Influence of Novel KGFR Tyrosine Kinase Inhibitors on KGF-mediated Proliferation of Breast Cancer

MEGHNA MEHTA¹, JASON W. KESINGER¹, XIAO-PING ZANG¹, MEGAN L. LERNER², DANIEL J. BRACKETT², ROBERT W. BRUEGGEMEIER³, PUI-KUI LI³ and J. THOMAS PENTO¹

¹Department of Pharmaceutical Sciences, College of Pharmacy, and
²VA Medical Center, University of Oklahoma, Health Sciences Center, Oklahoma City, OK 73117, U.S.A.;
³Division of Medicinal Chemistry and Pharmacognosy, College of Pharmacy,
The Ohio State University, Columbus, OH 43210, U.S.A.

Abstract. Background: Keratinocyte growth factor (KGF) acts at the KGF receptor (KGFR) to produce a rapid stimulation of breast cancer cell proliferation and motility which is mediated via the Erk signaling pathway. Enhancement of KGF/KGFR signal transduction may be an early step in the metastatic progression of breast cancer. Receptor modeling of KGFR was used to identify selective KGFR tyrosine kinase (TK) inhibitor molecules that have the potential to bind selectively to the KGFR. The present study evaluated the biological activity of 57 of these KGFR TK inhibitor compounds on breast cancer cells. Materials and Methods: These compounds were tested for their ability to inhibit KGF-mediated breast cancer cell proliferation in MCF-7 breast cancer cells. Furthermore, the effects of the most effective proliferation inhibitors were examined on Erk signaling and on the relative density of cell membrane KGFR. Results: It was observed that 27 of the 57 compounds tested produced a 20% or greater reduction in KGF-mediated proliferation; while five compounds produced greater than 50% inhibition. In addition, the most potent inhibitors also reduced Erk signaling and cell membrane density of the KGFR. Conclusion: The compounds examined appear to be selective KGFR inhibitors which inhibit KGFmediated activity and reduce the expression of KGFR on cancer cells. These results may lead to the development of a novel class of anticancer agents for the prevention of metastatic cancer progression.

Correspondence to: J. Thomas Pento, Department of Pharmaceutical Sciences, College of Pharmacy, University of Oklahoma Health Sciences Center, 1110 N. Stonewall Ave., Oklahoma City, OK 73117, U.S.A. Tel: +1 4052716593 ex.47244, Fax: +1 4052717505, e-mail: tom-pento@ouhsc.edu

Key Words: Cell proliferation, Erk signaling, keratinocyte growth factor, KGFR tyrosine kinase inhibitor, breast cancer, metastasis prevention.

Keratinocyte Growth Factor (KGF, also designated FGF-7), a member of the fibroblast growth factor family, was originally isolated from human embryonic lung fibroblasts (1). KGF is produced by stromal cells (2) and acts at the KGF receptor (KGFR) found on epithelial cells. KGFR (also known as FGFR-2IIIb) is a splice variant of FGFR-2 encoded by the FGFR-2 gene (3). Thus, KGFR is a member of the fibroblast growth factor receptor (FGFR) family which are membrane-spanning tyrosine kinase receptors consisting of four known peptides whose sequences are highly conserved (4). KGF acts at the KGFR and stimulates epithelial cell DNA synthesis, proliferation and migration in breast and other tissues (2, 5, 6). Accordingly, it has been observed that elevated levels of KGF in female rodent species induced mammary epithelial hyperplasia and the eventual development of metastatic mammary carcinomas (7, 8).

It has previously been shown that KGF treatment upregulates KGFR gene expression in MCF-7 breast cancer cells (9), and induces rapid and direct motility enhancement in MCF-7 and other estrogen receptor positive breast cancer cell lines (10). In addition, it has been reported that KGF/KGFR-induced proliferation and motility is mediated *via* the Erk1,2 signaling pathway in MCF-7 human breast cancer cells (11).

