

Review

Epigenetic Corruption of VDR Signalling in Malignancy

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Abstract. *Background: The ligand-mediated switch from binding co-repressor to co-activator complexes is central to the transcriptional actions of the vitamin D receptor (VDR) and other nuclear receptors. The capacity of deregulated co-repressors to attenuate the responsiveness of VDR signalling in cancer models was examined. Materials and Methods: Proliferation and gene regulation studies were undertaken in non-malignant and malignant cell line and primary models. Results: Both primary tissue models and cancer cell lines displayed a spectrum of suppressed responsiveness towards 1 α , 25 hydroxy vitamin D₃ (1 α 25(OH)₂D₃) which correlated with elevated co-repressor content: specifically, elevated silencing mediator of retinoid and thyroid hormone receptors/nuclear co-repressor 2 (NCoR2/SMRT) in prostate cancer cell lines and primary tumour cultures, and elevated nuclear receptor co-repressor 1 (NCoR1) in breast cancer cell lines. Interestingly, whilst the cancer cell lines frequently also displayed reduced VDR content, the primary tumour material retained and/or elevated VDR mRNA, correlated with co-*

repressor content. Functional approaches towards NCoR2/SMRT (siRNA) in prostate cancer cells or NCoR1 (overexpression) in non-malignant breast epithelial cells confirmed a role in suppressing VDR transcriptional and cellular actions. Targeted co-treatments of 1 α 25(OH)₂D₃ plus HDAC inhibitors (TSA, NaB) resulted in re-expression of antiproliferative target genes (e.g., GADD45 α , p21^(waf1/cip1)) and synergistic inhibition of proliferation. Conclusion: These data suggest that VDR actions in solid tumours are retained, but were skewed by epigenetic mechanisms to suppress selectively antiproliferative target gene promoter responses. This molecular lesion provides a novel chemotherapy target for acceptable doses of 1 α 25(OH)₂D₃ plus HDAC inhibitors.

Breast, Prostate and Colon are Self-renewing Tissues which Give Rise to Common Cancers

The underlying causes for these three high profile cancers are still not clearly understood. Only the minority of cases are strongly associated with a positive family history. For example, about 5% of all breast cancer cases have been linked to high penetrant genetic mutations at the *BRCA1* and *BRCA2* gene loci. Similarly the *APC* gene, whose product is a key regulator of the Wnt/ β -catenin pathway, is frequently mutated in blood relatives with aggressive colon cancer (55). Historically, this exclusive genetic causality has provided a paradigm for investigating the aetiology of cancer, although other strong penetrance genes have not been easily identified.

An alternative contemporary view of these and other common cancers is that they include a contribution from an ill-defined combination of genetic factors with weak penetrance, acting in response to a multitude of environmental factors (59). Statistically, the single greatest risk factor for most cancers is age, with the average age of onset of breast, prostate and colon cancer being in the sixth

Abbreviations: AR, androgen receptor; ER, estrogen receptor; ERR, estrogen-related receptor; FXR, farnesoid X receptor; HDAC, histone deacetylase; LCA, lithocholic acid; LXR, liver X receptor; NaB, sodium butyrate; NCoR1, nuclear receptor co-repressor 1; NCoR2/SMRT, silencing mediator of retinoid and thyroid hormone receptors/nuclear receptor co-repressor 2; PPAR, peroxisome proliferator-activated receptor; PXR, pregnane X receptor; RAR, retinoic acid receptor; RE, response element; RXR, retinoid X receptor; TSA, trichostatin A; VDR, vitamin D receptor; 1 α ,25(OH)₂D₃, 1 α ,25dihydroxyvitaminD₃.

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and seventh decades of life. The sporadic, temporal acquisition of a cancer phenotype is compatible with multifactorial models which require disruption of mechanisms of cell restraint and tissue organisation (42).

Prostate and mammary glands and the lining of the gastro-intestinal tract, all typify self-renewing tissues containing stem cell populations, which give rise to committed progenitors and, in turn, the multiple cell lineages required for tissue function (23, 27, 85, 87). Stem cells are relatively rare and frequently quiescent, but long-lived. Furthermore, they are uniquely able to undergo asymmetric division, to give rise to both other stem cells and transiently amplify populations of progenitor cells, which in turn give rise to the differentiated cell types. By contrast, differentiated epithelial cells are functional, but short-lived, and are lost through programmed cell death processes, to be replaced by newly differentiated transiently amplifying cells. Control of the intricate balance of the processes of division, differentiation and programmed cell death in these different cell types include common roles for Wnt, Hedgehog and other developmental signal transduction processes (10, 85). Convergent targets for these signals include key regulators of cell proliferation such as the cyclin-dependent kinase inhibitor p21^(waf1/cip1).

As a result of their long life cycle and high proliferative capacity, stem cells, rather than the differentiated cells, are the candidates for tumorigenesis (86). To counter this, there appear to be a range of mechanisms in place in stem cells to maintain genomic integrity (reviewed in (90)). These controls notwithstanding, the transformation of stem cells has given rise to the concept of cancer stem cells. Accumulating evidence supports the presence of these cells in prostate, breast and colon cancers (4, 20, 28, 88).

Emerging Roles for Diet Impacting on Malignancy

Recently, the appreciation of the impact of diet on either the initiation or progression of cancer has come significantly to the fore (reviewed in (6, 44, 65)). Reflecting on the accumulation of these data, the World Health Organisation has now stated that diet forms the second most preventable cause of cancer (after smoking). This impact will increase due to demographic factors and, quite possibly, due to changing dietary habits worldwide.

Aspects of these relationships are found in breast, prostate and colon cancer, where the aetiology of the disease reflects the cumulative impact of dietary factors over an individual's life time (12). Equally, these relationships have the potential to be exploited clinically, for example in the SELECT trial to assess the chemopreventive potentials of vitamin E and selenium in prostate cancer (78).

Despite the significance and potential clinical benefits of these relationships the critical time frame remains unclear,

when dietary factors may be protective against cancer development, for example during embryogenesis, childhood development or adult life. Resolving this is, understandably, highly challenging. Considerable resources were required to elucidate what is now established as a clear causal relationship between cigarette smoke and lung cancer (26). To address this, the emerging field of nutrigenomics aims to dissect the impact of dietary factors on genomic regulation, and, thereby, physiology and pathophysiology utilising a range of post-genomic technologies (73).

The Vitamin D Receptor and Other Nuclear Receptors Allow a Local, Integrated Response to Lipophilic Nutrients

The nuclear receptors form one of the largest human families of transcription factors and frequently bind, with a range of affinities, to lipid-derived hormonal, dietary and environmental factors to regulate gene targets, and can be classified broadly according to ligand affinities. The first group of receptors bind ligands with high affinity and are typified by the sex steroid hormone receptors, *e.g.*, the estrogen receptors (ER α and β). Equally a number of micronutrient ligands are also bound with high affinity by specific receptors. For example, the active metabolites of vitamins A and D are bound, respectively, by the retinoic acid and retinoid X receptors (RARs and RXRs) (15) and vitamin D receptor (VDR) (91). The second group of receptors, the adopted orphan receptors (71), bind with broader affinity more abundant macronutrients such as polyunsaturated fatty acids and bile acids, for example the peroxisome proliferator activated receptors (PPARs) (72, 92), liver X receptors (LXRs) (31) and farnesoid X receptor (FXR). Finally, a group of orphan receptors exists, for which no ligands have been identified. Phylogenetic classification, by contrast, has defined seven subfamilies and, within these, the VDR is in the group I sub-family, sharing homology with the LXRs and FXR and more distantly with the PPARs.

