

Review

Pancreatic Cancer Stem Cells and Therapeutic Approaches

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Abstract. Pancreatic ductal adenocarcinoma (PDAC) is considered one of the deadliest human cancers, with 1-5% 5-year survival rates (~6-month median survival duration) despite therapy; thus, PDAC represents an unmet therapeutic challenge. PDAC is the major histological subtype, comprising 90% of all pancreatic cancers. It is a highly complex and aggressive malignancy, presenting with early local invasion and metastasis, and is resistant to most therapies, all of which are believed to contribute to its extremely poor prognosis. PDAC is characterized by molecular alterations, including mutations of *K-RAS* (~90% of cases), *TP53*, transforming growth factor- β , *Hedgehog*, *WNT* and *NOTCH* signaling pathways. Given that cancer stem cells have a crucial role not only in tumor initiation and progression, but also in drug resistance and relapse or recurrence of various cancer types, they may be excellent targets for effective novel therapeutic approaches. Here, we reviewed recent therapeutic strategies targeting pancreatic cancer stem cells using chemotherapeutics and targeted drugs, non-coding RNAs (i.e., siRNA and miRNAs), immunotherapy, and natural compounds.

Pancreatic Cancer

Pancreatic cancer (PaCa) is currently the third most common cause of cancer-related death in the United States (1-4).

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Pancreatic ductal adenocarcinoma (PDAC) is a major histological subtype, comprising of 90% of all PaCas (1). Although both the exocrine and endocrine cells of the pancreas can form cancer, tumors of the endocrine pancreas are less common; they are known as islet cell tumors (such as insulinomas, glucagonomas, and somatostatinomas) and neuroendocrine tumors (such as gastrinomas). PaCa formed by exocrine cells is much more common; nearly all of these tumors are adenocarcinomas (2).

PDAC has an extremely poor prognosis, with 5-year survival rates of 1-5% (~6-month median survival duration) with currently available therapies (5). The majority of patients with PDAC (80-90%) have local metastasis at the time of diagnosis. These patients cannot benefit from surgery and instead undergo standard-of-care medical treatment using chemotherapeutics, including gemcitabine, or a combination of 5-fluorouracil (5-FU) plus leucovorin (6). The high mortality rate of PDAC is attributed to its aggressive nature, early local and advanced metastasis, its intrinsic resistance to chemotherapeutics, and absence of effective therapies. In addition, lack of early diagnostic tests prevents early therapeutic interventions, and is considered to be an important factor that contributes to the poor prognosis of PDAC (4).

Biology and Genetics of PaCa

Pancreatic intra-epithelial neoplasms (PanINs) are the most common precursors of PDAC; they develop in a background of pancreatic inflammation. *K-RAS* is mutated in approximately 90% of all PDACs and is found in almost all PanINs (7). Although *K-RAS* mutation is necessary for carcinogenesis and subsequent PDAC tumor maintenance, it is not sufficient to drive PanINs to develop into PDAC (8). In addition to *K-RAS*, mutations in tumor suppressors, including cyclin-dependent kinase inhibitor 2A (*CDKN2A*) (encoding p16^{INK4A} protein), mothers against decapentaplegic homolog

4 (*SMAD4*), and *TP53*, are required for malignant transformation (9). More than 60 molecular alterations related to apoptotic pathways and Hedgehog (HH), transforming growth factor- β (TGF- β), WNT, and NOTCH signaling pathways have been identified in PDAC (10, 11).

PDAC is also characterized by an excessive desmoplastic response, with generation of dense fibrotic tissue, an altered extracellular matrix, and hypovascularity that may be caused by HHS signaling (12). Highly dense fibrotic tissue constitutes 90% of the tumor volume; it is considered a barrier to effective drug delivery and prevents the penetration of therapeutics into tumor tissues. The other characteristic feature of PDAC is that it is resistant to chemotherapy-induced apoptosis due to intrinsic and acquired resistance to such therapies, and the presence of PaCa cancer stem cells (CSCs) (13). Recent emerging data indicate that CSCs are an important factor that contributes to the complex biology, tumorigenesis, progression, and poor clinical outcome of PDAC (14).

Normal and Cancer Stem Cells

Normal adult stem cells are tissue-specific cells that exhibit unique biological properties, including the ability to self-renew and differentiate into various mature cells of specific tissues. Functionally, these cells undergo an unlimited number of asymmetrical cell divisions, and with each division, they produce at least one daughter progenitor cell that maintains their indefinite self-renewing capacity (15). When adult stem cells divide asymmetrically, they can also produce cells that have limited division capacity, ultimately differentiating into mature cells (16). Since adult cells such as epithelial and blood cells undergo continuous cellular turnover, adult stem cells form a pool of long-lived cells that provide a continuous supply of differentiated cells for the homeostatic control of specific tissue compartments (17).

Small subpopulation of cancer cells, CSCs, contribute to tumor initiation, growth, and metastasis; therapy resistance; and cancer recurrence (14, 18). They were first identified in acute myelogenous leukemia and subsequently in many solid cancer types, including PaCa (19). CSCs share core regulatory pathways with normal stem cells but have diversely distinct reprogrammed pathways. Like normal pluripotent stem cells, CSCs have both self-renewal and multilineage differentiation capacities (20). While normal pluripotent stem cells have tightly controlled self-renewal and committed lineage differentiation capacities, which prevent tumor development, CSCs have the capability to differentiate into multiple cell types, leading to tumor initiation and progression (21). CSCs can give rise to both more CSCs and progeny that ultimately differentiate into the different cell types in a tumor by dividing indefinitely. CSCs also give rise to tumors that phenotypically resemble their

tissue of origin, either in their morphological characteristics or in their expression of tissue-specific genes (17). CSCs have quiescent potential in a dormant state and may be involved in both resistance to apoptosis and induction of angiogenesis. Thus, CSCs are not only considered tumor-initiating cells but also as cells that promote tumor development and therapy resistance, leading to disease progression and relapse (14, 18, 19, 22).

According to the CSC hypothesis, cancer-initiating cells are transformed tissue stem cells that are normally found in low numbers among the heterogeneous cells that comprise the malignant tumor mass; they retain the essential characteristic feature of self-protection through the activity of multiple drug resistance (MDR) transporters (23). Understanding the central role of MDR transporters in the protection and self-renewal of normal stem cells and CSCs may lead to the characterization of their differences, which could be used to develop therapeutic approaches that target CSCs. Strategies that only target cancer cells would not be sufficient to eliminate cancer or control the growth and progression of the disease (24).

