Chromosomal Rearrangements in *PLAG1* of Myoepithelial Salivary Gland Tumours

REINHARD E. FRIEDRICH¹, JULIA DILCHER^{2,3}, MICHAEL JAEHNE³ and THOMAS LÖNING⁴

¹Department of Oral and Cranio-Maxillofacial Surgery, ²Department of Pathology, ³Department of Otorhinolaryngology, Eppendorf University Hospital, University of Hamburg, Germany; ⁴Department of Pathology, Albertinen-Hospital, Hamburg, Germany

Abstract. PLAG1 mutations are related to the development of pleomorphic adenomas. A specific aspect of PA is the histological diversity of this entity, containing cells with mesenchymal, epithelial and myoepithelial differentiation. Evidence for myoepithelial cells in PA raises the question whether the very rare entity of pure myoepithelial salivary gland tumours shows chromosomal translocations and rearrangements and whether activation of PLAG1 can be detected. Materials and Methods: Fluorescence in-situ hybridisation (FISH) was established using the DNA-probes PLAG 233, PLAG 234, PLAG 235. The probes were generated from plasmids. Standardization of FISH was achieved in human lymphocytes. Routinely formalin-fixed, paraffin-embedded slices of myoepithelial salivary gland tumours were available for study. In some cases isolated nuclei were investigated. Isolation of the nuclei was performed according to Hedley. Scoring of the FISH was done with a Laser-scanning microscope (spot-counting: fluorescence signals/100 cells/slice). The number of signal variants was determined. All evaluated regions were registered on microphotographs. Results: PLAG1 was only rarely detected. PLAG1 is evidently not involved in the development of myoepithelial tumours. The proportion of 8q12-alterations in myoepithelial tumours was very low. Conclusion: PLAG1 is an insufficient marker to differentiate between benign and malignant myoepithelial tumours.

PLAG1 is the acronym for 'pleomorphic adenoma gene 1', a developmental gene localized on chromosome 8q12 (1,

Correspondence to: Prof. Dr. R.E. Friedrich, Oral and Cranio-Maxillofacial Surgery, Eppendorf University Hospital, University of Hamburg, Martinistr. 52, D-20246 Hamburg, Germany. Tel: +49 40741054885, e-mail: rfriedrich@uke.de

Key Words: PLAG1, myoepihelioma, myoepithelial carcinoma, FISH, chromosomal rearrangements.

2). PLAG1 is expressed during the foetal period (3). PLAG1 is the predominant locus of chromosomal translocations identified in pleomorphic adenoma (PA) of the salivary gland (1-7). Translocation partners are often gene sequences encoding for growth factors (8-11). During the process of chromosomal rearrangement an exchange of promotor sequences takes place resulting in the activation of the PLAG1 gene (12). PLAG1 mutations are related to the development of pleomorphic adenomas. A specific aspect of PA is the histological diversity of this entity, containing cells with mesenchymal, epithelial and myoepithelial differentiation (12-14). Evidence for myoepithelial cells in PA (15) raises the question whether the very rare entity of pure myoepithelial salivary gland shows chromosomal translocations rearrangements and whether activation of PLAG1 can be detected (16-21).

Materials and Methods

Fluorescence *in situ* hybridisation (FISH) was established using the DNA-probes *PLAG 233*, *PLAG 234*, PLAG 235 generated from plasmids (Figures 1 and 2). Standardization of FISH was achieved in human lymphocytes. Routinely formalin-fixed, paraffinembedded slices of myoepithelial salivary gland tumours were pretreated with proteinase K (100 μm/ml, 15 min). In some cases isolated nuclei were investigated. Isolation of the nuclei was performed according to Hedley. Secondary antibodies were labelled with Alexa Fluor (Molecular Probes, Eugene, OR, USA). Scoring of the FISH was done with a Laser-scanning microscope (spot-counting: fluorescence signals/100 cells/ slice). The number of signal variations was determined. All evaluated regions were registered on microphotographs. The technical details and evaluation standards are described elsewhere in detail (22, 23).