Both high and broad affinity receptors appear to work in concert. For example, in the case of the group I receptors, originally the VDR had been described for its central endocrine role in the maintenance of serum calcium levels. Similarly, the FXR and LXRs were described for their roles in regulating cholesterol and fatty and bile acids metabolism in the enterohepatic system. However, these and the related RXRs can equally mediate the local response to fatty and bile acids (39). Thus, the VDR can also respond to the secondary bile acid lithocholic acid (LCA), to induce the cytochrome P450, CYP3A4 (1, 67). Thus, the finding that oxysterols, bile acids, and some polyunsaturated fatty acids can bind and collectively activate LXRs, FXR, PPARs, pregnane X receptor (PXR), RXR and VDR providing a strong molecular link between tissue homeostasis and the

type and availability of nutrients, at least in the colon. The protection provided by PXR, FXR and VDR activation may become compromised when the detoxification pathway is overwhelmed (e.g., by increased levels of LCA due to sustained high-fat diets), or where other nuclear receptor ligands are deficient, for example reduced signalling via the VDR in cases of vitamin D deficiency and where diets have reduced levels of dietary-derived fatty acids which are known to be ligands for the PPARs. Equally the ERs and the estrogen-related receptors (ERRs) respond to estrogenic hormones and a range of phytoestrogens and flavonoids, with different affinities, and collectively play a role in the maintenance of the mammary gland (9, 11, 17, 36).

Examination of VDR, RARs, PPARs, FXR and LXR signalling reveals that they have in common target genes (5) that regulate the cell cycle (e.g., p21^(waf1/cip1) (18, 50, 66, 93) and xenobiotic clearance via cytochrome P450s (e.g., CYP3A4 (30, 38, 46, 56)). Furthermore, there appears to be co-regulation of the receptors. Thus, FXR has been shown to induce a range of target genes that may play other roles in controlling cell proliferation, such as the related nuclear receptor, PPAR α , thus providing molecular evidence for a local, paracrine, cross-talk between the bile acid-sensing receptor and cell proliferation (80). A similar co-regulatory relationship exists between the VDR and PPAR (29).

The post-genomic description of the nuclear receptor superfamily conjoined with profiling approaches (61) reveals that not only colon epithelial cell, but also breast myoepithelial and prostate epithelial cells express a rich cohort of nuclear receptors, many of which display overt nutrient sensing capacity for micro- and macro-nutrients (2, 25, 49, 62). The expression of these receptors in tissues, such as prostate and breast (Table I), suggests a broader and integrated role as a network or conduit in the local sensing of dietary lipid molecules, providing a functional link between both hormonal and environmental and dietary cues, and tissue homeostasis (30).

Local Remodelling of Chromatin is Central to Nuclear Receptor Transcriptional Functions

The nuclear receptors share a common architecture, which includes defined regions for DNA recognition, ligand binding and co-factor interactions. The DNA-binding domain recognises specific response elements (RE) in target gene enhancer/promoter regions. Most receptors preferentially form homo- or heterodimeric complexes; RXR is a central partner for VDR, PPARs, LXRs and FXR. Therefore, simple REs are formed by two recognition motives and their relative distance and orientation contribute to receptor binding specificity and, more recently, composite elements have been identified.

Table I. Relative expression of dietary sensing nuclear receptors in normal breast, prostate and colon epithelium and examples of dietary ligands. The relative abundance was determined *in silico* from publicly deposited SAGE data(61) and the gradation of colour indicates the relative mRNA abundance. The darker shades indicate higher expression.

Nuclear receptors	Relative abundance			Dietary-derived ligand
	Breast	Prostate	Colon	
High affinity ER α				Phytoestrogens, e.g., genistein and flavonoids,
ER β				
ERR α	■	■	■	
ERR β	■	■	■	
RAR α	■	■	■	All <i>trans</i> retinoic acid
RAR β		■		
RAR γ		■		
RXR α		■	■	9- <i>cis</i> retinoic acid, Docosahexanoic acid
RXR β	■	■	■	
RXR γ		■	■	
VDR	■		■	1 α 25(OH) $_2$ D $_3$ and LCA
Broad affinity PPAR α	■			Eicosapentaenoic acid
PPAR δ ,			■	Linoleic acid
PPAR γ			■	5,8,11,14-eicosatetraenoic acid
LXR α		■		24(S) hydroxycholesterol and cholesterol derivatives
LXR β	■	■	■	Oxysterols
FXR	■	■	■	Chenodeoxycholic acid and other bile acids

In the absence of ligand, the VDR-RXR dimer exists in an ‘apo’ state, as part of large complexes (~2.0 MDa) (63, 101), associated with co-repressors (e.g., NCoR2/SMRT) and bound to RE sequences. These complexes actively recruit a range of enzymes that post-translationally modify histone tails, for example histone deacetylases (HDACs) and methyltransferase and, thereby, maintain a locally closed chromatin structure, around RE sequences (75). Ligand binding induces a so-called *holo* state, facilitating the association of the VDR-RXR dimer with co-activator complexes. A large number of interacting co-activator proteins have been described, which can be divided into multiple families including the p160 family, the non-p160 members and members of the large ‘bridging’ DRIP/TRAP/ARC complex, which links the receptor complex to the co-integrators CBP/p300 and basal transcriptional machinery (68, 84, 96). These receptor co-activator complexes co-ordinate the activation of an antagonistic battery of enzymes such as histone acetyltransferases and, thereby, induce the reorganization of local chromatin regions at the response element (RE) of the target gene promoter. The

complex choreography of this event has recently emerged and involves cyclical rounds of promoter-specific complex assembly, gene transactivation, complex disassembly and proteasome-mediated receptor degradation (52, 84).

The expression, localisation and isoforms of co-repressor complexes have emerged as being critical to determine the spatio-temporal equilibrium between the antagonistic actions of the *apo* and *holo* nuclear receptor complexes and, thus, determine target gene promoter responsiveness in a range of physiological and pathological settings. For example, in regulating nuclear receptor function during neural cell differentiation, in determining cell-specific responses to steroid hormones and in the inappropriate silencing of nuclear receptor actions associated with neoplasia (45, 58, 89).

It remains unclear to what extent the various histone modifications initiated by the *apo* and *holo* nuclear receptor mega-complexes around gene promoter regions influence the subsequent transcriptional responsiveness of the promoter. It has been proposed that these modifications may form a stable and heritable "histone code", that determines the assembly of factors upon the chromatin template and controls individual promoter transcriptional responsiveness (51, 95). Functional studies of the SANT motif contained in the co-repressor NCoR2/SMRT supports this latter idea (43), thus recognising and sustaining specific histone modifications.

Cellular Resistance to the Actions of the VDR

The role for VDR actions in non-calcaemic tissues has been the subject of intensive investigation and a consistent theme that emerges is the regulation of target genes, which subsequently control cell growth, differentiation and programmed cell death. *In vitro* $1\alpha,25(\text{OH})_2\text{D}_3$ is able to regulate the proliferation of a wide range of normal tissues including epithelial cells from the prostate, breast and colon and myeloid CD34-positive precursors (Table II). Complimentary studies using the background of the *vdr*-deficient mice has revealed an important role for this receptor to influence mammary gland morphology by regulating differentiation (105) and integrate with estrogenic programs. Importantly, the *vdr* ablated backgrounds have now been crossed into a range of tumorigenic models and support anti-tumour chemoprevention roles for VDR (106, 107).

