Review

Utilization of Macrophages in Anticancer Therapy: The Macrophage Network Theory

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Abstract. Appropriate and rational modulation of innate immunity may enhance the therapeutic efficacy of emerging immune therapies for treating cancer. One of the crucial cells of innate immunity is the macrophage. The purpose of this article was to review those issues that suggest ways of exploiting macrophage local functions in immune therapy, and to discuss the suitability of low molecular-weight lipopolysaccharides as potent modulators of macrophage functions for immune therapy of cancer.

Development of immune therapy

At present, therapeutic interventions for various types of cancer are divided into four categories, namely: (1) surgical operation, (2) chemotherapy, (3) radiation therapy, and (4) immune therapy. Immune therapy, especially focusing on the acquired properties of the immune system, is the newest antitumor therapy.

It is well established *in vivo* that tumor cells are foreign targets of the immune system. During the last decade, several tumor-associated antigens (TAA) such as MAGE for melanoma, HER-2/neu for breast cancer and CEA for colorectal cancer have been identified(1-3). Currently, a promising hypothesis suggests that certain tumor cells can

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be eliminated with immune therapies that use antigens to generate cytotoxic T lymphocytes (CTL)(4, 5). The following immune therapies are being developed to target a number of cancers (Figure 1). (i) Lymphokine-activated killer cells (LAK), CTL, or tumor infiltrating lymphocytes (TIL) are passively administered to the patient's body after activation in vitro by stimulation with IL-2 and/or TAA (6). (ii) Dendritic cells (DCs) belong to the myeloid lineage and possess the strongest antigen presentation activity to helper T cells. They have been incorporated in therapies with the purpose of generating CTL more efficiently (7, 8). (iii) Vaccine therapy using killed tumor cells or TAA as a vaccine has also been tested for generating CTL endogenously in a patient's body (9, 10). Each technique augments a specific tumor-killing activity through proper activation of adaptive immunity. Other mechanisms are often combined to kill tumor cells and/or to maximize the antitumor effects of a host immune system. For example, (iv) biological response modifiers (BRMs) are used to activate the patient's immune system, which might be crucial as an adjuvant(11). (v) The drug delivery system, such as a missile therapy in which a chemotherapeutic drug is conjugated or encapsuled with a suitable antibody against TAAs, might be crucial for the reduction of side-effects(12).

These immune therapies are now combined with geneengineering techniques which make it possible to express TAAs or major histocompatiblity antigen classI (MHC classI) or accessory molecules in patient's tumor cells, or to express cytokines by possible effector cells *in vivo* and/or *ex vivo* (13-15). These advances in immune therapy are significant and suggest that future successes in establishing new therapeutic options will complement chemotherapy or radiation therapy in cancer.

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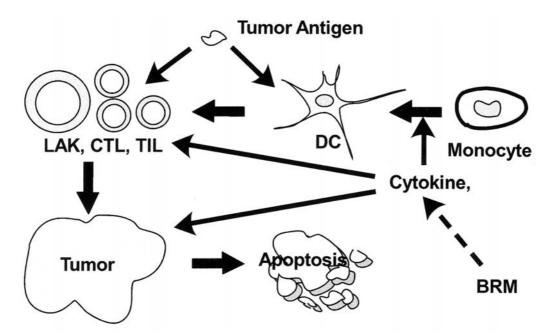


Figure 1. Scheme for immune therapy based on acquired-immunity principles. (1) LAK, CTL, or TIL are passively administered to the patient's body after activation in vitro by stimulation with cytokine and/or TAA. (2) DC cells possess strong antigen presentation activity. (3) Vaccine therapy using killed tumor cells or TAA as a vaccine for generating CTL. (4) BRMs and cytokine are also used to activate the patient's immune system.

However, in clinical settings, the efficacy of the immune therapies described above are still limited. There may be a number of factors that contribute to the discrepancy in the efficacy of immune therapy between clinical settings and model systems (including *in vivo* and *in vitro* systems). A major factor appears to be that, after escaping the host immune surveillance, the antigenecity of tumor cells is not necessarily high. It has been shown that most tumor cells lack MHC classI molecules, which is essential for inducing CTL(16). This might be a reason why experimental results focusing on adaptive immunity are not particularly effective in clinical settings.

Macrophages in immune therapy

The molecular basis of innate immunity is being elucidated by recent progress in immunology. The significance of innate immunity, either by itself and/or in combination with adaptive immunity, has encouraged the re-evaluation of innate immunity (17).

Macrophages are classified into the major cell types involved in the innate immune system. Until recently, manipulation of macrophages has not been considered important for immune therapies.

Macrophages have pleiotropic immune regulatory functions. The main functions are: (i) phagocytosis of foreign molecules or damaged cells, (ii) activation of the acquired immune system by antigen presentation, and (iii) secretion of multiple cytokines to act as regulators and effectors in immune responses (18, 19). Thus, macrophages are involved in almost all aspects of immune therapy. Also, macrophages are found in or around almost all tumors, although the role of tumor-associated macrophages is still controversial (20). Consequently, if it were possible to activate appropriate functions of the macrophages at a local tumor site, it would provide a new strategy for immunotherapy that is based not only on adaptive immunity but also on another modality.

