Review

# Antineoplastic Activity of Zoledronic Acid and Denosumab

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Abstract. Cancer patients suffer from cancer-induced bone pain, hypercalcemia, and reduced quality of life caused by pathological fractures. Many of these complications related to cancer can be treated, or at least controlled, using new anticancer agents. Recently, two agents used initially to treat osteoporosis demonstrated direct and indirect anticancer activity. In this review, we summarize current knowledge about direct and indirect anticancer activity of zoledronic acid (a third-generation bisphosphonate), and denosumab antibody against RANKL. Zoledronic acid influences the proliferation and viability of tumor cells in vitro, and effectively reduces tumor burden, tumor-induced pain, and tumor growth in vivo. Denosumab is a fully human monoclonal antibody preventing the binding of RANKL to its receptor on osteoclasts' membrane, and through this mechanism inhibits the resorption of the bone. Furthermore, this agent demonstrates direct anticancer activity through the RANKL signaling pathway. Because of these features both drugs may gain broader application for the treatment of cancer patients. However, further pre-clinical and clinical evaluation is needed for both agents to fully assess the antineoplastic mechanisms of activity of both agents.

Long bones are the most common site of spread of tumors such as breast, prostate, and kidney cancer (1-5) and hematological cancers such as multiple myeloma (6, 7). Patients suffering from these types of cancers experience cancer-induced bone pain, hypercalcemia, and loss of function from pathological fractures (8-10). All these complications related to cancer have a major influence on a patient's quality of life. There is, therefore, an emerging need to develop and introduce into clinical practice agents which could more effectively control tumor growth, and its burden in bone and distant sites. In recent years, two agents were

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introduced to the therapy of skeletal metastases. Besides inhibition of osteoclastogenesis and bone resorption, these two agents possess a direct antineoplastic activity. In this review we describe the direct and indirect effect of the *bis*phosphonate zoledrenic acid (ZA) and the (RANKL)binding antibody denosumab on tumor growth.

ZA, a third-generation bisphosphonate, has been widely used to treat osteoporosis (11, 12), and skeletal metastases; recent reports also suggest direct activity against cancer cells (13-16). Bisphosphonates directly inhibit osteoclast activity through selective affinity to the site of increased bone turnover e.g. fracture sites. In addition, the latest studies demonstrated the induction of apoptosis of cancer cells in vitro in prostate cancer, breast cancer and multiple myeloma cell lines (17-20). Moreover, ZA successfully inhibits angiogenesis in the tumor environment, and reduces the invasive ability of cancer cells (21, 22), including the one of soft tissue tumors (13, 17, 23). ZA influences the proliferation and viability of tumor cells in vitro, and effectively reduces tumor burden, tumor-induced pain, and tumor growth in vivo. ZA may decrease tumor growth indirectly by inhibition of tumor-induced osteolysis in bone, and down-regulation of expression and secretion of cytokines and other growth factors in the tumor environment (24, 25). In addition, it reduces new blood vessel formation at the tumor site (26). ZA also exhibits a direct effect on cancer cells through induction of apoptosis, and reduction of migration and adhesion ability (17, 19).

Denosumab is an inhibitor of the receptor activator of nuclear factor-KB (RANK)/RANKL signaling pathway. It was developed using XenoMouse transgenic mouse technology (27). It is a fully human monoclonal antibody which prevents the binding of RANKL to its receptor on osteoclast membrane, and through this mechanism it inhibits the resorption of bone. Recent reports suggest the direct antitumor activity of this antibody (28, 29).

# Mechanism of Action of Zoledronic Acid

*Bis*phosphonates accumulate in the mineralized bone matrix and are released during bone resorption. Bone has a very high affinity for *bis*phosphonates, this is possible only because of very specific uptake of these agents by activated osteoclasts (30). The nitrogen-containing bisphosphonates (N-BPs), which include ZA affect osteoclast activity and survival through inhibition of farnesyl diphosphonate (FPP) synthase, in the biosynthetic mevalonate pathway. This disrupts many cellular functions which are of great importance for osteolytic activity of the osteoclasts and their survival (31-33). Furthermore, ZA affects several signaling proteins such as: Rat sarcoma (RAS), Ras homology (RHO), and Ras-related C3 botulinum toxin substrate (RAC) involved in cytoskeleton and cellular motility (34-36). Selective inhibition of RAS signaling within osteoclasts causes disruption of intracellular vesicle transport, which reduces the ability of the cells to migrate and aggregate at the tumor bone border (30, 37). In addition, ZA may induce intracellular production of triphosphoric acid 1-adenosin-5'vl ester 3-[3-methylbut-3-envl] ester (ApppI), which can directly induce apoptosis (38, 39). This is induced by ZA by inhibition of farnesyl pyrophosphate synthase (FPPS), which causes down regulation of the mevalonate pathway, and secondary accumulation of isopentenyl pyrophosphate (IPP), which in turn, is conjugated to AMP to form a novel ATP analog (ApppI). ZA has a high potency at inhibiting FPPS activity and increase synthesis of ApppI in osteoclasts. Induction of synthesis of ApppI directly influences the proliferation, migration and apoptosis of osteoclasts, as well as tumor cells (39-42). Correlation between the osteolytic activity of osteoclasts and increased inhibition of FPPS was demonstrated in vitro and in vivo by Dunford et al. (43). Moreover, inhibition of signaling protein prenylation in cancer cells may produce the direct antineoplastic effect of ZA (Figure 1 a). In addition, ZA may directly influence the survival of cancer cells, by activation of caspases. ZA may directly enhance production of ATP analogs which interfere with mitochondrial ATP/ADP translocase. Treatment of MCF-7 breast cancer cells with ZA increased the release of cytochrome c from mitochondria and activation of caspase-3 which was associated with increased apoptosis (14, 44, 45). Furthermore, regulation of expression of B-cell lymphoma 2 (BCL-2) through ZA induces release of cytochrome c (46). Inhibition of activation of RAS signaling pathway may modulate both apoptotic pathways (47). ZA significantly inhibits the ability of tumor cells to invade and migrate into surrounding tissue. In in vitro experiments, ZA demonstrated dose-dependent reduction of invasion of tumor cells through the extracellular matrix. The inhibition of matrix metalloproteinase (MMP) activity may be one reason for this reduction in invasion ability of tumor cells (24, 48, 49). In addition, the induction of caspase activity and inhibition of RAS signaling pathway are very important mechanisms inhibiting tumor cell adhesion to extracellular matrix and affecting invasiveness. In some pre-clinical studies, ZA was found to have potent antiangiogenic effect.