In summary, tumor initiation, propagation, metastasis, and relapse are caused by a tiny subpopulation of cells called CSCs that are localized among cancer cells and have stem cell-like properties, with clonal long-term repopulation and self-renewal capacities. Due to their hierarchical organization, CSCs and non-CSC tumor cells share the same mutational and genetic background; however, CSCs are exclusively tumorigenic and therapy resistant (25). Thus, while developing therapeutic agents that are aimed to target and eliminate CSCs successfully, one should also consider the heterogeneity of CSCs (26). CSCs carry more differentiated cell features and characteristics among different cancer subtypes even in the same tumor and at different stages of progression. Thus, while CSCs undergo a differentiation process and give rise to tumor cells, there may be different markers at different stages of the cancer.

PaCa Stem Cells

Several common stem cell markers have been identified as CSC-specific markers in CSC subpopulations isolated from a wide variety of human cancer types and cell lines. These include octamer-binding transcription factor 4 (OCT4), sex determining region Y-box 2 (SOX2), homeobox protein (NANOG), tyrosine-protein kinase Kit (C-KIT), ATP-binding cassette sub-family G member 2 (ABCG2), cluster of differentiation (CD) CD34, CD44, CD123, CD133, CD44V6, tyrosine-protein kinase Met (C-MET), tetraspanin-8 (TSPAN8), integrin (A6B4), C-X-C chemokine receptor type 4 (CXCR4), claudin7, epithelial-specific antigen (ESA), and aldehyde dehydrogenase 1 (ALDH1) (18,19,22). Classically, CSCs are first isolated using techniques such as

flow cytometry or magnetic bead sorting and then implanted in immunocompromised mice to determine their efficiency in tumor initiation and progression *in vivo* (18).

Although many markers have been identified for pancreatic CSCs, which represent fewer than 1% of all PaCa cells, a universal marker is still lacking. Markers that have been used in identifying neoplastic and non-neoplastic stem cells include CD44, CD24, CD133, ESA, ALDH1, and Hoechst dye exclusion (side population) (18). CD133, ALDH1, and the triplet combination of CD24⁺CD44⁺ESA⁺ are the best established PaCa CSC markers (18, 19, 22). CD44⁺/CD24⁺/ESA⁺ cells were found to be much more tumorigenic than were marker-negative PaCa cells and possessed the capacity to self-renew and differentiate into different progeny cancer cell types. When injected into non-obese diabetic (NOD)/severe combined immunodeficiency (SCID) mice, these CSCs self renewed and produced differentiated progeny. They had a 100-fold increased tumorigenic potential and maintained their surface marker phenotype after repeated passages as xenografts. In concordance with these findings, clinical data suggest that CD44 positivity is a poor prognostic indicator in patients with PaCa (17).

A pancreatic CSC population with CD133 expression exhibited increased tumorigenicity, a metastatic phenotype and more resistance to chemotherapy than did CD133⁻ cells. During serial passages, CD133⁺ cells retained their ability to self-renew and generate differentiated non-tumorigenic descendants to reconstruct hierarchically organized tumors in an orthotopic mouse model. (27) In PaCa, a CD44⁺/CD133⁺ cell population has also been demonstrated in both tumor specimens and PaCa cell lines. Meanwhile, CD44⁺/CD133⁺ MiaPaCa2 cells, which express high levels of NOTCH and BCL2, were found to have a strong capacity to form tumorspheres *in vitro* and initiated tumors in a mouse xenograft model *in vivo* (28). CD44⁺/CD133⁺ cells isolated from the PANC-1 cell line were also capable of forming tumorspheres *in vitro*, exhibited tumor-initiating potential *in vivo*, and profoundly responded to WNT pathway inhibition (22). Moreover, high expression levels of both CD44 and CD133 were also found to be associated with a poor prognosis in patients with PaCa (22).

CD133⁺ (cytokeratin⁻) cells from human PaCa tissues displayed high tumorigenic potential (14). In a series of experiments, CD44⁺CD24⁺ESA⁺ cells (comprising 0.2-0.8% of the total cell population), identified by fluorescence-activated cell sorting from primary human cancer tissues, were implanted in immune-compromised mice. A small volume of tumor cells was consistently observed (18). When engrafted into nude mice, an even smaller number of CD133⁺ cells (fewer than 500) routinely led to the formation of tumors; although these cells were highly tumorigenic, CD133⁻ cells were not able to induce tumor formation (18).

In addition to cell surface markers, the cellular molecule ALDH1, which catalyzes the oxidation of intracellular aldehydes and converts retinol to retinoic acid, was also implicated as a stem cell marker in PaCa due to its role in the early differentiation of stem cells (18, 22). ALDH1⁺ cells, which have a higher clonogenic capacity than do ALDH1⁻ cells, were shown to be highly tumorigenic, able to initiate tumor development at low cell numbers, and undergo epithelial–mesenchymal transition (EMT) (22). Recently, CD133⁺/CXCR4⁺ cells were found to exert a more invasive and metastatic phenotype (15, 22). CD133⁺/CXCR4⁺ CSCs were detected in the invasive margin of pancreatic tumors, and removal of this subpopulation was shown to prevent tumor metastasis (14, 22). Moreover, the ligand of CXCR4, stromal cell-derived factor-1, was shown to enhance the *in vitro* chemotactic migration of CD133⁺/CXCR4⁺ PaCa cells, while this migration was blocked by CXCR4 blocking/inhibiting antibodies (17). In addition, eliminating CD133⁺/CXCR4⁺ cells by AMD-3100, which targets CXCR4, prevented the development of liver metastases in mice without affecting tumor-initiating ability (17). Since CD133⁺ PaCa cells represent distinct CSC subpopulations, all these markers (CD24⁺/CD44⁺/ESA⁺/CD133⁺/CXCR4⁺, or even more markers, such as ALDH1) may be required for CSC enrichment (4, 19, 22).

Signaling Pathways Important for Pancreatic CSCs

Signaling pathways have been implicated in PaCa tumorigenesis, prognosis, and resistance to therapy; thus, the identification of these pathways is considered a promising therapeutic strategy for the development of molecularly targeted therapies (18). CSCs are likely regulated by developmental pathways similar to those that regulate normal stem cells, including NOTCH, HH, WNT, polycomb complex protein (BMI1), and phosphatase and tensin homolog (PTEN) (17-19). Understanding the role of these key signaling pathways in CSC maintenance may lead to promising therapeutic targets in the treatment of PaCa and other types of cancer (22).

