

Review

***H19* in Endocrine System Tumours**

MAŁGORZATA ROLLA, ALEKSANDRA JAWIARCZYK-PRZYBYŁOWSKA,
KATARZYNA KOLAČKOV and MAREK BOLANOWSKI

Department of Endocrinology, Diabetes and Isotope Therapy, Wrocław Medical University, Wrocław, Poland

Abstract. Long non-coding RNAs (lncRNAs) are over 200 nucleotides long recently discovered RNA molecules that are not involved in the translation process. Accumulating evidence shows that *H19* lncRNA is an important regulator of gene expression and its altered expression contributes to carcinogenesis. The aim of this review was to reveal current knowledge about *H19* lncRNA and its impact on tumours of the endocrine system. We present findings about *H19* altered regulation and its association with tumorigenesis, cancer progression and differentiation, and its potential use in diagnostics, prognostics and therapy. The mechanism and molecular pathways involved in these processes are discussed.

Non-coding RNAs (ncRNAs) are recently discovered molecules, which do not participate in the translation process and do not have their own protein product (1-3). Approximately 80% of human genome is transcribed into functional RNA, but less than 2% is involved in translation and has protein-coding capacity (4). Therefore, ncRNAs are an abundant group of transcripts that can be divided according to their length or function. According to their length, we can distinguish them into small ncRNAs (less than 200 nucleotides long) and long non-coding RNAs (lncRNAs) (1-3). ncRNAs are divided according to their function into housekeeping ncRNAs and regulatory ncRNAs (2, 5). Ribosomal (r-), transfer (t-), small nuclear (sn-) and small nucleolar (sno-) ncRNAs are housekeeping, whereas micro (mi-), small interfering (si-), piwi-interacting (pi-) and

long non-coding (lnc-) ncRNAs are regulatory (2, 5). Up to November 2020, over 260,000 types of human lncRNAs had been identified (6). lncRNAs can be located in the nucleus or cytoplasm (5). Their function is still poorly understood, but their biological roles seem to be more crucial than it was initially hypothesized (1, 3, 5). Accumulating evidence shows that lncRNAs are important regulators of gene expression (3). They play roles in regulation and modification of transcription, post-transcription and epigenetic processes (2, 3). Evidence has revealed that they are involved in the development of diabetes (7, 8) and neurological diseases (9-11). Recent studies have shown that aberrant expression of lncRNAs may also contribute to carcinogenesis (2, 3, 12, 13).

***H19* RNA**

H19 lncRNA was the first discovered lncRNA; it was initially classified as an mRNA with unknown protein product and was extracted from a mouse liver (14). A few years later, Brannan *et al.* isolated *H19* gene from human tissues and stated that the only final product of *H19* gene may be an mRNA transcript, located in the cytoplasm (15). The full length of *H19* RNA chain is 2.3 kb (16). In human, the gene is mapped on chromosome 11p15.5 (17). The expression of *H19* is high during embryonic development (14, 18), mainly in the endoderm and mesoderm (19), and maximum expression has been observed in the liver, muscles and adrenals (19, 20). After birth, it is down-regulated in most tissues, but its expression is still detectable in inter alia, skeletal muscle, myocardium and mammary gland tissues (19, 21).

***H19* – Contribution to Carcinogenesis**

The linkage between *H19* and cancer development was the subject of many studies since the 1990s (18, 22, 23). Bartolomei *et al.* first discovered that *H19* is expressed exclusively from the maternal allele, due to the imprinting process (24). Knowledge about the influence of imprinting alterations on carcinogenesis led to search for an association between *H19* gene and its

This article is freely accessible online.

Correspondence to: Małgorzata Rolla, Department of This article is freely accessible online.

Endocrinology, Diabetes and Isotope Therapy, Wrocław Medical University, Wybrzeże L. Pasteura 4, 50-367, Wrocław, Poland.
e-mail: małgorzata.rolla@student.umed.wroc.pl

Key Words: *H19*, lncRNA, pituitary adenoma, thyroid cancer, adrenal tumour, neuroendocrine tumour, review.

potential role in cancer development (18). Probable mechanisms are loss of imprinting (LOI) – an epigenetic event resulting in biallelic gene expression, and changes in the methylation pattern of promoters sequences, which regulate the levels of gene expression (25). It has been proposed that these two mechanisms are strongly related, due to the involvement of methylation in the inactivation of the paternal allele (26). It is important to note that *H19* and *IGF2* genes are commonly imprinted interdependently due to their close location on 11p15.5 (27). Association between LOI of *H19* gene and tumorigenesis was described inter alia, for oesophageal (28), colorectal (28) and lung cancers (29). Additionally, in Wilms' tumour, LOI of *IGF2* gene contributes to methylation of *H19* promoter, resulting in the down-regulation of *H19* expression (30). Nevertheless, LOI does not always directly correspond to a methylation pattern and level of gene expression, as it has been shown by Byun *et al.* in a study on bladder cancer (25). In contrast to the above studies, Yballe *et al.* have shown no connection between LOI of *H19* gene and the occurrence of breast cancer (31). Similar results were obtained for neuroblastoma by Wada *et al.* (32). Further studies have been performed on the mechanisms of *H19* contribution in carcinogenesis. *H19* has been proposed as an oncogene (33, 34), tumour suppressor (35, 36) or as an oncofoetal RNA, associated with germ cell tumours (18, 19, 37).

The oncogenic properties of *H19* may be due to its increased expression in neoplasm tissues. Over-expression of *H19* RNA has been shown to contribute to the carcinogenesis and progression of tumours of the breast (34, 38), lung (39, 40), oesophagus (28, 41), stomach (42-44), colon (28, 45), liver (46), pancreas (47, 48), kidney (49), bladder (46, 50), cervix (51), ovary (52, 53), as well as in glioma (54, 55), leukaemia (56), oral squamous cell carcinoma (57), cholangiocarcinoma (58), osteosarcoma (59, 60) and melanoma (61). On the other hand, in some tumours down-regulation of *H19* expression was observed [inter alia in Wilms' tumour (30, 62)], which means that *H19* may be classified also as a tumour suppressor.

The mechanisms through which *H19* is involved in the process of cancer development include promotion of gene mutations, cell proliferation, invasion, migration and angiogenesis, immune and pro-apoptotic factors modulation and growth suppressor expression regulation (63, 64). Additionally, some studies have shown that *H19* RNA functions through sponging mi-RNAs including miR-675 (45, 48), miR-107 (39, 40), miR-370-3p (65), miR-106a-5p (61), miR-29a (54), miR-29a-3p (49) and miR-138-5p (51).

In recent meta-analyses (64, 66) the prognostic and clinicopathological values of *H19* in different types of cancers were explored. Both studies demonstrated that high levels of *H19* RNA contribute to shorter overall survival and associate with more advanced clinical stage of tumours and lymph node metastasis. Additionally, *H19* RNA positively correlates with poor tumour differentiation, earlier distant

metastasis (64), as well as with poorer histological tumour grade and disease-free survival (66). Summarizing, *H19* RNA has been demonstrated as a potential marker for tumour progression and patient's prognosis.