ZA exhibited in vitro dose-dependent inhibition of proliferation of human umbilical vein endothelial cells (HUVECs) (50, 51). Similar results have been observed in animal studies: in the 5T2 myeloma model, ZA significantly reduced tumor-associated angiogenesis. The antiangiogenic effect of ZA is mediated at least in part by modulation of  $\alpha v\beta 3$  and  $\alpha v\beta 5$  integrins, which are also required for osteoclasts for attachment to the bone at the resorption site (52). The integrins ( $\alpha v\beta 3$ ) also play an important role in the metastatic ability of tumor cells (53-55). This suggests that ZA directly affects the ability of tumor cells to spread, invade and migrate, as well as directly inhibit adhesion and resorption ability of osteoclasts. Moreover, ZA down regulates the release of some pro-angiogenic growth factors, such as vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (BFGF) in patients with cancer (24, 56).

## Mechanism of Action of Denosumab

Denosumab is a human monoclonal antibody possessing a very high affinity for RANKL. The monoclonal antibody can directly modulate the RANK/RANKL signaling pathway (57). This pathway is very important for maturation, activation, and survival of osteoclasts and tumor cells (Figure 1 b), and tumor cell ability to spread to distant sites (58-60). Specific binding of denosumab to RANKL prevents stimulation of RANK receptor, which is expressed on the surface of osteoclasts (61, 62). The balance between osteoclastic and osteoblastic activity is directly related to remodeling of bone and its resorption. The osteoclasts differentiate from monocyte-macrophage lineage, after maturation and final differentiation; they resorb bone in physiological and pathological states. Furthermore, some cytokines and growth factors such as RANKL modulate this process. RANKL is a tumor necrosis factor (TNF) family member released by activated T-cells (63). Binding of RANKL to RANK, expressed on osteoclasts and their precursors, increases their activity, migration and adhesion, and reduces apoptosis. This activation of osteoclasts causes osteolysis in bone. T-Cells and osteoblast lineage cells produce RANKL. The soluble and membrane-bound forms are produced by activated T-cells. The osteoblast lineage cells express RANKL on their membrane surface. Many cytokines, growth factors and hormones such 1,25(OH)2 vitamin D3, parathyroid hormone (PTH), TNF, and corticosteroids are involved in the modulation of the RANKL/RANK pathway (64-66). By binding to RANKL, denosumab neutralizes its action. Similar activity to denosumab is demonstrated by osteoprotegerin (OPG) (a decoy receptor for RANKL). Because of this, OPG has been widely used in animal models to study inhibition of RANKL. The reports showed that OPG can significantly reduce osteolysis in the bone (67-69).

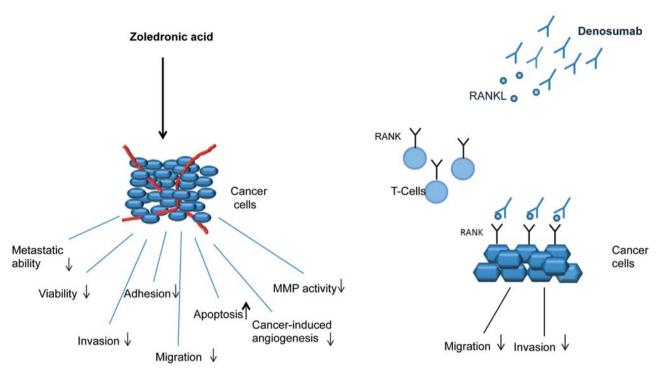


Figure 1. Direct anticancer activity of zoledronic acid (a) and denosumab (b).