NOTCH signaling pathway. The NOTCH signaling pathway is considered a potential contributor to CSC maintenance and plays an important role in regulating the balance between self-renewal and differentiation during normal pancreatic tissue development (19, 22). Several studies have shown that pancreatic CSCs express high levels of NOTCH1 and NOTCH2 (22). NOTCH signaling is inhibited by the γ -secretase inhibitor (γ -SI), which leads to the depletion of pancreatic CSCs and impaired function (22). Up-regulation of several NOTCH pathway components in pancreatic CSCs compared to PaCa cells has been shown, and the inhibition of Hes family BHLH transcription factor 1 (HES1) (a key

NOTCH target gene) by a γ -SI or small interfering RNA (siRNA) reduces pancreatic CSC self-renewal and tumorigenicity (22). Interestingly, although the NOTCH pathway has been shown to regulate the HH pathway by repressing γ -SI by HES1, no significant changes were observed in HH signaling components when NOTCH signaling was inhibited (29).

NOTCH signaling is also known to contribute to the regulation of EMT. Acquisition of the EMT phenotype in gemcitabine-resistant PaCa cells was found to be consistent with the up-regulation of *NOTCH2*, *NOTCH4*, and *CD339* (*JAGGED-1*). Inhibition of NOTCH signaling using siRNA partially reversed the EMT phenotype. Collectively, these findings indicate that the NOTCH pathway is involved in the self-renewal of pancreatic CSCs and the EMT process (22). Therefore, the NOTCH signaling pathway represents a promising therapeutic target for targeting pancreatic CSCs and metastasis.

Hedgehog signaling. The HH pathway is an evolutionarily conserved pathway essential for self-renewal of CSCs. It is required for normal stem and progenitor functions as well as pancreatic morphogenesis and cellular differentiation; thus, dysregulation of HH signaling is considered one of the key events in PaCa pathogenesis (30). Transgenic mice that overexpressed Sonic HH (SHH), a ligand of HH signaling, in the pancreatic endoderm developed PanIN-like lesions that contained *K-RAS* mutations and overexpressed human epidermal growth factor receptor 2 (HER2/neu), revealing that the HH signaling pathway plays an early role in pancreatic tumorigenesis. Recent evidence also demonstrated that SHH and the other HH signaling components are highly expressed in pancreatic CSCs, but not in normal pancreatic stem cells or pancreatic ductal epithelial cells (22). The natural compounds sulforaphane, epigallocatechin-3 gallate (EGCG), and quercetin have been reported to impede the self-renewal capacity of pancreatic CSCs by attenuating the HH signaling pathway (31), indicating that HH signaling plays an important role in pancreatic CSCs.

WNT/ β -catenin pathway. Activation of the WNT/ β -catenin pathway has been shown to be involved in PaCa development and progression (32). Although aberrant cytoplasmic and nuclear expression of β -catenin is frequently found in PaCa and PanIN tissue samples, it is not found in normal pancreatic tissues. High transcriptional activity of WNT/ β -catenin in PaCa is associated with poor disease-specific patient survival, indicating the clinical significance of this pathway (22). Activation of the WNT/ β -catenin signaling pathway in CSCs leads to resistance to conventional therapies. Increased activation of WNT/ β -catenin signaling results in an enhanced stem cell-like phenotype of PaCa (33).

Other pathways important to pancreatic CSCs. In addition to the above-mentioned embryonic signaling pathways, other signaling pathways, such as autophagy, forkhead box protein M1 (FOXO1) signaling, interleukin 8 (IL8/CXCR1), mechanistic target of rapamycin (mTOR), and NODAL/ACTIVIN signaling pathways, and the *K-RAS/c-Jun*-NH2-kinase (JNK) axis have also been shown to be involved in the regulation of PaCa CSC activity (22); however, the significance of these signaling pathways has not been clarified.

Major Obstacles and Potential Strategies for Targeting Pancreatic CSCs

CSC drug resistance. CSCs are thought to be intrinsically resistant to chemotherapeutic agents and radiation therapy, although the mechanism of this resistance is not entirely clear (18). Cancer cells that acquire resistance to chemotherapeutic agents may display cross-resistance to a broad spectrum of structurally-unrelated drugs due to MDR (34). The most common drug resistance mechanisms include metabolic inactivation, efflux of the drug from the cells, and mutation or overexpression of the drug target (35). Numerous studies have revealed that the MDR phenotype in tumors is associated with the overexpression of certain drug efflux transporter proteins, the ATP-binding cassette (ABC) transporters (36). These proteins bind ATP and use the energy to transport various molecules across cell membranes, reducing the effects of the drugs in cancer cells. This is also one of the principal mechanisms for protecting stem cells. Therefore, there is renewed attention being paid to the role of ABC transporters in CSC biology (35).

Transporters such as P-glycoprotein (ABCB1), breast cancer resistance protein (ABCG2), and the MDR-associated protein (ABCC1) have been identified in CSCs (37) and are considered to be important targets for overcoming drug resistance. Among these, P-gp, an ABC transporter with broad substrate specificity, is considered to be the most significant factor in the failure of treatment for leukemia and many solid tumors. ABCG2 seems to play an important role in CSC resistance to anticancer drugs in clinical applications (38). These mechanisms have been shown to be responsible for resistance in brain, colon, and pancreatic cancer (18). In drug-resistant cancer cells, increased CD133 expression and the regulation of protein kinase B (AKT) signaling were shown to be associated with high ABCG2 levels (36). Improved DNA repair, a low mitotic rate, increased ALDH oxidation of aldehydes or detoxification, and resistance to apoptosis are also thought to be involved in mediating chemotherapeutic resistance, while increased repair capacity of the DNA damage checkpoint is involved in radiotherapy resistance in CSCs. (39)

Quiescence of CSCs. CSCs are presumably arrested at a G_0 -like cell-cycle phase or checkpoint. Since the quiescent state

of these CSCs may account for many treatment failures, the only effective way to treat cancer is to target CSCs by blocking their ability to self-renew and generate multilineage differentiation (40).

Stem-cell 'niche'. An *in vivo* or *in vitro* stem cell 'niche', a specific microenvironment for CSCs, has also been proposed to contribute to resistance *via* various mechanisms, including MDR and ABC transporters. EMT, inflammation, hypoxia, and angiogenesis further contribute to the regulation of CSCs. Thus, the CSC niche needs to be targeted since this specific microenvironment plays important roles in tumor growth, metastasis, and response to standard therapies.

