7R and IL-2 by T cells is blocked (down-regulation) (10).

The FOXP3 Expression in Human Cancers

a) Colorectal Cancer

Patients with colorectal cancer (CRC) have higher levels of T lymphocytes in their blood than healthy control subjects. Moreover, FOXP3 level is higher in colorectal cancer tissues than in normal colorectal tissues (20). Additionally, according to Grimmig *et al.*, the function of the immune reaction depends on the stage of cancer (2, 20). The highest level of CD4/CD25+ is observed in early lesions (UICC I/II) than in advanced lesions (UICC III/IV). High level of FOXP3, IL-10 and transforming growth factor-beta (TGF-β)



Figure 1. The scheme of the structure of the FOXP3 protein.

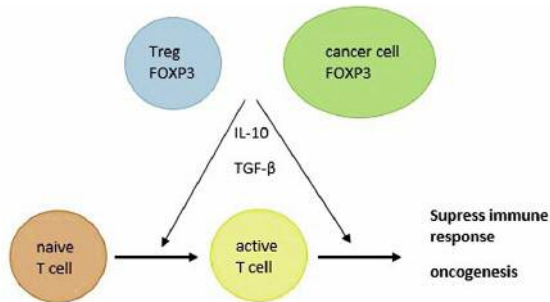


Figure 2. Cytokines stimulatory effect on oncogenesis. FOXP3 from cancer cells and Treg cells inhibit immune response by interacting with phosphodiesterase 3B, cGMP-inhibited PDE3B and the transcription factor NFκB. For this reason, the inflammation is inhibited by specific cytokines, such as IL-10 and TGF-β (5, 7).

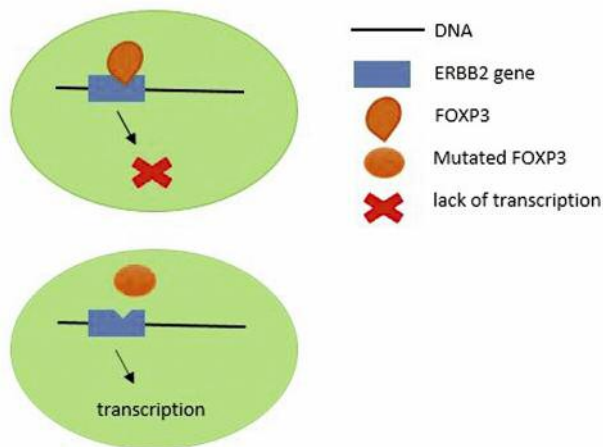


Figure 3. The effect of the mutation in FOXP3 in breast cancer.

are characteristic for malignant tumors (7). This finding can indicate that patients with lymph node metastasis have higher level of FOXP3 (UICC III/IV) than patients without metastasis. FOXP3+ Treg cells inhibit immune reaction resulting in immune escape by the tumor (2, 20). Treg cells (CD4⁺, CD25⁺) inhibit immune reaction. Treg cells and cancer cells produce cytokines, such as IL-10 and TGF-β (Figure 2). Cytokines inhibit the proliferation of naïve T cells (*in vivo*) and active T cells (*in vitro*) (7). As a result, tumor cell lymph node metastasis is promoted (20).

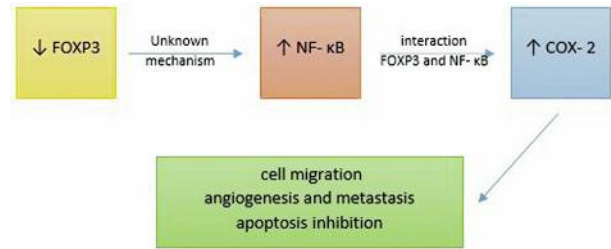


Figure 4. The mechanism of carcinogenesis in gastric cancer (19).