#### In Vitro Studies of ZA

Recent reports suggest a direct antineoplastic activity of ZA on variety of tumor cells e.g. multiple myeloma, lung cancer, and renal cancer. Pre-clinical studies with MDA-MB-231 and MCF-7 breast cell lines demonstrated dose-dependent reduction of proliferation, and survival by treatment with ZA (14, 47). Moreover, breast cancer cells treated with ZA exhibited an increased apoptosis rate, which could, however, be reversed by geranylgeraniol, which suggests that inhibition of prenylation in cancer cells may induce apoptosis (70, 71). In addition, PC-3 prostate cancer cells treated with ZA exhibited significantly increased apoptosis and reduced proliferation in vitro (72). This suggests the direct antitumor activity of ZA against tumor cells in in vitro studies. In addition, many preclinical studies demonstrated additive or synergistic effect of ZA with other anticancer drugs e.g. paclitaxel, and doxorubicin (73-76).

# In Vivo Models - Studying ZA

Pre-clinical data from animal studies support the antineoplastic effect of ZA observed in many *in vitro* studies. N-BPs very efficiently reduce osteolytic lesions in bone caused by tumor. In addition, they reduce formation of metastases and tumor-induced bone pain, and induce apoptosis of tumor cells (77-81). Some antineoplastic effect

has also been observed in soft tissue and visceral models (13, 17, 82). Most of these experiments have been carried out with breast and prostate cancer. The activity of ZA on tumor cells was documented indirectly by reduction of tumor-induced pain, or by reduced bone osteolysis around tumor. Direct antitumor activity of ZA on tumor cells has been assessed using pathohistological analysis. A breast cancer model with MDA-MB-231 cells treated with ZA showed significantly reduced osteolytic lesions as compared to non-treated animals (83). Another study with 4T1 murine breast cancer cells treated with ZA at the time of inoculation of the tumor cells demonstrated significantly lower metastatic ability of the cells (84). In addition, the histomorphorogical analysis showed increased apoptosis of osteoclasts and tumor cells. Similarly to in vitro experiments, ZA demonstrated additive effect in combination with conventional anti-neoplastic drugs, e.g., doxorubicin. Animal models carried out with prostate cancer PC-3 and LuCaP 23.1 reported an anti-neoplastic effect on tumor burden (85). After injection of these cells into tibia of mice and treatment with ZA either at the time of inoculation of tumor cells or after establishing the tumors, mice demonstrated fewer osteolytic lesions of tibia by x-ray analysis. Moreover, prostate-specific antigen in serum of LuCaP 23.1-bearing mice significantly decreased after treatment with ZA, which provides additional evidence of the direct antitumor activity of ZA. In some animal models ZA had a preventive effect in terms of metastatic ability of tumor cells. Mice injected with PC-3 cells and treated with ZA had a significantly lower incidence of bone metastases (86). Some animal models suggested that ZA can inhibit development of visceral metastases. Animals injected with 4T1 murine breast cancer and treated with ZA demonstrated a lower tumor burden in bone and reduced the number of lung and liver metatstases (84). The presented data suggest that ZA can significantly reduce osteolytic lesions in bone, tumor burden in soft tissue, and also reduce development of metastases through direct and indirect antitumor activity.

# In Vitro Activity of Denosumab

Recent reports interestingly demonstrated no antiangiogenic activity of denosumab in in vitro assays as compared to ZA (21). HUVECs treated with 0.31 to 160  $\mu$ M of denosumab demonstrated no decrease in viability and no effect on invasion and tubule formation. In addition, the treatment did not affect the viability of MDA-MB-436 and CG5 breast cancer cells. Mice treated with 10 mg/kg of denosumab twice a week for four consecutive weeks did not show reduced angiogenesis or any effect on growth of xenograft in that model. This clearly demonstrates that ZA has an additional antitumor effect through antiangiogenic activity as compared to denosumab. There are some data available suggesting that RANKL can modulate the migration of different tumor cells (87, 88). Human breast cell lines (MDA-MB-231, MCF-7 and Hs578T) treated with RANKL demonstrated dose-dependent increased migration. This increase in migration was significantly reduced after treatment with OPG. In the same study, OPG treatment of colon cancer cells (Colo205), which did not express RANK, did not affect migration. Migration of B16F10 melanoma cells increased in a dose-dependent manner after stimulation with RANKL and was again significantly inhibited with OPG (87). In addition, colony-stimulating factor-1 (CSF-1) did not stimulate B16F10 migration in the presence of RANKL. This additionally provides evidence that RANK stimulation has a direct effect on tumor cells in the context of the significant role of CSF-1 in oncogenesis.