These results suggest that KGF-mediated stimulation of breast epithelial cell proliferation and migration may be an important early event in the molecular cascade, which leads to breast cancer progression and metastasis (12). Thus, the inhibition of KGF/KGFR signaling may be an important therapeutic target to selectively retard the metastatic progression of breast cancer with few, if any, adverse side effects. The objective of the present study was to evaluate the biological activity of a novel group of KGFR TK inhibitors that have the potential to selectively reduce KGF-mediated cancer progression.

0250-7005/2010 \$2.00+.40 4883

Table I. Effect of KGFR TK inhibitors on KGF-mediated proliferation. Each % reduction value represents the mean of the 3-5 observations.

KGFR TK Inhibitor	Molecular Weight	% Reduction in KGF-Mediated Proliferation	<i>p</i> -Value	Chemical Structure
minotor	Wolcodiai Weight	Tomeration	p-value	Onemical outdetate
NSU-1	223	23.13	0.02690	CT _N T.
1,00		20.10	0.02000	
NSU-2	221	7.58	0.33141	L N L O
NSU-5	318	11.48	0.05247	OSO ₂ NH ₂
NSU-6	251	23.21	0.04854	Стросн,
NSU-7	237	49.05	0.00041	CTT.
NSU-8	316	25.4	0.00308	OSO ₂ NH ₂
NSU-9	253	44.77	0.00224	CH ₃ O N O
NSU-11	318	6.11	0.01198	H ₂ NO ₂ SO
NSU-13	237	23.23	0.35975	HO CINTO
NSU-17	318	35.93	0.00805	OSO ₂ NH ₂
NSU-21	253	22.95	0.00859	N o och,
NSU-22	239	5.84	0.03702	CTH OH
NSU-23	318	30.73	0.23864	N OSO ₂ NH ₂
NSU-32	411	13.5	0.45981	H ₂ NO ₂ SO OSO ₂ NH ₂
				сн,о СН,о осн,
NSU-33	283	13.25	0.00598	ОН
NSU-34	255	22.1	0.01914	OH CHICAGO
NSU-35	413	26.61	0.01620	H ₂ NO ₂ SO COSO ₂ NH ₂
				CH ₃ O CH ₃
NSU-36	281	27.72	0.00477	H

continued

Table I. continued

KGFR TK		% Reduction in KGF-Mediated		
Inhibitor	Molecular Weight	Proliferation	<i>p</i> -Value	Chemical Structure
NSU-37	253	29.64	0.06970	HO CINTO
				H ₂ NO ₂ SO OSO ₂ NH ₂
NSU-38	411	5.6	0.0920	H *
NSU-39	253	31.8	0.04195	CH ₃ O N
NSU-41	316	38.4	0.01470	H ₂ NO ₂ SO N
NSU-43	309	3.44	0.15273	оснасоосна
NSU-49	235	83.53	0.00647	
NSU-54	251	30.82	0.04312	CH.
NSU-57	281	45.04	0.00559	Hooc
NSU-61	279	26.9	0.00126	HOOC
NSU-71	281	3.6	0.0101	сньо Сньо
NSU-73	237	31.15	0.00028	HO CINTO
	4.2.53.4	200000000000000000000000000000000000000		NSU - 73
NSU-80	252	54.72	0.00197	он до
NSU-118	225	61.23	0.00098	\$
NSU-129	238	47.52	0.23442	HOLD HA
L-14	264	9.65	0.00896	
L-21	230.2	37.7	0.26520	H C NH
L-22	260.2	61.6	0.00066	The H-cooch
L-22	200.2	01.0	0.0000	N-(CH ₂) ₂ NH ₂
L-27	231.3	78.62	0.000009	N O H
L-31	280.1	37.45	0.13937	

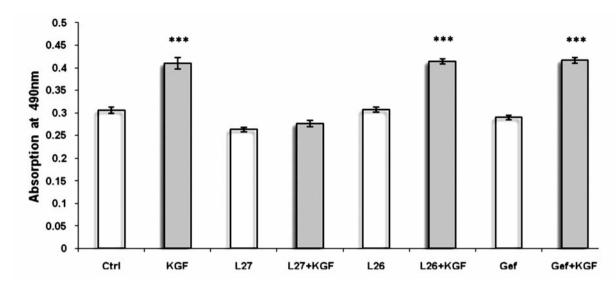


Figure 1. Effect of KGFR TK inhibitors (20 μ M) on KGF-mediated proliferation of MCF-7 at 48 hrs. The bars represent the mean of 3-5 observations \pm SEM. Statistical significance; ***p<0.005.