So far, *H19* has been introduced as an intriguing figure in neoplasms' origin and development. But what is its impact on tumours of the endocrine system?

Pituitary Adenomas

In the study by Lu *et al.*, significantly higher expression of *H19* was observed in aggressive growth hormone-secreting pituitary adenomas compared to non-invasive growth hormone-secreting tumours (67). A similar observation was made for oesophageal cancer (*H19* correlated positively with tumours' depth, stage and metastasis) (41), lung cancer (*H19* enhanced cell proliferation, migration, and invasion of a cell line) (68), glioblastoma (in cell line and xenograft mouse model, *H19* promoted invasion, angiogenesis and tumour growth) (55), cholangiocarcinoma (*H19* positively correlated with tumour size, cell migration and invasiveness in tissues and cell lines) (58). Thus, we could hypothesize that in invasive pituitary adenomas *H19* might be a potential marker of malignancy and patients' prognosis.

On the other hand, Wu *et al.* (69) and Zhang *et al.* (70) observed down-regulation of *H19* expression in pituitary tumour tissues and in the plasma obtained from patients with pituitary adenomas in comparison to normal pituitary glands and healthy controls. In *in vitro* and *in vivo* models, an increase in cell proliferation after knockdown of *H19* gene was observed (69). Furthermore, in mouse models injection of *H19* lentivirus led to shrinkage of tumour volumes. *H19* expression levels negatively correlated with tumour volumes. Antitumor effects were induced by inhibiting 4E-BP1 phosphorylation in the mTORC1/4E-BP1 pathway. Moreover, in xenograft experiments *H19* overexpression was more effective than cabergoline in suppressing tumour growth (69). Additionally, the investigators revealed that cabergoline stimulated *H19* expression and *H19* and dopamine agonists exerted a synergistic therapeutic effect. These results indicate that increasing *H19* expression can be a potential therapy for pituitary adenomas. The mechanism of the synergistic action of *H19* and dopamine agonists in prolactinomas was investigated in a recent study by Wu *et al.* (71). It was revealed that *H19* promotes the effects of dopamine agonists by inhibiting miRNA-93a and stimulating ATG7 expression, and this is another example of *H19* action by sponging mi-RNA. *H19*/miRNA-93a/ATG7 axis was elucidated as a potential target of therapy, especially in drug-resistant prolactinomas.

Opposite results regarding the influence of *H19* on drug resistance, but also describing *H19* impact on ATG7, were demonstrated by Pan *et al.* (72). In non-small cell lung

cancer cell lines and xenograft models, they observed that *H19* sponges miRNA-615-3p and regulates ATG7 expression, and that this mechanism is probably involved in erlotinib resistance.

Thyroid Cancer

Ambiguous associations between *H19* expression and tumour development have also been illustrated for thyroid cancer. In thyroid cancer samples and cell lines, Liu *et al.* (73) observed over-expression of *H19*. *H19* enhanced tumour growth by inhibiting apoptosis and promoting progression, migration and invasion. Moreover, the researchers found that *H19* affects miR-17-5p and antagonizes its effect on YES1 expression. The association between *H19* and miR-17-5p has also been illustrated in gastric cancer cells (74), whereas a positive correlation between the levels of these two RNA was determined. In that study, *H19* was associated with larger tumour size, more advanced TNM stage and lymph node metastases. Corresponding outcomes, but for thyroid cancer, were exemplified in the study by Liu *et al.* (75). Moreover, higher *H19* expression was related with lower 5-year survival rate.

The mechanism through which *H19* contributes to thyroid cancer development was the subject of the studies of Li *et al.* (76) and Wang *et al.* (77). In the Li *et al.* study (76), *H19* was found to function through the PI3K/AKT signalling pathway, which plays an important role in carcinogenesis. Similarly, the association of *H19* with PI3K/AKT was illustrated in colorectal cancer cell lines (78) and melanoma (79). An additional finding of Li *et al.* was over-expression of *H19* in thyroid cancer tissues compared to adjacent healthy thyroid tissues (76). Moreover, *H19* expression was higher in poorly differentiated thyroid cancer tissues. In an *in vitro* model, knockdown of *H19* resulted in cancer cell viability inhibition and induction of apoptosis (76).

In contrary, Wang *et al.* (77) showed that *H19* overexpression inhibits viability, migration and invasion and induces tumour cells apoptosis and these effects might be mediated *via* down-regulating the expression of IRS-I (insulin receptor substrate I). Moreover, IRS-I expression might be induced also by PI3/AKT signalling pathway. The results of Wang *et al.* suggest that *H19* could be potentially used in thyroid cancer treatment.

The *H19* effect on the development of specific types of thyroid cancer was the subject of several studies presented below. For papillary thyroid cancer (PTC), higher tissue expression of *H19* was observed in the studies of Liang *et al.* (80) and Li *et al.* (81). Different mechanisms were proposed for expounding *H19* involvement in PTC development. In the first study, higher expression was positively correlated with mesenchymal phenotype biomarkers (vimentin, ZEB2, Twist, Snail2), which indicates

that *H19* RNA induces epithelial–mesenchymal transition (EMT) process. EMT has been described to play a critical role in cancer invasiveness and metastasis (82). A similar effect of *H19* on EMT was depicted for ovarian (65), oesophageal (41) cancers and cholangiocarcinoma (58). Moreover, in the ovarian cell line, *H19* was shown to promote EMT-related activity and contribute to cisplatin resistance (83). Li *et al.* (81) proposed a mechanism that was related to ER β (oestrogen receptor beta). Oestradiol enhanced *H19* expression by ER β whereas high expression of *H19* promoted expression of ER β (as a positive feedback). Additionally, *H19* acted through miR-3126-5p and this is another example of sponging mi-RNA by *H19*.

In the study by Liang *et al.*, *H19* expression was positively correlated with tumour size and grade, as well as with lymph node metastases (80). The opposite results were obtained by Lan *et al.* (84). Jiao *et al.* (85) observed down-regulation of *H19* in papillary thyroid cancer tissues compared to paracancerous or benign nodes. Additionally lower expression of *H19* coincided with the presence of lymph node metastasis (84, 85), as well as with other features of poorer prognosis, such as higher tumour size, more aggressive histological type and poorer diseases-free survival (85).

For minimally invasive follicular thyroid cancer, Dai *et al.* examined whether *H19* could be a marker of distant metastasis and patients' prognosis (86). The study revealed low expression of *H19* in cancer tissues and *H19* levels were negatively correlated with tumour size, vascular invasion, distant metastasis and poorer overall survival.

Zhang *et al.* demonstrated that *H19* RNA is over-expressed in anaplastic thyroid carcinoma tissues and cell lines (87). Moreover, they showed that reduction of *H19* expression can be a potential target of molecular therapy – it decreased cell proliferation, migration and invasion *in vitro* as well as inhibited tumorigenesis and metastasis *in vivo*.

