Review

The NFkB Signaling Pathway in Papillomavirus-induced Lesions: Friend or Foe?

RUI M. GIL DA COSTA^{1,2}, MARGARIDA M.S.M. BASTOS¹, RUI MEDEIROS^{2,3,4,5}, PAULA A. OLIVEIRA⁶

¹Laboratory for Process Engineering, Environment, Biotechnology and Energy (LEPABE),
Chemical Engineering Department, Engineering Faculty, University of Porto, Porto, Portugal;

²Molecular Oncology Group, IPO-Porto Research Centre (CI-IPOP) and Virology Laboratory,
Portuguese Oncology Institute, Porto, Portugal;

³Abel Salazar Institute for Biomedical Sciences, University of Porto, Porto, Portugal;

⁴Biomedicine Research Centre, Health Sciences Faculty, Fernando Pessoa University, Porto, Portugal;

⁵Research Department, Portuguese League against Cancer, Porto, Portugal;

⁶Biological and Agroenvironmental Technology Research Centre (CITAB),
University of Trás-os-Montes and Alto Douro, Vila Real, Portugal

Abstract. Papillomaviruses induce a range of benign and malignant lesions in their hosts, including cervical cancer, that is associated with high-risk human papillomavirus (HPV) types. The nuclear factor kappa-light-chain-enhancer of activated B-cells (NFkB) plays a pivotal role in HPV-infected cells, and its expression and activity are modulated by several viral oncoproteins. NFkB modulation seems to first facilitate viral persistence and immune evasion, and later to drive tumour progression, but the many conflicting results and the complexity of its signaling networks require great prudence while interpreting the role of NFkB in papillomaviral lesions. Accordingly, the pharmacological targeting of the NFkB pathway in HPV-induced lesions is a complex and currently unmet challenge. This review deals with recent findings concerning NFkB activation in HPVinfected cells, its role in viral persistence, cell transformation and tumour progression, and with current efforts to target this pathway for cancer prevention and therapy.

Human papillomaviruses (HPV) are associated with a variety of benign and malignant lesions. In particular, HPV types 16

This article is freely accessible online.

Correspondence to: Professor Paula A. Oliveira, Departamento de Ciências Veterinárias, Universidade de Trás-os-Montes e Alto Douro (UTAD), Quinta de Prados, 5000-801, Vila Real, Portugal. Tel: +351 259350000, Fax: +351 259350480, e-mail: pamo@utad.pt

Key Words: Apoptosis, human papillomavirus, HPV, IKK, inflammation, NFκB, targeted therapy, review.

(HPV16) and 18 (HPV18), have long been recognized as important biological carcinogens, and are particularly associated with cervical cancer and other anogenital malignancies (1). Despite recent advances in cancer prevention, cervical cancer remains a major public health problem, with an estimated worldwide incidence of 527,624 new cases in 2012 (2). For 2013, 12,340 new cervical cancer cases were expected in the United States, as well as 4,030 deaths (3). Although HPV vaccination is expected to considerably reduce the burden of cervical cancer in developed countries, its implementation is slow in the developing world, maintaining high disease burdens (2). Another reason for concern is the increasing worldwide incidence of non-genital HPV-associated cancer, particularly HPV-associated oropharyngeal cancer (4). High-risk HPVs encode several oncogenes, most notably E6 and E7, which promote the neoplastic transformation of infected cells (5). The viral oncoprotein E6 leads to the ubiquitination and subsequent proteasomal degradation of p53, while E7 blocks the function of retinoblastoma protein (pRb), deregulating the cell cycle and blocking DNA-repair mechanisms and apoptosis.

Advanced cervical cancer poses a particularly difficult therapeutic challenge. In particular, there is a clear need for improvement in therapy for recurrent or metastatic cancer, associated with short survival periods. The standard treatment for metastatic and recurrent cervical cancer has long been cisplatin, as monotherapy or combined with radiotherapy, with a median survival of 6 months (6). However, cancer cells develop cisplatin resistance, requiring combination therapies with paclitaxel, topotecan, gemcitabine, vinorelbine or ifosfamide, in order to achieve improved median survival of 10 to 12.9 months. Monoclonal antibodies and inhibitors of

0250-7005/2016 \$2.00+.40

receptor tyrosine kinases targeting vascular endothelial growth factor receptor (VEGFR), stem-cell growth factor receptor (c-KIT) and platelet-derived growth factor receptor (PDGFR) are also under study in clinical trials (6).

In many types of cancers, including cervical cancer, resistance to chemotherapy has been found to be strongly dependent on the activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB) (7-12). The NFkB pathway and its regulation are highly complex, and may lead to cancer invasion and chemoresistance in multiple ways, including resistance to apoptosis, metabolic changes and modulation of the tumoural microenvironment through the production of inflammatory mediators (13).

This review deals with recent findings concerning NF κ B activation in HPV-infected cells, its role in viral persistence, cell transformation and tumour progression, and with current efforts to target this pathway for cancer prevention and therapy.

The NFkB Pathway

The NFκB transcription factor is composed of homoor heterodimers formed by five proteins belonging to the avian reticuloendotheliosis viral oncogene homolog (REL) which share a DNA-binding domain and a dimerization domain, the REL homology domain. These REL proteins include p50 (NFκB1), p52 (NFκB2), p65 (RELA), RELB and c-REL (REL), as reviewed by Perkins *et al.* (13). p50 and p52 are translated in the form of two protein precursors, known as p105 and p100. Proteasomal processing of these two proteins produces p50 and p52 (14). Binding of NFκB to DNA leads to transcription of its target genes, thereby modulating numerous cell functions.

Cells can rapidly activate NFkB signaling as a first line of defence against infection or otherwise stressful conditions. This requires most REL proteins to be pre-synthesized and kept in the cytoplasm in an inactive form, ready to be activated in response to adequate stimuli. REL proteins are bound by and sequestered in the cytoplasm by a family of inhibitors of NFkB (IkBs) proteins which include IkB α , IkB β and IkB ϵ (15). The ankyrin repeat motifs found in IkB proteins allow them to bind the REL subunits and are also present in the C-termini of p100 and p105, which also act as IkB-like NFkB inhibitors (16). IkB proteins also have other functions (17), although these are outside the scope of the present review.

The NFκB pathway is highly complex and subject to multiple regulatory factors that help shape its signalling activity to each cell's conditions. Figure 1 depicts certain essential aspects of the NFκB pathway, including its canonical and non-canonical activation routes. As mentioned above, the canonical pathway provides a rapid responds to stressful stimuli by activating pre-existing REL proteins. The

non-canonical pathway is involved in lymphoid organogenesis, B-cell maturation and survival, and bone metabolism, and requires the production of p52 from its inactive precursor p100, as reviewed by Sun (16). In the canonical pathway, the binding of a ligand to a receptor on the cell membrane, such as a tumour necrosis factor (TNF) receptor or a member of the interleukin-1 receptor/toll-like receptor (TLR) superfamily, allows the recruitment of adaptor proteins such as TNF receptor-associated proteins (TRAFs).

