Sulindac Sulfone Induces a Decrease of β-Catenin in HNSCC

ALEXANDER SAUTER¹, HANNAH SOULSBY², KARL HORMANN¹ and RAMIN NAIM³

¹Department of Otolaryngology, Head and Neck Surgery Ruprecht-Karls University Heidelberg, Faculty of Medicine Mannheim, Germany; ²Warwick Medical School, University of Warwick, Coventry CV4 7AL, U.K.; ³Department of Otolaryngology, Head and Neck Surgery, Dr. Horst Schmidt Kliniken, Academic Teaching Hospital of the University of Mainz, Wiesbaden, Germany

Abstract. Background: The most common neoplasm arising in the upper gastrointestinal tract is head and neck squamous cell carcinoma (HNSCC). This is an aggressive epithelial malignancy. Many growth factors and cytokines have been discovered that are responsible for the growth and formation of tumours. Among these factors, β -catenin is considered to be the most important for reducing cell-cell adhesions in malignant tissue. The degradation of β -catenin triggers apoptosis by different routes. Sulindac sulfone has been shown to induce apoptosis in several different tumours. In the present study, we surveyed the concentration of B-catenin in an HNSCC line after incubation with different concentrations of sulindac sulfone. Materials and Methods: Immunohistochemical and Western blot analyses were performed after treatment of the UMSCC 11A cell line with different concentrations of sulindac sulfone (100, 200, 400, 600 and 800 μMol) for 48 hours. Results: At 100 μMol of sulindac sulfone, a decrease in β -catenin concentration of 5% was observed; increasing concentrations of sulindac sulfone resulted in >70% reduction in secreted β -catenin. Thus in conclusion, incubation with sulindac sulfone seemed to stop proliferation. With respect to the controls, there was no greater reduction in total protein. Conclusion: In this study, sulindac sulfone reduced levels of secreted β -catenin in the HNSCC cell line UM-SCC 11A after 48 hours of incubation. It is presumed that reduction of cell-cell adhesion, which is predominately affected by β -catenin, is an essential step in the progression from localized malignancy to stromal and vascular invasion and ultimately metastatic disease. The reduction in the level of mural expression of β -catenin has been associated with loss of differentiation in

Correspondence to: Priv.-Doz. Dr. med. Alexander Sauter, Universitaets-HNO-Klinik, Theodor-Kutzer-Ufer, D-68135 Mannheim, Germany. e-mail: alexander.sauter@gmail.com

Key Words: HNSCC, UM-SCC, β -catenin, sulindac sulfone, cell line, cell culture, apoptosis.

laryngeal carcinomas. Thus, prevention of intracellular β -catenin accumulation is regarded as an attractive target for chemopreventive agents.

The formation of tumors is accompanied by genetic changes. Tumour-forming cells appear to be less genetically stable than normal cells (1). The malignant tumor cells are characterized by their ability to cross tissue barriers and to invade local tissue. Epithelial tumour cells migrate by overcoming adherent cell cell contact and by creating a pathway through basement membrane and stroma (2). In recent decades, many growth factors and cytokines have been discovered that are responsible for the growth and formation of different tumors. Among these factors, β-catenin is considered to be the most important factor for reducing cell cell adhesion in malignant tissue (3). β-Catenin is a multifunctional protein. Appearing as a free cytosolic protein, it serves as a component of the cytoskeleton in a multi-protein complex at the plasma membrane where it binds E-cadherin to the actin cytoskeleton (2). The accumulation of β-catenin results in increased transcription of Tcf/Lef-regulated genes, including cyclin D1 and c-Myc, and thus provides growth advantage to many tumours (4, 5). However, in normal tissue, cytosolic β-catenin is rapidly phosphorylated at multiple serine and theronine sites near the NH₂-terminal region of the protein by a multi-protein complex including adenomatosis polyposis coli (APC), glycogen synthase kinase 3- β (GSK-3 β) and axin/conductin (6). Aberle et al. (7) reported that after obligatory phosphorylation, βcatenin is targeted for ubiquitination and subsequent degradation by proteosomes, and apoptosis of the cell is initiated. The second physiological way of initiating β-catenindependant apoptosis of the cell involves the activation of the caspase family of proteases. Common cleavage sites for caspase-3, -6 and -8 have been recognized in the β -catenin protein (8). Sulindac sulfone is an active metabolite of the prodrug sulindac. A non steroidal anti-inflammatory drug, it was initially developed to prevent polyp formation and promote regression of existing polyps in patients with familial adenomatous polyposis (FAP). Recently, sulindac sulfone has

0250-7005/2010 \$2.00+.40

been shown to induce apoptosis selectively in precancerous and cancerous tissue when applied to models of haematological malginancies and solid cancer such as colon, prostate, bladder, mammary and lung cancer (9, 10). Intracellularly, sulindac sulfone inhibits the cyclic guanosine monophosphate (cGMP) phosphodiesterase (PDE), which results in sustained increase of cGMP and activation of protein kinase G (PKG) (Figure 1). Interestingly, β -catenin is a substrate of PKG, thus, phosphorylation of β -catenin initiates the degradation of β -catenin via, proteosomes. The degradation of β -catenin thus triggers apoptosis by different routes.