Alike divergences of *H19* expression levels in different types of thyroid cancer samples were observed by Wächter *et al.* (88). In anaplastic carcinoma, it was upregulated in six cases, down-regulated in two and was similar to healthy thyroid tissue in four. In follicular thyroid cancer, it was down-regulated in five samples and was the same in three cases. In papillary thyroid cancer it was overexpressed in five samples, down-regulated in two and stable in four. Thus, no association was observed between *H19* levels and type of thyroid cancer. In summary, in thyroid cancer, *H19* was found to act both as an oncogene as well as a suppressor.

Adrenals

During embryonic and foetal life adrenal expression of *H19* is very high (89, 90). In adulthood it remains highly expressed – it shows approximately 50% of the foetal

expression (91). Gao *et al.* (92) and Liu *et al.* (91) showed that in benign adrenal adenomas and hyperplastic adrenals, *H19* is expressed at about the same level as in healthy glands. However, similarly to Glover *et al.* they showed that in adrenocortical carcinomas the expression was reduced and it was significantly lower than in normal adrenals (91-93), whereas in pheochromocytomas the expression was variable, but generally decreased (91). Upon further investigation, Liu *et al.* showed that *H19* expression was also decreased in virilizing adrenal adenomas (94). The proposed mechanism causing the low *H19* expression in adrenocortical carcinomas was methylation of the promoter area (92). The degree of methylation of the promoter CpG regions in patients with adrenocortical cancers and adenomas was the subject of the study of Barreau *et al.* (95). The characterized cancers had a higher degree of methylation compared to adenomas that corresponded to patients' poorer prognosis. *H19* was found to be one of the genes with a hypermethylated promoter region leading to its down-regulation. Moreover, it showed the strongest observed inverse correlation between methylation levels and gene expression in this study, leading to a conclusion that *H19* plays a role as a suppressor. A comparable effect of methylation of the *H19* promoter on carcinogenesis was shown for bladder cancer (25) and Wilms' tumour (30). Additionally, Creemers *et al.* (96) proposed that the methylation status of *IGF2* and *H19* regulatory regions as useful markers in distinguishing malignant adrenocortical carcinomas from benign adenomas. Thus, we could conclude that *H19* expression levels and the methylation pattern of its regulatory regions could be promising tools in the diagnosis of adrenal tumours.

In addition, the various degrees of *H19* promoter methylation in benign ovarian teratomas (97) as well as in different types of germ cell tumours (GCTs) (98, 99), illustrated the diversity in origin and processes involved in the development of these neoplasms. Hence, reduced methylation in adrenocortical carcinomas may reflect their primordial features, however, further investigations are needed to evaluate this hypothesis.

Neuroendocrine Tumours

In the Ji *et al.* study, aberrant expression of *H19* was described as an important element in the development of non-functional pancreatic neuroendocrine neoplasms (pNENs) (100). In primary tumours as well as in metastatic tumours, the levels of *H19* expression were variable. However, after evaluation of the association between *H19* and tumour's malignancy, the researchers revealed that non-malignant tumours were characterized by low expression of *H19*, whereas in malignant pNENs as well as in liver metastases its expression was high. Moreover, high expression correlated positively with tumour size, lymph

node and liver metastasis, local invasion, TNM stage, tumour-related death, poorer progression free and overall survival. In the cell line models, the authors showed that silencing *H19* led to inhibition of cell proliferation, growth and colony formation and the opposite effects were observed after *H19* over-expression. Additionally, overexpression of *H19* promoted tumour growth and Ki67 expression in xenograft mouse models. The paper illustrates the possible association between high expression of *H19* and VGF (neuropeptide precursor) in neoplasms origin, progression and poorer patient prognosis. Additionally, similarly to Li *et al.* (76), *H19* was shown to be involved in the activation of PI3K/Akt signalling pathway.

Ramnarine *et al.* showed that *H19* was an epigenetic regulator, which contributed to neuroendocrine transdifferentiation (NETD) – a transformation from prostate cancer to neuroendocrine prostate cancer (101). In addition, high expression of *H19* was presented as a practical tool in distinguishing neuroendocrine prostate cancers from prostate adenocarcinomas.

Conclusion

The aim of this review was to demonstrate the current knowledge about *H19* lncRNA and its impact on tumours of the endocrine system. The collected data shed light on the mechanisms and molecular pathways involved in tumorigenesis. *H19* was determined to be involved in epigenetic regulation and in miRNA expression control. Moreover, *H19* may be a useful factor in differentiating malignancies from benign lesions, as it was demonstrated in aggressive pituitary adenomas (67), adrenocortical carcinomas (91-93) and pNENs (100). Another promising aspect is the down-regulation of *H19* as a purpose of targeted therapy, which was illustrated in cell line models of thyroid cancer (76, 87) and pNENs (100). On the other hand, upregulation of *H19* has also been proposed as a therapeutic tool (69, 70, 77). In addition, *H19* may improve the effects of treatment, like it was illustrated for dopamine agonists in prolactinomas (69, 71). Furthermore, abnormal expression of *H19* RNA in different types of malignancies makes it a potential biomarker for cancer diagnosis, prognosis and monitoring. In some reports correlation between *H19* expression and clinicopathological features was observed, which highlights the prognostic value of this RNA (75, 80, 84-86, 95).

Nevertheless, there are still many questions without unequivocal answers and are subjects for further investigation. First, studies concerning tumours of the endocrine system are limited. Particularly, there is a lack of studies exemplifying a connection between *H19* and parathyroid tumours. Additionally, only few studies concerned the potential association between *H19* and hormonal function of tumours. Second, there are

contradictory reports regarding *H19* expression in most of the described pathologies. Similarly, to outcomes obtained for other neoplasms, opposite effects of *H19* on tumorigenesis in endocrine gland tumours were demonstrated. *H19* was proposed to act as an oncogene as well as a suppressor. Currently, the possibility to use its levels as a simple tumour marker is limited. In addition, most presented results were obtained using cancer cell lines and xenograft mouse models. Further investigations on human tumour tissues and plasma concentrations are needed. Finally, the samples of the groups were small in some studies, sometimes due to the rare occurrence of the specific pathology. Therefore, studies with larger sample size are necessary.

In conclusion, *H19* is a novel and intriguing factor, which may allow elucidation of processes involved in carcinogenesis and tumour progression. Nevertheless, further investigation of its biological role in endocrine system tumours are still needed.

Conflicts of Interest

The Authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Authors' Contributions

MR, AJP and MB contributed to the article's conception and design. MR, AJP and KK collected the literature sources. The first draft of the manuscript was written by MR and corrected by AJP, KK and MB. All Authors contributed to the final version of the manuscript and approved it for publication.