A major limitation in the therapeutic exploitation of these anti-proliferative and pro-differentiative signals of $1\alpha,25(\text{OH})_2\text{D}_3$ is the resistance of cancer cells towards $1\alpha,25(\text{OH})_2\text{D}_3$; cancer and leukemic cell lines often display a spectrum of sensitivities including complete insensitivity to $1\alpha,25(\text{OH})_2\text{D}_3$, irrespective of VDR expression (Table II) (69, 104). The molecular mechanisms for $1\alpha,25(\text{OH})_2\text{D}_3$ -insensitivity in cancer are emerging. We and others have demonstrated that the VDR is neither mutated, nor is there a clear relationship between VDR expression and growth inhibition by $1\alpha,25(\text{OH})_2\text{D}_3$ (13, 14, 58). The lack of an anti-proliferative

Table II. Cellular responsiveness towards $1\alpha,25(\text{OH})_2\text{D}_3$ and HDAC inhibitors.

Cell line model	ED ₅₀ (nM) ¹	Co-operative anti-proliferative effects with HDACi ²	Co-operative gene regulatory effects with HDACi ³
Myeloid (74)			
U937	4	nd	nd
HL-60	80	++ (NaB)(103)	VDR (NaB)
KG-1a	>1000	nd	nd
Breast epithelial (14)			
MCF-12A	20	-/+ (NaB, TSA)	nd
T-47D	15	+ (NaB, TSA)	nd
MCF-7	100	+ (NaB, TSA)	nd
MDA-MB-231	>1000	++ (NaB, TSA)	<i>p21(waf1/cip1)</i> (TSA)
Prostate epithelial (58)			
RWPE-1	10	nd	nd
PrEC	15	-/+ (NaB, TSA)	nd
LNCaP	100	+ (NaB, TSA)	nd
PC-3	450	++ (NaB, TSA)	<i>GADD45a</i> , <i>MAPK-APK2 (TSA)</i>
DU 145	>1000	++ (NaB, TSA)	nd
Colon cancer			
CaCo2	100	++ (NaB)	<i>p21(waf1/cip1)</i> , VDR (NaB) (34)

¹ED₅₀ = the estimated dose required to inhibit cell proliferation by 50%.

²Co-operative antiproliferative interactions with HDAC inhibitors, either NaB or TSA as indicated).

+/- = moderate enhancement of effects only observed at low nM doses of $1\alpha,25(\text{OH})_2\text{D}_3$.

+ = moderate enhancement of effects observed across a dose response range of $1\alpha,25(\text{OH})_2\text{D}_3$.

++ = strong or synergistic enhancement of effects observed across a dose response of $1\alpha,25(\text{OH})_2\text{D}_3$.

³Gene targets identified to be co-ordinately regulated by co-treatment of HDAC with $1\alpha,25(\text{OH})_2\text{D}_3$.

nd = not determined.

response is reflected by a suppression of the transcriptional responsiveness of anti-proliferative target genes such as *p21(waf1/cip1)*, *p27(kip1)*, *GADD45a* and *BRCA1* (24, 85, 103). It is interesting to note that the levels of *p21(waf1/cip1)* and *p27(kip1)* mRNA expression play roles in the terminal differentiation of committed progenitor cells and thus $1\alpha,25(\text{OH})_2\text{D}_3$ may play an integrated role in regulating self-renewal.

Paradoxically, VDR transactivation is sustained or even enhanced, as measured by induction of the highly $1\alpha,25(\text{OH})_2\text{D}_3$ -inducible *CYP24* gene (70, 83). Together, these data suggest that the VDR transcriptome is skewed in cancer cells to disfavour antiproliferative target genes, and that a lack of functional VDR alone cannot explain resistance.

Epigenetic Resistance

The gene encoding the VDR protein is known to display a number of polymorphic variations, which are associated variously with cancer incidence, degree of aggressiveness and metastasis (reviewed in (41, 47)). To date, no cytogenetic abnormalities of the *VDR* have been reported.

By contrast we and others have begun to explore epigenetic mechanisms which disrupt VDR signalling. For example, we have proposed that apparent $1\alpha,25(\text{OH})_2\text{D}_3$ insensitivity is not determined solely by a linear relationship between the levels of $1\alpha,25(\text{OH})_2\text{D}_3$ and the VDR, but rather epigenetic events skew the responsiveness of target gene promoters. We have investigated these possibilities in breast and prostate cancer.

Prostate cancer: We found frequently elevated co-repressor mRNA expression, most commonly involving *NCoR2/SMRT*, in malignant primary cultures and cell lines, with reduced the $1\alpha,25(\text{OH})_2\text{D}_3$ antiproliferative response, but not normal or benign disease, indicating that the ratio of VDR to co-repressor may determine $1\alpha,25(\text{OH})_2\text{D}_3$ responsiveness in cancer cells. We explored the significance of elevated co-repressor levels in both cancer cell lines and primary cultures, and reasoned that this lesion could be targeted by co-treatment of ligand ($1\alpha,25(\text{OH})_2\text{D}_3$) plus the HDAC inhibitor trichostatin A (TSA). Supportively, we demonstrated that the $1\alpha,25(\text{OH})_2\text{D}_3$ -response of the androgen-independent PC-3 cells was restored to levels indistinguishable from control normal prostate epithelial cells, by co-treatment with low doses of TSA (58, 82).

This reversal of $1\alpha,25(\text{OH})_2\text{D}_3$ insensitivity provided the opportunity to examine gene expression patterns. Microarray studies demonstrated that $1\alpha,25(\text{OH})_2\text{D}_3$ plus TSA uniquely up-regulated a group of 'repressed' gene targets associated with the control of proliferation and induction of apoptosis, notably *GADD45a* and *MAPK-APK2*, a mediator of the p38 stress response pathway (58, 82).

A siRNA approach towards *NCoR2/SMRT* demonstrated the significant role this co-repressor plays in regulating this response, with its repression resulting in profound enhancement of the induction of *GADD45a* in response to $1\alpha,25(\text{OH})_2\text{D}_3$ (98). These data support a central role for elevated *NCoR2/SMRT* levels in suppressing the induction of key target genes resulting in loss of sensitivity to the antiproliferative action of $1\alpha,25(\text{OH})_2\text{D}_3$.

Roles for both *GADD45a* and *MAPK-APK2* have been demonstrated in cell lines that retain sensitivity to $1\alpha,25(\text{OH})_2\text{D}_3$ signalling. For example, p38/MAPK-APK2 activation regulating $1\alpha,25(\text{OH})_2\text{D}_3$ -induced HL-60 myeloid differentiation and up-regulation of *GADD45a* is a functional part of the antiproliferative action of EB1089 [an analogue of $1\alpha,25(\text{OH})_2\text{D}_3$] in SCC25 squamous carcinoma