TRAF proteins recruit and activate the IkB kinases alpha $(IKK\alpha)$ and beta $(IKK\beta)$, which together with a scaffold protein known as NFkB essential modulator (NEMO or IKKγ), form the IKK complex (18). The activated IKK complex phosphorylates two serine residues (ser²³/ser³⁶) in the IκBα regulatory domain, targeting it for ubiquitination and proteasomal degradation (15). IkB phosphorylation allows the release of REL proteins and their translocation into the cell nucleus, where they may bind DNA. The noncanonical pathway is activated by non-inflammatory stimuli, such as the binding of B cell-activating factor (BAFF) to its receptor (BAFFR) (16). The non-canonical pathway is critically mediated by NFkB-inducing kinase (NIK) which activates IKKα (but not IKKβ or IKKγ), leading to p100 phosphorylation, ubiquitination and consequent proteasomal processing to generate the active p52. Processing of p100 also abolishes its IkB-like inhibitory activity. The RELB/p52 dimer is then translocated to the nucleus, where it exerts its activity as a transcription factor. NFkB signaling may also proceed through atypical pathways initiated in response to DNA damage, which provide the link between genotoxic anticancer agents and NFkB activation (19). Poly(ADPribose) polymerase 1 (PARP1) senses DNA damage and mediates phosphorylation of the NFkB essential modulator (NEMO) and its binding to small ubiquitin-like modifier (SUMO, SUMOylation) through the ataxia telangiectasia mutated (ATM) kinase, ultimately leading to NFkB activation.

HPV-Induced Carcinogenesis

High-risk HPVs, most commonly HPV-16 and HPV-18, are the aetiological agents of most anogenital (20, 21) and of a significant number of oropharyngeal (4, 22) carcinomas. The involvement of HPV in malignancies such as oesophageal cancer remains a matter of debate. Other papillomavirus induce a range of benign and malignant lesions in animal species (23). As previously mentioned, the genome of high-risk HPVs encodes a number of oncogenes, expressed early after viral infection. These *early* genes are located in the so-called *early* region of the viral genome, as opposed to the late region, encoding structural proteins. Additionally, the HPV genome also contains regulatory elements, namely the long control region (LCR), involved in regulating gene expression (24). HPV oncogenes, most notably the *E6* and *E7* genes,

promote the proliferation of infected cells, while blocking DNA repair and apoptosis, thus potentially leading to the accumulation of genetic mutations and to carcinogenesis, as reviewed by Klingelhutz and Roman (25). In fact, the E6 protein interacts with the critical tumour-suppressor p53, efficiently promoting its ubiquitination and subsequent proteasomal degradation (26). p53 protein is normally present at low levels, which are raised in response to genotoxic or cytotoxic stress. Up-regulation and activation of p53 activates multiple pathways leading to DNA repair, cell-cycle arrest and apoptosis. Apart from its central role in inactivating p53, the E6 oncoprotein also contributes to tumourigenesis by activating telomerase, disrupting epithelial-cell adhesion, polarity and differentiation, altering gene transcription and reducing immune recognition of infected cells, as reviewed by Howie et al. (27). The E7 protein binds to and induces degradation of pRb, thus deregulating the cell cycle and allowing suprabasal cells to maintain unchecked proliferative activity (28, 29).

Initially, HPVs invade mitotically active basal cells of stratified squamous epithelia, presumably through microwounds, using specific cell-surface receptors (30). Viral genomes are produced at 50-100 copies per cell, maintained in episomal form and segregated to daughter cells with support from the E1 and E2 early viral proteins. As cells migrate to suprabasal epithelial strata, they differentiate but are kept in a proliferative state by the E6 and E7 oncoproteins. This allows the expression of the late genes L1and L2, encoding the capsid proteins, which is dependent on squamous cell differentiation, and the replication of viral DNA, culminating in virion assembly and release with mature squamous cells (5). Integration of viral DNA into host chromosomes is a comparatively infrequent but critical event, as it leads to E6 and E7 overexpression, and is believed to be the fundamental event for HPV-induced cell transformation (31). HPV-driven carcinogenesis is a well-characterized multistep process that typically develops from pre-malignant intraepithelial hyperplastic and dysplastic lesions, through in situ carcinoma, to invasive squamous cell carcinoma. Advanced and metastatic cancer poses significant therapeutic challenges due to the development of chemoresistance, a trait that is often dependent on NFkB signaling (12).

NFkB Modulation by HPV Oncoproteins

Early studies suggested that high-risk HPV oncoproteins modulate the expression of NFκB-responsive genes, pointing towards their intervention in NFκB activation (32-34). James *et al.* reported that HPV16 E6 activates NFκB and induces the expression of its anti-apoptotic target, inhibitor of apoptosis C2 (cIAP2) (35). The authors also concluded that this effect depended on the presence of the interaction domain named PSD95/Dlg/ZO1 (PDZ) binding motif of the E6

protein, which mediates its interaction with other PDZ-containing proteins. Activation of NFκB p52-containing complexes, presumably activated through the non-canonical pathway, was demonstrated, resulting in *cIAP2* up-regulation and abrogation of TNF-induced apoptosis. Importantly, cIAP2 is considered a critical antiapoptotic factor in cells expressing HPV16 oncoproteins and its knockdown is sufficient to induce apoptosis in HeLa cells or in HPV16-immortalized human oral keratinocytes (36). Taken together, these findings suggest that NFκB is activated by E6 and that abrogation of NFκB signaling is an interesting therapeutic target in HPV-induced cancer.

Additional details concerning the mode of NFkB activation by E6 were provided by later studies (Figure 2). Under hypoxic conditions, the E6 oncoprotein of high-risk HPVs was found to stimulate the ubiquitination and proteasomal degradation of cylindromatosis (CYLD) lysine 63 deubiquitinase, a negative regulator of the NFkB pathway that blocks TRAF-mediated IKK recruitment and activation (37). CYLD acts to deubiquitinate TRAF2, TRAF6 and NEMO, and CYLD inactivation up-regulates NFkB signalling in a sustained way, both in vitro and in vivo, under hypoxic conditions, such as those commonly observed in cervical or head-and-neck cancer. These findings are in accordance with the established link between hypoxia, aggressive biological behaviour and chemoresistance. Two years later, Xu et al. suggested that the E6-mediated activation of NFkB might also be mediated by the inactivation of the nuclear transcription factor X-box binding 1 (NFX1), which acts to inhibit NFKB (38). The authors also showed that NFX1 up-regulates the expression of p105, which acts to inhibit NFkB before it is cleaved to generate active p50. As E6 expression also resulted in p105 down-regulation and NFκB activation, the authors suggested this occurs through inhibition of NFX1-mediated p105 expression.