In the present study, we surveyed the concentration of β -catenin in an HNSCC line after incubation with different concentrations of sulindae sulfone.

Materials and Methods

Cell culture. The UMSCC cell lines are well-described human HNSCC cell lines obtained from Dr. T. E. Carey (The University of Michigan, Ann Arbor, Michigan, USA). We used the HNSCC cell line UMSCC 11A. They are gained from laryngeal carcinomas and are capable of growth in conventional semi-solid media (11). Cell cultures were carried out in Falcon petri-dishes at 37°C in a 5% CO₂ fully humidified atmosphere using Dulbecco's modified minimum essential medium (DMEM) (Fisher Scientific Co., Pittsburgh, PA, USA) supplemented with 10% fetal calf serum (FCS) and antibiotics [Life Technologies, Inc. (Gibco BRL), Gainthersburg, MD, USA]. Sulindac sulfone was dissolved in dimethylsulfide (DMS) and added to the cell culture medium.

β-Catenin-ELISA principle. Cell culture supernatants were collected in sterile test tubes and stored at -20°C until used. Secreted βcatenin was measured in the supernatant of the cell lines using an ELISA technique (R&D Systems, Wiesbaden, Germany). The system uses a solid-phase monoclonal antibody and an enzymelinked polyclonal antibody raised against β-catenin. The specificity of anti-human β- catenin antibodies used in the ELISA kit was examined by sodium dodecylsulphate polyacrylamide gel electrophoresis (SDS-PAGE) followed by Western blotting. A volume of 100 µl of supernatant were used for each ELISA assay. After 8, 24, and 48 hours of incubation with 0, 100, 200, 400, 600 and 800 μMol sulindac sulfone, the expression of the β-catenin protein in the supernatants of the treated and untreated culture cells was analyzed. All analyses and calibrations were carried out in duplicate. The calibrations on each microtitre plate included βcatenin standards provided in the kit. Optical density was determined using a microplate reader at a wavelength of 450 nm. Wavelength correction was set to 540 nm and concentrations were reported in µg/ml. Controls were performed using DMS without sulindac sulfone during incubation.

Characterisation of the cell lines (immunohistochemistry). Immunohistochemical analysis was performed using a monoclonal mouse anti- human antibody directed against β -catenin (β -catenin: C19220; Transduction Laboratories, Lexington, KY, USA). Immunostaining was performed using the alkaline phosphatase-antialkaline phosphatase method (APAAP). Therefore the HNSCC cells were cultured on glass coverslips overnight before immunhisto-

chemistry. When confluent, cells underwent a fixation with acetone and alcohol (2:1), were washed with PBS and received a microwave pre-treatment, which required boiling for 15 min at 600 W using citrate buffer for β-catenin. The following steps were performed by an automated staining system, DAKO TechMate 500 (DAKO, Hamburg, Germany). Sections were incubated with the primary antibody solution for 25 min at room temperature, using a working dilution of the antibody 1:300 for β-catenin. Slides were rinsed once in buffer (Buffer Kit, DAKO). Immunoreaction was demonstrated with the DAKO ChemMate Detection Kit (APAAP, Mouse, Code No. K 5000; DAKO) according to the specifications of the manufacturer. Sections were incubated with the chromogen alkaline phosphatase substrate (Neufuchsin; DAKO) for 20 min at room temperature. Finally, sections were counterstained by Mayer'shematoxylin for 3 min, dehydrated in graded ethanol, and a coverslip applied. Negative controls used all reagents except the primary antibody. The results of the immunohistochemically obtained rates of expression were analysed semi-quantitatively. The stain intensity was noted as follows: strong reactivity >80% of the epithelium cells were positive; moderate reactivity 50-80% reactive; weak reactivity <50% reactive; and no positive cells.