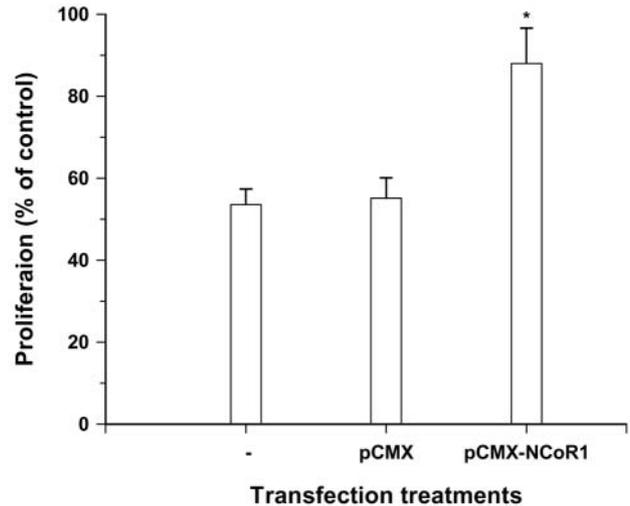


Figure 1. Transient transfection of *NCoR1* in MCF-12A cells. MCF-12A cells were plated in 96-well white-walled tissue culture-treated plates at either 4×10^3 cells/well, and allowed to adhere overnight. Prior to transfection, the cells were exposed to a short trypsin exposure to enhance transfection efficiency. Briefly, immediately prior to transfection the cells were exposed to $40 \mu\text{l}$ trypsin-EDTA (0.05% trypsin, 0.53 mM EDTA)/well for 60 seconds and then the reaction was stopped by the addition of FCS (200 μl /well), aspirated off and then washed with PBS (200 μl /well) prior to proceeding with plasmid transfection. The cells were subsequently transfected with either pCMX-*NCoR1* or pCMX (empty vector control). Plasmids (200 ng/well) were mixed in a 8:1 ratio with Lipofectamine and incubated with the cells for 18 hours in a final volume of 30 μl /well final. Subsequently, the transfection mix was removed and replaced with normal MCF-12A media and the cells were allowed to recover for a subsequent 6 hour prior to dosage with $1\alpha,25(\text{OH})_2\text{D}_3$ or left untreated (control). Liquid proliferation, as measured by the changes in cellular ATP as described previously, was measured after a further 96 hours with a redose after 48 hours. ATP levels were recorded in relative luciferase units and inhibition of proliferation was expressed as a percentage of control. All experiments were repeated in triplicate wells in three separate experiments. * $p < 0.05$.

cells (3) and $1\alpha,25(\text{OH})_2\text{D}_3$ in ovarian cancer cell lines (53). These studies and our own re-expression data highlight these targets as key mediators in the anti-proliferative actions of $1\alpha,25(\text{OH})_2\text{D}_3$.

Breast cancer: In parallel studies, we have demonstrated a similar spectrum of reduced $1\alpha,25(\text{OH})_2\text{D}_3$ -responsiveness between non-malignant breast epithelial cells and cancer cell lines. Again, this was not determined solely by a linear relationship between the levels of $1\alpha,25(\text{OH})_2\text{D}_3$ and *VDR* mRNA expression. Rather elevated co-repressors mRNA levels, notably *NCoR1*, in breast cancer cell lines was common and determined sensitivity towards $1\alpha,25(\text{OH})_2\text{D}_3$ (7, 8). Transient overexpression of *NCoR1* was performed in MCF-12A cells, which display an acute response to $1\alpha,25(\text{OH})_2\text{D}_3$, to assess whether this co-repressor influences cell responsiveness to $1\alpha,25(\text{OH})_2\text{D}_3$. Transfection efficiency was routinely approximately 40% (data not shown). In untreated and mock-transfected controls the antiproliferative

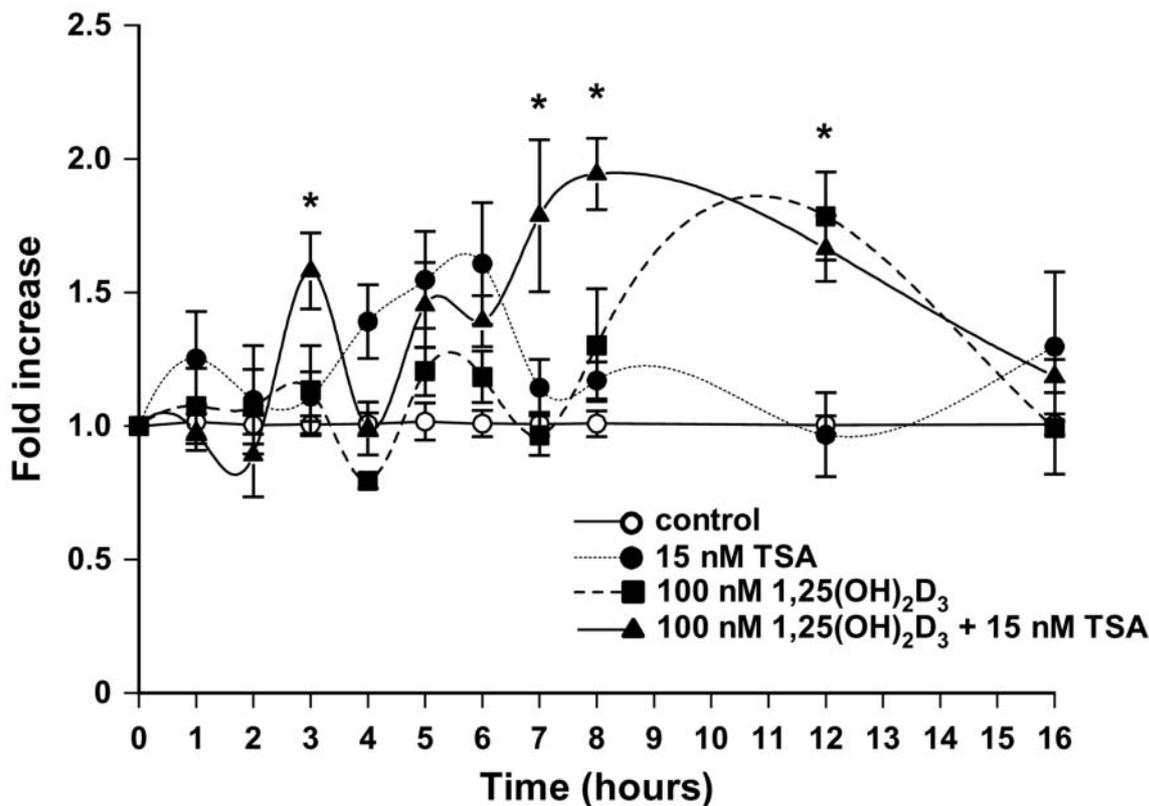


Figure 2. Regulation of *p21(waf1/cip1)* in MDA-MB-231 cells treated with $1\alpha25(\text{OH})_2\text{D}_3$ plus TSA. $2 \times 10^4/\text{cm}^2$ cells were plated into 6-well plates and allowed to grow for 36 hours to ensure that the cells were in the mid-exponential phase. Total RNA was isolated after the indicated time-periods, reverse transcribed and the target genes amplified and the fold increase calculated using quantitative reverse transcription PCR, as described previously. Each data-point represents the mean of three separate experiments amplified in triplicate wells \pm S.E.M. * $p < 0.05$.

sensitivity in liquid culture was equal [$52\% \pm 5$ and $54\% \pm 4$ of control proliferation in response to $1\alpha25(\text{OH})_2\text{D}_3$ (200 nM), respectively]. In contrast, *NCoR1*-transfected cultures displayed increased resistance to 200 nM $1\alpha25(\text{OH})_2\text{D}_3$, having $80\% \pm 9.4$ proliferation compared to untreated controls ($p < 0.01$ compared to both mock and non-transfected controls) (Figure 1).

We have attempted to target this molecular lesion through co-treatments of $1\alpha25(\text{OH})_2\text{D}_3$ with HDAC inhibitors. These data also support the concept that elevated co-repressors such as *NCoR1* reduce the capacity of the VDR to act as a transcription factor for antiproliferative target genes. Thus, treatment of MDA-MB-231 cells with $1\alpha25(\text{OH})_2\text{D}_3$ plus TSA appears to co-ordinately regulate the *p21(waf1/cip1)* mRNA expression; notably up-regulating the target in a unique manner between 5 and 10 hours post-treatment (Figure 2).