Other oncoproteins were suggested to be implicated in NFkB activation by HPV. Kim et al. reported that the HPV16 E5 oncoprotein up-regulated cyclo-oxygenase-2 (COX2) by activating NFkB (and, to a lesser extent, activator protein-1, API) signalling (39). The E2 protein of α , β , and μ -HPV types was shown to enhance the activation of NFkB stimulated by TNF but not by IL1 (40). This was mediated by direct TRAF5 binding and activation and was independent of the NFκB regulator TAX1 binding protein-1 (TAX1BP1). The authors speculated that this mechanism might be involved in the differentiation of infected keratinocytes, allowing the implementation of the productive viral cycle. These findings are particularly relevant as they point towards a general mechanism common to many HPV types and also because they suggest that NFkB activation may be an early event after HPV infection and not necessarily confined to advanced lesions. Hussain et al. demonstrated that both the E6 and E7 proteins from cutaneous HPV38 activate the

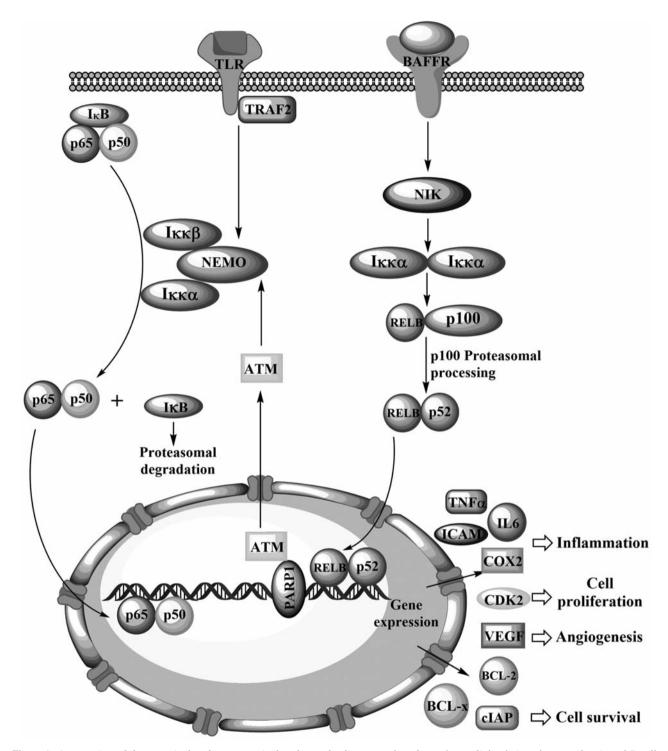


Figure 1. An overview of the canonical and non-canonical pathways leading to nuclear factor kappa-light-chain-enhancer of activated B-cells (NFkB) activation.

canonical (but not the non-canonical) NF κ B signalling pathway, up-regulating the expression of anti-apoptotic genes cIAP1, cIAP2 and xIAP, and effectively inhibiting TNF or UV-mediated apoptosis (41).

The close relationship between NF κ B and cancer was suggested by early studies showing that the turkey retrovirus REV-T encodes the *v-rel* oncogene, a homologue of the NF κ B DNA-binding subunits (42). Numerous epithelial and lymphoid

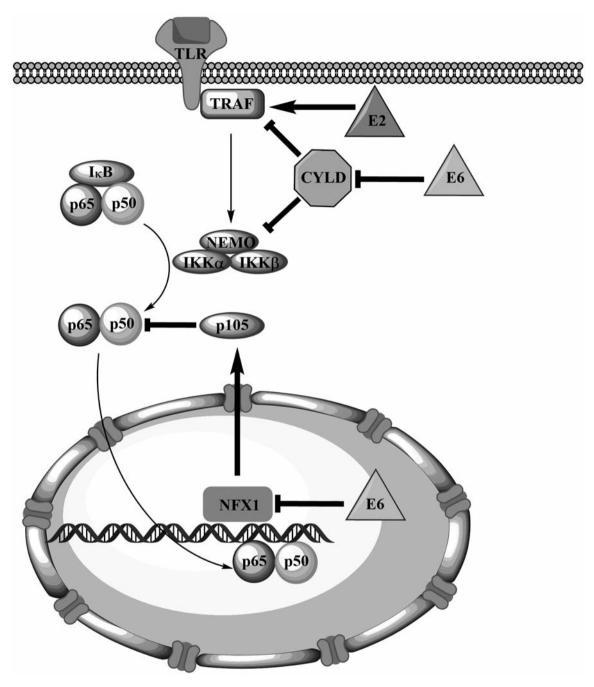


Figure 2. Activation of the canonical nuclear factor kappa-light-chain-enhancer of activated B-cells (NFkB) pathway by human papillomavirus (HPV) oncoproteins E2 and E6. Normal arrows depict the normal signaling pathway, while arrows in bold depict the action of HPV oncoproteins (straight arrows indicate activation, blunt arrows indicate inhibition).

malignancies show NF κ B activation, either due to mutations leading to high NF κ B signalling, or driven by continuous NF κ B stimulation (*e.g.* by cytokines produced by tumourassociated macrophages), or even in response to common chemotherapeutic agents, such as cisplatin (13, 43-46).

NFκB activation regulates the transcription of a number of diverse genes encoding proteins and miRNAs, thereby regulating inflammation, cell survival, proliferation, energy metabolism and adhesion, as well as the cellular microenvironment (47, 48). However, the effects of NFκB

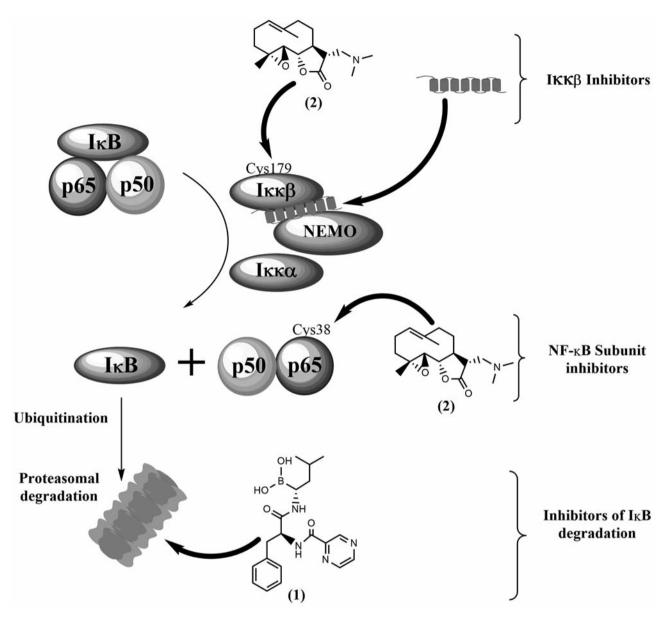


Figure 3. Some mechanisms of nuclear factor kappa-light-chain-enhancer of activated B-cells (NF κ B) inhibition, including: inhibition of inhibitor of NF κ B (I κ B) degradation by proteasomal inhibitor bortezomib (1), I κ B kinase (IKK β) inhibition by covalent binding of dimethylaminoparthenolide (2) to Cys179 or interaction of an inhibitory peptide with the NF κ B essential modulator (NEMO)-binding domain, and direct inhibition of p65 activity by covalent binding of dimethylaminoparthenolide (2) to Cys38. Normal arrows depict the normal signaling pathway, while arrows in bold depict the action of NF κ B inhibitors.