Materials and Methods

Development of small-molecule ATP-competitive inhibitors as selective KGFR TK inhibitors. A homology model of the KGFR tyrosine kinase domain using 2FGI, the crystal structure of FGFR-1, was previously constructed as a template (16). The protein structure of FGFR-1 has 86% homology with the KGFR tyrosine kinase domain and has been used to guide the design of novel ATP site-directed ligands (13-15). In silico site-directed mutagenesis was used to generate a model of the KGFR TK domain as previously described (16).

Using this model, a series of 57 potential inhibitors of KGFR was identified. The series contains compounds with indolinone and quinolinone structural core and their conformationally restricted analogues. Synthesis of these compounds was previously described (16).

Cell culture. MCF-7 human breast cancer cells obtained from the Michigan Cancer Foundation were maintained as monolayer cultures in RPMI 1640 media (without phenol red) supplemented with 2 mM L-glutamine, gentamicin (50 μg/ml), penicillin (100 units/ml), streptomycin (100 μg/ml), estradiol (10⁻¹¹ M) (all from Sigma, St. Louis, MO, USA) and bovine calf serum (Hyclone, Logan, UT, USA) (5%) as previously reported (11).

Cell proliferation assay. One day before treatment, approximately 1000 cells per well were seeded in 96 well plates and allowed to attach overnight. The treatment vehicle contained 0.5% DMSO. Treatment groups consisted of 3-5 wells each. The cells were treated for 48 h with either human recombinant KGF (R&D Systems, Minneapolis, NM, USA) at 50 ng/ml, KGFR TKI at 20 µM or a combination of KGF and KGFR TK at the same concentrations or vehicle alone in the control group. At the end of the 48 h treatment period, the viable cell number was determined using an MTS assay according to the manufacturer's protocol (Promega, Madison, WI, USA).

Erk signaling assay. Two days before treatment, approximately 3000 cells per well were seeded in 96 well plates and allowed to attach overnight. Treatment groups consisted of 3-5 wells each. The cells are treated for 10 min with either KGF at 500 ng/ml, KGFR TK inhibitor at 100 μM or a combination of KGF and KGFR TK inhibitor at the same concentrations or vehicle in the control group. At the end of the treatment period the relative Erk 1/2 phosphorylation was measured using a cell based ELISA assay according to the manufacturer's protocol (Ray Biotech, Norcross, GA, USA).

Immunocytochemistry. Cells were placed on glass slides and air dried for 24 h at room temperature and prepared for immuocytochemistry as previously described (17). The cells were stained for KGFR using the KGFR Bek rabbit polyclonal antibody (1:200; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA). The slides were counterstained with hematoxylin (Vector Laboratories, Burlingmane, CA, USA), cleared with xylene, and coverslipped with Acrymount (StatLab, Lewisville, TX, USA). Cells were processed in the same manner, except without KGFR primary antibody, were included to exclude negative immunoreactivity.

Statistical analysis. Multiple group comparisons were conducted using ANOVA and Student's *t*-test for pair-wise comparisons. Group differences resulting in *p*-values of less than 0.05 were considered to be statistically significant.