To fully and appropriately utilize the functions of macrophages locally at tumor sites, it is necessary to understand the tissue-specific characteristics of macrophages, and the regulatory mechanisms involved in the recognition and elimination of the tumor cells.

Tissue specificity of macrophages

Macrophages exist in a variety of tissues as tissue-specific macrophages (18). They are called microglias in brain, alveolar macrophages in lung, intestinal macrophages in intestine, Kupffer cells in liver, and so on (Figure 2). Of the tissue macrophages, it is the alveolar macrophages and intestinal macrophages that are exposed directly to the external environment. However, the response of each type of macrophage to lipopolysaccharide (LPS) can be quite

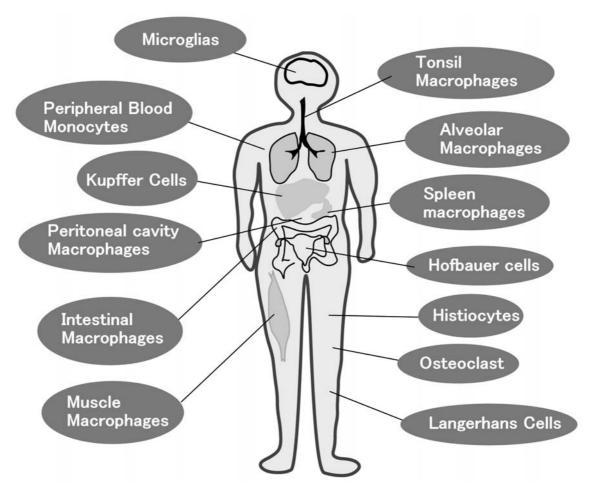


Figure 2. Distribution of macrophages. Tissue-specific macrophages exist in a variety of tissues.

different. We have recently examined the production of tumor necrosis factor (TNF) and nitric oxide (NO) by rat intestinal macrophages, alveolar macrophages and peritoneal macrophages in response to LPS. Production of TNF and NO was not observed in intestinal macrophages even with a high concentration of LPS (100 μ g/ml). On the other hand, alveolar macrophages responded to LPS and produced both molecules. However, the amount of NO produced by alveolar macrophages is lower than the amount produced by peritoneal macrophages (manuscript in preparation). These results clearly show that responses to external stimuli, as exemplified by LPS, are not identical for each type of macrophage. Thus, the tissue specificity of macrophages must be considered when they are utilized for the purpose of regressing tumors.

Recently, there has been remarkable progress in the study of the recognition of foreign substances, namely the pathogen-associated molecular pattern (PAMP) receptors (21). It has been shown that LPS provokes a cellular

response by attaching to toll-like receptors (TLR) 4 (one of the PAMP receptors), which is followed by clustering of several cytoplasmic adaptor molecules and, finally, by induction of activation of transcription factors NF-kB and/or IRF3(22). Inductions of TNF and iNOS, which is an enzyme catalyzing the oxidative deamination of L-arginine to produce NO, depends on NF-kB activation(23).

We examined the expression of the mRNAs of several receptors and adaptor molecules that are involved with LPS responses in intestinal macrophages, alveolar macrophages and peritoneal macrophages by real-time PCR to determine the regulatory mechanism in which TNF and NO are produced in response to LPS. The level of expression of the mRNAs corresponding to each molecule is different for each type of tissue macrophage. This differential expression of mRNAs is relevant to the signal transduction of LPS, and may be partly responsible for the different LPS-susceptibilities for each type of tissue macrophage. However, the expression profile of these mRNAs was not

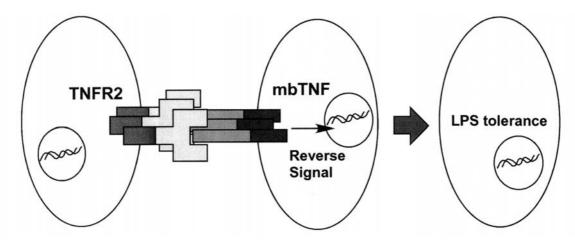


Figure 3. Induction of an LPS-tolerance state in macrophages using a reverse-signal mechanism. The state of LPS tolerance or priming in macrophages can be modified by cell-to-cell contact. The crosslink of membrane-bound TNF reverse signal with TNFR2 leads to an LPS-tolerance state in THP-1 cells.

necessarily coordinated with the expression of the effector molecules such as TNF and NO, for each type of macrophage. In some instances, the mRNA expression levels of TLR4, CD14, Myd88 and TRAF6 (which are all considered to be molecules essential to signal transduction from LPS) in intestinal macrophages are comparable or higher than the expression by alveolar macrophages. This suggests that other regulatory mechanisms may be involved in the production of TNF and NO, and that the mechanisms might also be regulated differently for each type of tissue macrophage (manuscript in preparation). These results clearly show that tissue macrophages have different characteristics depending on the tissue where they exist.