Furthermore, stemness markers have been shown to be linked to chemoresistance in PaCa. CD133⁺ cells isolated from patients with PaCa have been shown to be significantly more resistant to gemcitabine than are CD133⁻ cells (41). Prolonged gemcitabine exposure leads to a decrease in tumor size in xenograft models while causing an increase in the CD133⁺ cell fraction in a dose-dependent manner. Similar results have been observed for CSCs after 5-FU exposure. On the other hand, stem cell markers such as CD44⁺CD24⁺ESA⁺ have also been found to be expressed in cells that are resistant to both gemcitabine therapy and radiotherapy. Interestingly, gemcitabine treatment eliminates only the bulk of tumor cells while leaving both the CD44⁺-enriched cell population and a small subset of CD44⁺CD24⁺ESA⁺ cells that are capable of proliferating and regenerating tumor. In an *in vitro* study, it was shown that high-dose gemcitabine eliminated most PaCa cells while failing to eliminate CD44⁺ and CD44⁺/CD24⁺CSCs; proliferating CD44⁺ CSCs generated resistant PaCa cells (42). Clinical data confirmed this finding, showing that patients with PaCa with a poor prognosis have CD44 positivity (43). These findings suggest that PaCa CD44⁺/CD24⁺CSCs are responsible for gemcitabine resistance and, thus, poor prognosis. A study also demonstrated that gemcitabine-resistant human PaCa tissues, which are rich in PaCa CD133⁺ CSCs, are highly tumorigenic, as they are capable both of forming colonies and resisting conventional chemotherapy (44). A cell adhesion molecule called CXCR4, the receptor for CXCL12 (stromal cell-derived factor-1, SDF1) that was found only at the invasive margin of the tumor, was shown to be responsible for the difference in response to therapies, leading to metastasis in a murine model (18).

Potential Therapies Targeting CSCs

Reactive oxygen species (ROS) inhibitors. While investigating the molecular mechanisms underlying chemoresistance to 5-FU and gemcitabine therapies in PaCa, Suzuki *et al.* discovered that the JNK pathway, through

suppression of intracellular ROS levels, plays a critical role in the development of resistance in pancreatic CSCs (45). Targeting the JNK-ROS axis, in combination with 5FU or gemcitabine-based regimens, may be a promising approach to eliminating pancreatic CSCs for the more successful treatment of PaCa.

Acetyl salicylic acid. Aspirin was shown to inhibit cell signaling for self-renewal capacity, tumor growth, and invasion by inhibiting markers of inflammation, and it sensitized CSCs to gemcitabine-mediated cytotoxicity both *in vitro* and *in vivo*. It has been demonstrated that aspirin can inhibit the characteristic features of CSCs through inhibition of inflammatory activity, self-renewal potential, stem cell marker expression, tumor growth, metastasis and stromal reaction, and can enhance the efficacy of gemcitabine (46).

mTOR inhibitors. Since the mTOR signaling pathway is aberrantly activated in various human malignancies and plays a crucial role in CSCs, mTOR inhibitors have been proposed and used as a novel strategy for eliminating CSCs. Various mTOR inhibitors, including rapamycin, everolimus, temsirolimus, and ridaforolimus, have been used to target CSCs (47). Gemcitabine treatment of PaCa cell lines or cells derived from primary human PaCa tumors leads to enrichment of the CD133⁺ CSC population. Rapamycin reduces the viability of CD133⁺ PaCa cells and sphere formation, which is indicative of the self-renewal of stem-like cells, indicating that the mTOR pathway maintains cancer stem-like cells (48, 49). Combining gemcitabine with rapamycin has been found to highly suppress CSC survival, suggesting that mTOR inhibitors may be used especially in combination with standard therapies to target CSCs (48, 49).

Metformin. Metformin is an anti-diabetic drug that has been implicated in targeting CSCs, including pancreatic CSCs; it reduces the expression of self-renewal-associated genes such as *NANOG*, *OCT4*, and *SOX2* in cells that are positive for CD133, CD44, CXCR4, and stage-specific embryonic antigen 1 (SSEA1). Metformin exerts its action by activating the liver kinase B1- AMP-activated protein kinase (LKB1-AMPK) axis and thereby indirectly inhibiting the mechanistic target of rapamycin complex 1 (mTORC1) (52). Mohammed *et al.* investigated the effect of metformin on PanIN, and found that the incidence of PDAC was reduced by about 20% with metformin treatment; while tumor weight and tumor metastasis were also decreased in a dose-dependent manner in mice (53).

The expression of CSC markers in pancreatic tissue were found to be significantly reduced; as indicated by lower levels of CD44-, CD133-, ALDH1- and epithelial cell adhesion molecule (EPCAM)-positive cells. These results suggest that the biological effects of metformin are mediated

by a decreased number of CSCs, indicating that metformin has significant potential for targeting pancreatic CSCs (54).

Metformin was also found to increase ROS production in CSCs and reduce mitochondrial transmembrane potential by hampering the cells' self-renewal capacity; it significantly reduced the size and number of tumor spheres and delayed the formation of secondary and tertiary tumorspheres. In cancer cells, metformin reduces cell proliferation and protein synthesis (55).

Glucose transporter 1 (GLUT1) inhibitors. CSCs are dependent on glycolysis for their survival and growth; thus, glucose metabolism is even more active in CSCs. Such increase in glucose metabolism is now known to be a hallmark of cancer (54). GLUT1 is a facilitative glucose transporter that is essential for the maintenance of pancreatic CSCs, and a specific GLUT1 inhibitor, WZB117, was found to be capable of suppressing the self-renewal and tumor-initiating capacities of CSCs without compromising their proliferative potential *in vitro* via inhibition of GLUT1. The systemic administration of WZB117 also inhibited tumor initiation *in vivo* after CSC implantation without causing significant side-effects in host animals. Since these results indicate that GLUT1-dependent glucose metabolism has a crucial role not only in the growth and survival of CSCs but also in the maintenance of their stemness, GLUT1 may also be considered a promising target for CSC-directed cancer therapy (56).

Histone deacetylase (HDAC) inhibitors. HDAC inhibitors, including suberylanilidehydroxamic acid (SAHA), reactivate aberrantly silenced genes by restoring histone acetylation. SAHA has been shown to inhibit cell growth and induce apoptosis in many types of human cancer by activating caspase-3, caspase-9, peroxisome proliferator-activated receptors (PPAR) cleavage, cytochrome *c* release, and the up-regulation of FAS and FAS ligand expression. A combination of SAHA and the SMOOTHENED antagonist SANT1 was shown to suppress cell growth and induce apoptosis in gemcitabine-resistant PaCa cell lines. Moreover, SAHA was shown to be capable of inhibiting cell growth and inducing apoptosis, differentiation, and cell cycle arrest by up-regulating *p21*, CCAAT/enhancer binding protein alpha (*C/EBPα*), retinoic acid receptor alpha (*RARA*), and E-cadherin while down-regulating cyclin B1, cyclin D1, and c-MYC, suggesting that it is a promising therapeutic agent for human pancreatic CSCs (54).