Results

Incubation up to 48 hours with sulindac sulfone. After 48 hours of incubation with 100 μ Mol sulindac sulfone, the average level of β -catenin was 0.00166 μ g/ml. The level of β -catenin after 8 hours was 0.00175 μ g/ml. This is a decrease of 5% in growth rate. At 200 μ Mol of sulindac sulfone, average β -catenin was 0.00193 μ g/ml after 8 hours and after 48 hours β -catenin had dropped to 0.00173 μ g/ml, a 12% decrease in growth rate. At 400 μ Mol of sulindac sulfone, a decrease of 21% was observed (0.00149 μ g/ml); at 600 μ Mol the decrease was 44% (0.00079 μ g/ml) and incubation with 800 μ Mol resulted in a 73% reduction (0.00030 μ g/ml). Both control cell cultures, 0 μ Mol sulindac sulfone and DMS with 0 μ Mol sulindac sulfone, revealed an increase of β -catenin of 9% and 4%, respectively (Figure 2 and 3).

The total protein concentration also decreased after incubation with sulindac sulfone. Incubation with sulindac sulfone seemed to stop proliferation; however, as compared to the controls, there was no greater reduction of the total protein. Overall, the decrease of the total protein was approximately 5 to 11% after 48 hours (p>0.05) (Figure 4).

Immunohistochemistry. At increasing concentrations of sulindac sulfone, intracellular β -catenin immunostaining decreased (Table I). The controls showed highest reactivity for intracellular β -catenin (Figure 5 A and B).

Discussion

In this study, sulindac sulfone decreased levels of β -catenin in the HNSCC cell line UM-SCC 11A. As a metabolite of sulindac, sulindac sulfone represents a new class of pro-

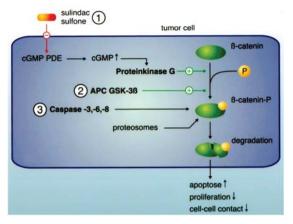


Figure 1. Different mechanisms of β -catenin-dependent induction of apoptosis.

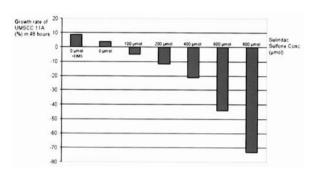


Figure 2. Growth rate of the cells after incubation with different levels of sulindac sulfone.

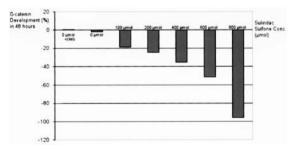


Figure 3. Concentration of secreted β -catenin after incubation with different levels of sulindac sulfone.

apoptotic drugs, the so-called selective apoptotic antineoplastic drugs (SAAND). Piazza $et\ al.$ (9) reported that the mechanism of action of SAANDs is completely independent of cyclooxygenase (COX)-1 or COX-2 inhibition and p53 pathways. They showed that in proliferating HT-29 cells, sulindac sulfone was able to block G_1 cell cycle progression but also induced apoptosis.

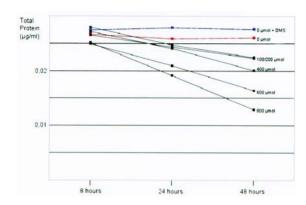


Figure 4. Total protein concentration after incubation of cells with different levels of sulindac sulfone.

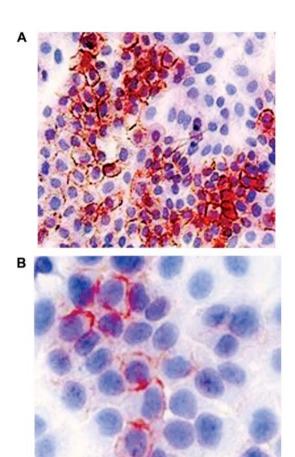


Figure 5. Immunohistochemical reactivity for cells treated with different levels of sulindac sulfone. A, 0 μ Mol (×10); B, 600 μ Mol (×20).

However, in non- proliferating cells, sulindac sulfone did not reduce cell growth. Piazza *et al.* (9) concluded that sulindac sulfone resulted in growth inhibition and apoptosis only in cells without cell cycle arrest. Lim *et al.* (12)

Table I. Immunohistochemical score (IHS): Percentage grading of the immunostaining of β -catenin in UMSCC cell line after treatment with different concentrations of sulindac sulfone (n=100%).