References

- Ma L, Bajic VB and Zhang Z: On the classification of long non-coding RNAs. *RNA Biol* 10: 924-933, 2013. PMID: 23696037. DOI: 10.4161/rna.24604
- Zhang P, Wu W, Chen Q and Chen M: Non-Coding RNAs and their Integrated Networks. *J Integr Bioinform* 16: 20190027, 2019. PMID: 31301674. DOI: 10.1515/jib-2019-0027
- Shi X, Sun M, Liu H, Yao Y and Song Y: Long non-coding RNAs: a new frontier in the study of human diseases. *Cancer Lett* 339: 159-166, 2013. PMID: 23791884. DOI: 10.1016/j.canlet.2013.06.013
- ENCODE Project Consortium: An integrated encyclopedia of DNA elements in the human genome. *Nature* 489: 57-74, 2012. PMID: 22955616. DOI: 10.1038/nature11247
- Ponting CP, Oliver PL and Reik W: Evolution and functions of long noncoding RNAs. *Cell* 136: 629-641, 2009. PMID: 19239885. DOI: 10.1016/j.cell.2009.02.006
- Ma L, Cao J, Liu L, Du Q, Li Z, Zou D, Bajic VB and Zhang Z: Lncbook: a curated knowledgebase of human long non-coding RNAs. *Nucleic Acids Res* 47: D128-D134, 2019. PMID: 30329098. DOI: 10.1093/nar/gky960
- Mirza AH, Kaur S and Pociot F: Long non-coding RNAs as novel players in β cell function and type 1 diabetes. *Hum Genomics* 11: 17, 2017. PMID: 28738846. DOI: 10.1186/s40246-017-0113-7
- Motterle A, Gattesco S, Peyot ML, Esguerra JLS, Gomez-Ruiz A, Laybutt DR, Gilon P, Burdet F, Ibberson M, Eliasson L, Prentki M and Regazzi R: Identification of islet-enriched long non-coding RNAs contributing to β -cell failure in type 2 diabetes. *Mol Metab* 6: 1407-1418, 2017. PMID: 29107288. DOI: 10.1016/j.molmet.2017.08.005
- Idda ML, Munk R, Abdelmohsen K and Gorospe M: Noncoding RNAs in Alzheimer's disease. *Wiley Interdiscip Rev RNA* 9: e1463, 2018. PMID: 29327503. DOI: 10.1002/wrna.1463
- Luo Q and Chen Y: Long noncoding RNAs and Alzheimer's disease. *Clin Interv Aging* 11: 867-872, 2016. PMID: 27418812. DOI: 10.2147/CIA.S107037
- Hashemian F, Ghafouri-Fard S, Arsang-Jang S, Mirzajani S, Fallah H, Mehvari Habibabadi J, Sayad A and Taheri M: Epilepsy is associated with dysregulation of long non-coding RNAs in the peripheral blood. *Front Mol Biosci* 6: 113, 2019. PMID: 31709263. DOI: 10.3389/fmolb.2019.00113
- Gibb EA, Brown CJ and Lam WL: The functional role of long non-coding RNA in human carcinomas. *Mol Cancer* 10: 38, 2011. PMID: 21489289. DOI: 10.1186/1476-4598-10-38
- Tsai KW, Tsai CY, Chou NH, Wang KC, Kang CH, Li SC, Lao YH and Chang HT: Aberrant DNA hypermethylation silenced lncRNA expression in gastric cancer. *Anticancer Res* 39: 5381-5391, 2019. PMID: 31570433. DOI: 10.21873/anticancer.13732
- Pachnis V, Belayew A and Tilghman SM: Locus unlinked to alpha-fetoprotein under the control of the murine raf and Rif genes. *Proc Natl Acad Sci USA* 81: 5523-5527, 1984. PMID: 6206499. DOI: 10.1073/pnas.81.17.5523
- Brannan CI, Dees EC, Ingram RS and Tilghman SM: The product of the H19 gene may function as an RNA. *Mol Cell Biol* 10: 28-36, 1990. PMID: 1688465. DOI: 10.1128/mcb.10.1.28
- Gabory A, Ripoché MA, Yoshimizu T and Dandolo L: The H19 gene: regulation and function of a non-coding RNA. *Cytogenet Genome Res* 113: 188-193, 2006. PMID: 16575179. DOI: 10.1159/000090831
- Yoshimura H, Matsuda Y, Yamamoto M, Kamiya S and Ishiwata T: Expression and role of long non-coding RNA H19 in carcinogenesis. *Front Biosci (Landmark Ed)* 23: 614-625, 2018. PMID: 28930564. DOI: 10.2741/4608
- Ariel I, Ayesh S, Perlman E, Pizov G, Tanos V, Schneider T, Erdmann V, Podeh D, Komitowski D, Quasem A, de Groot N and Hochberg A: The product of the imprinted H19 gene is an oncofetal RNA. *Mol Pathol* 50: 34-44, 1997. PMID: 9208812. DOI: 10.1136/mp.50.1.34
- Poirier F, Chan CT, Timmons PM, Robertson EJ, Evans MJ and Rigby PW: The murine H19 gene is activated during embryonic stem cell differentiation in vitro and at the time of implantation in the developing embryo. *Development* 113: 1105-1114, 1991. PMID: 1811930.
- Goshen R, Rachmilewitz J, Schneider T, De-Groot N, Ariel I, Palti Z and Hochberg AA: The expression of the H-19 and IGF-2 genes during human embryogenesis and placental development. *Mol Reprod Dev* 34: 374-379, 1993. PMID: 7682421. DOI: 10.1002/mrd.1080340405
- Dugimont T, Cury JJ, Wernert N, Delobelle A, Raes MB, Joubel A, Stehelin D and Coll J: The H19 gene is expressed within both epithelial and stromal components of human