Salinomycin. Salinomycin is another drug that is used to target CD133⁺ pancreatic CSCs; it was shown to be effective at eradicating PaCa xenografts in mice when used in combination with gemcitabine (57). The mechanism of action by which salinomycin kills CSCs specifically is not well established but it is thought to function as a potassium

ionophore. Salinomycin and its derivatives exert significant antiproliferative activity towards drug-resistant cancer cell lines. Salinomycin can induce apoptosis of human cancer cells and effectively eliminate CSCs and induce partial clinical regression of heavily pretreated and therapy-resistant cancer (58).

Sorafenib. Sorafenib is an Food and Drug Administration (FDA)-approved small inhibitor of several tyrosine protein kinases, such as vascular endothelial growth factor receptor, platelet-derived growth factor, and serine/threonine-protein kinase (RAF) family kinases. Sorafenib is used for the treatment of advanced kidney and liver cancer and has been shown to target pancreatic CSCs and induce apoptosis and decrease proliferation, spheroid formation, clonogenicity, ALDH1 activity, and angiogenesis. Its use leads to an increase in survival (59, 60). Sulforaphane, an isothiocyanate found in broccoli, was shown to eliminate pancreatic CSCs by down-regulating NF-κB activity without inducing toxic side-effects; this activity was enhanced when it was combined with sorafenib (61).

Irinotecan plus gemcitabine. Recently, Qin *et al.* developed an electrospun scaffold made of polycaprolactone and gelatin to facilitate the survival and tumorigenesis of CD24⁺CD44⁺ CSCs in PaCa tumor murine models *in vivo*; they found increases in both tumor formation incidence and *in vivo* tumor growth and concluded that the homogeneous nature of CD24⁺CD44⁺ CSCs significantly reduced the biological variation of PaCa tumor masses harvested from patients. When they evaluated the therapeutic effect of irinotecan plus gemcitabine on murine models of pancreatic tumor *in vivo*, they discovered that it was a promising chemotherapy for PaCa because of its devastating influence on CD24⁺CD44⁺ pancreatic CSCs (62).

Carbon ion beam. Combined with gemcitabine, carbon ion beam therapy has a superior effect on pancreatic CSCs and tumor growth *in vitro* and *in vivo* at relatively low doses in comparison to carbon ion beam alone or conventional x-ray irradiation. Carbon ion beam combined with gemcitabine synergistically enhanced the death of pancreatic CSCs by inhibiting DNA repair, as well as by inducing irreparable complex DNA damage *via* increasing apoptosis and autophagy and inhibiting cell proliferation at relatively low doses compared to carbon ion beam alone. Taken together, these results demonstrate the benefits of carbon ion beams in combination with chemotherapy in targeting conventional radioresistant locally advanced PaCa (63).

Inhibitors of facilitates chromatin transcription (FACT). CBL0137 is a member of a new class of recently discovered candidate anticancer agents that target FACT (SSRP1 and

SPT16 subunits), as monotherapy or in combination with gemcitabine. In patient-derived PDAC xenografts and PANC-1 orthotopic tumors, CBL0137 inhibited gemcitabine-induced CSC enrichment; this, together with the *in vivo* data, make CBL0137 a reasonable and promising adjuvant treatment for gemcitabine since CBL0137 kills the CSCs while gemcitabine eliminates the remaining bulk population. As a result, CBL0137 is effective at preventing cancer recurrence (64).

Insulin-like growth factor-1 receptor (IGF1R) and epidermal growth factor receptor (EGFR) inhibitors. The impact of the concurrent inhibition of IGF1R and EGFR (HER2) has been shown in PaCa cell lines and particularly in pancreatic CSCs. NVP-AEW541 and lapatinib in combination simultaneously inhibited IGF1R and ERBB receptors/HER2 and thus prevented the resistance observed at the molecular level with individual treatments. Interestingly, these inhibitors eliminated PaCa cells and overcame their resistance to conventional chemotherapy. Thus, the synergy observed with this combined treatment indicates that the appropriate combination of currently known anticancer agents is efficient at maximizing patient benefit (65).

Carboxypeptidase inhibitors. Latexin is the only known mammalian carboxypeptidase inhibitor; it is an antigen expressed in a subset of neurons in the rat cerebral cortex, as well as in various types of non-neural tissues, such as heart, prostate, ovary, kidney, pancreas and colon tissue. Latexin shares 30% sequence similarity with tazarotene-induced gene 1, which is down-regulated or absent in many tumor subtypes. Xue *et al.* investigated the differential expression of latexin in both CD133⁺ PaCa stem-like cells and CD133⁻ PaCa cells. They tested the effect of exogenous latexin in inducing apoptosis and inhibiting proliferation in CD133⁺ PaCa stem-like cells and studied the underlying mechanisms. It was demonstrated that latexin induces apoptosis in and inhibits the proliferation of CD133⁺ pancreatic CSCs derived from MiaPaca-2 by modulating the BCL2 family and c-MYC; thus, targeting latexin may be a new therapeutic strategy for PaCa (66).

Role of MicroRNA (miRNA, miR) in Pancreatic CSCs

MicroRNAs (miRNAs are endogenous non-coding short RNAs (length of 21-23 nucleotides) encoded by nuclear DNA that are involved in the posttranscriptional regulation of gene expression. By binding to specific sequences of messenger RNA (mRNA) complementarily, they lead to gene silencing *via* translational repression or target degradation. miRNAs play a crucial role in various developmental, metabolic, and cellular processes, such as apoptosis, cell

proliferation and differentiation, by altering the expression of proto-oncogenes or tumor-suppressor genes and thus regulating their levels in various tumor tissues (67). Moreover, miRNAs have a crucial role in tumorigenesis: an important alteration is found in the expression of proto-oncogenes or tumor-suppressor genes in cancer, including PaCa, which is associated with clinical prognosis, therapy resistance and tumor recurrence or relapse. By affecting signaling pathways and CSC signature genes, miRNAs are also able to regulate CSC characteristics (68). Indeed, several microRNAs have already been implicated in the regulation of normal stem cells and CSCs.