Sulindac sulfone (μMol)						
IHS	0	100	200	400	600	800
0	7	5	17	20	14	17
I	16	36	22	33	35	52
II	14	28	61	47	51	31
III	63	31	0	0	0	0

0: No positive cells; I: weak reactivity (<50% positive cells); II: moderate (50%-80% positive cells); III: strong reactivity (>80% positive cells).

showed sulindac sulfone induced inhibition of growth in human prostate cancer. Soriano et al. (13) reported chemopreventive effects in human lung cancer; several other types of solid tumor cells including bladder, cervical and hepatic were also reported to show apoptosis after treatment with sulindac sulfone (9, 14, 15). The authors also reported that the targeted enzyme cGMP PDE was overexpressed in these tumors. Franchi et al. (16) showed that in HNSCC, cGMP PDE was significantly increased. The present study showed a small reduction in total protein levels (p>0.05), which indirectly served to reflect the number of all incubated cells. This result showed that the proliferation rate of the treated UM-SCC 11A cells is possibly affected by increased cell death. Thus, as a therapeutic agent, sulindac sulfone inhibited cell growth in HNSCC. It is presumed that a reduction of cell-cell adhesion, which is predominately affected by β -catenin, is an essential step in the progression from localized malignancy to stromal and vascular invasion, and ultimately metastatic disease (17). Cytoplasmic accumulation of βcatenin is considered as an oncogene factor (3). Lopez-Gonzalez et al. reported that the reduction in the level of mural expression of β-catenin was associated with loss of differentiation in laryngeal carcinomas (18). Hirvikoski et al. reported that the percentage of tumours showing increased cytoplasmic reactivity for β-catenin increased with histological dedifferentiation in laryngeal cancer (19). Increased cytoplasmic β-catenin seems to be a pivotal feature of malignancy. Untreated cultured UM-SCC 11A cells showed increased reactivity against cytoplasmic βcatenin, however membranous reactivity was relatively weak (Figure 5A). After incubation with sulindac sulfone, cytosolic β-catenin expression significantly decreased. In previous studies, diminished cytoplasmic β-catenin concentration seemed to reflect stabilized proliferation and reduced invasiveness of cells. Accumulated cytosolic βcatenin translocates to the nucleus, where it regulates oncogenic transcription factors (20). Thus, prevention of βcatenin accumulation is regarded as an attractive mode of action for chemopreventive agents. In this study, increased levels of sulindac sulfone resulted in greater reductions of secreted β-catenin in the HNSCC cell line. It is possible that by treatment with sulindac sulfone, PKG is activated in these cells (Figure 1). Thompson et al. (21) reported that PKG can phosphorylate β-catenin in vitro. However, it remains possible that other protein kinases could also phosphorylate additional sites of the β-catenin protein, leading to ubiquitation and protozomal degradation. Fukuda et al. (22) reported that inhibition of caspase activity did not return β-catenin protein levels to control levels, indicating that caspases are not completely responsible for loss of β -catenin protein expression by sulindac sulfone. Further research is necessary to reveal the mechanisms of sulindac sulfone-induced apoptosis in different HNSCC cell lines. Regarding the clinical application of sulindac sulfone, it is difficult to directly compare our short-term, high-dose in vitro results with longer term human and animal studies: In this in vitro study the concentration of sulindac sulfone used was markedly higher than has been measured in vivo in the plasma of humans, which Davies et al. (23) reported to be approximately 50 µMol in human plasma.

Acknowledgements

We would like to thank Petra Prohaska for her excellent technical assistance.

References

- 1 Loeb LA: Mutator phenotype may be required for multistage carcinogenesis. Cancer Res 51(12): 3075-3079, 1991.
- 2 Aberle H, Schwartz H and Kemler R: Cadherin-catenin complex: protein interactions and their implications for cadherin function. J Cell Biochem 61(4): 514-523, 1996.
- 3 Rubinfeld B, Robbins P, El Gamil M, Albert I, Porfiri E and Polakis P: Stabilization of beta-catenin by genetic defects in melanoma cell lines. Science *275(5307)*: 1790-1792, 1997.
- 4 Shtutman M, Zhurinsky J, Simcha I, Albanese C, D'Amico M, Pestell R and Ben Ze'ev A: The cyclin D1 gene is a target of the beta-catenin/LEF-1 pathway. Proc Natl Acad Sci USA 96(10): 5522-5527, 1999.
- 5 He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B and Kinzler KW: Identification of c-MYC as a target of the APC pathway. Science 281(5382): 1509-1512, 1998.
- 6 Polakis P, Hart M and Rubinfeld B: Defects in the regulation of beta-catenin in colorectal cancer Adv Exp Med Biol pp. 47023-47032, 1999.
- 7 Aberle H, Bauer A, Stappert J, Kispert A and Kemler R: Betacatenin is a target for the ubiquitin-proteasome pathway EMBO J 16(13): 3797-3804, 1997.
- 8 Herren B, Levkau B, Raines EW and Ross R: Cleavage of betacatenin and plakoglobin and shedding of VE-cadherin during endothelial apoptosis: evidence for a role for caspases and metalloproteinases. Mol Biol Cell 9(6): 1589-1601, 1998.