These findings compliment a number of parallel studies undertaken by others, which have established cooperativity between $1\alpha25(\text{OH})_2\text{D}_3$ and butyrate compounds, such as sodium butyrate (NaB) (21, 22, 32, 33, 35, 76, 94) (Table II). These compounds are short-chain fatty acids produced

during fermentation by endogenous intestinal bacteria and have the capacity to act as HDAC inhibitors (16). Stein and co-workers have also identified the effects in colon cancer cells of $1\alpha25(\text{OH})_2\text{D}_3$ plus NaB co-treatments to include the co-ordinate regulation of the VDR itself. In our studies, in the time-frame studied (0-24 hours), no evidence for changes in VDR mRNA levels upon co-treatment with $1\alpha25(\text{OH})_2\text{D}_3$ plus TSA were seen. However, together these studies further underscore the importance of the dietary-derived milieu to regulate epithelial proliferation and differentiation beyond classic sites of action in the gut.

Summary

These findings suggest that in cancer the VDR is not overtly disrupted by either genetic mechanisms, such as mutation, or appears to be the subject of cytogenetic re-arrangements. Rather, epigenetic mechanisms attenuate and selectively skew the transcriptional responsiveness. The combination of either VDR or other nuclear receptor ligands with potent HDAC inhibitors has the potential to deliver a more focused and sustained 'anticancer' regime for a range of solid tumours.

Thus, we propose that deregulated, possibly tissue-specific, patterns of co-repressors inappropriately sustain histone deacetylation around the VDRE or target gene promoter regions, and shifts the dynamic equilibrium between *apo* and *holo* receptor conformations to favour transcriptional repression of key target genes such as *p21^(waf1/cip1)* or *GADD45a*. Thus, VDR gene targets are less responsive in $1\alpha,25(\text{OH})_2\text{D}_3$ -insensitive cancer cells compared to non-malignant counterparts. Furthermore, targeting this molecular lesion with co-treatments of vitamin D_3 compounds plus HDAC inhibitors generates a temporal window where the equilibrium point between *apo* and *holo* complexes is shifted to favour a more transcriptionally permissive environment and allows gene targets to be modulated in a unique and significantly greater manner than either agent alone; notably *p21^(waf1/cip1)* and *GADD45a*. The *apo*-VDR complex may, in turn, form a template for subsequent more stable epigenetic silencing of these regions. Equally, the silencing of nuclear transcriptional actions may allow cytoplasmic VDR actions to be sustained, some of which have recently been shown to suppress apoptosis *via* non-transcriptional interactions (97). These studies add to a growing body of data which places the expression of the CoA/CoR milieu as being critical to determine nuclear receptor actions (19, 37, 40, 45, 48, 54, 57, 60, 64, 79, 81, 100).

Epigenetic mechanisms which disrupt VDR signalling most probably disrupt other receptors resulting in reduced sensitivity to a wide range of dietary-derived macro- and micro-nutrient ligands. It is interesting to note that the stem cells in prostate and breast tissues do not appear to express either the androgen receptor (AR) or ER α , respectively. It remains unknown to what extent these cells express the VDR or its related receptors and, if so, what their function is. Addressing these questions will therefore address what role deregulated co-repressor function may play in cancer initiation.

Future Perspectives

Historically, researchers have studied the abilities of single nuclear receptors to regulate a discrete group of gene targets and influence cell function. This has led to substantial knowledge concerning many of these receptors, individually. Cell and organism function, however, depends on the dynamic interaction of a collection of receptors, through the networks that link them. The current lack of an integral view of how these interactions bring about function and dysfunction of the aging human individual can be attributed to the relatively recent limitations of available techniques and tools to undertake such studies. The implementation of the new post-genomic techniques, together with bioinformatics and systems biology methodology, will generate an integral view of the processes by which cells, tissues and organisms interact with diet (99). This transition will allow VDR

processes to be described in the dynamic interaction with other nuclear receptors and cell signal transduction pathways to identify critical modes of control. An equally important component will be to define how these receptors are expressed and regulated during epithelial tissue self-renewal.

A common goal of many researchers around the world is to understand how dietary intervention, based on the knowledge of nutritional requirements, nutritional status and genotype, can be used to prevent, ameliorate or cure chronic disease. Thus, as pharmacogenomics has led to the concept of "personalized medicine", so nutrigenomics may open the way for "personalized nutrition". A critical step to promote this understanding is to construct mathematical models, using system biology approaches, to define how these interactions can work. Ultimately, this will deliver a predicative, preventative and personalized understanding of the dietary interactions in the individual.

References

- 1 Adachi R, Shulman AI, Yamamoto K, Shimomura I, Yamada S, Mangelsdorf DJ and Makishima M: Structural determinants for vitamin D receptor response to endocrine and xenobiotic signals. *Mol Endocrinol* 18: 43-52, 2004.
- 2 Adams JY, Leav I, Lau KM, Ho SM and Pflueger SM: Expression of estrogen receptor beta in the fetal, neonatal and prepubertal human prostate. *Prostate* 52: 69-81, 2002.
- 3 Akutsu N, Lin R, Bastien Y, Bestawros A, Enepekides DJ, Black MJ and White JH: Regulation of gene expression by $1\alpha,25$ -dihydroxyvitamin D_3 and its analog EB1089 under growth-inhibitory conditions in squamous carcinoma cells. *Mol Endocrinol* 15: 1127-1139, 2001.
- 4 Al Hajj M and Clarke MF: Self-renewal and solid tumor stem cells. *Oncogene* 23: 7274-7282, 2004.
- 5 Anderson SP, Dunn C, Laughter A, Yoon L, Swanson C, Stulnig TM, Steffensen KR, Chandraratna RA, Gustafsson JA and Corton JC: Overlapping transcriptional programs regulated by the nuclear receptors peroxisome proliferator-activated receptor $\{\alpha\}$, retinoid X receptor and liver X receptor in mouse liver. *Mol Pharmacol*, 2004.
- 6 Astorg P: Dietary n - 6 and n - 3 polyunsaturated fatty acids and prostate cancer risk: a review of epidemiological and experimental evidence. *Cancer Causes Control* 15: 367-386, 2004.
- 7 Banwell CM, O'Neill LP, Uskokovic MR and Campbell MJ: Targeting $1\alpha,25$ -dihydroxyvitamin D_3 antiproliferative insensitivity in breast cancer cells by co-treatment with histone deacetylation inhibitors. *J Steroid Biochem Mol Biol* 89-90: 245-249, 2004.
- 8 Banwell CM, Singh R, Stewart PM, Uskokovic MR and Campbell MJ: Antiproliferative signalling by $1,25(\text{OH})_2\text{D}_3$ in prostate and breast cancer is suppressed by a mechanism involving histone deacetylation. *Rec Res Cancer Res* 164: 83-98, 2003.
- 9 Barkhem T, Nilsson S and Gustafsson JA: Molecular mechanisms, physiological consequences and pharmacological implications of estrogen receptor action. *Am J Pharmacogenomics* 4: 19-28, 2004.
- 10 Beachy PA, Karhadkar SS and Berman DM: Tissue repair and stem cell renewal in carcinogenesis. *Nature* 432: 324-331, 2004.