- invasive adenocarcinomas. *Biol Cell* 85: 117-124, 1995. PMID: 8785513. DOI: 10.1016/0248-4900(96)85272-5
- 22 Biran H, Ariel I, de Groot N, Shani A and Hochberg A: Human imprinted genes as oncodevelopmental markers. *Tumour Biol* 15: 123-134, 1994. PMID: 8073225. DOI: 10.1159/000217882
- 23 Ariel I, Lustig O, Schneider T, Pizov G, Sappir M, De-Groot N and Hochberg A: The imprinted H19 gene as a tumor marker in bladder carcinoma. *Urology* 45: 335-338, 1995. PMID: 7855987. DOI: 10.1016/0090-4295(95)80030-1
- 24 Bartolomei MS, Zemel S and Tilghman SM: Parental imprinting of the mouse H19 gene. *Nature* 351: 153-155, 1991. PMID: 1709450. DOI: 10.1038/351153a0
- 25 Byun HM, Wong HL, Birnstein EA, Wolff EM, Liang G and Yang AS: Examination of IGF2 and H19 loss of imprinting in bladder cancer. *Cancer Res* 67: 10753-10758, 2007. PMID: 18006818. DOI: 10.1158/0008-5472.CAN-07-0329
- 26 Li E, Beard C and Jaenisch R: Role for DNA methylation in genomic imprinting. *Nature* 366: 362-365, 1993. PMID: 8247133. DOI: 10.1038/366362a0
- 27 Thorvaldsen JL, Duran KL and Bartolomei MS: Deletion of the H19 differentially methylated domain results in loss of imprinted expression of H19 and Igf2. *Genes Dev* 12: 3693-3702, 1998. PMID: 9851976. DOI: 10.1101/gad.12.23.3693
- 28 Hibi K, Nakamura H, Hirai A, Fujikake Y, Kasai Y, Akiyama S, Ito K and Takagi H: Loss of H19 imprinting in esophageal cancer. *Cancer Res* 56: 480-482, 1996. PMID: 8564957.
- 29 Kondo M, Suzuki H, Ueda R, Osada H, Takagi K and Takahashi T: Frequent loss of imprinting of the H19 gene is often associated with its overexpression in human lung cancers. *Oncogene* 10: 1193-1198, 1995. PMID: 7700644.
- 30 Steenman MJ, Rainier S, Dobry CJ, Grundy P, Horon IL and Feinberg AP: Loss of imprinting of IGF2 is linked to reduced expression and abnormal methylation of H19 in Wilms' tumour. *Nat Genet* 7: 433-439, 1994. PMID: 7920665. DOI: 10.1038/ng0794-433
- 31 Yballe CM, Vu TH and Hoffman AR: Imprinting and expression of insulin-like growth factor-II and H19 in normal breast tissue and breast tumor. *J Clin Endocrinol Metab* 81: 1607-1612, 1996. PMID: 8636375. DOI: 10.1210/jcem.81.4.8636375
- 32 Wada M, Seeger RC, Mizoguchi H and Koeffler HP: Maintenance of normal imprinting of H19 and IGF2 genes in neuroblastoma. *Cancer Res* 55: 3386-3388, 1995. PMID: 7614476.
- 33 Vernucci M, Cerrato F, Besnard N, Casola S, Pedone P V, Bruni CB and Riccio A: The H19 endodermal enhancer is required for Igf2 activation and tumor formation in experimental liver carcinogenesis. *Oncogene* 19: 6376-6385, 2000. PMID: 11175353. DOI: 10.1038/sj.onc.1204024
- 34 Berteaux N, Lottin S, Monté D, Pinte S, Quatannens B, Coll J, Hondermarck H, Cury JJ, Dugimont T and Adriaenssens E: H19 mRNA-like noncoding RNA promotes breast cancer cell proliferation through positive control by E2F1. *J Biol Chem* 280: 29625-29636, 2005. PMID: 15985428. DOI: 10.1074/jbc.M504033200
- 35 Moulton T, Crenshaw T, Hao Y, Moosikasuwan J, Lin N, Dembitzer F, Hensle T, Weiss L, McMorro L, Loew T, Kraus W, Gerald W and Tycko B: Epigenetic lesions at the H19 locus in Wilms' tumour patients. *Nat Genet* 7: 440-447, 1994. PMID: 7920666. DOI: 10.1038/ng0794-440
- 36 Hao Y, Crenshaw T, Moulton T, Newcomb E and Tycko B: Tumour-suppressor activity of H19 RNA. *Nature* 365: 764-767, 1993. PMID: 7692308. DOI: 10.1038/365764a0
- 37 Verkerk AJ, Ariel I, Dekker MC, Schneider T, van Gurp RJ, de Groot N, Gillis AJ, Oosterhuis JW, Hochberg AA and Looijenga LH: Unique expression patterns of H19 in human testicular cancers of different etiology. *Oncogene* 14: 95-107, 1997. PMID: 9010236. DOI: 10.1038/sj.onc.1200802
- 38 Adriaenssens E, Dumont L, Lottin S, Bolle D, Leprêtre A, Delobelle A, Bouali F, Dugimont T, Coll J and Cury JJ: H19 overexpression in breast adenocarcinoma stromal cells is associated with tumor values and steroid receptor status but independent of p53 and Ki-67 expression. *Am J Pathol* 153: 1597-1607, 1998. PMID: 9811352. DOI: 10.1016/S0002-9440(10)65748-3
- 39 Qian B, Wang DM, Gu XS, Zhou K, Wu J, Zhang CY and He XY: LncRNA H19 serves as a ceRNA and participates in non-small cell lung cancer development by regulating microRNA-107. *Eur Rev Med Pharmacol Sci* 22: 5946-5953, 2018. PMID: 30280776. DOI: 10.26355/eurrev_201809_15925
- 40 Cui J, Mo J, Luo M, Yu Q, Zhou S, Li T, Zhang Y and Luo W: c-Myc-activated long non-coding RNA H19 downregulates miR-107 and promotes cell cycle progression of non-small cell lung cancer. *Int J Clin Exp Pathol* 8: 12400-12409, 2015. PMID: 26722426.
- 41 Huang C, Cao L, Qiu L, Dai X, Ma L, Zhou Y, Li H, Gao M, Li W, Zhang Q, Han K and Lv H: Upregulation of H19 promotes invasion and induces epithelial-to-mesenchymal transition in esophageal cancer. *Oncol Lett* 10: 291-296, 2015. PMID: 26171017. DOI: 10.3892/ol.2015.3165
- 42 Song H, Sun W, Ye G, Ding X, Liu Z, Zhang S, Xia T, Xiao B, Xi Y and Guo J: Long non-coding RNA expression profile in human gastric cancer and its clinical significances. *J Transl Med* 11: 225, 2013. PMID: 24063685. DOI: 10.1186/1479-5876-11-225
- 43 Li H, Yu B, Li J, Su L, Yan M, Zhu Z and Liu B: Overexpression of lncRNA H19 enhances carcinogenesis and metastasis of gastric cancer. *Oncotarget* 5: 2318-2329, 2014. PMID: 24810858. DOI: 10.18632/oncotarget.1913
- 44 Arita T, Ichikawa D, Konishi H, Komatsu S, Shiozaki A, Shoda K, Kawaguchi T, Hirajima S, Nagata H, Kubota T, Fujiwara H, Okamoto K and Otsuji E: Circulating long non-coding RNAs in plasma of patients with gastric cancer. *Anticancer Res* 33: 3185-3194, 2013. PMID: 23898077.
- 45 Tsang WP, Ng EK, Ng SS, Jin H, Yu J, Sung JJ and Kwok TT: Oncofetal H19-derived miR-675 regulates tumor suppressor RB in human colorectal cancer. *Carcinogenesis* 31: 350-358, 2010. PMID: 19926638. DOI: 10.1093/carcin/bgp181
- 46 Matouk IJ, DeGroot N, Mezan S, Ayesb S, Abu-lail R, Hochberg A and Galun E: The H19 non-coding RNA is essential for human tumor growth. *PLoS One* 2: e845, 2007. PMID: 17786216. DOI: 10.1371/journal.pone.0000845
- 47 Ma L, Tian X, Wang F, Zhang Z, Du C, Xie X, Kornmann M and Yang Y: The long noncoding RNA H19 promotes cell proliferation via E2F-1 in pancreatic ductal adenocarcinoma. *Cancer Biol Ther* 17: 1051-1061, 2016. PMID: 27573434. DOI: 10.1080/15384047.2016.1219814
- 48 Ma L, Tian X, Guo H, Zhang Z, Du C, Wang F, Xie X, Gao H, Zhuang Y, Kornmann M, Gao H and Yang Y: Long noncoding RNA H19 derived miR-675 regulates cell proliferation by