Results

Effects of KGFR TK inhibitors on KGF-mediated cell proliferation. It was observed that 27 of the 57 KGFR TK inhibitors tested produced greater than 20% inhibition of KGF-mediated proliferation, while five compounds (NSU-49, NSU-80, NSU-118, L-22 and L-27) produced greater

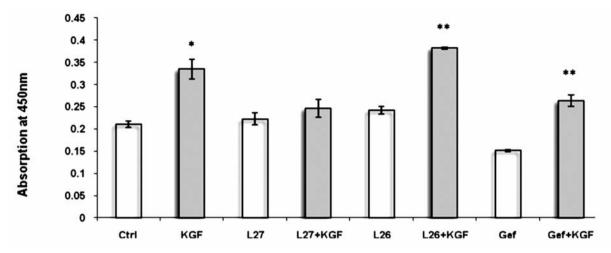


Figure 2. Effect of KGFR TK inhibitors (100 μ M) on KGF-mediated Erk phosphorylation in MCF-7 at 10 min. The bars represent the mean of 3-5 observations \pm SEM. Statistical significance; *p<0.05, **p<0.01.

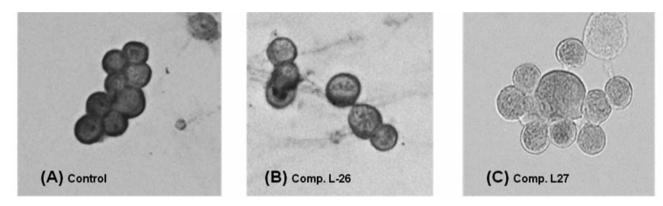


Figure 3. Immuno-localization of KGFR in MCF-7 cells. In these photomicrographs ($\times 200$) MCF-7 cells were treated with either (A) vehicle or (B) compound L-26 (60 μ M, negative control) or (C) L27 (60 μ M) for 24 h before processing by immuno-cytochemistry. Increased density represents KGFR immuno-localization.

than 50% inhibition (p<0.05) in the breast cancer cells (see Table I). The effects of an active inhibitor (L-27), inactive compound (L-26) and EGFR selective inhibitor (gefitnib) are presented in Figure 1. One of the KGFR TK inhibitors (NSU-129) which inhibited KGF-mediated proliferation (see Table I), produced a 31% increase (p<0.05) in proliferation when administered in the absence of KGF (data not shown).

Effects of KGFR TK inhibitors on KGF-mediated Erk signaling. The influence of the five compounds found to be the most potent in reducing KGF-mediated proliferation were examined for their effects on KGF-mediated phosphorylation or activation of Erk. The results indicated that three of the active KGFR TK inhibitors (NSU-80, L-22 and L-27) reduced

(*p*<0.05) KGF-mediated phosphorylation, while the inactive KGFR TK inhibitors, those that did not alter KGF-mediated proliferation, did not alter KGF-mediated phosphorylation. The results of an experiment with the active KGFR TK inhibitor (L-27), inactive KGFR TK inhibitor (L-26) and EGFR selective TK inhibitor (gefitnib) are illustrated in Figure 2.

Effects of KGFR TK inhibitors on KGFR immuno-localization. Compounds which were found to be the most potent KGF inhibitors in the cell proliferation assays (NSU-49, NSU-80, NSU-118, L-22 and L-27) produced a marked reduction in KGFR density on the MCF-7 cells as determined by immunocytochemistry. Figure 3 illustrates the effects of L-27 on KGFR receptor density as compared to inactive compound L-26 and a vehicle treated control.

Discussion

Up-regulation of KGF and KGFR expression has been observed in human primary breast tumor specimens (9, 18). Furthermore, there is evidence that KGF acts as a paracrine growth factor in breast cancer (19). Moreover, KGF and KGFR have been reported to enhance the progression of breast cancer by inhibiting normal apoptosis (20).

KGF treatment of ER-positive breast cancer cells *in vitro* has been observed to produce a rapid increase in the proliferation and motility and an increased metastatic potential (10, 21). Furthermore, this KGF-mediated effect appears to be mediated primarily by activation of KGFR *via* the Erk1,2 signal transduction pathway (11). Thus, the enhancement of KGF/KGFR signaling may be an early event in breast cancer metastatic progression (22). Accordingly, specific inhibition of KGF-mediated receptor signaling at the KGFR TK receptor is expected to reduce or eliminate KGF-associated effects on breast cancer motility and metastatic progression.