signaling depend on the cell type involved: neoplastic cells often exhibit a 'malignant' version of the protective role played by NFκB in normal cells (13). In fact, NFκB activity closely recalls all the hallmarks of cancer (49). Among the many genes transcriptionally influenced by NFκB, some encode proteins involved in inflammation (*e.g.* COX2, TNF, IL6, and intercellular adhesion molecule, (ICAM)), cell survival (cIAP, XIAP, B-cell lymphoma-2 (BCL2) and BCL-x), proliferation

(cyclin-dependent kinase 2, CDK2), and angiogenesis (VEGF). This central position of NFκB, between inflammatory, proliferative and survival pathways, marks it as a key player in the link between inflammation and cancer. NFκB can also promote the metabolic switch from oxidative phosphorylation to glycolysis even in the presence of oxygen, known as the Warburg effect, as reviewed by Tornatore *et al.* (50). In the presence of functional p53 protein (*e.g.* in normal cells) p65

interacts with p53. Both proteins enter the nucleus to upregulate the expression of metabolic genes which encode, for instance, the complex IV subunit of cytochrome c oxidase, increasing aerobic metabolism and reducing glycolysis (51). In parallel, p53 also down-regulates expression of the glucose receptor GLUT3. However, in the absence of p53 (e.g. in $p53^{-/-}$ or HPV-induced cancer), p65 interacts with mortalin which mediates its translocation into the mitochondria, suppressing mitochondrial function (52). Simultaneously, NFkB up-regulates GLUT3 expression, thus elevating intracellular glucose levels and favouring glycolysis. This is an example of how the outcome of NFkB signalling depends on the cellular context, in particular on the status of tumour suppressors such as p53 and phosphatase and tensin homolog (PTEN). The inactivation of tumour suppressors can drive NFkB signalling towards an oncogenic, tumour-promoting outcome (13). This is of particular significance in HPV-induced tumours, which depend on p53 degradation by E6 and commonly bear wild-type p53. Due to the complexity of the NFkB pathway, it is even possible that NFkB activation may result in a pro-apoptotic affect, depending on several factors such as cell type, the nature of the stimulus, presence of tumour suppressors, and histone acetylation status, as reviewed by Perkins (13) and Godwin et al. (53). For instance, NFkB was shown to mediate apoptosis triggered by wild-type p53 in vitro (54). It was also demonstrated that NFkB can activate p53 and target polo-like kinase 3 (PIK3) to phosphorylate p53, thus increasing its half-life and potency (55).

A well-established line of evidence shows that some types of cancer induced by high-risk HPV display NFkB activation. NFkB was reported to be constitutively activated in cervical cancer and intraepithelial lesions (56, 57). NFkB DNAbinding activity progressively increased from low-grade towards high-grade lesions and invasive cancer. Interestingly, the formation of p50-p50 homodimers, rather than the common p50-p65 heterodimers was reported. Later, Li et al. described a statistically significant association between NFkB activation and tumour progression, aggressive biological behaviour (higher histological grade, lymphatic metastasis, interstitial invasion and larger tumour size) and poor prognosis (lower survival rates) in cervical cancer, suggesting NFkB as a potential therapeutic target (58). Constitutive NFkB activation was also reported for HPV-positive oral cancer, showing p50-p65 heterodimerization (59). However, a recent study suggests that NFkB activation is more frequently observed in HPV-negative than in HPV-positive head-and-neck cancers (60). While there seems to be a clear role for NFkB in cervical cancer, the case for HPV-positive head-and-neck cancer requires additional efforts before any definite conclusions may be drawn.

A second line of evidence shows that NF κ B signaling triggers important protective antiviral responses and that under certain conditions, viral oncoproteins inactivate NF κ B,

averting such responses (61). Early this century, Spitkovsky et al. showed that E7 associates with and represses the IkB complex, effectively down-regulating NFkB (62). Signalling by NFkB is essential in order to activate dendritic cells and trigger protective immune responses (63). High-risk HPV types were found to impair the innate immune response usually triggered by infected keratinocytes, by up-regulating ubiquitin carboxyl-terminal hydrolase L1 and downregulating NEMO (61). Recently, Nakahara et al. showed that HPV16 E1 protein activates NFkB, triggering a feedback loop which limits viral replication (64). These findings suggest that circumventing NFkB activation may be a valid strategy to promote viral replication and persistence. This line of evidence draws attention to the protective role of NFkB signalling during the early phases of viral infection, and its importance for establishing a protective immune response, able to eradicate HPV infection, preventing viral persistence and potential cell transformation.

Targeting NFkB

In 2006, over 750 NFκB inhibitors are known, including antioxidants, small molecules, peptides, small RNA/DNA, microbial and viral proteins (65). The present review focuses on general NFkB targeting mechanisms, while presenting some of the most promising molecules, in the context of HPV-induced cancer. The rationale underlying NFκB inhibition for treating cancer is clear: inhibiting the antiapoptotic and cancer-promoting functions of NFkB will counteract the aggressive biological behaviour of cancer and, if included in combination therapies, will sensitize them to conventional drugs. However, the complexity of the NFkB pathway and its regulatory mechanisms - discussed below make it necessary to carefully design appropriate targeting strategies in order to avoid off-target effects and associated toxicities. Inhibitors of the NFkB pathway are numerous and variably specific, but can be grouped into those targeting IKK activation, IkB degradation and NFkB DNA-binding (Figure 3), as proposed by Nakanishi and Toi (66).

Several highly specific small-molecule IKK β inhibitors have been developed which have interesting anti-inflammatory and anti-neoplastic activity (67-70). In general, these compounds act as ATP analogues with some degree of specificity for IKKs, molecules with allosteric effects on IKK structure, and compounds that interact with Cys179 in the IKK β activation loop (65). However, inhibiting IKKs is likely to have severe physiological implications, *e.g.* in the regulation of inflammatory processes, and such drugs may exert significant off-target effects *in vivo* (71,72). A cell-permeable peptide spanning the IKK β NEMO-binding domain was found to disrupt TNF α -induced NF κ B activation *in vivo*, preventing inflammation in animal models (73). Interestingly, this peptide was reported to allow the

maintenance of basal IKK activity while suppressing enhanced activation by pro-inflammatory cytokines, which may help minimizing the side-effects associated with more drastic NFkB inhibition.

Inhibiting IkB degradation by blocking the proteasome has been a successful strategy in treating certain haematopoietic malignancies. Bortezomib (1 in Figure 3), a proteasome inhibitor, has been approved for clinical use in newly diagnosed and relapsed/refractory multiple myeloma and multiple mantle cell lymphoma. However, proteasome inhibitors are expected to have other effects besides inhibiting NFkB, and significant toxicity has been reported with bortezomib use, namely peripheral neuropathy (74). Despite its success with haematopoietic malignancies, single-agent or combination therapies including bortezomib have shown only limited efficacy against solid tumours such as melanoma (phase II clinical trial NCT00512798), metastatic breast cancer (75), and urothelial cancer (76), as reviewed by Cao et al. (77). A second generation of proteasome inhibitors (e.g. marizomib, oprozomib and delanzomib) holds the promise of providing greater efficacy with reduced toxicity, as well as more flexible dosing schedules (78).