Acquired functional diversity of macrophages

As described above, the responses of macrophages to LPS are different depending on tissue type. In addition, macrophages exposed to LPS show reduced responses to a second stimulation with LPS, which is termed "LPS tolerance" (24). On the other hand, IFN-γ pretreatment appears to augment TNF production by macrophages in response to LPS, although IFN-γ by itself is not capable of stimulating TNF production. This phenomenon is termed "priming" (25).

Interestingly, LPS tolerance or priming could be modified by cell-to-cell contact. We tested this phenomenon using THP-1 cells, a human acute-monocytic-leukemia cell line. THP-1 cells grow in suspension. After co-culture with cells adherently grown, THP-1 cells were re-collected and stimulated with LPS. When, THP-1 cells were co-cultured with NIH 3T3 cells, the production of TNF in response to LPS was repressed. On the other hand, when THP-1 cells were co-cultured with COS-1 cells, the production of TNF

in response to LPS was enhanced. These results suggest that macrophages could be induced into opposite states, LPS tolerance or priming, by cell-to-cell contact. This phenomenon is of interest, because this functional diversity is acquired by contiguous substances, *i.e.* self and non-self.

Although the molecular mechanism for LPS tolerance and priming are not fully understood, down- and upregulation of receptors and/or adaptor molecules for LPS signal transduction appeared to be involved (26-29).

On the other hand, a "reverse signal" mechanism may be involved in the induction of LPS tolerance by cell-to-cell contact. We first mentioned the reverse signal, by which TNFR could act as a ligand to transmit some signals through membrane-bound TNF (30-34). THP-1 cells co-cultured with NIH3T3 cells up-regulate TNFR2 expression, but not THP-1 cells co-cultured with COS-1 cells. Between the two TNF receptors, TNFR2 is known to be the principle receptor to bind membrane-bound TNF (35). Our preliminary experiment suggested that crosslinking of membrane-bound TNF with TNFR2 led to the LPS tolerance state in THP-1 cells (manuscript in preparation). Thus, there is a possibility that TNFR2 might be involved in the reductive expression of TNF through a reverse-signal mechanism (Figure 3).

Both LPS tolerance and priming are phenomena reflecting elaborative regulation of immune and inflammatory processes *in vivo*. An understanding of the mechanism would facilitate the use of macrophage functions in immune therapy.

Suitability of lipopolysaccharide as a potent modulator of macrophage function

One of the strongest activators for macrophages is LPS (36, 37), although there are exceptional cases as observed for

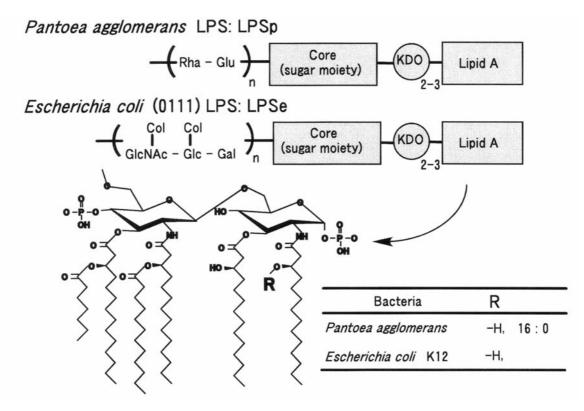


Figure 4. Structure of LPSp and LPSe. LPSp is distinguished from LPSe by (1) the number of fatty acid chains in the lipid A moiety, (2) component of sugar moiety, and (3) the average molecular weight.

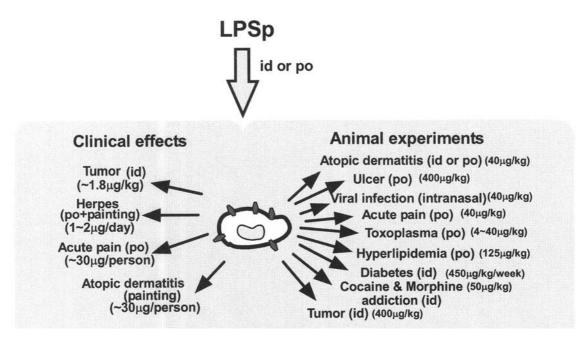


Figure 5. Therapeutic effects of LPSp. LPSp appeared to have therapeutic effects on various diseases in the clinical settings as well as in animal models, when administered orally or intradermally.