- 11 Bentrem D, Fox JE, Pearce ST, Liu H, Pappas S, Kupfer D, Zapf JW and Jordan VC: Distinct molecular conformations of the estrogen receptor alpha complex exploited by environmental estrogens. *Cancer Res* 63: 7490-7496, 2003.
- 12 Boyle P, Severi G and Giles GG: The epidemiology of prostate cancer. *Urol Clin North Am* 30: 209-217, 2003.
- 13 Campbell MJ, Elstner E, Holden S, Uskokovic M and Koeffler HP: Inhibition of proliferation of prostate cancer cells by a 19-nor-hexafluoride vitamin D₃ analogue involves the induction of p21^{waf1}, p27^{kip1} and E-cadherin. *J Mol Endocrinol* 19: 15-27, 1997.
- 14 Campbell MJ, Gombart AF, Kwok SH, Park S and Koeffler HP: The anti-proliferative effects of 1alpha,25(OH)₂D₃ on breast and prostate cancer cells are associated with induction of BRCA1 gene expression. *Oncogene* 19: 5091-5097, 2000.
- 15 Campbell MJ, Park S, Uskokovic MR, Dawson MI and Koeffler HP: Expression of retinoic acid receptor-beta sensitizes prostate cancer cells to growth inhibition mediated by combinations of retinoids and a 19-nor hexafluoride vitamin D₃ analog. *Endocrinology* 139: 1972-1980, 1998.
- 16 Chen JS, Faller DV and Spanjaard RA: Short-chain fatty acid inhibitors of histone deacetylases: promising anticancer therapeutics? *Curr Cancer Drug Targets* 3: 219-236, 2003.
- 17 Cheng G, Weihua Z, Warner M and Gustafsson JA: Estrogen receptors ER alpha and ER beta in proliferation in the rodent mammary gland. *Proc Natl Acad Sci USA* 101: 3739-3746, 2004.
- 18 Chiba H, Itoh T, Satohisa S, Sakai N, Noguchi H, Osanai M, Kojima T and Sawada N: Activation of p21^{CIP1/WAF1} gene expression and inhibition of cell proliferation by overexpression of hepatocyte nuclear factor-4alpha. *Exp Cell Res* 302: 11-21, 2005.
- 19 Chien Y and White MA: RAL GTPases are linchpin modulators of human tumour-cell proliferation and survival. *EMBO Rep* 4: 800-806, 2003.
- 20 Clarke MF: Epigenetic regulation of normal and cancer stem cells. *Ann NY Acad Sci* 1044: 90-93, 2005.
- 21 Costa EM and Feldman D: Modulation of 1,25-dihydroxyvitamin D₃ receptor binding and action by sodium butyrate in cultured pig kidney cells (LLC-PK1). *J Bone Miner Res* 2: 151-159, 1987.
- 22 Daniel C, Schroder O, Zahn N, Gaschott T and Stein J: p38 MAPK signaling pathway is involved in butyrate-induced vitamin D receptor expression. *Biochem Biophys Res Commun* 324: 1220-1226, 2004.
- 23 De Marzo AM, Nelson WG, Meeker AK and Coffey DS: Stem cell features of benign and malignant prostate epithelial cells. *J Urol* 160: 2381-2392, 1998.
- 24 Devgan V, Mammucari C, Millar SE, Brisken C and Dotto GP: p21^{WAF1/Cip1} is a negative transcriptional regulator of Wnt4 expression downstream of Notch1 activation. *Genes Dev* 19: 1485-1495, 2005.
- 25 Di Leo A, Messa C, Cavallini A and Linsalata M: Estrogens and colorectal cancer. *Curr Drug Targets Immune Endocr Metabol Disord* 1: 1-12, 2001.
- 26 Doll R, Peto R, Boreham J and Sutherland I: Mortality from cancer in relation to smoking: 50 years observations on British doctors. *Br J Cancer* 92: 426-429, 2005.
- 27 Dontu G, Al Hajj M, Abdallah WM, Clarke MF and Wicha MS: Stem cells in normal breast development and breast cancer. *Cell Prolif* 36 Suppl 1: 59-72, 2003.
- 28 Dontu G, El-Ashry D and Wicha MS: Breast cancer, stem/progenitor cells and the estrogen receptor. *Trends Endocrinol Metab* 15: 193-197, 2004.
- 29 Dunlop TW, Vaisanen S, Frank C, Molnar F, Sinkkonen L and Carlberg C: The human peroxisome proliferator-activated receptor delta gene is a primary target of 1alpha,25-dihydroxyvitamin D₃ and its nuclear receptor. *J Mol Biol* 349: 248-260, 2005.
- 30 Francis GA, Fayard E, Picard F and Auwerx J: Nuclear receptors and the control of metabolism. *Annu Rev Physiol* 65: 261-311, 2003.
- 31 Fukuchi J, Kokontis JM, Hiipakka RA, Chuu CP and Liao S: Antiproliferative effect of liver X receptor agonists on LNCaP human prostate cancer cells. *Cancer Res* 64: 7686-7689, 2004.
- 32 Gaschott T and Stein J: Short-chain fatty acids and colon cancer cells: the vitamin D receptor--butyrate connection. *Rec Res Cancer Res* 164: 247-257, 2003.
- 33 Gaschott T, Steinhilber D, Milovic V and Stein J: Tributyrin, a stable and rapidly absorbed prodrug of butyric acid, enhances antiproliferative effects of dihydroxycholecalciferol in human colon cancer cells. *J Nutr* 131: 1839-1843, 2001.
- 34 Gaschott T, Wächtershauser A, Steinhilber D and Stein J: 1,25-Dihydroxycholecalciferol enhances butyrate-induced p21^(Waf1/Cip1) expression. *Biochem Biophys Res Commun* 283: 80-85, 2001.
- 35 Gaschott T, Werz O, Steinmeyer A, Steinhilber D and Stein J: Butyrate-induced differentiation of Caco-2 cells is mediated by vitamin D receptor. *Biochem Biophys Res Commun* 288: 690-696, 2001.
- 36 Giguere V: To ERR in the estrogen pathway. *Trends Endocrinol Metab* 13: 220-225, 2002.
- 37 Gnanapragasam VJ, Leung HY, Pulimood AS, Neal DE and Robson CN: Expression of RAC 3, a steroid hormone receptor co-activator in prostate cancer. *Br J Cancer* 85: 1928-1936, 2001.
- 38 Gnerre C, Blattler S, Kaufmann MR, Looser R and Meyer UA: Regulation of CYP3A4 by the bile acid receptor FXR: evidence for functional binding sites in the CYP3A4 gene. *Pharmacogenetics* 14: 635-645, 2004.
- 39 Goldstein JT, Dobrzyn A, Clagett-Dame M, Pike JW and DeLuca HF: Isolation and characterization of unsaturated fatty acids as natural ligands for the retinoid-X receptor. *Arch Biochem Biophys* 420: 185-193, 2003.
- 40 Grignani F, De Matteis S, Nervi C, Tomassoni L, Gelmetti V, Ciocce M, Fanelli M, Ruthardt M, Ferrara FF, Zamir I, Seiser C, Grignani F, Lazar MA, Minucci S and Pelicci PG: Fusion proteins of the retinoic acid receptor-alpha recruit histone deacetylase in promyelocytic leukaemia. *Nature* 391: 815-818, 1998.
- 41 Guy M, Lowe LC, Bretherton-Watt D, Mansi JL, Peckitt C, Bliss J, Wilson RG, Thomas V and Colston KW: Vitamin D receptor gene polymorphisms and breast cancer risk. *Clin Cancer Res* 10: 5472-5481, 2004.
- 42 Hanahan D and Weinberg RA: The hallmarks of cancer. *Cell* 100: 57-70, 2000.
- 43 Hartman HB, Yu J, Alenghat T, Ishizuka T and Lazar MA: The histone-binding code of nuclear receptor co-repressors matches the substrate specificity of histone deacetylase 3. *EMBO Rep* 6: 445-451, 2005.
- 44 Henderson MM: Nutritional aspects of breast cancer. *Cancer* 76: 2053-2058, 1995.