Targeting NFkB subunits is a strategy that may provide more specific effects with less toxicity. A peptide spanning the Ser536 p65 phosphorylation site was shown to inhibit NFkB function selectively in response to some inflammatory stimuli in vitro and in vivo (79-80). The activity of NFkB can be inhibited by some natural products, such as sesquiterpene lactones found in Asteraceae plants (81), which possess a reactive α-methylene-γ-lactone group and target a highly conserved cysteine residue in the REL homology domain (e.g. Cys38 in p65) (82, 83). Inhibition of IKKB by targeting a similar cysteine residue (Cys179) is also thought to play a secondary role in NFkB inhibition by sesquiterpene lactones. Parthenolide is one such NFkB inhibitor, well-known for its anti-inflammatory properties, and has been tested in vitro and in vivo against a range of malignancies, as reviewed by Amorim et al. (84). However, as with many natural products, the pharmacokinetic properties of parthenolide hamper its in vivo efficacy. This problem has been solved through the development of a semi-synthetic aminoderivative, dimethylaminoparthenolide (2 in Figure 3), which provided a favourable pharmacokinetic profile, retaining NFkBinhibitory properties, and had significant anti-neoplastic activity in vivo, coupled with minimal toxicity (85). In view of these favourable results, dimethylaminoparthenolide is reported to have entered phase I clinical trials against haematological malignancies (86).

When targeting NF κ B, it is also important to consider that its outcome is largely determined by interactions with other signalling pathways. As previously mentioned, the cell's p53 status critically determines the effects of NF κ B activation. Here again, sesquiterpene lactones have promising characteristics by

targeting multiple, potentially synergic pathways. Parthenolide releases p53 from murine double minute 2 (MDM2)-mediated inhibition, thus raising the intracellular levels of active p53, while also generating reactive oxygen species to increase oxidative stress and trigger apoptosis (81). This ability to raise the levels of active p53 in conjunction with NFkB inhibition makes parthenolide and its derivative dimethylamino-parthenolide particularly interesting in the context of HPV-induced cancer, which rely on E6 to degrade wild-type p53. It appears that parthenolide is able to sufficiently up-regulate the levels of functional p53 in the presence of E6, removing one of the fundamental basis for HPV-induced cancer and triggering apoptosis (87).

Curcumin, found in the Indian spice turmeric, is another natural compound, with pleiotropic biological activities, which also acts to inhibit NFkB. Although this is not a targeted therapy, curcumin and other natural antiinflammatory and antioxidant compounds remain a persistent focus of scientific interest. The chemopreventative and chemotherapeutic effects of these compounds on cervical cancer, largely mediated through inhibition of the NFkB pathway, were recently reviewed (88). In order to obtain effective tissue concentrations, high oral doses of curcumin are required, but gastrointestinal toxicity (fullness, abdominal pain) may be dose-limiting (89). Several strategies have been proposed in order to increase the oral bioavailability of curcumin, including the use of adjuvants, curcumin nanoparticles and analogues and phase II/III clinical trials on patients with cervical cancer are reported to be under way (90).

Conclusion

There exists accumulating evidence on the roles of NFkB in papillomavirus-induced lesions but experimental findings often seem contradictory. This is in line with the complex signaling network involving this transcription factor and the different settings in which it has been studied. In many instances, it seems plausible that NFkB plays a protective role during the early phases of HPV infection and persistence, while promoting tumour progression and resistance to radiotherapy and chemotherapy in advanced lesions. Inhibiting NFkB signaling is a tempting strategy to fight some types of HPV-induced cancer, especially cervical cancer. In fact, some targeted and non-targeted NFkB inhibitors have been successfully tested against HPV induced cancer or cancer cell lines. The development of second-generation proteasome inhibitors, novel formulations for polyphenolic compounds with enhanced bioavailability, and the development of compounds able to simultaneously target both NFkB and p53 makes this a rapidly evolving field of research, likely to contribute with improved therapies for patients with cancer in the near future.

Conflicts of Interest

The Authors declare that they have no conflict of interest in regard to this article.

Acknowledgements

This work was financially supported by: Project POCI-01-0145-FEDER-006939 (Laboratory for Process Engineering, Environment, Biotechnology and Energy – LEPABE funded by FEDER funds through COMPETE2020 - Programa Operacional Competitividade e Internacionalização (POCI) – and by national funds through FCT - Fundação para a Ciência e a Tecnologia, and also by European Investment Funds by FEDER/COMPETE/POCI – Operational Competitiveness and Internationalization Programme, under Project POCI-01-0145-FEDER-006958 and National Funds by FCT, under the project UID/AGR/04033/2013. Rui M. Gil da Costa is supported by postdoctoral research grant SFRH/BPD/85462/2012, from FCT, funded by the Portuguese Government and the Social European Fund.

References

- 1 zur Hausen H: Papillomaviruses in the causation of human cancers - a brief historical account. Virology 384: 260-265, 2009.
- 2 Ferlay J, Soerjomataram I and Ervik M: GLOBOCAN, cancer incidence and mortality worldwide: IARC cancer base no. 11. International Agency for Research on Cancer, Lyon, France, 2013.
- 3 Siegel R, Naishadham D and Jemal A: Cancer statistics, 2013. CA Cancer J Clin *63*: 11-30, 2013.
- 4 Zandberg DP, Bhargava R, Badin S and Cullen KJ: The role of human papillomavirus in nongenital cancers. CA Cancer J Clin 63: 57-81, 2013.
- 5 Narisawa-Saito M and Kiyono T: Basic mechanisms of high-risk human papillomavirus-induced carcinogenesis: roles of E6 and E7 proteins. Cancer Sci 98: 1505-1511, 2007.
- 6 Mountzios G, Soultati A, Pectasides D, Pectasides E, Dimopoulos M-A and Papadimitriou CA: Developments in the systemic treatment of metastatic cervical cancer. Cancer Treat Rev 39: 430-443, 2013.
- 7 Shen W, Liang B, Yin J, Li X and Cheng J: Noscapine increases the sensitivity of drug-resistant ovarian cancer cell line SKOV3/DDP to cisplatin by regulating cell cycle and activating apoptotic pathways. Cell Biochem Biophys 72: 202-213, 2015.
- 8 Jung H, Kim JS, Kim WK, Oh K-J, Kim J-M, Lee HJ, Han BS, Kim DS, Seo YS, Lee SC, Park SG and Bae K-H: Intracellular annexin A2 regulates NF-κB signalling by binding to the p50 subunit: implications for gemcitabine resistance in pancreatic cancer. Cell Death Dis 6: e1606, 2015.
- 9 D'Amato NC, Rogers TJ, Gordon MA, Greene LI, Cochrane DR, Spoelstra NS, Nemkov TG, D'Alessandro A, Hansen KC, Richer JK: A TDO2-AhR signalling axis facilitates anoikis resistance and metastasis in triple-negative breast cancer. Cancer Res 75: 4651-4664, 2015.
- 10 Sreekanth CN, Bava SV, Sreekumar E and Anto RJ: Molecular evidence for the chemosensitizing efficacy of liposomal curcumin in paclitaxel chemotherapy in mouse models of cervical cancer. Oncogene 30: 3139-3152, 2011.