miR-1181 was recently shown to inhibit stemness and tumorigenicity. *miR-1181* expression suppressed the CSC phenotypes of PaCa by targeting SOX2 and signal transducer and activator of transcription 3 (STAT3) (69). SOX2 has an important function during embryonic development and is involved in the maintenance of CSC phenotypes by directly targeting the genes that control tumor survival, proliferation, stemness, and invasion. SOX2 is overexpressed in both poorly differentiated PaCa and highly invasive PaCa. STAT3 is frequently overexpressed in PaCa and plays an important role in its pathogenesis. Since *miR-1181* was found to be down-regulated in PaCa cell lines and clinical patient tissues, its therapeutic delivery and expression in PaCa tumors may be an important strategy in targeting pancreatic CSCs and cancer cells (69). *miR-21* and *miR-221* are up-regulated in pancreatic CSCs and contribute to important biological functions in cancer progression by promoting cell proliferation and migration and chemotherapy resistance. Inhibition of *miR-21* and *miR-221* by targeting CSCs may be a therapeutic strategy for PaCa (70).

miR-17-92 cluster: Epigenetic mechanisms may also account for the strong phenotypical differences seen in CSCs. The miR-17-92 cluster is down-regulated in chemoresistant CSCs. NODAL/ACTIVIN/TGFβ1 signaling, which promotes chemoresistance in CSCs, is inhibited by the miR-17-92 cluster. Overexpression of *miR-17-92* leads to the abrogation of CSC phenotypes and the eventual loss of the *in vivo* tumorigenicity of CSCs (71).

LIN28B is an RNA-binding protein that regulates cell growth and differentiation. A novel CSC subpopulation that overexpresses both CD44 and LIN28B at the cell surface is found in human primary PaCa tissues; this CD44⁺/LIN28B⁺ pancreatic CSC subpopulation proliferates rapidly and exhibits MDR, highly invasive ability, and high adherin levels. Therefore, CD44⁺/LIN28B⁺ pancreatic CSCs may represent a powerful *in vitro* model, either to study cancer cell metastasis, invasion, and self-renewal or to assess the effectiveness of novel therapeutics for PDAC. CD44⁺/LIN28B⁺ pancreatic CSCs were more resistant to growth inhibition when cisplatin and gemcitabine hydrochloride were used as chemotherapeutic drugs and

rapidly and easily formed tumors *in vivo*. siRNA interference in endogenous *LIN28b* gene expression in these CD44⁺/LIN28B⁺ pancreatic CSCs not only reduced their proliferation but also inhibited the cell cycle due to the suppression of cyclin D1 expression after the stimulation of miRNA *LET-7B* expression (72).

miR-1246 has been shown to induce chemoresistance and contributes to CSC stemness in PaCa cell lines. *miR-1246* targets and controls the function of the cyclin-G2 (*CCNG2*) tumor suppressor gene. Given that *CCNG2* participates in inhibiting cancer proliferation, invasion, differentiation, and chemoresistance, which characterize CSCs, *CCNG2* inhibition may be at least partially responsible for the maintenance of CSC-like spheroid cells. *CCNG2* expression is negatively correlated with *miR-1246* expression, suggesting that the *miR-1246*–*CCNG2* axis is critical for chemoresistance and can be used to control CSCs (73).

miR-335: An miRNA array assay showed that *miR-335* is linked to OCT4 expression in PaCa. When OCT4-overexpressing cells were infected with LV-*miR-335*, migration and invasion decreased as did levels of the mesenchymal markers fibronectin, vimentin, α -Smooth muscle actin, and snail family transcriptional repressor 1, while those of the epithelial marker E-cadherin increased in PaCa cells, suggesting that *miR-335* can inhibit metastasis and EMT in PaCa cells. Systemically delivered *miR-335* was found to inhibit PaCa metastasis and extend animal survival (74).

Immunotherapy for Pancreatic CSCs

Immunotherapy is a promising strategy that offers a complementary method for successfully treating cancer. The current strategies consist of cancer vaccines as active immunization, monoclonal antibodies as passive immunization, and cellular therapies that often include lymphocytes and dendritic cells. Cancer vaccines may be effective at preventing cancer by attacking various tumor cells, inducing an immune response (75).

Interferon- γ (IFN γ) belongs to a group of type-I interferons and exerts its cellular activity by binding to a specific membrane receptor on the surface of many types of cells, including malignant cells. IFN γ shows its therapeutic effects by improving the differentiation, maturation, and function of dendritic cells, enhancing the survival of T-cells through the expression of anti-apoptotic genes, generating CD8⁺ memory cells, enhancing macrophage activities, and activating natural killer cells and has already been approved for clinical cancer therapies of melanoma and renal cell carcinoma. Interestingly, IFN γ up-regulates the expression of the CSC markers CD24, CD44, and CD133 in *in vitro* and *in vivo* models of PDAC in correlation with the original level of surface marker expression. While IFN γ has a certain cytotoxic effect on PDAC cells and reduces their numbers,

it conversely enhances the enrichment of PDAC CSCs. Given that IFN γ 's effects on the migration and invasion of PDAC cells are associated with the level of CSC marker expression *in vivo*, IFN may promote metastasis in the early stage of tumor growth while inhibiting tumor growth; thus, it needs to be carefully considered as a therapeutic option in PDAC (76).

EpCAM is a marker of poor prognosis and is overexpressed in many types of human cancer, including PaCa. Therapies targeting EpCAM can inhibit various types of cancer. Recently, catumaxomab, a bi-specific antibody that binds to EpCAM on tumor cells and CD3 on T-cells for activation, was approved by the FDA to treat malignant ascites. The combination of catumaxomab and activated T-cells has been shown to eradicate pancreatic CSCs. Pre-treatment with catumaxomab, followed by the addition of IL2/OKT3-activated autologous T-cells, eliminated CSCs during a short incubation period. Moreover, the CSCs, which became more aggressive when MU-PK1 cells were cultured under hypoxic conditions were successfully lysed by the combination of cytokine-activated killer T-cells with catumaxomab. In conclusion, catumaxomab combined with activated T-cells may be a potent therapeutic modality for eradicating chemoresistant pancreatic CSCs (77).

CSC vaccines: Pancreatic CSCs that were isolated from tumor specimens and cultured were used to produce a vaccine that was evaluated for its safety and efficacy in low-, medium-, and high-dose groups (78). When comparison of pre- and post-vaccination immunity was made; it was found to be significantly strengthened in the high-dose group with no side-effects, suggesting that the pancreatic CSC vaccine is safe and effective. However, from the viewpoint of a long-term curative effect, the benefit of the CSC vaccine on progression-free survival and overall survival requires further refinement (78).