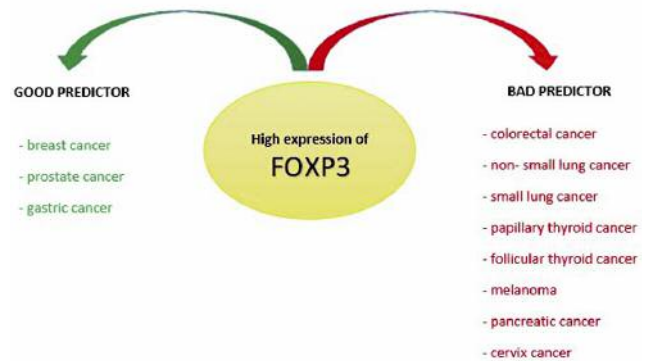


Figure 5. FOXP3 as a prognostic factor.

b) Non- small Lung Cancer

Dimitrakopoulos *et al.* analyzed the FOXP3 level in patients with and without non-small lung cancer (NSCLC). They demonstrated low expression of nuclear FOXP3 in normal bronchial epithelium and normal lymphocytes. In contrast, in NSCLC cells and in tumor-infiltrating lymphocytes, FOXP3 is overexpressed (12). Additionally, He *et al.* described dependence between FOXP3 promoter rs3761548 mutation and NSCLC (14). Moreover, it is possible that, in carcinogenesis, two additional factors participate, prostaglandin E2 (PGE2) and TGF-β. High expression of these factors induces high level of FOXP3 by CD4⁺/CD25⁻ and CD4⁺/CD25⁺ T- cells. Additionally, PGE2 is necessary to tumor progression, angiogenesis and inhibition of apoptosis (12).

Previous studies show that nuclear FOXP3 level in cancer cells does not correlate with age or sex of the patients, histologic type, stage and tumor grade. However, lymphocyte FOXP3 level is correlated with patients' age (it is higher in patients under 64 years old) (12).

c) Papillary and Follicular Thyroid Cancers

According to Chu *et al.*, high level of FOXP3 is associated with thyroid carcinoma. In nodular goiters and follicular adenomas, the level of FOXP3 is low (1, 11, 21). Cunha *et*

al. investigated the association between thyroid cancer and levels of FOXP3. They demonstrated higher level of nuclear FOXP3 in metastatic cancer (1, 21, 22).

Some authors demonstrated that high levels of FOXP3 inhibit expression of PPAR γ (nuclear hormone receptor) and caspase-3 (a key pro-apoptotic molecule) and increase expression of nuclear factor- κ B (NF- κ B) and cyclin D1 (Table IV). The details of this mechanism are unknown. Moreover, high levels of NF- κ B and cyclin D induce cell proliferation and migration in thyroid cancer. Additionally, low levels of PPAR γ and caspase-3 block apoptosis in cancer cells (11). In conclusion, it could be argued that high levels of FOXP3 decrease PPAR γ and indirectly reduce apoptosis of cancer cells.

d) Breast and Prostate Cancer

In mammary tumors, in the process of carcinogenesis, two most important genes participate, *HER-2* and *SKP2*. *HER-2* regulates growth and survival. *SKP2* regulates cell cycle during S and G2 phases by regulated degradation of P27 (9). The high expression of these two genes is related with poor prognosis in patients with mammary cancer (10). On the contrary, high expression of FOXP3 in breast cancer is a good predictor of patients survival (9, 10, 23, 24).

FOXP3 is expressed in normal breast tissues (10). Wild-type *FOXP3*, in normal mammary cells, is bound to and represses *HER-2* and *SKP2* genes. It represses transcription of *HER-2* by binding to the specific locus in the 5' *ERBB2* gene promoter (Figure 3) (9, 23). This indicates that FOXP3 is an important tumor suppressor in breast cancer (9). In contrast, mutated *FOXP3* in cancer cells does not play suppressor function and, as a result, *HER-2* and *SKP2* oncogene expression is missing (10, 23).