- 9 Piazza GA, Rahm AK, Finn TS, Fryer BH, Li H, Stoumen AL, Pamukcu R and Ahnen DJ: Apoptosis primarily accounts for the growth-inhibitory properties of sulindac metabolites and involves a mechanism that is independent of cyclooxygenase inhibition, cell cycle arrest, and p53 induction. Cancer Res 57(12): 2452-2459, 1997.
- 10 Goluboff ET: Exisulind, a selective apoptotic antineoplastic drug. Expert Opin Investig Drugs 10(10): 1875-1882, 2001.
- 11 Grenman R, Burk D, Virolainen E, Buick RN, Church J, Schwartz DR and Carey TE: Clonogenic cell assay for anchorage-dependent squamous carcinoma cell lines using limiting dilution. Int J Cancer 44(1): 131-136, 1989.
- 12 Lim JT, Piazza GA, Han EK, Delohery TM, Li H, Finn TS, Buttyan R, Yamamoto H, Sperl GJ, Brendel K, Gross PH, Pamukcu R and Weinstein IB: Sulindac derivatives inhibit growth and induce apoptosis in human prostate cancer cell lines. Biochem Pharmacol 58(7): 1097-1107, 1999.
- 13 Soriano AF, Helfrich B, Chan DC, Heasley LE, Bunn PA Jr and Chou TC: Synergistic effects of new chemopreventive agents and conventional cytotoxic agents against human lung cancer cell lines. Cancer Res 59(24): 6178-6184, 1999.
- 14 Han EK, Arber N, Yamamoto H, Lim JT, Delohery T, Pamukcu R, Piazza GA, Xing WQ and Weinstein IB: Effects of sulindac and its metabolites on growth and apoptosis in human mammary epithelial and breast carcinoma cell lines. Breast Cancer Res Treat 48(3): 195-203, 1998
- 15 Rahman MA, Dhar DK, Masunaga R, Yamanoi A, Kohno H and Nagasue N: Sulindac and exisulind exhibit a significant antiproliferative effect and induce apoptosis in human hepatocellular carcinoma cell lines. Cancer Res 60(8): 2085-2089, 2000.
- 16 Franchi A, Santucci M, Masini E, Sardi I, Paglierani M and Gallo O: Expression of matrix metalloproteinase 1, matrix metalloproteinase 2, and matrix metalloproteinase 9 in carcinoma of the head and neck. Cancer 95(9): 1902-1910, 2002.

- 17 Wijnhoven BP, Dinjens WN and Pignatelli M: E-Cadherincatenin cell-cell adhesion complex and human cancer. Br J Surg 87(8): 992-1005, 2000.
- 18 Lopez-Gonzalez JS, Cristerna-Sanchez L, Vazquez-Manriquez ME, Jimenez-Orci G and Aguilar-Cazares D: Localization and level of expression of beta-catenin in human laryngeal squamous cell carcinoma. Otolaryngol Head Neck Surg 130(1): 89-93, 2004
- 19 Hirvikoski P, Kumpulainen EJ, Virtaniemi JA, Helin HJ, Rantala I, Johansson RT, Juhola M and Kosma VM: Cytoplasmic accumulation of alpha-catenin is associated with aggressive features in laryngeal squamous-cell carcinoma. Int J Cancer 79(5): 546-550, 1998.
- 20 Behrens J, von Kries JP, Kuhl M, Bruhn L, Wedlich D, Grosschedl R and Birchmeier W: Functional interaction of betacatenin with the transcription factor LEF-1. Nature 382(6592): 638-642, 1996.
- 21 Thompson WJ, Piazza GA, Li H, Liu L, Fetter J, Zhu B, Sperl G, Ahnen D and Pamukcu R: Exisulind induction of apoptosis involves guanosine 3',5'-cyclic monophosphate phosphodiesterase inhibition, protein kinase G activation, and attenuated beta-catenin. Cancer Res 60(13): 3338-3342, 2000.
- 22 Fukuda K: Apoptosis-associated cleavage of beta-catenin in human colon cancer and rat hepatoma cells. Int J Biochem Cell Biol 31(3-4): 519-529, 1999.
- 23 Davies NM and Watson MS: Clinical pharmacokinetics of sulindac. A dynamic old drug. Clin Pharmacokinet 32(6): 437-459, 1997.

Received June 18, 2009 Revised January 18, 2010 Accepted January 19, 2010