Dendritic cells (DCs) are potent antigen-presenting cells that play a crucial role in inducing primary immune responses against tumor-associated antigens. To generate anti-tumor immune responses, DCs have been charged with pancreatic CSC antigens (79). After being co-cultured with lymphocytes at different ratios, the lysate-exposed DCs effectively promoted lymphocyte proliferation and induced the secretion of high levels of INF γ and IL2, which are strong antitumor cytokines. The DCs had significant cytotoxic effects on Panc-1 CSCs and parental Panc-1 cells.

Natural Dietary Compounds Against Pancreatic CSCs

Recently, dietary compounds have gained substantial attention for their potential therapeutic applications in cancer, leading to the novel term 'nutraceutical', generated by combining the words nutrition and pharmaceutical. Indeed,

nutraceuticals, particularly soy isoflavone genistein, curcumin, resveratrol, quercetin, EGCG, and lycopene, can inhibit not only cancer cells but also CSCs in PaCa and other cancer types (80).

Genistein. Soybeans are rich in natural phytoestrogenic isoflavones, particularly genistein, daidzein, and glycitein. Genistein (4,5,7-trihydroxyisoflavone) has multiple biological effects in various human cancer types, with low toxicity towards normal cells (81). Genistein inhibits cell growth, migration, invasion, angiogenesis, and metastasis through the regulation of multiple cellular signaling pathways, including the inhibition of NF- κ B, WNT, NOTCH1, and HH pathways, and by acting as a protein tyrosine kinase inhibitor (mostly of EGFR) in human cancer (82). Genistein inhibits cell growth and pancreatosphere formation and reduces the expression of CSC surface markers, mainly by down-regulating the NOTCH pathway. Moreover, it has been shown to induce apoptosis in PaCa cell lines by inhibiting NF- and NOTCH1 signaling. Genistein attenuates β -catenin-mediated expression of WNT downstream target genes in mammary epithelial cells by up-regulating E-cadherin. It modulates the expression of genes involved in cellular functions, such as proliferation, apoptosis, and angiogenesis, particularly by inhibiting or down-regulating the AKT and NF- κ B pathways (83). Genistein is also capable of potentiating the antitumor effects of chemotherapeutic agents (*e.g.*, gemcitabine, cisplatin, and oxaliplatin) by modulating the apoptotic pathways (84). In orthotopic animal models, genistein combined with gemcitabine synergistically increased the growth inhibition of PaCa cells through NF- κ B inhibition. It also significantly improved the outcome of patients with advanced PaCa treated with erlotinib, an inhibitor of EGFR signaling, and gemcitabine (84). Genistein can overcome cancer drug resistance and inhibit cancer relapse and recurrence (85). Overall data suggest that genistein potentiates anticancer effects by promoting both apoptotic and autophagic cell death and inhibiting multiple signaling pathways in PaCa and PaCa-derived CSCs (86).

Curcumin. Curcumin (diferuloylmethane) is present in the rhizome of turmeric (*Curcuma longa*) (87, 88). Curcumin exerts anticarcinogenic, antioxidant, antimicrobial, and anti-inflammatory activities, has hepatoprotective and renoprotective properties, and has hypoglycemic effects. In preclinical studies using both *in vitro* and *in vivo* models, curcumin exhibited antiproliferative, antioxidant, anti-inflammatory, and pro-apoptotic effects, leading to antitumor effects in various cancer types, including thyroid, lung, and breast cancer, hepatocellular carcinoma, and PaCa (89, 88). Moreover, curcumin inhibited the growth, migration, angiogenesis, invasion, and metastasis of PaCa cells by

suppressing multiple cellular signaling pathways, such as AKT, NF- κ B, and NOTCH (87). It was also found to inhibit the activity of distinct signaling pathways, including mTOR, HH, EGFR, STAT3, and multidrug transporters such as MDR-associated protein 5 (ABCC5). It modulates the expression levels of different oncogenic and tumor suppressor microRNAs (90). In addition, curcumin has synergistic effects with gemcitabine (91). By modulating the activation of various transcription factors, curcumin regulates the expression of inflammatory enzymes, cytokines, adhesion molecules, and cell survival proteins (92). Curcumin also down-regulates cyclin D1, cyclin E, and mouse double minute 2 homolog (MDM2) and up-regulates *p21*, *p27*, and *p53* (93).

Recent preclinical and clinical studies have demonstrated the antitumor and anti-angiogenic properties of curcumin (94). Curcumin enhanced the effects of 5-FU and oxaliplatin in mediating the growth inhibition of colon cancer cells by modulating EGFR and IGFR (95, 96). The efficacy of curcumin in the treatment of PaCa was also assessed: both phase I and II clinical trials have yielded promising results on the use of curcumin in PaCa therapy. More importantly, curcumin was safe and non-toxic, even at high doses, in clinical trials and preclinical models (97, 98). Overall, data from pre-clinical and clinical studies suggest that curcumin is a safe PaCa therapeutic agent due to its broad effects on PaCa cells, the tumor microenvironment, and pancreatic CSCs.

Resveratrol. Resveratrol (trans-3,5,4'-tri-hydroxystilbene) is a natural polyphenol that is mainly found in the skins of red grapes, red wine, berries, and peanuts (99-101). Resveratrol has received a great deal of attention as a cancer-chemopreventive agent. It has been demonstrated to have an antitumorigenic capacity by inducing growth inhibition, cell-cycle arrest, apoptosis, and changes in biomarker expression in more than 30 types of tumor cells, including those originating from the pancreas (100). Resveratrol inhibits cell growth and prevents metastasis in PaCa through the induction of mitochondrial dysfunction, cytochrome *c* release, caspase activation, and apoptosis (100). Resveratrol can suppress PaCa cell migration, invasion, and the progression of EMT through inhibition of the PI3K/AKT/NF- κ B signaling pathway. It has been found to suppress the growth and self-renewal capacity of pancreatic CSCs derived from *k-ras* transgenic mice and human primary tumors. Therefore, the potential effect of resveratrol against CSCs needs to be further investigated, especially its effects on signaling pathways (102). Resveratrol has been shown to directly inhibit the proliferation and viability of human PaCa cells *in vitro* in a dose- and time-dependent manner (100, 101). Resveratrol treatment may thus be a novel therapeutic option for PaCa by inhibiting the HH signaling pathway (101).