- down-regulating E2F-1 in human pancreatic ductal adenocarcinoma. *J Cancer* 9: 389-399, 2018. PMID: 29344285. DOI: 10.7150/jca.21347
- 49 He H, Wang N, Yi X, Tang C and Wang D: Long non-coding RNA H19 regulates E2F1 expression by competitively sponging endogenous miR-29a-3p in clear cell renal cell carcinoma. *Cell Biosci* 7: 65, 2017. PMID: 29214011. DOI: 10.1186/s13578-017-0193-z
 - 50 Ariel I, Sughayer M, Fellig Y, Pizov G, Ayesh S, Podeh D, Libdeh BA, Levy C, Birman T, Tykocinski ML, de Groot N and Hochberg A: The imprinted H19 gene is a marker of early recurrence in human bladder carcinoma. *Mol Pathol* 53: 320-323, 2000. PMID: 11193051. DOI: 10.1136/mp.53.6.320
 - 51 Ou L, Wang D, Zhang H, Yu Q and Hua F: Decreased expression of miR-138-5p by lncRNA H19 in cervical cancer promotes tumor proliferation. *Oncol Res* 26: 401-410, 2018. PMID: 28797320. DOI: 10.3727/096504017X15017209042610
 - 52 Tanos V, Prus D, Ayesh S, Weinstein D, Tykocinski ML, De-Groot N, Hochberg A and Ariel I: Expression of the imprinted H19 oncofetal RNA in epithelial ovarian cancer. *Eur J Obstet Gynecol Reprod Biol* 85: 7-11, 1999. PMID: 10428315. DOI: 10.1016/S0301-2115(98)00275-9
 - 53 Zhu Z, Song L, He J, Sun Y, Liu X and Zou X: Ectopic expressed long non-coding RNA H19 contributes to malignant cell behavior of ovarian cancer. *Int J Clin Exp Pathol* 8: 10082-10091, 2015. PMID: 26617715.
 - 54 Jia P, Cai H, Liu X, Chen J, Ma J, Wang P, Liu Y, Zheng J and Xue Y: Long non-coding RNA H19 regulates glioma angiogenesis and the biological behavior of glioma-associated endothelial cells by inhibiting microRNA-29a. *Cancer Lett* 381: 359-369, 2016. PMID: 27543358. DOI: 10.1016/j.canlet.2016.08.009
 - 55 Jiang X, Yan Y, Hu M, Chen X, Wang Y, Dai Y, Wu D, Wang Y, Zhuang Z and Xia H: Increased level of H19 long noncoding RNA promotes invasion, angiogenesis, and stemness of glioblastoma cells. *J Neurosurg* 124: 129-136, 2016. PMID: 26274999. DOI: 10.3171/2014.12.JNS1426
 - 56 Guo G, Kang Q, Chen Q, Chen Z, Wang J, Tan L and Chen JL: High expression of long non-coding RNA H19 is required for efficient tumorigenesis induced by Bcr-Abl oncogene. *FEBS Lett* 588: 1780-1786, 2014. PMID: 24685695. DOI: 10.1016/j.febslet.2014.03.038
 - 57 Zhang DM, Lin ZY, Yang ZH, Wang YY, Wan D, Zhong JL, Zhuang PL, Huang ZQ, Zhou B and Chen WL: lncRNA H19 promotes tongue squamous cell carcinoma progression through β -catenin/GSK3 β /EMT signaling via association with EZH2. *Am J Transl Res* 9: 3474-3486, 2017. PMID: 28804564.
 - 58 Xu Y, Wang Z, Jiang X and Cui Y: Overexpression of long noncoding RNA H19 indicates a poor prognosis for cholangiocarcinoma and promotes cell migration and invasion by affecting epithelial-mesenchymal transition. *Biomed Pharmacother* 92: 17-23, 2017. PMID: 28528181. DOI: 10.1016/j.biopha.2017.05.061
 - 59 Chan LH, Wang W, Yeung W, Deng Y, Yuan P and Mak KK: Hedgehog signaling induces osteosarcoma development through Yap1 and H19 overexpression. *Oncogene* 33: 4857-4866, 2014. PMID: 24141783. DOI: 10.1038/onc.2013.433
 - 60 Zhao J and Ma ST: Downregulation of lncRNA H19 inhibits migration and invasion of human osteosarcoma through the NF- κ B pathway. *Mol Med Rep* 17: 7388-7394, 2018. PMID: 29568924. DOI: 10.3892/mmr.2018.8746
 - 61 Luan W, Zhou Z, Ni X, Xia Y, Wang J, Yan Y and Xu B: Long non-coding RNA H19 promotes glucose metabolism and cell growth in malignant melanoma via miR-106a-5p/E2F3 axis. *J Cancer Res Clin Oncol* 144: 531-542, 2018. PMID: 29350287. DOI: 10.1007/s00432-018-2582-z
 - 62 Cui H, Hedborg F, He L, Nordenskjöld A, Sandstedt B, Pfeifer-Ohlsson S and Ohlsson R: Inactivation of H19, an imprinted and putative tumor repressor gene, is a preneoplastic event during Wilms' tumorigenesis. *Cancer Res* 57: 4469-4473, 1997. PMID: 9377554.
 - 63 Lecerf C, Le Bourhis X and Adriaenssens E: The long non-coding RNA H19: an active player with multiple facets to sustain the hallmarks of cancer. *Cell Mol Life Sci* 76: 4673-4687, 2019. PMID: 31338555. DOI: 10.1007/s00018-019-03240-z
 - 64 Yu H, Li S, Wu SX, Huang S, Li S and Ye L: The prognostic value of long non-coding RNA H19 in various cancers: A meta-analysis based on 15 studies with 1584 patients and the Cancer Genome Atlas data. *Medicine (Baltimore)* 99: e18533, 2020. PMID: 31914026. DOI: 10.1097/MD.00000000000018533
 - 65 Li J, Huang YY, Deng XJ, Luo ML, Wang XF, Hu HY, Liu C Di and Zhong M: Long noncoding RNA H19 promotes transforming growth factor- β -induced epithelial-mesenchymal transition by acting as a competing endogenous RNA of miR-370-3p in ovarian cancer cells. *Onco Targets Ther* 11: 427-440, 2018. PMID: 29403287. DOI: 10.2147/OTT.S149908
 - 66 Liu FT, Pan H, Xia GF, Qiu C and Zhu ZM: Prognostic and clinicopathological significance of long noncoding RNA H19 overexpression in human solid tumors: evidence from a meta-analysis. *Oncotarget* 7: 83177-83186, 2016. PMID: 27825121. DOI: 10.18632/oncotarget.13076
 - 67 Lu T, Yu C, Ni H, Liang W, Yan H and Jin W: Expression of the long non-coding RNA H19 and MALAT-1 in growth hormone-secreting pituitary adenomas and its relationship to tumor behavior. *Int J Dev Neurosci* 67: 46-50, 2018. PMID: 29604339. DOI: 10.1016/j.ijdevneu.2018.03.009
 - 68 Liao S, Yu C, Liu H, Zhang C, Li Y and Zhong X: Long non-coding RNA H19 promotes the proliferation and invasion of lung cancer cells and regulates the expression of E-cadherin, N-cadherin, and vimentin. *Onco Targets Ther* 12: 4099-4107, 2019. PMID: 31190899. DOI: 10.2147/OTT.S185156
 - 69 Wu ZR, Yan L, Liu YT, Cao L, Guo YH, Zhang Y, Yao H, Cai L, Shang HB, Rui WW, Yang G, Zhang XB, Tang H, Wang Y, Huang JY, Wei YX, Zhao WG, Su B and Wu ZB: Inhibition of mTORC1 by lncRNA H19 via disrupting 4E-BP1/Raptor interaction in pituitary tumours. *Nat Commun* 9: 4624, 2018. PMID: 30397197. DOI: 10.1038/s41467-018-06853-3
 - 70 Zhang Y, Liu YT, Tang H, Xie WQ, Yao H, Gu WT, Zheng YZ, Shang HB, Wang Y, Wei YX, Wu ZR and Wu ZB: Exosome-transmitted lncRNA H19 inhibits the growth of pituitary adenoma. *J Clin Endocrinol Metab* 104: 6345-6356, 2019. PMID: 31369093. DOI: 10.1210/je.2019-00536
 - 71 Wu ZR, Zheng Y, Xie W, Li Q, Zhang Y, Ren B, Cai L, Cheng Y, Tang H, Su Z and Wu ZB: The long noncoding RNA-H19/miRNA-93a/ATG7 axis regulates the sensitivity of pituitary adenomas to dopamine agonists. *Mol Cell Endocrinol* 518: 111033, 2020. PMID: 32946927. DOI: 10.1016/j.mce.2020.111033
 - 72 Pan R and Zhou H: Exosomal transfer of lncRNA H19 promotes erlotinib resistance in non-small cell lung cancer via miR-615-3p/ATG7 axis. *Cancer Manag Res* 12: 4283-4297, 2020. PMID: 32606925. DOI: 10.2147/CMAR.S241095