- 45 Hermanson O, Jepsen K and Rosenfeld MG: N-CoR controls differentiation of neural stem cells into astrocytes. *Nature* *419*: 934-939, 2002.
- 46 Hukkanen J, Vaisanen T, Lassila A, Piipari R, Anttila S, Pelkonen O, Raunio H and Hakkola J: Regulation of CYP3A5 by glucocorticoids and cigarette smoke in human lung-derived cells. *J Pharmacol Exp Ther* *304*: 745-752, 2003.
- 47 Ingles SA, Ross RK, Yu MC, Irvine RA, La Pera G, Haile RW and Coetzee GA: Association of prostate cancer risk with genetic polymorphisms in vitamin D receptor and androgen receptor. *J Natl Cancer Inst* *89*: 166-170, 1997.
- 48 Ishii S, Yamada M, Satoh T *et al*: Aberrant dynamics of histone deacetylation at the thyrotropin-releasing hormone gene in resistance to thyroid hormone. *Mol Endocrinol* *18*: 1708-1720, 2004.
- 49 Jarred RA, McPherson SJ, Bianco JJ, Couse JF, Korach KS and Risbridger GP: Prostate phenotypes in estrogen-modulated transgenic mice. *Trends Endocrinol Metab* *13*: 163-168, 2002.
- 50 Jarvis MC, Gray TJ and Palmer CN: Both PPAR γ and PPAR δ influence sulindac sulfide-mediated p21^(WAF1/CIP1) up-regulation in a human prostate epithelial cell line. *Oncogene* *24*: 8211-8215, 2005.
- 51 Jenuwein T and Allis CD: Translating the histone code. *Science* *293*: 1074-1080, 2001.
- 52 Jepsen K and Rosenfeld MG: Biological roles and mechanistic actions of co-repressor complexes. *J Cell Sci* *115*: 689-698, 2002.
- 53 Jiang F, Li P, Fornace AJ Jr, Nicosia SV and Bai W: G2/M arrest by 1,25-dihydroxyvitamin D₃ in ovarian cancer cells mediated through the induction of GADD45 *via* an exonic enhancer. *J Biol Chem* *278*: 48030-48040, 2003.
- 54 Jiang WG, Douglas-Jones A and Mansel RE: Expression of peroxisome-proliferator activated receptor-gamma (PPAR γ) and the PPAR γ co-activator, PGC-1, in human breast cancer correlates with clinical outcomes. *Int J Cancer* *106*: 752-757, 2003.
- 55 Jo WS and Chung DC: Genetics of hereditary colorectal cancer. *Semin Oncol* *32*: 11-23, 2005.
- 56 Jurutka PW, Thompson PD, Whitfield GK *et al*: Molecular and functional comparison of 1,25-dihydroxyvitamin D(3) and the novel vitamin D receptor ligand, lithocholic acid, in activating transcription of cytochrome P450 3A4. *J Cell Biochem* *94*: 917-943, 2005.
- 57 Kawashima H, Takano H, Sugita S, Takahara Y, Sugimura K and Nakatani T: A novel steroid receptor co-activator protein (SRAP) as an alternative form of steroid receptor RNA-activator gene: expression in prostate cancer cells and enhancement of androgen receptor activity. *Biochem J* *369*: 163-171, 2003.
- 58 Khanim FL, Gommersall LM, Wood VH, Smith KL, Montalvo L, O'Neill LP, Xu Y, Peehl DM, Stewart PM, Turner BM and Campbell MJ: Altered SMRT levels disrupt vitamin D(3) receptor signalling in prostate cancer cells. *Oncogene* *23*: 6712-6725, 2004.
- 59 Kotnis A, Sarin R and Mulherkar R: Genotype, phenotype and cancer: role of low penetrance genes and environment in tumour susceptibility. *J Biosci* *30*: 93-102, 2005.
- 60 Krogsdam AM, Nielsen CA, Neve S, Holst D, Helledie T, Thomsen B, Bendixen C, Mandrup S and Kristiansen K: Nuclear receptor corepressor-dependent repression of peroxisome-proliferator-activated receptor delta-mediated transactivation. *Biochem J* *363*: 157-165, 2002.
- 61 Lal A, Lash AE, Altschul SF, Velculescu V, Zhang L, McLendon RE, Marra MA, Prange C, Morin PJ, Polyak K, Papadopoulos N, Vogelstein B, Kinzler KW, Strausberg RL and Riggins GJ: A public database for gene expression in human cancers. *Cancer Res* *59*: 5403-5407, 1999.
- 62 Lau KM, LaSpina M, Long J and Ho SM: Expression of estrogen receptor (ER)-alpha and ER-beta in normal and malignant prostatic epithelial cells: regulation by methylation and involvement in growth regulation. *Cancer Res* *60*: 3175-3182, 2000.
- 63 Li J, Wang J, Wang J, Nawaz Z, Liu JM, Qin J and Wong J: Both corepressor proteins SMRT and N-CoR exist in large protein complexes containing HDAC3. *EMBO J* *19*: 4342-4350, 2000.
- 64 Lin RJ, Nagy L, Inoue S, Shao W, Miller WH Jr and Evans RM: Role of the histone deacetylase complex in acute promyelocytic leukaemia. *Nature* *391*: 811-814, 1998.
- 65 Lipkin M, Reddy B, Newmark H and Lamprecht SA: Dietary factors in human colorectal cancer. *Annu Rev Nutr* *19*: 545-586, 1999.
- 66 Liu M, Lee MH, Cohen M, Bommakanti M and Freedman LP: Transcriptional activation of the Cdk inhibitor p21 by vitamin D₃ leads to the induced differentiation of the myelomonocytic cell line U937. *Genes Dev* *10*: 142-153, 1996.
- 67 Makishima M, Lu TT, Xie W, Whitfield GK, Domoto H, Evans RM, Haussler MR and Mangelsdorf DJ: Vitamin D receptor as an intestinal bile acid sensor. *Science* *296*: 1313-1316, 2002.
- 68 Metivier R, Penot G, Hubner MR, Reid G, Brand H, Kos M and Gannon F: Estrogen receptor-alpha directs ordered, cyclical and combinatorial recruitment of cofactors on a natural target promoter. *Cell* *115*: 751-763, 2003.
- 69 Miller CW, Morosetti R, Campbell MJ, Mendoza S and Koeffler HP: Integrity of the 1,25-dihydroxyvitamin D₃ receptor in bone, lung and other cancers. *Mol Carcinog* *19*: 254-257, 1997.
- 70 Miller GJ, Stapleton GE, Hedlund TE and Moffat KA: Vitamin D receptor expression, 24-hydroxylase activity and inhibition of growth by 1alpha,25-dihydroxyvitamin D₃ in seven human prostatic carcinoma cell lines. *Clin Cancer Res* *1*: 997-1003, 1995.
- 71 Mohan R and Heyman RA: Orphan nuclear receptor modulators. *Curr Top Med Chem* *3*: 1637-1647, 2003.
- 72 Mueller E, Smith M, Sarraf P, Kroll T, Aiyer A, Kaufman DS, Oh W, Demetri G, Figg WD, Zhou XP, Eng C, Spiegelman BM and Kantoff PW: Effects of ligand activation of peroxisome proliferator-activated receptor gamma in human prostate cancer. *Proc Natl Acad Sci USA* *97*: 10990-10995, 2000.
- 73 Muller M and Kersten S: Nutrigenomics: goals and strategies. *Nat Rev Genet* *4*: 315-322, 2003.
- 74 Munker R, Norman A and Koeffler HP: Vitamin D compounds. Effect on clonal proliferation and differentiation of human myeloid cells. *J Clin Invest* *78*: 424-430, 1986.
- 75 Nagy L and Schwabe JW: Mechanism of the nuclear receptor molecular switch. *Trends Biochem Sci* *29*: 317-324, 2004.
- 76 Newmark HL and Young CW: Butyrate and phenylacetate as differentiating agents: practical problems and opportunities. *J Cell Biochem Suppl* *22*: 247-253, 1995.
- 77 Palmer HG, Gonzalez-Sancho JM, Espada J, Berciano MT, Puig I, Baulida J, Quintanilla M, Cano A, de Herreros AG, Lafarga M and Munoz A: Vitamin D(3) promotes the differentiation of colon carcinoma cells by the induction of E-cadherin and the inhibition of beta-catenin signaling. *J Cell Biol* *154*: 369-387, 2001.