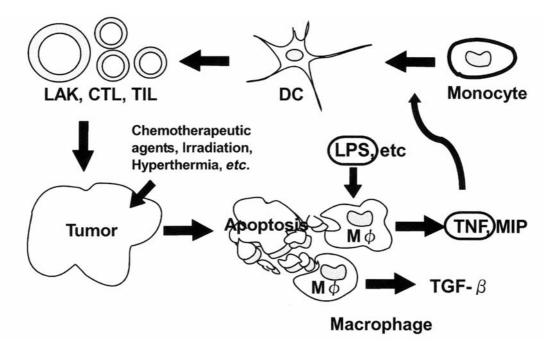


Figure 6. Importance of macrophages in immune therapy. Macrophages stimulated with LPS during phagocytosis of apoptotic bodies induced by chemotherapy or radiation therapy, secrete cytokines such as TNF or MIP. TNF promotes differentiation of monocytes to dendritic cells which have strong antigen presentation ability. As a result, the acquired immune system can be efficiently invoked specifically around tumor lesions.

intestinal macrophages (38). We discovered that LPS derived from Pantoea agglomerans (LPSp) has an interesting feature related to macrophage activation when compared to standard LPS obtained from E.coli (LPSe) (39). P. agglomerans is a Gram-negative symbiotic bacteria isolated from wheat, fruit and other plants (39-41). LPSp is distinguished from LPSe in the following ways: (i) number of fatty acid chains in lipid A moiety: seven in LPSp vs. six in LPSe, (ii) component of sugar moiety; combination with rhamnose and glucose is the unit in LPSp vs. combination with N-acetylglucosamine, glucose, galactose and 2 colitoses is the unit in LPSe; (iii) the average molecular weight of LPSp is about 5kDa, which is smaller than that of LPSe, which is about 20kDa (Figure 4) (42). Each of these physical differences in LPSp would be reflected in the biological characteristics of LPSp and would explain how it functions differently than LPSe.

Although LPS is known to be highly toxic when it is in the blood stream, it was not toxic when it was administered orally, percutaneously, or intradermally. We have tested the therapeutic effects of LPSp administered orally or intradermally on various diseases in clinical settings as well as in animal models. It has been shown that LPSp appeared to have therapeutic effects on various diseases including cancer (Figure 5) (39, 43-57). Depending on the usage, LPS

can be a useful potent drug because it activates macrophages that regulate the immune system. Of the various types of LPS, it appears that LPSp, with its small molecular weight, might have an advantage in the activation of mucosal macrophages through oral or intradermal administration.

In our preliminary testing using an *in vitro* model, our results show a unique therapeutic potential for LPSp compared to that of LPSe. An alveolar macrophage cell line, NR8383, was treated either by LPSp or LPSe, followed by coculture with A549, a human lung adenocarcinoma cell line, or SLC, a rat lung carcinoma cell line. After a 4-h treatment with LPSp or LPSe, NR8383 was co-cultured with A549 or SLC for 24h, and apoptosis of lung carcinoma cells was measured by the secretion of lactate dehydrogenase. Cytotoxic activity to A549 or SLC was significantly augmented by treatment with LPSp compared to that with LPSe.

Usefulness of macrophage activation in immune therapy

Activation of macrophages may affect not only tumor cells but also host cells *in vivo*. Luca *et al.* (58) demonstrated that signals generated by apoptotic cells are integrated with signals from LPS to promote macrophage responses that are

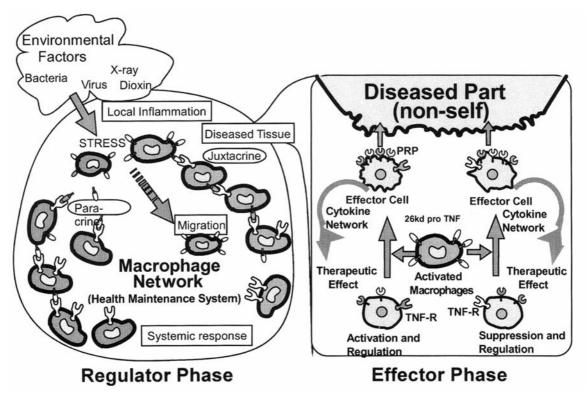


Figure 7. Macrophage network. Initially, tissue macrophages respond to external environmental stimuli and can transmit this information throughout the body by paracrine, juxtacrine, through production of cytokines, and/or by macrophage migration. The formation of the macrophage network functions to maintain homeostasis in the body. Macrophages also function for host defenses in diseased tissues both directly and indirectly.