- 73 Liu L, Yang J, Zhu X, Li D, Lv Z and Zhang X: Long noncoding RNA H19 competitively binds miR-17-5p to regulate YES1 expression in thyroid cancer. *FEBS J* 283: 2326-2339, 2016. PMID: 27093644. DOI: 10.1111/febs.13741
- 74 Jia J, Zhang X, Zhan D, Li J, Li Z, Li H and Qian J: LncRNA H19 interacted with miR-130a-3p and miR-17-5p to modify radio-resistance and chemo-sensitivity of cardiac carcinoma cells. *Cancer Med* 8: 1604-1618, 2019. PMID: 30843379. DOI: 10.1002/cam4.1860
- 75 Liu N, Zhou Q, Qi YH, Wang H, Yang L and Fan QY: Effects of long non-coding RNA H19 and microRNA let7a expression on thyroid cancer prognosis. *Exp Mol Pathol* 103: 71-77, 2017. PMID: 28655518. DOI: 10.1016/j.yexmp.2017.06.004
- 76 Li X, Li Q, Jin X, Guo H and Li Y: Long non-coding RNA H19 knockdown inhibits the cell viability and promotes apoptosis of thyroid cancer cells through regulating the PI3K/AKT pathway. *Exp Ther Med* 18: 1863-1869, 2019. PMID: 31410148. DOI: 10.3892/etm.2019.7720
- 77 Wang P, Liu G, Xu W, Liu H, Bu Q and Sun D: Long noncoding RNA H19 inhibits cell viability, migration, and invasion via downregulation of IRS-1 in thyroid cancer cells. *Technol Cancer Res Treat* 16: 1102-1112, 2017. PMID: 29332545. DOI: 10.1177/1533034617733904
- 78 Zhong ME, Chen Y, Zhang G, Xu L, Ge W and Wu B: LncRNA H19 regulates PI3K-Akt signal pathway by functioning as a ceRNA and predicts poor prognosis in colorectal cancer: integrative analysis of dysregulated ncRNA-associated ceRNA network. *Cancer Cell Int* 19: 148, 2019. PMID: 31164794. DOI: 10.1186/s12935-019-0866-2
- 79 Liao Z, Zhao J and Yang Y: Downregulation of lncRNA H19 inhibits the migration and invasion of melanoma cells by inactivating the NF- κ B and PI3K/Akt signaling pathways. *Mol Med Rep* 17: 7313-7318, 2018. PMID: 29568965. DOI: 10.3892/mmr.2018.8782
- 80 Liang WQ, Zeng D, Chen CF, Sun SM, Lu XF, Peng CY and Lin HY: Long noncoding RNA H19 is a critical oncogenic driver and contributes to epithelial-mesenchymal transition in papillary thyroid carcinoma. *Cancer Manag Res* 11: 2059-2072, 2019. PMID: 30881130. DOI: 10.2147/CMAR.S195906
- 81 Li M, Chai HF, Peng F, Meng YT, Zhang LZ, Zhang L, Zou H, Liang QL, Li MM, Mao KG, Sun DX, Tong MY, Deng ZQ, Hou ZJ, Zhao Y, Li J, Wang XC, Lv SS, Zhang QQ, Yu X, Lam EW, Liu Q, Cui XN and Xu J: Estrogen receptor β upregulated by lncRNA-H19 to promote cancer stem-like properties in papillary thyroid carcinoma. *Cell Death Dis* 9: 1120, 2018. PMID: 30389909. DOI: 10.1038/s41419-018-1077-9
- 82 Lu W and Kang Y: Epithelial-mesenchymal plasticity in cancer progression and metastasis. *Dev Cell* 49: 361-374, 2019. PMID: 31063755. DOI: 10.1016/j.devcel.2019.04.010
- 83 Wu Y, Zhou Y, He J, Sun H and Jin Z: Long non-coding RNA H19 mediates ovarian cancer cell cisplatin-resistance and migration during EMT. *Int J Clin Exp Pathol* 12: 2506-2515, 2019. PMID: 31934077.
- 84 Lan X, Sun W, Dong W, Wang Z, Zhang T, He L and Zhang H: Downregulation of long noncoding RNA H19 contributes to the proliferation and migration of papillary thyroid carcinoma. *Gene* 646: 98-105, 2018. PMID: 29287713. DOI: 10.1016/j.gene.2017.12.051
- 85 Jiao X, Lu J, Huang Y, Zhang J, Zhang H and Zhang K: Long non-coding RNA H19 may be a marker for prediction of prognosis in the follow-up of patients with papillary thyroid cancer. *Cancer Biomark* 26: 203-207, 2019. PMID: 31403942. DOI: 10.3233/CBM-190273
- 86 Dai Y, Miao Y, Zhu Q, Gao M and Hao F: Expression of long non-coding RNA H19 predicts distant metastasis in minimally invasive follicular thyroid carcinoma. *Bioengineered* 10: 383-389, 2019. PMID: 31791180. DOI: 10.1080/21655979.2019.1658489
- 87 Zhang H, Yu Y, Zhang K, Liu X, Dai Y and Jiao X: Targeted inhibition of long non-coding RNA H19 blocks anaplastic thyroid carcinoma growth and metastasis. *Bioengineered* 10: 306-315, 2019. PMID: 31299871. DOI: 10.1080/21655979.2019.1642722
- 88 Wächter S, Damanakis AI, Elxnat M, Roth S, Wunderlich A, Verburg FA, Fellingner SA, Bartsch DK and Di Fazio P: Epigenetic modifications in thyroid cancer cells restore NIS and radio-iodine uptake and promote cell death. *J Clin Med* 7: 61, 2018. PMID: 29561759. DOI: 10.3390/jcm7040061
- 89 Voutilainen R, Ilvesmäki V, Ariel I, Rachmilewitz J, de Groot N and Hochberg A: Parallel regulation of parentally imprinted H19 and insulin-like growth factor-II genes in cultured human fetal adrenal cells. *Endocrinology* 134: 2051-2056, 1994. PMID: 7512497. DOI: 10.1210/endo.134.5.7512497
- 90 Lustig O, Ariel I, Ilan J, Lev-Lehman E, De-Groot N and Hochberg A: Expression of the imprinted gene H19 in the human fetus. *Mol Reprod Dev* 38: 239-246, 1994. PMID: 7917273. DOI: 10.1002/mrd.1080380302
- 91 Liu J, Kahri AI, Heikkilä P, Ilvesmäki V and Voutilainen R: H19 and insulin-like growth factor-II gene expression in adrenal tumors and cultured adrenal cells. *J Clin Endocrinol Metab* 80: 492-496, 1995. PMID: 7531713. DOI: 10.1210/jcem.80.2.7531713
- 92 Gao ZH, Supola S, Liu J, Heikkilä P, Jänne J and Voutilainen R: Association of H19 promoter methylation with the expression of H19 and IGF-II genes in adrenocortical tumors. *J Clin Endocrinol Metab* 87: 1170-1176, 2002. PMID: 11889182. DOI: 10.1210/jcem.87.3.8331
- 93 Glover AR, Zhao JT, Ip JC, Lee JC, Robinson BG, Gill AJ, Soon PS and Sidhu SB: Long noncoding RNA profiles of adrenocortical cancer can be used to predict recurrence. *Endocr Relat Cancer* 22: 99-109, 2015. PMID: 25595289. DOI: 10.1530/ERC-14-0457
- 94 Liu J, Kahri AI, Heikkilä P and Voutilainen R: Ribonucleic acid expression of the clustered imprinted genes, p57KIP2, insulin-like growth factor II, and H19, in adrenal tumors and cultured adrenal cells. *J Clin Endocrinol Metab* 82: 1766-1771, 1997. PMID: 9177379. DOI: 10.1210/jcem.82.6.3968
- 95 Barreau O, Assié G, Wilmot-Roussel H, Ragazzon B, Baudry C, Perlemonne K, René -Corail F, Bertagna X, Dousset B, Hamzaoui N, Tissier F, de Reynies A and Bertherat J: Identification of a CpG island methylator phenotype in adrenocortical carcinomas. *J Clin Endocrinol Metab* 98: E174-184, 2013. PMID: 23093492. DOI: 10.1210/jc.2012-2993
- 96 Creemers SG, van Koetsveld PM, van Kemenade FJ, Papathomas TG, Franssen GJ, Dogan F, Eekhoff EM, van der Valk P, de Herder WW, Janssen JA, Feelders RA and Hofland LJ: Methylation of IGF2 regulatory regions to diagnose adrenocortical carcinomas. *Endocr Relat Cancer* 23: 727-737, 2016. PMID: 27535174. DOI: 10.1530/ERC-16-0266
- 97 Miura K, Obama M, Yun K, Masuzaki H, Ikeda Y, Yoshimura S, Akashi T, Niikawa N, Ishimaru T and Jinno Y: Methylation imprinting of H19 and SNRPN genes in human benign ovarian