- 11 Dijkgraaf EM, Heusinkveld M and Tummers B: Chemotherapy alters monocyte differentiation to favor generation of cancersupporting M2 macrophages in the tumor microenvironment. Cancer Res 73: 2480-2492, 2013.
- 12 Park JH, Yoon DS, Choi HJ, Hahm DH and Oh SM: Phosphorylation of IκBα at serine 32 by T-lymphokine-activated killer-cell-originated protein kinase is essential for chemoresistance against doxorubicin in cervical cancer cells. J Biol Chem 288: 3585-3593, 2013.
- 13 Perkins ND: The diverse and complex roles of NFκB subunits in cancer. Nat Rev Cancer 12: 121-132, 2012.
- 14 Sun SC and Ley SC: New insights into NFκB regulation and function. Trends Immunol 29: 469-478, 2008.
- 15 Hayden MS and Ghosh S: Shared principles in NFκB signalling. Cell 732: 344-362, 2008.
- 16 Sun SC: Non-canonical NFκB signalling pathway. Cell Res 21: 71-85, 2011.
- 17 Rao P, Hayden MS, Long M, Scott ML, West AP, Zhang D, Oeckinghaus A, Lynch C, Hoffmann A, Baltimore D and Ghosh S: IκBβ acts to inhibit and activate gene expression during the inflammatory response. Nature 466: 1115-1119, 2010.
- 18 Devin A, Lin Y, Yamaoka S, Li Z, Karin M and Liu Z-G: The α and β subunits of IκB kinase (IKK) mediate TRAF2-dependent IKK recruitment to tumor necrosis factor (TNF) receptor 1 in response to TNF. Mol Cell Biol 21: 3986-3994, 2001.
- 19 Stilmann M, Hinz M, Arslan SC, Zimmer A, Schreiber V and Scheidereit C: A nuclear poly(ADP-ribose)-dependent signalosome confers DNA damage-induced IkB kinase activation. Mol Cell 36: 365-378, 2009.
- 20 Egawa N, Egawa K, Griffin H and Doorbar J: Human papillomaviruses: epithelial tropisms, and the development of neoplasia. Viruses 7: 3863-3890, 2015.
- 21 Steenbergen RD, de Wilde J, Wilting SM, Brink AA, Snijders PJ and Meijer CJ: HPV-mediated transformation of the anogenital tract. J Clin Virol 32(Suppl 1): S25-S33, 2005.
- 22 Gillison ML, Chaturvedi AK, Anderson WF and Fakhry C: Epidemiology of human papillomavirus-positive head and neck squamous cell carcinoma. J Clin Oncol 33: 3235-3242, 2015.
- 23 Gil da costa RM and Medeiros R: Bovine papillomavirus: opening new trends for comparative pathology. Arch Virol 159: 191-198, 2014.
- 24 Tommasino M: The human papillomavirus family and its role in carcinogenesis. Semin Cancer Biol 26: 13-21, 2014.
- 25 Klingelhutz AJ and Roman A: Cellular transformation by human papillomaviruses: lessons learned by comparing high- and lowrisk viruses. Virology 424: 77-98, 2012.
- 26 Werness BA, Levine AJ and Howley PM: Association of human papillomavirus types 16 and 18 E6 proteins with p53. Science 248: 76-79, 1990.
- 27 Howie HL, Katzenellenbogen RA and Galloway DA. Papillomavirus E6 proteins. Virology *384*: 324-334, 2009.
- 28 Dyson N, Guida P, Münger K and Harlow E: Homologous sequences in adenovirus E1A and human papillomavirus E7 proteins mediate interaction with the same set of cellular proteins. J Virol 66: 6893-6902, 1992.
- 29 McLaughlin-Drubin ME and Münger K: The human papillomavirus E7 protein. Virology 384: 335-344, 2009.
- 30 Yoon CS, Kim KD, Park SN and Cheong SW: Alpha(6) integrin is the main receptor of human papillomavirus type 16 VLP. Biochem Biophys Res Commun 283: 668-673, 2001.

- 31 Jeon S and Lambert PF: Integration of human papillomavirus type 16 DNA into human genome leads to increased stability of E6 and E7 mRNAs: implications for cervical carcinogenesis. Proc Natl Acad Sci USA 92: 1654-1658, 1995.
- 32 Nees M, Geoghegan JM, Hyman T, Frank S, Miller L and Woodworth CD: Papillomavirus type 16 oncogenes downregulate expression of interferon-responsive genes and upregulate proliferation-associated and NFκB-responsive genes in cervical keratinocytes. J Virol 75: 4283-4296, 2001.
- 33 Havard L, Delvenne P, Frare P, Boniver J and Giannini SL: Differential production of cytokines and activation of NFκB in HPV-transformed keratinocytes. Virology 298: 271-285, 2002.
- 34 Havard L, Rahmouni S, Boniver J and Delvenne P: High levels of p105 (NFKB1) and p100 (NFKB2) proteins in HPV-transformed keratinocytes: role of E6 and E7 oncoproteins. Virology 331: 357-366, 2005.
- 35 James MA, Lee JH and Klingelhutz AJ: Human papillomavirus type 16 E6 activates NFκB, induces cIAP2 expression and protects against apoptosis in a PDZ binding motif-dependent manner. J Virol 80: 5301-5307, 2006.
- 36 Yuan H, Fu F, Zhuo J, Wang W, Nishitani J, An DS, Chen IS and Liu X: Human papillomavirus type 16 E6 and E7 proteins upregulate *c-IAP2* gene expression and confer resistance to apoptosis. Oncogene 24: 5069-5078, 2005.
- 37 An J, Mo D, Liu H, Veena MS, Srivatsan ES, Massoumi R and Rettig MB: Inactivation of the CYLD deubiquitinase by HPV E6 mediates hypoxia-induced NFκB activation. Cancer Cell 14: 394-407, 2008.
- 38 Xu M, Katzenellenbogen RA, Grandori C and Galloway DA: NFX1 plays a role in human papillomavirus type 16 E6 activation of NFκB activity. J Virol 84: 11461-11469, 2010.
- 39 Kim S-H, Oh J-M, No J-H, Bang Y-J, Juhnn Y-S and Song Y-S: Involvement of NFkB and AP1 in COX2 up-regulation by human papillomavirus 16 E5 oncoprotein. Carcinogenesis 30: 753-757, 2009.
- 40 Boulabiar M, Boubaker S, Favre M and Demeret C: Keratinocyte sensitization to tumour necrosis factor-induced nuclear factor kappa B activation by the E2 regulatory protein of human papillomaviruses. J Gen Virol 92: 2422-2427, 2011.
- 41 Hussain I, Fathallah I, Accardi R, Yue J, Saidi D, Shukla R, Hasan U, Gheit T, Niu Y, Tommasino M and Sylla BS: NFκB protects human papillomavirus type 38 E6/E7-immortalized human keratinocytes against tumor necrosis factor alpha and UV-mediated apoptosis. J Virol 85: 9013-9022, 2011.
- 42 Gilmore TD: Role of rel family genes in normal and malignant lymphoid cell growth. Cancer Surv 15: 69-87, 1992.
- 43 Shukla S, Shankar E, Fu P, MacLennan GT and Gupta S: Suppression of NFκB and NFκB-regulated gene expression by apigenin through IKBa and IKK pathway in TRAMP mice. PLoS One 10: A1293, 2015.
- 44 de Donatis GM, Pape EL, Pierron A, Cheli Y, Hofman V, Hofman P, Allegra M, Zahaf K, Bahadoran P, Rocchi S, Bertolotto C, Ballotti R and Passeron T: NFkB induces senescence bypass in melanoma *via* a direct transcriptional activation of EZH2. Oncogene (in press), 2015.
- 45 Giopanou I, Lilis I, Papaleonidopoulos V, Marazioti A, Spella M, Vreka M, Papadaki H and Stathopoulos GT: Comprehensive evaluation of nuclear factor-κB expression patterns in non-small cell lung cancer. PLoS One 10: A132527, 2015.