qualitatively different from responses to stimuli given individually. Additionally, Ohno et al. (59) demonstrated that the number of macrophages that migrated into a tumor mass was a good prognosis factor in gastric cancer or in uterine cancer(60). Taking this data into consideration, we hypothesize that macrophage activation would benefit immune therapy (see Figure 6). Both chemotherapy and radiation therapy partially induce apoptosis in tumor cells, and the destroyed tumor cells (apoptotic bodies) are phagocytosed by macrophages. Usually, macrophages that are involved in phagocytosing apoptotic bodies secrete TGF-β and this causes the inflammatory process to proceed to down-regulation(61). However, macrophages stimulated with pathogen-associated molecules, such as LPS, during phagocytosis, secrete cytokines such as TNF or macrophageinflammatory protein (MIP); these chemicals are not secreted when only phagocytosis has occurred (58). Among the cytokines, TNF has been reported to promote differentiation of monocytes to DCs (62). The DCs show the strongest antigen-presenting function. Thus, these cells are now the focus of attention in finding ways to establish

immunotherapy in conjunction with adaptive immunity. For example, if the macrophages around and/or in tumor lesions are simultaneously manipulated to secrete TNF through PAMP receptors by LPSp, the acquired immune system could be efficiently invoked specifically around tumor lesions. Previously, we had reported that a combination of cyclophosphamide (CY), a chemotherapeutic drug, along with LPSp induced an inflammatory state around tumor lesions, and effectively induced CTL (63). Although, CY is not known to bind PAMP receptors, the drug can induce TNF specifically around tumor lesions (64). Also, CY could dramatically augment the antitumor effects of endogenous TNFinducing therapy, which is one of the immune therapies (65). Our results can be adequately explained by this hypothesis. Thus, in developing an immune therapy for cancer, local macrophages could play a pivotal role. In particular, the TNF secreted from activated macrophages appears to be very important. However, a method for local application of activated macrophages is still missing. For this, a suitable drug delivery system will be required.

Macrophage network

The innate immune system has an important role in the maintenance of homeostasis in the body. One aspect of homeostasis is adapting to the external environment. Macrophages, especially mucosal macrophages, initially respond to the external environmental stimuli and transmit this information throughout the body, by paracrine, juxtacrine, through production of cytokines, and/or migration of the macrophages (Figure 7) (18). This action of macrophages seems to be pivotal in the maintenance of homeostasis. We can tentatively call the global regulation in the body by the tissue macrophages a "macrophage network." Understanding the molecules and mechanisms involved in this network would help to explain the basic regulatory mechanisms that maintain homeostasis. Macrophages are also well-known to function for host defense in diseased tissues directly and indirectly (66, 67). Thus, the utilization of macrophage regulator and effector functions as targets of immune therapy would open new strategies against cancer.

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References

- 1 Kuroki M, Shibaguchi H, Imakiire T, Uno K, Shirota K, Higuchi T, Shitama T, Yamada H, Hirose Y and Nagata A: Immunotherapy and gene therapy of cancer using antibodies or their genes against tumor-associated antigens. Anticancer Res 23: 4377-4381, 2003.
- 2 Mami-Chouaib F, Echchakir H, Dorothee G, Vergnon I and Chouaib S: Antitumor cytotoxic T-lymphocyte response in human lung carcinoma: identification of a tumor-associated antigen. Immunol Rev 188: 114-121, 2002.
- 3 Yoshino I, Goedegebuure PS, Peoples GE, Parikh AS, DiMaio JM, Lyerly HK, Gazdar AF and Eberlein TJ: HER2/neu-derived peptides are shared antigens among human non-small cell lung cancer and ovarian cancer. Cancer Res 54: 3387-3390, 1994.
- 4 Boon T, Coulie PG and Van den Eynde B: Tumor antigens recognized by T cells. Immunol Today *18*: 267-268, 1997.
- 5 Van den Eynde BJ and van der Bruggen P: T cell defined tumor antigens. Curr Opin Immunol 9: 684-693, 1997.
- 6 Hoffman DM, Gitlitz BJ, Belldegrun A and Figlin RA: Adoptive cellular therapy. Semin Oncol 27: 221-233, 2000.
- 7 Conrad C and Nestle FO: Dendritic cell-based cancer therapy. Curr Opin Mol Ther 5: 405-412, 2003.
- 8 Wiethe C, Dittmar K, Doan T, Lindenmaier W and Tindle R: Provision of 4-1BB ligand enhances effector and memory CTL responses generated by immunization with dendritic cells expressing a human tumor-associated antigen. J Immunol 170: 2912-2922, 2003.