The present study was designed to examine the effectiveness of a novel group of KGFR TK inhibitors and to identify the most potent and selective inhibitors of KGF-mediated breast cancer cell progression. The results of the biological testing conducted in this study revealed that approximately half of these compounds produced more that 20% inhibition of KGF activity and that five compounds produced greater than 50% inhibition. These biological results compared favorably with the estimated free binding energy of these molecules obtained through molecular screening (16). Furthermore, the reduction of KGFR density on the surface of the cancer cells suggests that interaction of the inhibitor compounds at the ATP binding site of KGFR produces a down-regulation of KGFR expression in the breast cancer cell. This agrees with an earlier observation that KGF treatment enhanced KGFR expression using a cDNA expression array (23). Further analysis of structure-activity relationships, based on these biological results, should permit the creation of a more accurate homology model and result in the identification of even more effective KGFR TK inhibitors.

It is possible that an increased release of KGF from breast stromal tissue, an up-regulation of KGFR or receptor signaling in developing breast cancer tissue may represent an early enabling step in the initiation of metastatic progression (20). Thus, therapeutic approaches such as selective inhibition of KGFR mediated activity may effectively inhibit the growth and early progression of breast cancer to a more malignant and metastatic phenotype with fewer adverse side effects than current chemotherapy.

In conclusion, these results demonstrate that modeling of the KGFR is capable of creating highly effective and selective KGFR TK inhibitors. Furthermore, these compounds may have the potential to be used therapeutically for the reduction or prevention of breast cancer metastatic progression.

Acknowledgements

This study was supported in part by an NIH/NCI grant (CA-125493).

References

- 1 Rubin JS, Osada H, Finch PW, Taylor WG, Rudikoff S and Aaronson SA: Purification and characterization of a newly identified growth factor specific for epithelial cells. Proc Natl Acad Sci USA 86: 802-806, 1989.
- 2 Rubin JS, Bottaro DP, Chedid M, Miki T, Ron D, Cheon G, Taylor WG, Fortney E, Sakata H, Finch PW and LaRochelle WJ: Keratinocyte growth factor. Cell Biol Int 19: 399-411, 1995.
- 3 Miki T, Bottaro DP, Fleming TP, Smith CL, Burgess WH, Chan AM and Aaronson SA: Determination of ligand-binding specificity by alternative splicing: Two distinct growth factor receptors encoded by a single gene. Proc Natl Acad Sci USA 89: 246-250, 1992.
- 4 Celli G, LaRochelle WJ, Mackem S, Sharp R and Merlino G: Soluble dominant-negative receptor uncovers essential roles for fibroblast growth factors in multi-organ induction and patterning. Embo J 17: 1642-1655, 1998.
- 5 Aaronson SA, Bottaro DP, Miki T, Ron D, Finch PW, Fleming TP, Ahn J, Taylor WG and Rubin JS: Keratinocyte growth factor. A fibroblast growth factor family member with unusual target cell specificity. Ann N Y Acad Sci 638: 62-77, 1991.
- 6 Bottaro DP, Rubin JS, Ron D, Finch PW, Florio C and Aaronson SA: Characterization of the receptor for keratinocyte growth factor. Evidence for multiple fibroblast growth factor receptors. J Biol Chem 265: 12767-12770, 1990.
- 7 Ulich TR, Yi ES, Cardiff R, Yin S, Bikhazi N, Biltz R, Morris CF and Pierce GF: Keratinocyte growth factor is a growth factor for mammary epithelium in vivo. The mammary epithelium of lactating rats is resistant to the proliferative action of keratinocyte growth factor. Am J Pathol 144: 862-868, 1994.
- 8 Kitsberg DI and Leder P: Keratinocyte growth factor induces mammary and prostatic hyperplasia and mammary adenocarcinoma in transgenic mice. Oncogene 13: 2507-2515, 1996.
- 9 Zang XP, Lerner ML, Brackett DJ and Pento JT: Keratinocyte growth factor-mediated pattern of gene expression in breast cancer cells. Cancer Genomics Proteomics 1: 339-344, 2004.
- 10 Zang XP and Pento JT: Keratinocyte growth factor-induced motility of breast cancer cells, Clin Exp Metastasis 18: 573-580, 2001.
- 11 Zang XP, Siwak D, Nguyen TX, Tari AM and Pento JT: KGF-induced motility of breast cancer cells is dependent on grb2 and erk1,2. Clin Exptl Metastasis 21: 437-443, 2004.
- 12 Taniguchi F, Harada T, Sakamoto Y, Yamauchi N, Yoshida S, Iwabe T and Terakawa N: Activation of mitogen-activated protein kinase pathway by keratinocyte growth factor or fibroblast growth factor-10 promotes cell proliferation in human endometrial carcinoma cells. J Clin Endocrinol Metab 88: 773-780, 2003.