The age of patients with myoepithelioma ranged between 17 and 79 years (mean: 51.8 years). Sex distribution of myoepithelioma patients was: male 59%, female 36%, unknown 5%. The age of patients with the diagnosis myoepithelial carcinoma ranged between 20 and 86 years (mean: 59.9 years). Sex distribution of myoepithelial carcinoma patients was: male 39%, female 49%, unknown 12%.

0250-7005/2012 \$2.00+.40

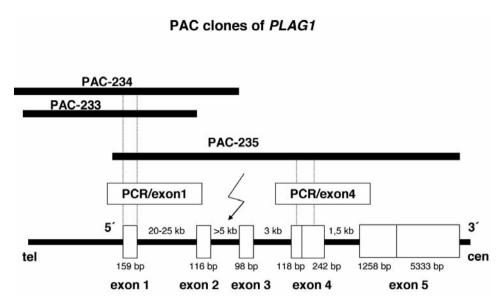


Figure 1. PAC clones of PLAG1 (PAC-233, PAC-234, PAC-235).

Results

Chromosomal translocations of *PLAG1* were detected in myoepithelioma and myoepithelial carcinoma at very low levels. Results for myoepithelioma are visualized in Figure 3.

Myoepithelioma. Twenty myoepithelioma cases were investigated. In three of these cases both isolated nuclei and paraffin slices were investigated by FISH. In 11 cases only paraffin embedded tissue was evaluated and in 6 cases solely the nuclei were studied. About 3.2% of cells showed positive signals. However, in 2.9% the signal constellation excluded translocation and only 0.3% was indicative for *PLAG1* translocations. This finding resulted in 4 myoepithelioma cases in minimum 9% and maximum 80% signal-positive cells. The ratio of cells with evidence for translocation was to low to be effective in the development of this tumour. Further, in these few cases no relation was found to age, sex and localisation of tumour.

Myoepithelial carcinoma. Forty-two patients with myoepithelial carcinoma were investigated. In 16 cases FISH was performed on isolated nuclei and paraffin embedded tissues. In 26 cases results were based on paraffin slices. Tagged nuclei were more frequently detected in paraffin embedded tissues than in extracted nuclei of single cells. This phenomenon is explained in part by differences in the sizes of the isolated nuclei and by the presence of contaminated nuclei of non-tumourous cells. Indeed, non-tumourous cells may even overlap with tumour cells and mask signals. Comparison of FISH analysis in paraffin embedded tissues and isolated nuclei of the same tumour

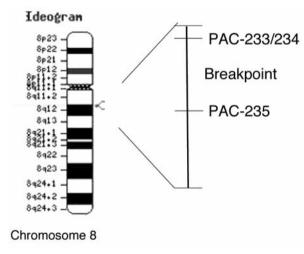


Figure 2. Ideogram of chromosome 8 and localization of PAC clones.

revealed hybridization signals indicative for translocations in 63% (10/16). However, nuclear translocations were not exceeding 27% and at least 4% in these cases. This findings support the hypothesis that the ratio of PLAG1 mutations falls short to effect the tumour development.

PLAG1 translocations were only rarely detected. PLAG1 is obviously not involved in the development of myoepithelial tumours. The proportion of 8q12-alterations in myoepithelial tumours was very low. PLAG1 is an insufficient marker to differentiate between benign and malignant myoepithelial tumours.

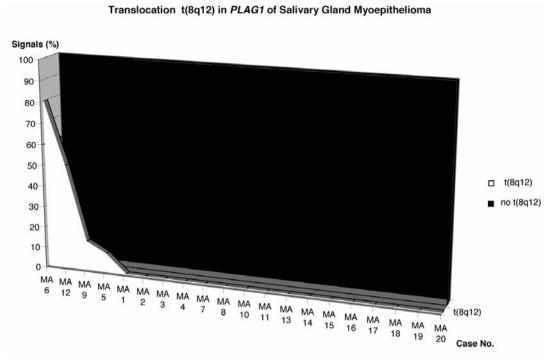


Figure 3. Translocation t(8q12) in salivary gland myoepithelioma: only a small number of patients are affected.