- 9 Rosenberg SA: Cancer vaccines based on the identification of genes encoding cancer regression antigens. Immunol Today 18: 175-182, 1997.
- 10 Gitlitz BJ, Belldegrun AS and Figlin RA: Vaccine and gene therapy of renal cell carcinoma. Semin Urol Oncol 19: 141-147, 2001.
- 11 Wimer BM: The ideal biological response modifier. Mol Biother 1: 311-317, 1989.
- 12 Palapattu GS and Reiter RE: Monoclonal antibody therapy for genitourinary oncology: promise for the future. J Urol *168*: 2615-2623, 2002.
- 13 Haupt K, Roggendorf M and Mann K: The potential of DNA vaccination against tumor-associated antigens for antitumor therapy. Exp Biol Med (Maywood) 227: 227-237, 2002.
- 14 Hodge JW, Grosenbach DW and Schlom J: Vector-based delivery of tumor-associated antigens and T-cell co-stimulatory molecules in the induction of immune responses and anti-tumor immunity. Cancer Detect Prev 26: 275-291, 2002.
- 15 Zhang L, Tang Y, Akbulut H, Zelterman D, Linton PJ and Deisseroth AB: An adenoviral vector cancer vaccine that delivers a tumor-associated antigen/CD40-ligand fusion protein to dendritic cells. Proc Natl Acad Sci USA 100: 15101-15106, 2003.
- 16 Chang CC, Campoli M and Ferrone S: HLA class I defects in malignant lesions: what have we learned? Keio J Med 52: 220-229, 2003.
- 17 Vercelli D: Innate immunity: sensing the environment and regulating the regulators. Curr Opin Allergy Clin Immunol 3: 343-346, 2003.
- 18 Ross JA and Auger MJ: The biology of the macrophage. *In*: The Macrophage. (Burke B and Lewis EC, eds). New York, Oxford University Press Inc. 2002, pp 1-57,
- 19 Paglia P and Colombo MP: Macrophage as antigen-presenting cells: relationship to dendritic cells and use in vaccination studies. *In*: The Macrophage. (Burke B and Lewis EC, eds). New York, Oxford University Press Inc. 2002, pp 103-125.
- 20 Ohno S, Suzuki N, Ohno Y, Inagawa H, Soma G and Inoue M: Tumor-associated macrophages: foe or accomplice of tumors? Anticancer Res 23: 4395-4409, 2003.
- 21 Medzhitov R and Janeway C Jr: The Toll receptor family and microbial recognition. Trends Microbiol 8: 452-456, 2000.
- 22 Takeda K and Akira S: TLR signaling pathways. Semin Immunol *16*: 3-9, 2004.
- 23 Mungrue IN, Bredt DS, Stewart DJ and Husain M: From molecules to mammals: what's NOS got to do with it? Acta Physiol Scand *179*: 123-135, 2003.
- 24 Ziegler-Heitbrock HW: Molecular mechanism in tolerance to lipopolysaccharide. J Inflamm *45*: 13-26, 1995.
- 25 Beutler B, Tkacenko V, Milsark I, Krochin N and Cerami A: Effect of gamma interferon on cachectin expression by mononuclear phagocytes. Reversal of the lpsd (endotoxin resistance) phenotype. J Exp Med 164: 1791-1796, 1986.
- 26 Mita Y, Dobashi K, Shimizu Y, Nakazawa T and Mori M: Toll-like receptor 2 and 4 surface expressions on human monocytes are modulated by interferon-gamma and macrophage colony-stimulating factor. Immunol Lett 78: 97-101, 2001.
- 27 Suzuki M, Hisamatsu T and Podolsky DK: Gamma interferon augments the intracellular pathway for lipopolysaccharide (LPS) recognition in human intestinal epithelial cells through coordinated up-regulation of LPS uptake and expression of the intracellular Toll-like receptor 4-MD-2 complex. Infect Immun 71: 3503-3511, 2003.