Quercetin. Flavonoids are one of the most actively studied classes of molecules for their potential to prevent cancer. Quercetin, 3,3',4',5,7-pentahydroxyflavone, is a natural dietary polyphenol and flavonol-type flavonoid that is ubiquitously present in fruits and vegetables, such as broccoli, onions, tea, apples, and berries (103-105). It has been well-documented that quercetin is a potential anticancer agent in *in vitro* and *in vivo* models. Quercetin exerts its anticancer effect through the inhibition of several intracellular pathways in cancer cells, including PI3K/Akt/mTOR, glycogen synthase kinase 3 (GSK3 β), NF- κ B, and heat-shock protein 70 (105, 106). It elicits antitumor effects by acting as an antioxidant; modulating signaling pathways; inducing apoptosis, autophagy, and cell cycle arrest; blocking cell migration protein kinase C inhibitory activity; and inhibiting the fatty acid synthesis required for *de novo* membrane synthesis (106-108). Quercetin also suppressed local and distant tumor growth and prolonged survival in murine PaCa models. Quercetin itself showed growth-inhibitory activity in both drug-sensitive and MDR1 cells. In addition, at a non-cytotoxic concentration, it enhanced the effect of chemotherapeutic drugs in MDR cells. Quercetin has also been shown to act as a chemosensitizer for ABC pump-proteins in MDR tumor cell lines. Furthermore, it interacts directly with transporter proteins to inhibit drug efflux, mediated by either MDR1, MRP1, or BCRP (109, 110). Quercetin also suppressed local and distant tumor growth and prolonged survival in murine PaCa models (104). Quercetin targets pancreatic CSCs by inhibiting the β -catenin signaling pathway (106). It reduced the self-renewal properties of CSCs and ALDH activity in PaCa (103, 107). More importantly, quercetin showed synergistic effects in eliminating pancreatic CSCs *in vitro* and *in vivo* when combined with sulforaphane (107). Quercetin may be an important modulator of cancer cell sensitivity to anticancer chemotherapeutic agents (106-108). Quercetin 3-O-glucoside and gemcitabine co-treatment was found to have an additive or synergistic anti-migratory effect on human PaCa cells, suggesting that this drug combination may reduce the risk of side-effects (111).

EGCG. As the most abundant catechin found in green tea and recognized for its potent chemopreventive properties, EGCG induces growth inhibition and apoptosis of various PaCa cell lines. *In vivo* studies have also demonstrated the inhibitory effect of green tea on tumorigenesis in nitrosamine-induced pancreatic tumors (112). EGCG inhibits angiogenesis, possibly through the inhibition of proangiogenic factors, including vascular endothelial growth factor (113). It has been found to be most effective against cancer of the gastrointestinal tract. EGCG has been shown to inhibit NF- κ B activity, the MAPK pathway, activator protein-1 activity, and EGFR-mediated downstream signaling pathways. The antiproliferative effects

of EGCG on PaCa cell growth *in vitro* are potentiated by treatment with pterostilbene (114).

Lycopene. One of the most extensively studied carotenoids in tomatoes, lycopene, possesses potent antioxidant activity due to its extended conjugated hydrocarbon chain. Lycopene is a acyclic isomer of β -carotene; it is synthesized by plants and microorganisms, not animals, and is a natural pigment. Lycopene has been shown to prevent carcinogenesis. Lycopene induces apoptosis and inhibits cell-cycle progression in various cancer cells, and its efficacy against xenograft tumors has been reported in a number of *in vivo* studies (115).

Propolis and caffeic acid phenethyl ester (CAPE). Propolis, a sticky hive product collected by bees from various plant sources, is known to have pharmacological activity, including anticancer, antioxidant and anti-inflammatory effects (116,117). Bioactive components from propolis such as CAPE have been studied and shown to have antioxidant and anti-inflammatory properties that involve the inhibition of enzyme activities, such as the activation of xanthine oxidase, cyclo-oxygenase, and transcriptional factor NF- κ B. CAPE inhibits EMT in PaCa and is a potent apoptosis-inducing agent (118). Its antiproliferative activity may involve the induction of mitochondrial dysfunction and the activation of caspase-3/caspase-7 (119). CAPE induces apoptosis after autophagy inhibition in a caspase-dependent and a caspase-independent manner in PaCa cells (120). Coral *et al.* investigated the effect of CAPE on breast CSCs derived from aggressive triple-negative breast cancer cells and showed CAPE has effects on CSC self-renewal, progenitor formation, CD44 cell marker phenotype, cell cycle, and apoptosis (121). Their results strongly suggest that CSCs are induced into a less malignant state after CAPE treatment and may terminally differentiate their progeny, making them more susceptible to chemotherapy.

Future Perspective and Conclusion

In addition to providing an elegant model of carcinogenesis, the CSC concept has important clinical implications since targeted therapies that eliminate CSCs offer the potential to improve therapy, maintain remission, or lead to a complete cure. Emerging evidence shows that CSC-targeted therapies are effective in a preclinical setting, with a marked survival benefit. Although further studies are needed to strengthen these findings, CSCs seem a powerful target for more effective therapies in cancer. Regarding implementing all of the information generated by the preclinical studies, a practical approach to clinical translational may involve systems biology and bioinformatics approach to pinpoint signal hubs, molecular mediators and cross-roads that are

Table I. Summary of compounds that have an effect on cancer stem cells.

	Pathway	Reference
Drugs		
Gemcitabine	JNK	45
Aspirin	NF-κB	46
Rapamycin/everolimus/temsirolimus/ridaforolimus	mTOR	48, 49
γ-Secretase	NOTCH	22
Metformin	AMPK/mTOR	52, 53
Salinomycin	NF-κB	57, 58
Sorafenib	NF-κB, HH	59, 60
WZB117	GLUT1	56
Natural compounds		
Genistein	NOTCH, WNT, NF-κB	82, 84
Curcumin	NF-κB, AKT, NOTCH, mTOR, STAT3, ERK	87
Resveratrol	PI3K/AKT/NF-κB	102
Quercetin	PI3K/AKT/mTOR, NF-κB, HH	105, 106
Epigallocatechin-3-gallate	NF-κB, MAPK, HH	114

common to all of the molecular signaling pathways required for CSC survival and maintenance indicated by the preclinical studies. These studies could provide a foundation for rationally designed molecularly therapies targeting CSCs. To succeed, future studies should focus on the identification of i) CSC-specific pathways that can be pharmacologically targeted, ii) CSC-specific surface markers for antibody therapy, iii) gene silencing approaches by siRNA or miRNAs, and iv) natural products that promote the differentiation of CSCs into progenitors that do not self-renew or that differentiate only into normal tissue cells (summarized in Table I) (17). Although CSCs represent a very intriguing target for therapy, the CSC concept still leads to many unanswered questions. A major future goal is to address the question of clonal evolution, particularly a monitoring of CSCs during cancer development and after treatment. While CSCs may be an important target for therapy, it remains to be determined whether targeting them is the best way to neutralize their ability to progress, expand, and resist treatment in the host environment (122).

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