Discussion

Myoepithelial tumours of the salivary glands show some parallel features with pleomorphic adenoma. These morphological affinities and the known pathogenetic association of pleomorphic adenoma and malignant salivary gland tumours with myoepithelial differentiation were the reasons to investigate tumours with prominent myoepithelial differentiation for PLAG1 mutations. Recently, rough calculations derived from comparative genomic hybridization studies had shown genetic alterations on a very low level in both benign and malignant myoepithelial tumours (24), probably due to the employed techniques. On the other hand a low rate of genetic aberrations is not precluding the diagnosis of a malignant myoepithelial tumour (24). Indeed, genetic alterations are not detectable in cases of balanced translocations. In this situation other hybridization techniques should be applied.

Balanced translocations are exceptional types of genomic aberrations in epithelial neoplasms and are detectable in mesenchymal tumours (25). Balanced translocations were revealed in pleomorphic adenomas of salivary glands, including recurrent translocations like t(3; 8) (p21; q12) affecting *PLAG1*. This type of translocations proved to be associated with a certain phenotype (patients of young age, special growth type of tumours and predominat epithelial

cellularity). This translocation causes an exchange of promotors resulting in an increased *PLAG1* expression (5). About 85% of pleomorphic adenoma of salivary glands show chromosomal alterations of the *PLAG 1*, localized on chromosome 8 (8q12) (13). During the last years more and more insights were obtained on the function of *PLAG1*. Activation of *PLAG1* is capable to suppress at least 47 genes and activates more than 10. The most frequent class of antigens activated *via PLAG1* are growth factors (8).

Pleomorphic adenomas may give rise to malignant myoepithelial tumours (26, 27). This transformation could be causally associated with PLAG1. In order to consider the hypothesis that chromosomal alterations may be dependent on the type of myoepithelial salivary gland tumour, 20 myoepithelioma and 42 myoepithelial carcinoma were investigated for PLAG1 with FISH. The number of counted signals was very low both in benign and malignant lesions. A translocation possibly related to t(8q12) was revealed in 0.3% of all myoepithelioma. In malignant myoepithelial tumours the constellation of signals were in 0.2% of cases consistent with a possible translocation t(8q12). Concerning the density of the signals both tumour entities showed no relevant t(8q12) translocations. There were no differences concerning the chromosomal alterations between both entities. It is likely that PLAG1 alterations are specific for pleomorphic adenoma and carcinomas derived thereof (13). Current studies are intended to investigate the role of other proteins/factors in the pathogenesis of myoepithelial salivary gland tumours (1, 28-36).