### In Vivo Studies of Denosumab

In another study, high doses of OPG and ZA were used to inhibit the progression of the MDA-231-B/LUC + breast cancer cells (80). Histological and radiographic analysis demonstrated a significant decrease of osteolysis in bone. Interestingly, no (TRAP)-positive osteoclasts were identified in animals treated with Fc-OPG. However, OPG did not significantly affect tumor growth itself. Osteolysis of the bone caused by tumor growth is modulated by dysregulation of the RANK/RANKL pathway. Mice bearing MDA-MB-231 breast cancer cells expressed higher levels of RANKL than control mice (59). In addition, mice treated with increasing doses of tumor bone border. Furthermore, this treatment significantly inhibited growth of tumor cells in preventive as well as therapeutic modes. The OPG-increased activation of caspase-3, through this apoptosis mechanism, reduced tumor burden and tumor growth. Osteoclastogenesis induced by prostatic cancer CaP in an intratibial model treated with OPG (89), was significantly inhibited. In this study, the authors showed that CaP cells produce a soluble form of RANKL which can modulate osteoclastogenesis. The number of osteoclasts at the tumor bone border decreased after treatment with OPG, however, there was no effect on viability, and proliferation of CaP cells. This demonstrates how important the role of osteoclastogenesis and osteoclast activity is in the development and growth of skeletal metastasis. This study also presented evidence that OPG has no direct activity on tumor cells themself. CaP cells treated with OPG in vitro, as well as in subcutaneous models, did not demonstrate increased apoptosis or reduced viability. There is no correlation of data of RANKL level in blood and its impact on tumor growth in bone. This may be due to local release of RANKL at the tumor bone site, or by limitation of detection by currently used assays. Furthermore, RANK receptor is present on many epithelial tissues and epithelial cells, however, not much is known about its role in migration and development of bone metastases. In vivo, intracardiac injection of B16F10 cells into C57BL/6 mice resulted in rapid development of multiorgan metastases, including, of long bones. Treatment of the mice with OPG significantly reduced the tumor burden in bone; however, it did not affect tumor burden and metastases in ovaries, adrenal glands, and brain (87).

OPG-Fc demonstrated significantly lower osteolysis of the

## **Clinical Trials**

Denosumab in a double-blind, randomized study, phase III trial in breast cancer, prostate and multiple myeloma showed significantly better control of first skeletal events compared to ZA treatment. Furthermore, denosumab more frequently caused hypocalcemia, and did not demonstrate any advantages in terms of tumor progression and overall survival (90). Patients with giant cell tumors of the bone had significantly reduced tumor size. The histopathological analysis demonstrated a reduced number of RANK-positive giant tumor cells. Moreover, the treatment reduced the density of proliferative stromal tumor cells. In a phase II clinical trial, patients with recurrent or unresectable giant cell tumor in bone were treated with 120 mg every four weeks. The histopathological analysis demonstrated 90% decrease in the number of giant cells and significant reduction of tumor stromal cells (91). Male patients with bone metastases from castration-resistant prostate cancer were treated with denosumab at a dose of 120 mg on a month, and compared to a ZA treatment group. Denosumab resulted in delay or

prevention of skeletal-related events in patients with advanced prostate cancer (92). Similarly, in female patients with advanced breast cancer treated either with denosumab (120 mg) or ZA (4 mg), there was a longer delay of time to diagnosis of the first bone metastases with use of denosumab (93). Another preventive study compared the efficacy of denosumab in preventing development of skeletal metastases in patients with advanced cancer, bone tumor or myeloma, excluding breast and prostate cancer. The patients were treated with subcutaneous denosumab or intravenous ZA. The study demonstated that treatment with denosumab was equally good compared to that with ZA in delaying the diagnosis of the first metastasis, with fewer side-effects. There was also no difference between groups in terms of disease progression or survival (94). A randomized, double-blind, double-dummy phase III study, also compared denosumab with ZA for pain alleviation in patients with advanced breast cancer and bone metastases. The denosumab treatment group had significantly improved pain prevention, with similar pain alleviation to that of ZA; in addition, fewer patients in the denosumab group switched to opioid therapy (95).

## Conclusion

Nowadays, patients with advanced cancer have longer survival, but have increased risk of bone-related events, such as pathological fractures, reduced ambulation and cancerinduced pain. Because of this, there is increasing need to develop new agents to reduce these complications and improve control of cancer growth in this population. In the past decade, ZA has been introduced into clinical practice as an antiosteoclastic agent. Many patients and physicians may prefer to use denosumab, because of its subcutaneous application compared to intravenous injections of ZA. Furthermore, aninitial evaluation showed that denosumab causes fewer complications related to treatment e.g. osteonecrosis of the jaw, and hypocalcemia. In addition, there are no limitations in the use of denosumab for patients with renal impairment. On the other hand, ZA demonstrates superiority in terms of its direct antineoplastic activity demonstrated both in vitro and in vivo compared to denosumab. Further pre-clinical and clinical evaluation is needed for both agents to fully assess the antineoplastic mechanisms of activity of both agents.

# References

- 1 Rosenberg A and Mathew P: Imatinib and prostate cancer: lessons learned from targeting the platelet-derived growth factor receptor. Expert Opin Investig Drugs 22: 787-794, 2013.
- 2 Aktas B, Kasimir-Bauer S, Lehmann N, Kimmig R and Tewes M: Validity of bone marker measurements for monitoring response to *bis*phosphonate therapy with zoledronic acid in metastatic breast cancer. Oncol Rep 30: 441-447, 2013.