- teratomas. *Am J Hum Genet* 65: 1359-1367, 1999. PMID: 10521301. DOI: 10.1086/302615
- 98 Sievers S, Alemazkour K, Zahn S, Perlman EJ, Gillis AJ, Looijenga LH, Göbel U and Schneider DT: IGF2/H19 imprinting analysis of human germ cell tumors (GCTs) using the methylation-sensitive single-nucleotide primer extension method reflects the origin of GCTs in different stages of primordial germ cell development. *Genes Chromosomes Cancer* 44: 256-264, 2005. PMID: 16001432. DOI: 10.1002/gcc.20237
- 99 Kawakami T, Zhang C, Okada Y and Okamoto K: Erasure of methylation imprint at the promoter and CTCF-binding site upstream of H19 in human testicular germ cell tumors of adolescents indicate their fetal germ cell origin. *Oncogene* 25: 3225-3236, 2006. PMID: 16434968. DOI: 10.1038/sj.onc.1209362
- 100 Ji M, Yao Y, Liu A, Shi L, Chen D, Tang L, Yang G, Liang X, Peng J and Shao C: lncRNA H19 binds VGF and promotes pNEN progression *via* PI3K/AKT/CREB signaling. *Endocr Relat Cancer* 26: 643-658, 2019. PMID: 31117050. DOI: 10.1530/ERC-18-0552
- 101 Ramnarine VR, Alshalalfa M, Mo F, Nabavi N, Erho N, Takhar M, Shukin R, Brahmabhatt S, Gawronski A, Kobelev M, Nouri M, Lin D, Tsai H, Lotan TL, Karnes RJ, Rubin MA, Zoubeidi A, Gleave ME, Sahinalp C, Wyatt AW, Volik S V, Beltran H, Davicioni E, Wang Y and Collins CC: The long noncoding RNA landscape of neuroendocrine prostate cancer and its clinical implications. *Gigascience* 7: giy050, 2018. PMID: 29757368. DOI: 10.1093/gigascience/giy050

Received December 21, 2020

Revised January 8, 2021

Accepted January 11, 2021