The same pathomechanism was found in prostate cancer (10). Studies on prostate cancer revealed chromosomal deletion, somatic mutation and epigenetic silencing of *FOXP3* (9, 17). In those cases, *FOXP3* is unable to repress an oncogene *c-MYC* (10). This causes prostate hyperplasia and prostate cancer. Overexpression of FOXP3 was found in 80% cases of prostate cancer. The correlation between FOXP3 and carcinogenesis was demonstrated by Wang *et al.* They revealed that normal prostate cells proliferated slowly and expressed low levels of *c-MYC*, in contrast to neoplastic cells (25).

e) Melanoma

The high expression of FOXP3 is an unfavorable survival factor in patients with melanoma (9). Low expression of FOXP3 is associated with longer patients' survival (26). FOXP3⁺/CD4⁺ T cells accumulate in the vicinity of tumor cells in higher level than in blood in the same patient. These T cells are similar in phenotype in the blood of healthy patients, patients with melanoma and melanoma tissue. The

only difference is the expression of CTLA-4 by T cells in melanoma tissues that results in increased suppression function (27). Tan *et al.*, however, revealed that FOXP3 in SK-MEL-28 cell line inhibits proliferation, clonogenicity (*in vitro*) and xenograft growth (*in vivo*). Additionally, FOXP3 stimulates expression of genes involved in pigmentation, namely *MLNA*, *TYR* and *TYRP*. Moreover, FOXP3 stimulates apoptosis (28). This was also confirmed by Redpath *et al.* According to their result, *FOXP3* represses the expression of *c-MYC*, which controls cell cycle and apoptosis. *FOXP3* indirectly, by *c-MYC* repression, represses *CDK4* and *CCND2* and activates CDK inhibitors, such as *CDKN1A* and *CDKN2B* (9). *CDKN1A* and *CDKN2B* are specific tumor suppressors (28). Finally, FOXP3 is related to better survival and higher degree of differentiation (9).

f) Gastric Cancer

In gastric cancer, high level of the FOXP3 is a good prognostic factor (18, 19). Ma *et al.* demonstrated that patients with FOXP3-positive tumors have longer survival than patients with FOXP3-negative tumors. Same phenomenon was observed in prostate and breast cancer as FOXP3 inhibits proliferation and migration of cells (11, 18, 19).

One of the key players in carcinogenesis, in gastric cancer, is cyclooxygenase-2 (COX-2) (Figure 4). COX-2 induces angiogenesis, metastasis and causes poor differentiation. Expression of this protein is controlled by NF- κ B levels. The details of this mechanism are unknown. Hao *et al.* demonstrated a relationship between COX-2, NF- κ B and FOXP3. Low level of FOXP3 is associated with high level of NF- κ B and, hence, high COX-2 levels. Some authors suggest that one of the possibilities of carcinogenic activity of this pathway is modification in the *COX-2* locus by interaction between FOXP3 and NF- κ B. Additionally, it is possible that NF- κ B is a COX-2 co-repressor. Interaction between FOXP3 and NF- κ B might reduce binding the NF- κ B to the *COX-2* promoter, thus inducing COX-2 expression (19).

g) Pancreatic Adenocarcinoma (Pancreatic Cancer)

There is no expression of FOXP3 in normal pancreatic duct cells (10, 29). However,, the expression of FOXP3 is an unfavorable survival factor in patients with pancreatic adenocarcinoma (10). This is related to a high level of Treg cells in patients' blood, cancer microenvironment and metastasis in local lymph nodes (29, 30). Treg cells mediate immune escape, thus allowing tumor progression (31). The level of the Treg cells correlates with tumor histologic grade and prognosis. Age, sex and tumor size are irrelevant (30). The mechanism of carcinogenesis in pancreatic cancer is depended on TGF- β 2 (but not TGF- β 1), which is expressed by cancer cells (29, 32). TGF- β 2 secretion results in

increased expression of Treg cells (conversion of naïve T cells into Treg cells (32)), FOXP3 and finally inhibition of the antitumor response (29). This is confirmed by Wang *et al.* in studies of pancreatic adenocarcinoma where the level of the FOXP3 in Treg cells is higher than in healthy people but the level of Th17 in peripheral blood is lower. This disturbed balance between FOXP3 and Th17 is associated with cancer progression (31).

h) Cervix Cancer

High expression of FOXP3 is a bad predictor of survival for patients with cervical cancer. Cervix cancer is preceded by cervical intraepithelial neoplasia/carcinoma *in situ* (CIN I-III/Cis). Mutations in normal cervix cells is caused by human papilloma virus (HPV) (33-35). The main epithelial cell infection by HPV marker is expression of p16INK4a (33, 36). Only a small percentage of cervix cancer is HPV-negative (35).