References

- 1 Abdollahi A, Roberts D and Godwin AK: Identification of a zinc-finger gene at 6q25: a chromosomal region implicated in development of many solid tumors. Oncogene 14: 1973-1979, 1997
- 2 Kas K, Voz ML, Roijer E, Astrom AK, Meyen E, Stenman G and van de Ven WJ: Promoter swapping between the genes for a novel zinc finger protein and β-catenin in pleomorphic adenomas with t(3;8)(p21;q12) translocations. Nature Genetics 15: 170-174, 1997.
- 3 Abdollahi A: LOT1 (ZAC1/PLAG1) and its family members: mechanisms and functions. J Cell Physiol 210: 16-25, 2007.
- 4 Asp J, Persson F, Kost–Alimova M and Stenman G: *CHCHD7-PLAG1* and *TCEA-PLAG1* gene fusions resulting from cryptic, intrachromosomal 8q12 rearrangements in pleomorphic salivary gland adenomas. Genes Chromo Cancer 45: 820-829, 2006.
- 5 Bullerdiek J, Wobst G, Meyer-Bolte K, Chilla R, Haubrich J, Thode B and Bartnitzke S: Cytogenetic subtyping of 220 salivary gland pleomorphic adenomas: correlation to occurrence, histological subtype and *in vitro* cellular behaviour. Cancer Genet Cytogenet 65: 27-31, 1993.
- 6 Debiec-Rychter M, Van Valckenborgh I, Van den Broeck C, Hagemeijer A, van de Ven WJ, Kas K, van Damme B and Voz ML: Histologic localization of *PLAG1* (pleomorphic adenoma gene 1) in pleomorphic adenoma of the salivary gland: cytogenetic evidence of common origin of phenotypically diverse cells. Lab Invest 81: 1289-1297, 2001.
- 7 Roijer E, Kas K, Klawitz I, Bullerdiek J, van de Ven W and Stenman G: Identification of a yeast artificial chromosome spanning the 8q12 breakpoint in pleomorphic adenomas with t(3;8) (p21;q12). Genes Chrom Cancer *17*: 166-171, 1996.
- 8 Voz ML, Mathys J, Hensen K, Pendeville H, Van Valckenborgh I, Van Huffel C, Chavez M, Van Damme B, De Moor B, Moreau Y and van de Ven WJ: Microarray screening for target genes of the proto-oncogene PLAG1. Oncogene 23: 179-191, 2004.
- 9 Voz ML, Astrom AK, Kas K, Mark J, Stenman G and van de Ven WJ: The recurrent translocation t(5;8)(p13;q12) in pleomorphic adenomas results in upregulation of *PLAG1* gene expression of the *LIFR* promoter. Oncogene 16: 1409-1416, 1998.
- 10 Voz ML, van de Ven WJ and Kas K: First insights into the molecular basis of pleomorphic adenomas of the salivary glands. Adv Dent Res 14: 81-83, 2000.
- 11 Voz ML, Agten NS, van de Ven WJ and Kas K: *PLAG1*, the main translocation target in pleomorphic adenoma of the salivary glands, is a positive regulator of *IGF-II*. Cancer Res *60*: 106-113, 2000.
- 12 Astrom AK, Voz ML, Kas K, Roijer E, Wedell B, Mandahl N, van de Ven W and Mark J: Conserved mechanism of *PLAG1*-activation in salivary gland tumors with and without chromosome 8q12 abnormalities: Identification of SII as a new fusion partner gene. Cancer Res *59*: 918-923, 1999.

- 13 Martins C, Fonseca I, Roque L, Pereira T, Ribeiro C, Bullerdiek J and Soares J: *PLAG1* gene alterations in salivary gland pleomorphic adenoma and carcinoma ex-pleomorphic adenoma: a combined study using chromosome banding, *in situ* hybridization and immunocytochemistry. Mod Pathol 18: 1048-55, 2005.
- 14 Rohen C, Rogalla P, Meyer-Bolte K, Bartnitzke S, Chilla R and Bullerdiek J: Pleomorphic adenomas of the salivary glands: absence of *HMGIY* rearrangements. Cancer Genet Cytogenet *111*: 178-181, 1999.
- 15 Ellis GL and Auclair PL: Atlas of Tumor Pathology: Tumors of the Salivary Glands, 3rd series. Armed Forces Institute of Pathology, Washington, DC, 1996
- 16 Simpson RH, Jones H and Beasley P: Benign myoepithelioma of the salivary glands: a true entity? Histopathol 27: 1-9, 1995.
- 17 Dardick I, Thomas MJ and van Nostrand AWP: Myoepithelioma new concepts of histology and classifications: a light and electron microscopy study. Ultrastructural Pathology 13: 187-224, 1989.
- 18 El-Naggar AK, Lovell M, Callender DL, Ordonez NG and Killary AM: Cytogenetic analysis of a primary salivary gland myoepithelioma. Cancer Genet Cytogenet 113: 49-53, 1999.
- 19 Savera AT, Sloman A, Huvos AG and Klimstra DS: Myoepithelial carcinoma of the salivary glands. Am J Surg Pathol 24: 761-774, 2000.
- 20 Sciubba JJ and Brannon RB: Myoepithelioma of salivary glands: report of 23 cases. Cancer 49: 562-572, 1982.
- 21 Skalova A, Michal M, Ryska A, Simpson RH, Kinkor Z, Walter J and Leivo I: Oncocytic myoepithelioma and pleomorphic adenoma of the salivary glands. Virchows Arch 434: 537-546, 1999.
- 22 Murga Penas EM, Hinz K, Röser K, Copie-Bergman C, Wlodarska I, Marynen P, Hagemeijer A, Gaulard P, Löning T, Hossfeld DK and Dierlamm J: Translocations t(11;18)(q21;q21) and t(14;18)(q32;q21) are the main chromosomal abnormalities involving MLT/MALT1 in MALT lymphomas. Leukemia *17*: 2225-2229, 2003.
- 23 Rigby PW, Dieckmann M, Rhodes C and Berg P: Labeling desoxyribonucleic acid to high specific activity in vitro by nick translation with DNA polymerase I. J Mol Biol 113: 237-251, 1977.
- 24 Hungermann D: Cytogenetische und immunphänotypische Charakterisierung von Myoepitheliomen und myoepithelialen Karzinomen der Kopfspeicheldrüsen, Dissertation, University of Münster, 2001.
- 25 Hungermann D, Roeser K, Buerger H, Löning T and Herbst H: Relative paucity of gross genetic alterations in myoepitheliomas and myoepithelial carcinomas of salivary glands. J Pathol 198: 487-494, 2002.
- 26 Friedrich RE, Li L, Knop J, Giese M and Schmelzle R: Pleomorphic adenoma of the salivary glands: analysis of 94 patients. Anticancer Res 25: 1703-1705, 2005.
- 27 Vékony H, Röser K, Löning T, Ylstra B, Meijer GA, van Wieringen WN, van de Wiel MA, Carvalho B, Kok K, Leemans CR, van der Waal I and Bloemena E: Copy number gain at 8q12.1-q22.1 is associated with a malignant tumor phenotype in salivary gland myoepitheliomas. Genes Chromo Cancer 48: 202-212, 2009.
- 28 Wanschura S, Dal Cin P, Kazmierczak B, Bartnitzke S, Van den Berghe H and Bullerdiek J: Hidden paracentric inversions of chromosome arm 12q affecting the HMGIC gene. Genes Chromo Cancer 18: 322-323, 1997.