- 28 Tamai R, Sugawara S, Takeuchi O, Akira S and Takada H: Synergistic effects of lipopolysaccharide and interferon-gamma in inducing interleukin-8 production in human monocytic THP-1 cells is accompanied by up-regulation of CD14, Toll-like receptor 4, MD-2 and MyD88 expression. J Endotoxin Res 9: 145-153, 2003.
- 29 Nomura F, Akashi S, Sakao Y, Sato S, Kawai T, Matsumoto M, Nakanishi K, Kimoto M, Miyake K, Takeda K and Akira S: Cutting edge: endotoxin tolerance in mouse peritoneal macrophages correlates with down-regulation of surface toll-like receptor 4 expression. J Immunol 164: 3476-3479, 2000.
- 30 Tanabe Y, Kitahara-Tanabe N, Mizuno D and Soma GI: Enhanced production of tumour necrosis factor alpha (TNFalpha) by its precursor on the cell surface of primed THP-1 cells. Cytokine 6: 337-348, 1994.
- 31 Soma IG, Nishizawa T, Inagawa H, Tanabe Y, Noguchi K, Goto S, Takagi K and Mizuno D: Bidirectional feedback regulation on 17 kD tumor necrosis factor (TNF) production by 26 kD membrane-bound TNF precursor. J Inflamm 47: 52-60, 1995.
- 32 Tanabe Y, Kohchi C, Kitahara-Tanabe N, Mizuno D and Soma G: Involvement of 26-kDa membrane-bound tumour necrosis factor precursor in bidirectional feedback regulation on 17-kDa tumour necrosis factor production after stimulation by lipopolysaccharide. Cytokine 10: 82-92, 1998.
- 33 Watts AD, Hunt NH, Wanigasekara Y, Bloomfield G, Wallach D, Roufogalis BD and Chaudhri G: A casein kinase I motif present in the cytoplasmic domain of members of the tumour necrosis factor ligand family is implicated in 'reverse signalling'. Embo J 18: 2119-2126, 1999.
- 34 Domonkos A, Udvardy A, Laszlo L, Nagy T and Duda E: Receptor-like properties of the 26 kDa transmembrane form of TNF. Eur Cytokine Netw 12: 411-419, 2001.
- 35 Grell M, Douni E, Wajant H, Lohden M, Clauss M, Maxeiner B, Georgopoulos S, Lesslauer W, Kollias G, Pfizenmaier K et al: The transmembrane form of tumor necrosis factor is the prime activating ligand of the 80 kDa tumor necrosis factor receptor. Cell 83: 793-802, 1995.
- 36 Guha M and Mackman N: LPS induction of gene expression in human monocytes. Cell Signal 13: 85-94, 2001.
- 37 Bancroft GJ, Kelly JP, Kaye PM, McDonald V and Cross CE: Pathways of macrophage activation and innate immunity. Immunol Lett 43: 67-70, 1994.
- 38 Smith PD, Smythies LE, Mosteller-Barnum M, Sibley DA, Russell MW, Merger M, Sellers MT, Orenstein JM, Shimada T, Graham MF and Kubagawa H: Intestinal macrophages lack CD14 and CD89 and consequently are down-regulated for LPS- and IgA-mediated activities. J Immunol 167: 2651-2656, 2001.
- 39 Nishizawa T, Inagawa H, Oshima H, Okutomi T, Tsukioka D, Iguchi M, Soma G and Mizuno D: Homeostasis as regulated by activated macrophage. I. Lipopolysaccharide (LPS) from wheat flour: isolation, purification and some biological activities. Chem Pharm Bull (Tokyo) 40: 479-483, 1992.
- 40 Asis CA Jr and Adachi K: Isolation of endophytic diazotroph Pantoea agglomerans and nondiazotroph Enterobacter asburiae from sweetpotato stem in Japan. Lett Appl Microbiol 38: 19-23, 2004.
- 41 Nunes C, Usall J, Teixido N, Fons E and Vinas I: Post-harvest biological control by *Pantoea agglomerans* (CPA-2) on Golden Delicious apples. J Appl Microbiol 92: 247-255, 2002.

- 42 Tsukioka D, Nishizawa T, Miyase T, Achiwa K, Suda T, Soma G and Mizuno D: Structural characterization of lipid A obtained from *Pantoea agglomerans* lipopolysaccharide. FEMS Microbiol Lett 149: 239-244, 1997.
- 43 Inagawa H, Saitoh F, Iguchi M, Nishizawa T, Okutomi T, Morikawa A, Soma GI and Mizuno D: Homeostasis as regulated by activated macrophage. III. Protective effect of LPSw (lipopolysaccharide (LPS) of wheat flour) on gastric ulcer in mice as compared with those of other LPS from various sources. Chem Pharm Bull (Tokyo) 40: 998-1000, 1992.
- 44 Okutomi T, Nishizawa T, Inagawa H, Morikawa A, Takeuchi S, Soma GI and Mizuno D: Homeostasis as regulated by activated macrophage. IV. Analgesic effect of LPSw, a lipopolysaccharide of wheat flour. Chem Pharm Bull (Tokyo) 40: 1001-1003, 1992.
- 45 Iguchi M, Inagawa H, Nishizawa T, Okutomi T, Morikawa A, Soma GI and Mizuno D: Homeostasis as regulated by activated macrophage. V. Suppression of diabetes mellitus in non-obese diabetic mice by LPSw (a lipopolysaccharide from wheat flour). Chem Pharm Bull (Tokyo) 40: 1004-1006, 1992.
- 46 Suzuki Y, Kobayashi A, Nishizawa T, Inagawa H, Morikawa A, Soma G and Mizuno D: Homeostasis as regulated by activated macrophage. VI. Protective effect of LPSw (a lipopolysaccharide from wheat flour) against acute infection by *Toxoplasma gondii* in mice. Chem Pharm Bull (Tokyo) 40: 1266-1267, 1992.
- 47 Okutomi T, Nishizawa T, Inagawa H, Takano T, Morikawa A, Soma G and Mizuno D: Homeostasis as regulated by activated macrophage. VII. Suppression of serum cholesterol level by LPSw (a lipopolysaccharide from wheat flour) in WHHL (Watanabe heritable hyperlipidemic) rabbit. Chem Pharm Bull (Tokyo) 40: 1268-1270, 1992.
- 48 Kawashima K, Endo H, Nishizawa T, Inagawa H, Okutomi T, Morikawa A, Soma G and Mizuno D: Homeostasis as regulated by activated macrophage. VIII. LPSw (a lipopolysaccharide from wheat flour) can regulate bone resorption of chick embryo. Chem Pharm Bull (Tokyo) 40: 1271-1273, 1992.
- 49 Suzuki J, Nishizawa T, Inagawa H, Okutomi T, Morikawa A, Soma G and Mizuno D: Homeostasis as regulated by activated macrophage. IX. Enhancement effect of LPSw (a lipopolysaccharide from wheat flour) on hen egg-laying and breaking strength of eggshell. Chem Pharm Bull (Tokyo) 40: 1274-1276, 1992.
- 50 Okutomi T, Nishizawa T, Inagawa H, Soma G, Minami M, Satoh M and Mizuno D: Inhibition of morphine dependence by a lipopolysaccharide from *Pantoea agglomerans*. Eur Cytokine Netw 3: 417-420, 1992.
- 51 Mizuno D and Soma G: Oral or percutaneous administration of lipopolysaccharide of small molecular size may cure various intractable diseases: a new version of Coley's toxin. Mol Biother 4: 166-169, 1992.
- 52 Suzuki T, Funada M, Sugano Y, Misawa M, Okutomi T, Soma G and Mizuno D: Effects of a lipopolysaccharide from *Pantoea* agglomerans on the cocaine-induced place preference. Life Sci 54: PL75-80, 1994.
- 53 Kamei J, Iwamoto Y, Suzuki T, Misawa M, Kasuya Y, Nagase H, Okutomi T, Soma G and Mizuno D: Antinociceptive effect of lipopolysaccharide from *Pantoea agglomerans* on streptozotocininduced diabetic mice. Eur J Pharmacol 251: 95-98, 1994.
- 54 Kamei J, Iwamoto Y, Suzuki T, Misawa M, Kasuya Y, Okutomi T, Soma G and Mizuno D: Suppression of hyperalgesia in streptozotocin-induced diabetic mice by a lipopolysaccharide from *Pantoea agglomerans*. Biol Pharm Bull 17: 632-634, 1994.