Luo *et al.* showed high expression of FOXP3 in cervix cancer tissues and metastatic lymph nodes. Additionally, FOXP3 and p16INK4a expression in healthy people is low or does not occur at all (33). This indicates that FOXP3 level is higher in advanced cancer (33, 36, 37). According to Luo *et al.*, high expression of FOXP3 is correlated with FIGO stage, lymph nodes metastasis and tumor size. No correlation was demonstrated between FOXP3 expression and differentiation degree, pathological type and patients' age (33, 37). In contrast, in Chao *et al.*'s study, the level of FOXP3 expression and FIGO stage and lymph node metastasis was no statistically significant (36).

The exact FOXP3 role in cervical epithelium carcinogenesis is unknown. It has been shown that *FOXP3* silencing reduces p16INK4a expression. Probably, *FOXP3* stimulates proliferation, inhibits apoptosis, increases cell invasion and reduces cells in S and G₂ phase of the cell cycle (33).

Final Remarks

There are plenty of evidences that *FOXP3* gene mutation can contribute to carcinogenesis. It can control many other proteins and may play important roles in various biological processes, particularly those of the immunological pathway. FOXP3 is a key factor in pathomechanism in which tumor escapes from immune response. This finding may be useful in developing new drugs as those currently employed, such as denileukin diftitox (ONTAK) (10), LMB-2 (10), cyclophosphamide (10), 5-aza-2'-deoxycytidine (Aza) (38), hydroxamates, cyclic tetrapeptides, aliphatic acids and electrophilic ketones (38) are under intensive research. Moreover, the level of FOXP3 may be used as a prognostic factor (Figure 5). In the nearest future, FOXP3 may prove to be a crucial protein that can bring interesting clinical implications.

References

- 1 Cunha L, Morari E, Nonogaki S, Soares F, Vassallo J and Ward L: FOXP3 expression is associated with aggressiveness in differentiated thyroid carcinomas. *Clinics* 67: 483-488, 2012.
- 2 Jiang L and Ruan L-W: Association between FOXP3 promoter polymorphisms and cancer risk: A meta-analysis. *Oncol Lett* 2795-2799, 2014.
- 3 Magg T, Mannert J, Ellwart JW, Schmid I and Albert MH: Subcellular localization of FOXP3 in human regulatory and nonregulatory T cells. *Eur J Immunol* 42: 1627-1638, 2012.
- 4 Müller S, Poehnert D, Müller JA, Scheumann GWF, Koch M and Lück R: Regulatory T cells in peripheral blood, lymph node, and thyroid tissue in patients with medullary thyroid carcinoma. *World J Surg* 34: 1481-1487, 2010.
- 5 Chen Y, Chen C, Zhang Z, Liu C-C, Johnson ME, Espinoza C a, Edsall LE, Ren B, Zhou XJ, Grant SFA, Wells AD and Chen L: DNA binding by FOXP3 domain-swapped dimer suggests mechanisms of long-range chromosomal interactions. *Nucleic Acids Res* 43: 1-15, 2015.
- 6 Waight JD, Takai S, Marelli B, Qin G, Hance KW, Zhang D, Tighe R, Lan Y, Lo K-M, Sabzevari H, Hofmeister R and Wilson NS: Cutting Edge: Epigenetic Regulation of FOXP3 Defines a Stable Population of CD4+ Regulatory T Cells in Tumors from Mice and Humans. *J Immunol* 194: 878-882, 2015.
- 7 Grimmig T, Kim M, Germer C-T, Gasser M and Waaga-Gasser AM: The role of FOXP3 in disease progression in colorectal cancer patients. *Oncoimmunology* 2: e24521, 2013.
- 8 Nik Tavakoli N, Hambly BD, Sullivan DR and Bao S: Forkhead box protein 3: essential immune regulatory role. *Int J Biochem Cell Biol* 40: 2369-2373, 2008.
- 9 Redpath M, Xu B, van Kempen LC and Spatz A: The dual role of the X-linked FOXP3 gene in human cancers. *Mol Oncol* 5: 156-163, 2011.
- 10 Martin F, Ladoire S, Mignot G, Apetoh L and Ghiringhelli F: Human FOXP3 and cancer. *Oncogene* 29: 4121-4129, 2010.
- 11 Chu R, Liu SYW, Vlantis AC, van Hasselt CA, Ng EKW, Fan MD, Ng SK, Chan ABW, Du J, Wei W, Liu X, Liu Z and Chen GG: Inhibition of FOXP3 in cancer cells induces apoptosis of thyroid cancer cells. *Mol Cell Endocrinol* 399: 228-234, 2015.
- 12 Dimitrakopoulos F-ID, Papadaki H, Antonacopoulou AG, Kottorou A, Gotsis AD, Scopa C, Kalofonos HP and Mouzaki A: Association of FOXP3 expression with non-small cell lung cancer. *Anticancer Res* 31: 1677-1683, 2011.
- 13 Ipex K, Liu R, Li S and Yang W: IPEX Syndrome, FOXP3 and Cancer. *I*: 1-7, 2013.
- 14 He Y-Q, Bo Q, Yong W, Qiu Z-X, Li Y-L and Li W-M: FOXP3 genetic variants and risk of non-small cell lung cancer in the Chinese Han population. *Gene* 531: 422-425, 2013.
- 15 Zhao J, Wang Z, Han J, Qiu X, Pan J and Chen J: Increased frequency of CD4+ CD25+ FOXP3+ cells correlates with the progression of 4-nitroquinoline-1-oxide-induced rat tongue carcinogenesis. *Clin Oral Invest* 25: 1-6, 2013.
- 16 Van Der Vliet HJJ and Nieuwenhuis EE: IPEX as a result of mutations in FOXP3. *Clin Dev Immunol* 2007: 3-8, 2007.
- 17 Teng P, Bateman NW, Darcy KM, Hamilton CA, Maxwell GL, Bakkenist CJ, Conrads TP, Service O, Reed W, Military N and Hospital IF: HHS Public Access. *Gynecol Oncol* 136: 554-561, 2015.