- 3 Slaney CY, Moller A, Hertzog PJ and Parker BS: The role of Type I interferons in immunoregulation of breast cancer metastasis to the bone. Oncoimmunology 2: 1-3, 2013.
- 4 Hagberg KW, Taylor A, Hernandez RK and Jick S: Incidence of bone metastases in breast cancer patients in the United Kingdom: Results of a multi-database linkage study using the General Practice Research Database. Cancer Epidemiol 37: 240-246, 2013.
- 5 Wedin R, Bauer HC and Rutqvist LE: Surgical treatment for skeletal breast cancer metastases: A population-based study of 641 patients. Cancer 92: 257-262, 2001.
- 6 Ria R, Reale A, Moschetta M, Mangialardi G, Dammacco F and Vacca A: A retrospective study of skeletal and disease-free survival benefits of zoledronic acid therapy in patients with multiple myeloma treated with novel agents. Int J Clin Exp Med 6: 30-38, 2013.
- 7 Rosen LS, Gordon D, Kaminski M, Howell A, Belch A, Mackey J, Apffelstaedt J, Hussein M, Coleman RE, Reitsma DJ, Seaman JJ, Chen BL and Ambros Y: Zoledronic acid *versus* pamidronate in the treatment of skeletal metastases in patients with breast cancer or osteolytic lesions of multiple myeloma: A phase III, double-blind, comparative trial. Cancer J 7: 377-387, 2001.
- 8 Morgans AK and Smith MR: Bone-targeted agents: Preventing skeletal complications in prostate cancer. Urol Clin North Am 39: 533-546, 2012.
- 9 Dore-Savard L, Beaudet N, Tremblay L, Xiao Y, Lepage M and Sarret P: A micro-imaging study linking bone cancer pain with tumor growth and bone resorption in a rat model. Clin Exp Metastasis 30: 225-236, 2013.
- 10 Ratasvuori M, Wedin R, Keller J, Nottrott M, Zaikova O, Bergh P, Kalen A, Nilsson J, Jonsson H and Laitinen M: Insight opinion to surgically treated metastatic bone disease: Scandinavian Sarcoma Group Skeletal Metastasis Registry report of 1195 operated skeletal metastases. Surg Oncol 22: 132-138, 2013.
- 11 Black DM, Delmas PD, Eastell R, Reid IR, Boonen S, Cauley JA, Cosman F, Lakatos P, Leung PC, Man Z, Mautalen C, Mesenbrink P, Hu H, Caminis J, Tong K, Rosario-Jansen T, Krasnow J, Hue TF, Sellmeyer D, Eriksen EF, Cummings SR and Trial HPF: Once-yearly zoledronic acid for treatment of postmenopausal osteoporosis. N Engl J Med 356: 1809-1822, 2007.
- 12 Reid IR, Brown JP, Burckhardt P, Horowitz Z, Richardson P, Trechsel U, Widmer A, Devogelaer JP, Kaufman JM, Jaeger P, Body JJ, Brandi ML, Broell J, Di Micco R, Genazzani AR, Felsenberg D, Happ J, Hooper MJ, Ittner J, Leb G, Mallmin H, Murray T, Ortolani S, Rubinacci A, Saaf M, Samsioe G, Verbruggen L and Meunier PJ: Intravenous zoledronic acid in postmenopausal women with low bone mineral density. N Engl J Med 346: 653-661, 2002.
- 13 Liu Q, Tao YH, Bai RZ, Chang SJ and Hua D: Zoledronic acid inhibits growth of hepatocellular carcinoma cells *in vitro* and *in vivo*. Chin Med J *126*: 1486-1490, 2013.
- 14 Ibrahim T, Mercatali L, Sacanna E, Tesei A, Carloni S, Ulivi P, Liverani C, Fabbri F, Zanoni M, Zoli W and Amadori D: Inhibition of breast cancer cell proliferation in repeated and nonrepeated treatment with zoledronic acid. Cancer Cell Int *12*: 48, 2012.
- 15 Chang J, Wang W, Zhang H, Hu Y and Yin Z: Bisphosphonates regulate cell proliferation, apoptosis and pro-osteoclastic expression in MG-63 human osteosarcoma cells. Oncol Lett 4: 299-304, 2012.