- 55 Inagawa H, Nishizawa T, Noguchi K, Minamimura M, Takagi K, Goto S, Soma G and Mizuno D: Anti-tumor effect of lipopolysaccharide by intradermal administration as a novel drug delivery system. Anticancer Res 17: 2153-2158, 1997.
- 56 Iwamoto I, Goto S, Kera J, Soma G, Takeuchi S and Nagata Y: Mechanistic analysis of high antitumor effect of intradermal administration of lipopolysaccharide from *Pantoea agglomerans*. Med Oncol 13: 103-109, 1996.
- 57 Goto S, Sakai S, Kera J, Suma Y, Soma GI and Takeuchi S: Intradermal administration of lipopolysaccharide in treatment of human cancer. Cancer Immunol Immunother 42: 255-261, 1996.
- 58 Lucas M, Stuart LM, Savill J and Lacy-Hulbert A: Apoptotic cells and innate immune stimuli combine to regulate macrophage cytokine secretion. J Immunol 171: 2610-2615, 2003.
- 59 Ohno S, Inagawa H, Dhar DK, Fujii T, Ueda S, Tachibana M, Suzuki N, Inoue M, Soma G and Nagasue N: The degree of macrophage infiltration into the cancer cell nest is a significant predictor of survival in gastric cancer patients. Anticancer Res 23: 5015-5022, 2003.
- 60 Ohno S, Ohno Y, Suzuki N, Inagawa H, Kohchi C, Soma G-I and Inoue M: Functional differences depending on histological localization of tumor-associated macrophages in endometrial cancer. Anticancer Res 24: in press, 2004.
- 61 Voll RE, Herrmann M, Roth EA, Stach C, Kalden JR and Girkontaite I: Immunosuppressive effects of apoptotic cells. Nature 390: 350-351, 1997.
- 62 Chomarat P, Dantin C, Bennett L, Banchereau J and Palucka AK: TNF skews monocyte differentiation from macrophages to dendritic cells. J Immunol 171: 2262-2269, 2003.

- 63 Inagawa H, Ohshiro S, Nishizawa T, Goto S, Soma G and Mizuno D: Augmentation of antitumor effect of endogenously induced tumor necrosis factor by cyclophosphamide. Anticancer Res 17: 55-60, 1997.
- 64 Inagawa H, Nishizawa T, Honda T, Nakamoto T, Takagi K and Soma G: Mechanisms by which chemotherapeutic agents augment the antitumor effects of tumor necrosis factor: involvement of the pattern shift of cytokines from Th2 to Th1 in tumor lesions. Anticancer Res 18: 3957-3964, 1998.
- 65 Takiguchi K, Nakamoto T, Inagawa H, Kohchi C, Nishizawa T, Nagasue N and Soma G-I: Profile of cytokine produced in tumor tissue after administration of cyclophosphamide in a combination therapy with tumor necrosis factor. Anticancer Res 24: 1823-1828, 2004.
- 66 Heale J-P and Speert DP: Macrophages in bacterial infection. In: The Macrophage. (Burke B and Lewis CE, eds). New York, Oxford University Inc. 2002, pp 212-237.
- 67 Kreutz M, Fritsche J and Andreesen R: Macrophage in tumor biology. *In*: The Macrophage. (Burke B and Lewis CE, eds).New York, Oxford University Press Inc. 2002, pp 457-474.

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