- 46 DiDonato JA, Mercurio F and Karin M: NFκB and the link between inflammation and cancer. Immunol Rev 246: 379-400, 2012.
- 47 Karin M: Nuclear factor-κB in cancer development and progression. Nature *441*: 431-436, 2006.
- 48 Iliopoulos D, Hirsch HA and Struhl K: An epigenetic switch involving NFκB, LIN28, LET-7 microRNA, and IL6 links inflammation to cell transformation. Cell 139: 693-706, 2009.
- 49 Hanahan D and Weinberg RA: Hallmarks of cancer: the next generation. Cell 144: 646-674, 2011.
- 50 Tornatore L, Thokatura AK, Bennett J, Moretti M and Franzoso G: The nuclear factor kappa B signaling pathway: integrating metabolism with inflammation. Trends Cell Biol 22: 557-566, 2012.
- 51 Mauro C, Leow SC, Anso E, Rocha S, Thotakura AK, Tornatore L, Moretti M, De Smaele E, Beq AA, Tergaonkar V, Chandel NS and Franzoso G: NFκB controls energy homeostasis and metabolic adaptation by up-regulating mitochondrial respiration. Nature Cell Biol 13: 1272-1279, 2011.
- 52 Johnson RF, Witzel I-I and Perkins ND: P53-dependent regulation of mitochondrial energy production by the RelA subunit of NFκB. Cancer Res 71: 5588-5597, 2011.
- 53 Godwin P, Baird AM, Heavey S, Barr MP, O'Byme KJ and Gately K: Targeting nuclear factor-kappa B to overcome resistance to chemotherapy. Front Oncol *3*: 120, 2013.
- 54 Ryan KM, Ernst MK, Rice NR and Vousden KH: Role of NF-kappaB in p53-mediated programmed cell death. Nature 404: 892-897, 2000.
- 55 Li Z, Niu J, Uwagawa T, Peng B and Chiao PJ: Function of pololike kinase 3 in NF-kappaB-mediated proapoptotic response. J Biol Chem 280: 16843-16850, 2005.
- 56 Prusty BK, Husain SA and Das BC: Constitutive activation of nuclear factor-κB: preferential homodimerization of p50 subunits in cervical carcinoma. Front Biosci *10*: 15010-15019, 2005.
- 57 Du CX and Wang Y: Expression of p-AKT, NFκB and their correlation with human papillomavirus infection in cervical carcinoma. Eur J Gynaecol Oncol *33*: 274-277, 2012.
- 58 Li J, Jia H, Xie L, Wang X, Wang X, He H, Lin Y and Hu L: Association of constitutive nuclear factor-κB activation with aggressive aspects and poor prognosis in cervical cancer. Int J Gynecol Cancer 19: 1421-1426, 2009.
- 59 Mishra A, Bharti AC, Varghese P, Saluja D and Das BC: Differential expression and activation of NFκB family proteins during oral carcinogenesis: role of high risk human papillomavirus infection. Int J Cancer 119: 2840-2850, 2006.
- 60 Gaykalova DA, Manola JB, Ozawa H, Zizkova V, Morton K, Bishop JA, Sharma R, Zhang C, Michailidi C, Considine M, Tan M, Fertig EJ, Hennessey PT, Ahn J, Koch WM, Westra WH, Khan Z, Chung CH, Ochs MF and Califano JA: NFκB and STAT3 transcription factor signatures differentiate HPV-positive and HPV-negative head and neck squamous cell carcinoma. Int J Cancer 137: 1879-1889, 2015.
- 61 Karim R, Tummers B, Meyers C, Biryukov JL, Alam S, Backendorf C, Jha V, Offringa R, van Ommen GB, Melief CJM, Guardavaccaro D, Boer JM and van der Burg S: Human papillomavirus (HPV) up-regulates the cellular deubiquitinase UCHL1 to suppress the keratinocyte's inate immune response. PLOS Pathog 9: e1003384, 2013.
- 62 Spitkovsky D, Hehners SP, Hofmann TG, Möller A and Schmitz ML: The human papillomavirus oncoprotein E7 attenuates NFκB activation by targeting the IκB kinase complex. J Biol Chem 277: 25576-25582, 2002.