- 16 Zwolak P, Manivel JC, Jasinski P, Kirstein MN, Dudek AZ, Fisher J and Cheng EY: Cytotoxic effect of zoledronic acidloaded bone cement on giant cell tumor, multiple myeloma, and renal cell carcinoma cell lines. J Bone Joint Surg Am 92: 162-168, 2010.
- 17 Okamoto S, Kawamura K, Li Q, Yamanaka M, Yang S, Fukamachi T, Tada Y, Tatsumi K, Shimada H, Hiroshima K, Kobayashi H and Tagawa M: Zoledronic acid produces antitumor effects on mesothelioma through apoptosis and S-phase arrest in p53-independent and Ras prenylation-independent manners. J Thorac Oncol 7: 873-882, 2012.
- 18 Ma YG, Liu WC, Dong S, Du C, Wang XJ, Li JS, Xie XP, Wu L, Ma DC, Yu ZB and Xie MJ: Activation of BK(Ca) channels in zoledronic acid-induced apoptosis of MDA-MB-231 breast cancer cells. PLoS One 7: e37451, 2012.
- 19 Koto K, Murata H, Kimura S, Horie N, Matsui T, Nishigaki Y, Ryu K, Sakabe T, Itoi M, Ashihara E, Maekawa T, Fushiki S and Kubo T: Zoledronic acid inhibits proliferation of human fibrosarcoma cells with induction of apoptosis, and shows combined effects with other anticancer agents. Oncol Rep 24: 233-239, 2010.
- 20 Yang TY, Chang GC, Chen KC, Hung HW, Hsu KH, Sheu GT and Hsu SL: Sustained activation of ERK and CDK2/cyclin-A signaling pathway by pemetrexed leading to S-phase arrest and apoptosis in human non-small cell lung cancer A549 cells. Eur J Pharmacol 663: 17-26, 2011.
- 21 Misso G, Porru M, Stoppacciaro A, Castellano M, De Cicco F, Leonetti C, Santini D and Caraglia M: Evaluation of the *in vitro* and *in vivo* antiangiogenic effects of denosumab and zoledronic acid. Cancer Biol Ther 13: 1491-1500, 2012.
- 22 Yamada J, Tsuno NH, Kitayama J, Tsuchiya T, Yoneyama S, Asakage M, Okaji Y, Shuno Y, Nishikawa T, Tanaka J, Takahashi K and Nagawa H: Antiangiogenic property of zoledronic acid by inhibition of endothelial progenitor cell differentiation. J Surg Res 151: 115-120, 2009.
- 23 Nagao S, Hattori N, Fujitaka K, Iwamoto H, Ohshimo S, Kanehara M, Ishikawa N, Haruta Y, Murai H and Kohno N: Regression of a primary pulmonary adenocarcinoma after zoledronic acid monotherapy. Hiroshima J Med Sci 60: 7-9, 2011.
- 24 Li XY, Lin YC, Huang WL, Hong CQ, Chen JY, You YJ and Li WB: Zoledronic acid inhibits proliferation and impairs migration and invasion through down regulating VEGF and MMPs expression in human nasopharyngeal carcinoma cells. Med Oncol 29: 714-720, 2012.
- 25 You Y, Liu J, Wang Z, Zhang Y, Ran Y, Guo X, Liu H and Wang H: The enhancement of radiosensitivity in human esophageal squamous cell carcinoma cells by zoledronic acid and its potential mechanism. Cytotechnology 2013.
- 26 Insalaco L, Di Gaudio F, Terrasi M, Amodeo V, Caruso S, Corsini LR, Fanale D, Margarese N, Santini D, Bazan V and Russo A: Analysis of molecular mechanisms and antitumoural effects of zoledronic acid in breast cancer cells. J Cell Mol Med *16*: 2186-2195, 2012.
- 27 Vassiliou V: Management of metastatic bone disease in the elderly with bisphosphonates and receptor activator of NF-kB ligand inhibitors: Effectiveness and safety. Clin Oncol 25: 290-297, 2013.
- 28 Santini D, Schiavon G, Vincenzi B, Gaeta L, Pantano F, Russo A, Ortega C, Porta C, Galluzzo S, Armento G, La Verde N, Caroti C, Treilleux I, Ruggiero A, Perrone G, Addeo R, Clezardin P, Muda AO and Tonini G: Receptor activator of NF-kB (RANK)

expression in primary tumors associates with bone metastasis occurrence in breast cancer patients. PLoS One 6: e19234, 2011.