- 29 Weber A, Langhanki L, Schütz A, Gerstner A, Bootz F, Wittekind C and Tannapfel A: Expression profiles of p53, p63 and p73 in benign salivary gland tumors. Virchows Arch 441: 428-436, 2002.
- 30 Bracken AP, Kleine-Kohlbrecher D, Dietrich N, Pasini D, Gargiulo G, Beekman C, Theilgaard-Mönch K, Minucci S, Porse BT, Marine JC, Hansen KH and Helin K: The polycomb group proteins bind throughout the INK4A-ARF locus and are disassociated in senescent cells. Genes Dev 21: 525-530, 2007.
- 31 Bracken AP, Kleine-Kohlbrecher D, Dietrich N, Pasini D, Gargiulo G, Beekman C, Theilgaard-Mönch K, Minucci S, Porse BT, Marine JC, Hansen KH and Helin K: The polycomb group proteins bind throughout the INK4A-ARF locus and are disassociated in senescent cells. Genes Dev 21: 525-530, 2007.
- 32 Bracken AP, Pasini D, Capra M, Prosperini E, Colli E and Helin K: EZH2 is downstream of the pRB–E2F pathway, essential for proliferation and amplified in cancer. EMBO J 22: 5323-5335, 2003.
- 33 Barbareschi M, Pecciarini L, Cangi MG, Macri E, Rizzo A, Viale G and Doglioni C: p63, a p53 homologue, is a selective nuclear marker of myoepithelial cells of the human breast. Am J Surg Pathol 25: 1054-1060, 2001.

- 34 Jacobs JJ, Kieboom K, Marino S, dePinho RA and van Lohuizen M: The oncogene and polycomb–group gene bmi–1 regulates cell proliferation and senescence through the ink4a–locus. Nature 397: 164-168, 1999.
- 35 Jacobs JJ, Scheijen B, Voncken JW, Kieboom K, Bems A and van Lohuizen M: Bmi-1 collaborates with c-myc in tumorigenesis by inhibiting c-myc-induced apoptosis *via* INK4a/ARF. Genes Dev *13*: 2678-2690, 1999.
- 36 Satijn DP and Otte AP: Polycomb group protein complexes: do different complexes regulate distinct target genes? Biochim Biophys Acta 1447: 1-16, 1999.

Received April 18, 2012 Revised April 24, 2012 Accepted April 24, 2012