- 18 Ma G-F, Miao Q, Liu Y-M, Gao H, Lian J-J, Wang Y-N, Zeng X-Q, Luo T-C, Ma L-L, Shen Z-B, Sun Y-H and Chen S-Y: High FOXP3 expression in tumour cells predicts better survival in gastric cancer and its role in tumour microenvironment. *Br J Cancer* 110: 1552-1560, 2014.
- 19 Hao Q, Zhang C, Gao Y, Wang S, Li J, Li M, Xue X, Li W, Zhang W and Zhang Y: FOXP3 inhibits NF- κ B activity and hence COX2 expression in gastric cancer cells. *Cell Signal* 26: 564-569, 2014.
- 20 Liu Z, Huang Q, Liu G, Dang L, Chu D, Tao K and Wang W: Presence of FOXP3 + Treg cells is correlated with colorectal cancer progression. 7: 1781-1785, 2014.
- 21 Szyllberg Ł, Bodnar M, Harasymczuk J and Marszałek A: Expression of FOXP3 Protein Plays a Key Role in Thyroid Tumors in Children. *Fetal Pediatr Pathol* 33: 84-91, 2014.
- 22 Hay ID, Gonzalez-Losada T, Reinalda MS, Honetschlager J a., Richards ML and Thompson GB: Long-term outcome in 215 children and adolescents with papillary thyroid cancer treated during 1940 through 2008. *World J Surg* 34: 1192-1202, 2010.
- 23 Zuo T, Wang L, Morrison C, Chang X, Zhang H, Li W, Liu Y, Wang Y, Liu X, Chan MWY, Liu J, Love R, Liu C, Godfrey V, Shen R, Huang TH, Yang T, Park K, Wang C, Zheng P and Liu Y: NIH Public Access 129: 1275-1286, 2008.
- 24 Fiori Lopes L, Losi Guembarovski R, Guembarovski AL, Okuyama Kishima M, Campos CZ, Oda JMM, Ariza CB, De Oliveira KB, Borelli SD and Watanabe MAE: FOXP3 transcription factor: A candidate marker for susceptibility and prognosis in triple negative breast cancer. *Biomed Res Int* 2014: 341654, 2014.
- 25 Wang L, Liu R, Li W, Chen C, Katoh H, Chen GY, McNally B, Lin L, Zhou P, Zuo T, Cooney KA, Liu Y and Zheng P: Somatic Single Hits Inactivate the X-Linked Tumor Suppressor FOXP3 in the Prostate. *Cancer Cell* 16: 336-346, 2009.
- 26 Gerber AL, Müntz A, Schlapbach C, Shafighi M, Kiermeir D, Hüsler R and Hunger RE: High expression of FOXP3 in primary melanoma is associated with tumour progression. *Br J Dermatol* 170: 103-109, 2014.
- 27 Ahmadzadeh M, Felipe-Silva A, Heemskerk B, Powell DJ, Wunderlich JR, Merino MJ and Rosenberg SA: FOXP3 expression accurately defines the population of intratumoral regulatory T cells that selectively accumulate in metastatic melanoma lesions. *Blood* 112: 4953-4960, 2008.