- 13 Mohammadi M, Froum S, Hamby JM, Schroeder MC, Panek RL, Lu GH, Eliseenkova AV, Green D, Schlessinger J and Hubbard SR: Crystal structure of an angiogenesis inhibitor bound to the FGF receptor tyrosine kinase domain. Embo J 17: 5896-5904, 1998.
- 14 Mohammadi M, McMahon G, Sun L, Tang C, Hirth P, Yeh BK, Hubbard SR and Schlessinger J: Structures of the tyrosine kinase domain of fibroblast growth factor receptor in complex with inhibitors. Science 276: 955-960, 1997.
- 15 Mohammadi M, Schlessinger J and Hubbard SR: Structure of the FGF receptor tyrosine kinase domain reveals a novel autoinhibitory mechanism. Cell 86: 577-587, 1996.
- 16 Hackett J, Xiao Z, Zang XP, Lerner ML, Brackett DJ, Brueggemeier RW, Li PK and Pento JT: Development of keratinocyte growth factor receptor tyrosine kinase inhibitors for the treatment of cancer. Anticancer Res 27: 3801-3806, 2007.
- 17 Zang XP, Lerner MR, Dunn ST, Brackett DJ and Pento JT: Antisense KGFR oligonucleotide inhibition of KGF-induced motility in breast cancer cells. Anticancer Res 23: 4913-4919, 2003.
- 18 Koos RD, Banks PK, Inkster SE, Yue W and Brodie AM: Detection of aromatase and keratinocyte growth factor expression in breast tumors using reverse transcriptionpolymerase chain reaction. J Steroid Biochem Mol Biol 45: 217-225, 1993.
- 19 Palmieri C, Roberts-Clark D, Assadi-Sabet A, Coope RC, O'Hare M, Sunters A, Hanby A, Slade MJ, Gomm JJ, Lam EW and Coombes RC: Fibroblast growth factor 7, secreted by breast fibroblasts, is an interleukin-1beta-induced paracrine growth factor for human breast cells. J Endocrinol 177: 65-81, 2003.

- 20 Hishikawa Y, Tamaru N, Ejima K, Hayashi T and Koji T: Expression of keratinocyte growth factor and its receptor in human breast cancer: Its inhibitory role in the induction of apoptosis possibly through the overexpression of bcl-2. Arch Histol Cytol 67: 455-464, 2004.
- 21 Zang XP, Bullen EC, Manjeshwar S, Jupe ER, Howard EW and Pento JT: Enhanced motility of KGF-transfected breast cancer cells. Anticancer Res 26: 961-966, 2006.
- 22 Lochter A, Galosy S, Muschler J, Freedman N, Werb Z and Bissell MJ: Matrix metalloproteinase stromelysin-1 triggers a cascade of molecular alterations that leads to stable epithelialto-mesenchymal conversion and a premalignant phenotype in mammary epithelial cells. J Cell Biol 139: 1861-1872, 1997.
- 23 Zang XP, Lerner ML, Brackett DJ and Pento JT: A comparison of keratinocyte growth factor receptor expression in breast and other cancer tissue. Breast Cancer Res Treat 82: 525, 2003.

Received August 13, 2010 Revised October 27, 2010 Accepted October 29, 2010