- 78 Pathak SK, Sharma RA and Mellon JK: Chemoprevention of prostate cancer by diet-derived antioxidant agents and hormonal manipulation. *Int J Oncol* 22: 5-13, 2003 (Review).
- 79 Picard F, Kurtev M, Chung N, Topark-Ngarm A, Senawong T, Oliveira RM, Leid M, McBurney MW and Guarente L: Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 429: 771-776, 2004.
- 80 Pineda T, I, Claudel T, Duval C, Kosykh V, Fruchart JC and Staels B: Bile acids induce the expression of the human peroxisome proliferator-activated receptor alpha gene *via* activation of the farnesoid X receptor. *Mol Endocrinol* 17: 259-272, 2003.
- 81 Planas-Silva MD, Shang Y, Donaher JL, Brown M and Weinberg RA: AIB1 enhances estrogen-dependent induction of cyclin D1 expression. *Cancer Res* 61: 3858-3862, 2001.
- 82 Rashid SF, Moore JS, Walker E, Driver PM, Engel J, Edwards CE, Brown G, Uskokovic MR and Campbell MJ: Synergistic growth inhibition of prostate cancer cells by 1alpha,25 dihydroxyvitamin D(3) and its 19-nor-hexafluoride analogs in combination with either sodium butyrate or trichostatin A. *Oncogene* 20: 1860-1872, 2001.
- 83 Rashid SF, Mountford JC, Gombart AF and Campbell MJ: 1alpha,25-Dihydroxyvitamin D(3) displays divergent growth effects in both normal and malignant cells. *Steroids* 66: 433-440, 2001.
- 84 Reid G, Hubner MR, Metivier R, Brand H, Denger S, Manu D, Beaudouin J, Ellenberg J and Gannon F: Cyclic, proteasome-mediated turnover of unliganded and liganded ERalpha on responsive promoters is an integral feature of estrogen signaling. *Mol Cell* 11: 695-707, 2003.
- 85 Reya T and Clevers H: Wnt signalling in stem cells and cancer. *Nature* 434: 843-850, 2005.
- 86 Reya T, Morrison SJ, Clarke MF and Weissman IL: Stem cells, cancer and cancer stem cells. *Nature* 414: 105-111, 2001.
- 87 Richardson GD, Robson CN, Lang SH, Neal DE, Maitland NJ and Collins AT: CD133, a novel marker for human prostatic epithelial stem cells. *J Cell Sci* 117: 3539-3545, 2004.
- 88 Schalken JA and van Leenders G: Cellular and molecular biology of the prostate: stem cell biology. *Urology* 62: 11-20, 2003.
- 89 Shang Y and Brown M: Molecular determinants for the tissue specificity of SERMs. *Science* 295: 2465-2468, 2002.
- 90 Sherley JL: Asymmetric cell kinetics genes: the key to expansion of adult stem cells in culture. *Scientific World J* 2: 1906-1921, 2002.
- 91 Skowronski RJ, Peehl DM and Feldman D: Vitamin D and prostate cancer: 1,25 dihydroxyvitamin D₃ receptors and actions in human prostate cancer cell lines. *Endocrinology* 132: 1952-1960, 1993.
- 92 Stephen RL, Gustafsson MC, Jarvis M, Tatoud R, Marshall BR, Knight D, Ehrenborg E, Harris AL, Wolf CR and Palmer CN: Activation of peroxisome proliferator-activated receptor delta stimulates the proliferation of human breast and prostate cancer cell lines. *Cancer Res* 64: 3162-3170, 2004.
- 93 Suzui M, Shimizu M, Masuda M, Lim JT, Yoshimi N and Weinstein IB: Acyclic retinoid activates retinoic acid receptor beta and induces transcriptional activation of p21^(CIP1) in HepG2 human hepatoma cells. *Mol Cancer Ther* 3: 309-316, 2004.
- 94 Tanaka Y, Bush KK, Klauck TM and Higgins PJ: Enhancement of butyrate-induced differentiation of HT-29 human colon carcinoma cells by 1,25-dihydroxyvitamin D₃. *Biochem Pharmacol* 38: 3859-3865, 1989.
- 95 Turner BM: Histone acetylation and control of gene expression. *J Cell Sci* 99(Pt 1): 13-20, 1991.
- 96 Vaisanen S, Dunlop TW, Frank C and Carlberg C: Using chromatin immunoprecipitation to monitor 1alpha,25-dihydroxyvitamin D(3)-dependent chromatin activity on the human CYP24 promoter. *J Steroid Biochem Mol Biol* 89-90: 277-279, 2004.
- 97 Vertino AM, Bula CM, Chen JR *et al*: Nongenotropic, anti-apoptotic signaling of 1alpha,25(OH)₂-vitamin D₃ and analogs through the ligand binding domain of the vitamin D receptor in osteoblasts and osteocytes: Mediation by Src, phosphatidylinositol-3 and JNK kinases. *J Biol Chem* 280: 14130-14137, 2005.
- 98 Wang X, Rao J and Studzinski GP: Inhibition of p38 MAP kinase activity up-regulates multiple MAP kinase pathways and potentiates 1,25-dihydroxyvitamin D(3)-induced differentiation of human leukemia HL60 cells. *Exp Cell Res* 258: 425-437, 2000.
- 99 Westerhoff HV and Palsson BO: The evolution of molecular biology into systems biology. *Nat Biotechnol* 22: 1249-1252, 2004.
- 100 Weston AD, Blumberg B and Underhill TM: Active repression by unliganded retinoid receptors in development: less is sometimes more. *J Cell Biol* 161: 223-228, 2003.
- 101 Yoon HG, Chan DW, Huang ZQ, Li J, Fondell JD, Qin J and Wong J: Purification and functional characterization of the human N-CoR complex: the roles of HDAC3, TBL1 and TBLR1. *EMBO J* 22: 1336-1346, 2003.
- 102 Yoshida M, Tanaka Y, Eguchi T, Ikekawa N and Saijo N: Effect of hexafluoro-1,25-dihydroxyvitamin D₃ and sodium butyrate combination on differentiation and proliferation of HL-60 leukemia cells. *Anticancer Res* 12: 1947-1952, 1992.
- 103 Yu H, Yuan Y, Shen H *et al*: Hematopoietic stem cell exhaustion impacted by p18INK4C and p21^{Cip1/Waf1} in opposite manners. *Blood* 107: 1200-1206, 2006.
- 104 Zhuang SH, Schwartz GG, Cameron D and Burnstein KL: Vitamin D receptor content and transcriptional activity do not fully predict antiproliferative effects of vitamin D in human prostate cancer cell lines. *Mol Cell Endocrinol* 126: 83-90, 1997.
- 105 Zinser G, Packman K and Welsh J: Vitamin D(3) receptor ablation alters mammary gland morphogenesis. *Development* 129: 3067-3076, 2002.
- 106 Zinser GM and Welsh J: Vitamin D receptor status alters mammary gland morphology and tumorigenesis in MMTV-neu mice. *Carcinogenesis* 25: 2361-2372, 2004.
- 107 Zinser GM and Welsh J: Accelerated mammary gland development during pregnancy and delayed postlactational involution in vitamin D₃ receptor null mice. *Mol Endocrinol* 18: 2208-2223, 2004.

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