- 28 Tan B, Anaka M, Deb S, Freyer C, Ebert LM, Chueh AC, Al-Obaidi S, Behren A, Jayachandran A, Cebon J, Chen W and Mariadason JM: FOXP3 over-expression inhibits melanoma tumorigenesis *via* effects on proliferation and apoptosis. *Oncotarget* 5: 264-276, 2014.
- 29 Hinz S, Pagerols-Raluy L, Oberg H-H, Ammerpohl O, Grüssel S, Sipos B, Grützmann R, Pilarsky C, Ungefroren H, Saeger H-D, Klöppel G, Kabelitz D and Kalthoff H: FOXP3 expression in pancreatic carcinoma cells as a novel mechanism of immune evasion in cancer. *Cancer Res* 67: 8344-8350, 2007.
- 30 Jiang Y, Du Z, Yang F, Di Y, Li J, Zhou Z, Pillarisetty VG and Fu D: FOXP3+lymphocyte density in pancreatic cancer correlates with lymph node metastasis. *PLoS One* 9: e106741, 2014.
- 31 Wang X, Wang L, Mo Q, Dong Y, Wang G and Jia A: Changes of Th17 / Treg cell and related cytokines in pancreatic cancer patients. *Int J Clin Exp Pathol* 8: 5702-5708, 2015.
- 32 Zeevaart JG, Wang L, Thakur V V, Leung CS, Tirado- J, Bailey CM, Domaol RA, Anderson KS and William L: NIH Public Access 130: 9492-9499, 2009.
- 33 Luo Q, Zhang S, Wei H, Pang X and Zhang H: Roles of FOXP3 in the occurrence and development of cervical cancer. *Int J Clin Exp Pathol* 8: 8717-8730, 2015.
- 34 Santesso N, Mustafa RA, Wiercioch W, Kehar R, Gandhi S, Chen Y, Cheung A, Hopkins J, Khatib R, Ma B, Mustafa AA, Lloyd N, Wu D, Broutet N and Schünemann HJ: Systematic reviews and meta-analyses of benefits and harms of cryotherapy, LEEP, and cold knife conization to treat cervical intraepithelial neoplasia. *Int J Gynecol Obstet* 132: 266-271, 2016.
- 35 Press D: Profile of bevacizumab and its potential in the treatment of cervical cancer. *Onco Targets Ther* 8: 3425-3431, 2015.
- 36 Zeng C, Yao Y, Jie W, Zhang M, Hu X, Zhao Y, Wang S, Yin J and Song Y: Up-regulation of FOXP3 participates in progression of cervical cancer. *Cancer Immunol Immunother* 62: 481-487, 2013.
- 37 Pang X, Zhang Y, Wei H, Zhang J, Luo Q, Huang C and Zhang S: Expression and effects of high-mobility group box 1 in cervical cancer. *Int J Mol Sci* 15: 8699-8712, 2014.
- 38 Lal G and Bromberg JS: Review article Epigenetic mechanisms of regulation of FOXP3 expression. *Blood* 114: 3727-3735, 2009.

Received May 22, 2016

Revised June 13, 2016

Accepted June 14, 2016