- 29 Jin R, Sterling JA, Edwards JR, Degraff DJ, Lee C, Park SI and Matusik RJ: Activation of NF-kappa B signaling promotes growth of prostate cancer cells in bone. PLoS One 8: e60983, 2013.
- 30 Peng H, Sohara Y, Moats RA, Nelson MD, Jr, Groshen SG, Ye W, Reynolds CP and DeClerck YA: The activity of zoledronic acid on neuroblastoma bone metastasis involves inhibition of osteoclasts and tumor cell survival and proliferation. Cancer Res 67: 9346-9355, 2007.
- 31 Van Beek E, Lowik C, van der Pluijm G and Papapoulos S: The role of geranylgeranylation in bone resorption and its suppression by bisphosphonates in fetal bone explants *in vitro*: A clue to the mechanism of action of nitrogen-containing bisphosphonates. J Bone Miner Res 14: 722-729, 1999.
- 32 Roelofs AJ, Thompson K, Ebetino FH, Rogers MJ and Coxon FP: Bisphosphonates: Molecular mechanisms of action and effects on bone cells, monocytes and macrophages. Curr Pharm Des *16*: 2950-2960, 2010.
- 33 Russell RG, Watts NB, Ebetino FH and Rogers MJ: Mechanisms of action of bisphosphonates: Similarities and differences and their potential influence on clinical efficacy. Osteoporos Int 19: 733-759, 2008.
- 34 Sorscher SM and Lockhart AC: Ras inhibition and the survival benefit favoring zoledronic acid compared with denosumab in patients with multiple myeloma. J Clin Oncol 29: 2735-2736, 2011.
- 35 Nogawa M, Yuasa T, Kimura S, Kuroda J, Segawa H, Sato K, Yokota A, Koizumi M and Maekawa T: Zoledronic acid mediates Ras-independent growth inhibition of prostate cancer cells. Oncol Res 15: 1-9, 2005.
- 36 Denoyelle C, Hong L, Vannier JP, Soria J and Soria C: New insights into the actions of bisphosphonate zoledronic acid in breast cancer cells by dual RhoA-dependent and -independent effects. Br J Cancer 88: 1631-1640, 2003.
- 37 Luckman SP, Hughes DE, Coxon FP, Russell GG and Rogers MJ: JBMR anniversary classic. Nitrogen-containing *bis*phosphonates inhibit the mevalonate pathway and prevent post-translational prenylation of GTP-binding proteins, including Ras. J Bone Miner Res 20: 1265-1274, 2005.
- 38 Ebert R, Zeck S, Meissner-Weigl J, Klotz B, Rachner TD, Benad P, Klein-Hitpass L, Rudert M, Hofbauer LC and Jakob F: Kruppel-like factors KLF2 and 6 and Ki-67 are direct targets of zoledronic acid in MCF-7 cells. Bone 50: 723-732, 2012.
- 39 Mitrofan LM, Pelkonen J and Monkkonen J: The level of ATP analog and isopentenyl pyrophosphate correlates with zoledronic acid-induced apoptosis in cancer cells *in vitro*. Bone 45: 1153-1160, 2009.
- 40 Raikkonen J, Crockett JC, Rogers MJ, Monkkonen H, Auriola S and Monkkonen J: Zoledronic acid induces formation of a proapoptotic ATP analogue and isopentenyl pyrophosphate in osteoclasts *in vivo* and in MCF-7 cells *in vitro*. Br J Pharmacol *157*: 427-435, 2009.
- 41 Monkkonen H, Kuokkanen J, Holen I, Evans A, Lefley DV, Jauhiainen M, Auriola S and Monkkonen J: *Bis*phosphonateinduced ATP analog formation and its effect on inhibition of cancer cell growth. Anticancer Drugs *19*: 391-399, 2008.
- 42 Monkkonen H, Ottewell PD, Kuokkanen J, Monkkonen J, Auriola S and Holen I: Zoledronic acid-induced IPP/ApppI production *in vivo*. Life Sci 81: 1066-1070, 2007.

- 43 Dunford JE, Thompson K, Coxon FP, Luckman SP, Hahn FM, Poulter CD, Ebetino FH and Rogers MJ: Structure activity relationships for inhibition of farnesyl *bis*phosphate synthase *in vitro* and inhibition of bone resorption *in vivo* by nitrogen-containing bisphosphonates. J Pharmacol Exp Ther 296: 235-242, 2001.
- 44 Senaratne SG, Mansi JL and Colston KW: The bisphosphonate zoledronic acid impairs Ras membrane [correction of impairs membrane] localisation and induces cytochrome *c* release in breast cancer cells. Br J Cancer *86*: 1479-1486, 2002.
- 45 Mitrofan LM, Castells FB, Pelkonen J and Monkkonen J: Lysosomal-mitochondrial axis in zoledronic acid-induced apoptosis in human follicular lymphoma cells. J Biol Chem 285: 1967-1979, 2010.
- 46 Tamura T, Shomori K, Nakabayashi M, Fujii N, Ryoke K and Ito H: Zoledronic acid, a third-generation *bis*phosphonate, inhibits cellular growth and induces apoptosis in oral carcinoma cell lines. Oncol Rep 25: 1139-1143, 2011.
- 47 Almubarak H, Jones A, Chaisuparat R, Zhang M, Meiller TF and Scheper MA: Zoledronic acid directly suppresses cell proliferation and induces apoptosis in highly tumorigenic prostate and breast cancers. J Carcinog *10*: 2, 2011.
- 48 Dedes PG, Gialeli C, Tsonis AI, Kanakis I, Theocharis AD, Kletsas D, Tzanakakis GN and Karamanos NK: Expression of matrix macromolecules and functional properties of breast cancer cells are modulated by the *bis*phosphonate zoledronic acid. Biochim Biophys Acta 1820: 1926-1939, 2012.
- 49 Mani J, Vallo S, Barth K, Makarevic J, Juengel E, Bartsch G, Wiesner C, Haferkamp A and Blaheta RA: Zoledronic acid influences growth, migration and invasive activity of prostate cancer cells *in vitro*. Prostate Cancer Prostatic Dis 15: 250-255, 2012.
- 50 Hasmim M, Bieler G and Ruegg C: Zoledronate inhibits endothelial cell adhesion, migration and survival through the suppression of multiple, prenylation-dependent signaling pathways. J Thromb Haemost 5: 166-173, 2007.
- 51 Bezzi M, Hasmim M, Bieler G, Dormond O and Ruegg C: Zoledronate sensitizes endothelial cells to tumor necrosis factorinduced programmed cell death: evidence for the suppression of sustained activation of focal adhesion kinase and protein kinase B/Akt. J Biol Chem 278: 43603-43614, 2003.
- 52 Bellahcene A, Chaplet M, Bonjean K and Castronovo V: Zoledronate inhibits ανβ3 and ανβ5 integrin cell surface expression in endothelial cells. Endothelium *14*: 123-130, 2007.
- 53 Maubant S, Leroy-Dudal J, Carreiras F, Deslandes E, Duigou F, Staedel C and Gauduchon P: Cell surface overexpression of  $\alpha\nu\beta5$ integrin impedes  $\alpha\nu\beta3$ -mediated migration of the human ovarian adenocarcinoma cell line IGROV1. Cell Biol Int *31*: 109-118, 2007.
- 54 Skuli N, Monferran S, Delmas C, Favre G, Bonnet J, Toulas C and Cohen-Jonathan Moyal E: ανβ3/ανβ5 Integrins-FAK-RhoB: A novel pathway for hypoxia regulation in glioblastoma. Cancer Res 69: 3308-3316, 2009.
- 55 Sun LC, Luo J, Mackey LV, Fuselier JA and Coy DH: A conjugate of camptothecin and a somatostatin analog against prostate cancer cell invasion via a possible signaling pathway involving PI3K/AKT,  $\alpha\nu\beta3/\alpha\nu\beta5$  and MMP-2/-9. Cancer Lett 246: 157-166, 2007.
- 56 Di Salvatore M, Orlandi A, Bagala C, Quirino M, Cassano A, Astone A and Barone C: Antitumour and antiangiogenetic effects of zoledronic acid on human non-small cell lung cancer cell line. Cell Prolif 44: 139-146, 2011.