- 63 Yan M, Peng J, Jabbar IA, Liu X, Filqueira L, Frazer IH and Thomas R: Activation of dendritic cells by human papillomavirus-like particles through TLR4 and NFκB-mediated signalling, moderated by TGFβ. Immunol Cell Biol 83: 83-91, 2005.
- 64 Nakahara T, Tanaka K, Ohno S, Egawa N, Yugawa T and Kiyono T: Activation of NFκB by human papillomavirus 16 E1 limits E1dependent viral replication through degradation of E1. J Virol 89: 5040-5059, 2015.
- 65 Gilmore TD and Herscovitch M: Inhibitors of NFκB signalling: 785 and counting. Oncogene 25: 6885-6899, 2006.
- 66 Nakanishi C and Toi M: Nuclear factor-κB inhibitors as sensitizers to anticancer drugs. Nat Rev Cancer 5: 297-309, 2005.
- 67 Mbalaviele G, Sommers CD, Bonar SL, Mathialagan S, Schindler JF, Guzova JA, Shaffer AF, Melton MA, Christine LJ, Tripp CS, Chiang PC, Thompson DC, Hu Y and Kishore N: A novel, highly selective, tight binding IκB kinase-2 (IKK2) inhibitor: a tool to correlate IKK2 activity to the fate and functions of the components of the nuclear factor-κB pathway in arthritis-relevant cells and animal models. J Pharmacol Exp Ther 329: 14-25, 2009.
- 68 Chiang PC, Kishore NN and Thompson DV: Combined use of pharmacokinetic modeling and a steady-state delivery approach allows early assessment of IκB kinase-2 (IKK2) target safety and efficacy. J Pharm Sci 99: 1278-1287, 2010.
- 69 Avila CM, Lopes AB, Gonçalves AS, da Silva LL, Romeiro NC, Miranda AL, Sant'Anna CM, Barreiro EJ and Fraga CA: Structure-based design and biological profile of (E)-N-(4-nitrobenzylidene)-2-naphtohydrazide, a novel small molecule inhibitor of IκB kinase-β. Eur J Med Chem 46: 1245-1253, 2011.
- 70 Kim S, Jung JK, Lee HS, Kim Y, Kim J, Baek DJ, Moon B, Oh KS, Lee BH, Shin KJ, Pae AN, Nam G, Roh EJ, Cho YS, and Choo H: Discovery of piperidinyl aminopyrimidine derivatives as IKK2 inhibitors. Bioorg Med Chem Lett 21: 3002-3006, 2011.
- 71 Greten FR, Arkan MC, Bollrath J, Hsu LC, Goode J, Miething C, Göktuna SI, Neuenhahn M, Fierer J, Paxian S, van Rooijen N, Xu Y, O'Cain T, Jaffee BB, Busch DH, Duyster J, Schmid RM, Eckmann L and Karin M: NFκB is a negative regulator of IL1β secretion as revealed by genetic and pharmacological inhibition of IKKβ. Cell 130: 918-931, 2007.
- 72 Pasparakis M: Regulation of tissue homeostasis by NFκB signaling: implications for inflammatory diseases. Nat Rev Immunol 9: 778-788, 2009.
- 73 Strickland I and Ghosh S: Use of cell permeable NBD peptides for suppression of inflammation. Ann Rheum Dis 65(Suppl 3): iii75-iii82, 2006.
- 74 Broyl A, Jongen JL and Sonneveld P: General aspects and mechanisms of peripheral neuropathy associated with bortezomib in patients with newly diagnosed multiple myeloma. Semin Hematol 49: 249-257, 2012.
- 75 Yang CH, Gonzalez-Angulo AM, Reuben JM, Booser DJ, Pusztai L, Krishnamurthy S, Esseltine D, Stec J, Broglio KR, Islam R, Hortobagyi GN and Cristofanili M: Bortezomib (VELCADE) in metastatic breast cancer: pharmacodynamics, biological effects and prediction of clinical benefits. Ann Oncol 17: 813-817, 2006.
- 76 Gomez-Abuin G, Winquist E, Stadler WM, Pond G, Degendorfer P, Wright J and Moore MJ: A phase II study of PS-341 (bortezomib) in advanced or metastatic urothelial cancer. A trial of the Princess Margaret Hospital and University of Chicago phase II consortia. Invest New Drugs 25: 181-185, 2007.

- 77 Cao B, Li J and Mao X: Dissecting bortezomib: development, application, adverse effects and future direction. Curr Pharm Des *19*: 190-200, 2013.
- 78 Allegra A, Alonci A, Gerace D, Russo S, Inao V, Calabró L and Musolino C: New orally active proteasome inhibitors in multiple myeloma. Leuk Res 38: 1-9, 2014.
- 79 Takada Y, Singh S and Aggarwal BB: Identification of a p65 peptide that selectively inhibits NFκB activation induced by various inflammatory stimuli and its role in down-regulation of NFκB-mediated gene expression and up-regulation of apoptosis. J Biol Chem 279: 15096-15194, 2004.
- 80 Oakley F, Teoh V, Ching-a-Sue G, Bataller R, Colmenero J, Jonsson LR, Eliopoulos AG, Watson MR, Manas D and Mann DA: Angiotensin II activates IκB kinase phosphorylation of RelA at Ser 536 to promote myofibroblast survival and liver fibrosis. Gastroenterology 136: 2334-2344, 2009.
- 81 Bastos MMSM, Kijjoa and Pinto MMM: Constituents of *Centaurea ornata* ssp. ornata. Fitoterapia 65: 191, 1994.
- 82 García-Piñeres AJ, Castro V, Mora G, Schmidt TJ, Strunck E, Pahl HL and Merfort I: Cysteine 38 in p65/NFκB plays a crucial role in DNA binding inhibition by sesquiterpene lactones. J Biol Chem 276: 39713-39720, 2001.
- 83 García-Piñeres AJ, Lindenmeyer MT and Merfort I: Role of cysteine residues of p65/NFκB on the inhibition by the sesquiterpene lactone parhenolide and N-ethyl-maleimide, and on its transactivating potential. Life Sci 75: 841-856, 2004.
- 84 Amorim MHR, Gil da Costa RM, Lopes C and Bastos MMSM: Sesquiterpene lactones: adverse health effects and toxicity mechanisms. Crit Rev Toxicol *43*: 559-579, 2013.
- 85 Guzmán ML, Rossi RM, Neelankantan S, Li X, Corbett CA, Hassane DC, Becker MW, Bennett JM, Sullivan E, Lachowicz JL; Vaughan A, Sweeney CJ, Matthews W, Liesveld JL, Crooks PA and Jordan CT: An orally bioavailable parthenolide analog selectively eradicates acute myelogenous leukemia stem and progenitor cells. Blood 10: 4427-4435, 2007.
- 86 Ghantous A, Sinjab A, Herceg Z and Darwiche N: Parthenolide: from plant shoots to cancer roots. Drug Discov Today 18: 894-905, 2013.
- 87 Mendonça MS, Chin-Sinex H, Gomez-Millan J, Datzman N, Hardacre M, Comerford K, Nakshatri H, Nye M, Benjamin L, Mehta S, Patino F and Sweeney C: Parthenolide sensitizes cells to X-ray-induced cell killing through inhibition of NFκB and split-dose repair. Radiat Res *168*: 689-697, 2007.
- 88 Di Domenico F, Foppoli C, Coccia R and Perluigi M: Antioxidants in cervical cancer: chemopreventive and chemotherapeutic effects of polyphenols. Biochim Biophys Acta 1822: 737-747, 2012.
- 89 Epelbaum R, Schaffer M, Vizel B, Badmaev V and Bar-Sela G: Curcumin and gemcitabine in patients with advanced pancreatic cancer. Nutr Cancer 62: 1137-1141, 2010.
- 90 Shehzad A, Wahid F and Lee YS: Curcumin in cancer chemoprevention: molecular targets, pharmacokinetics, bioavailability and clinical trials. Arch Pharm 9: 489-499, 2010.

Received February 7, 2016 Revised April 2, 2016 Accepted April 7, 2016