- 57 Terpos E, Efstathiou E, Christoulas D, Roussou M, Katodritou E and Dimopoulos MA: RANKL inhibition: Clinical implications for the management of patients with multiple myeloma and solid tumors with bone metastases. Expert Opin Biol Ther 9: 465-479, 2009.
- 58 Schmiedel BJ, Scheible CA, Nuebling T, Kopp HG, Wirths S, Azuma M, Schneider P, Jung G, Grosse-Hovest L and Salih HR: RANKL expression, function, and therapeutic targeting in multiple myeloma and chronic lymphocytic leukemia. Cancer Res 73: 683-694, 2013.
- 59 Canon JR, Roudier M, Bryant R, Morony S, Stolina M, Kostenuik PJ and Dougall WC: Inhibition of RANKL blocks skeletal tumor progression and improves survival in a mouse model of breast cancer bone metastasis. Clin Exp Metastasis 25: 119-129, 2008.
- 60 Armstrong AP, Miller RE, Jones JC, Zhang J, Keller ET and Dougall WC: RANKL acts directly on RANK-expressing prostate tumor cells and mediates migration and expression of tumor metastasis genes. Prostate *68*: 92-104, 2008.
- 61 Kostenuik PJ, Nguyen HQ, McCabe J, Warmington KS, Kurahara C, Sun N, Chen C, Li L, Cattley RC, Van G, Scully S, Elliott R, Grisanti M, Morony S, Tan HL, Asuncion F, Li X, Ominsky MS, Stolina M, Dwyer D, Dougall WC, Hawkins N, Boyle WJ, Simonet WS and Sullivan JK: Denosumab, a fully human monoclonal antibody to RANKL, inhibits bone resorption and increases BMD in knock-in mice that express chimeric (murine/human) RANKL. J Bone Miner Res 24: 182-195, 2009.
- 62 Ominsky MS, Stouch B, Schroeder J, Pyrah I, Stolina M, Smith SY and Kostenuik PJ: Denosumab, a fully human RANKL antibody, reduced bone turnover markers and increased trabecular and cortical bone mass, density, and strength in ovariectomized Cynomolgus monkeys. Bone *49*: 162-173, 2011.
- 63 Kuroda Y and Matsuo K: Molecular mechanisms of triggering, amplifying and targeting RANK signaling in osteoclasts. World J Orthop 3: 167-174, 2012.
- 64 McCoy EM, Hong H, Pruitt HC and Feng X: IL-11 produced by breast cancer cells augments osteoclastogenesis by sustaining the pool of osteoclast progenitor cells. BMC Cancer 13: 16, 2013.
- 65 Mancino AT, Klimberg VS, Yamamoto M, Manolagas SC and Abe E: Breast cancer increases osteoclastogenesis by secreting M-CSF and up regulating RANKL in stromal cells. J Surg Res 100: 18-24, 2001.
- 66 Taylor RM, Kashima TG, Knowles HJ and Athanasou NA: VEGF, FLT3 ligand, PIGF and HGF can substitute for M-CSF to induce human osteoclast formation: implications for giant cell tumour pathobiology. Lab Invest 92: 1398-1406, 2012.
- 67 Ryser MD, Qu Y and Komarova SV: Osteoprotegerin in bone metastases: Mathematical solution to the puzzle. PLoS Comput Biol 8: e1002703, 2012.
- 68 Yonou H, Horiguchi Y, Ohno Y, Namiki K, Yoshioka K, Ohori M, Hatano T and Tachibana M: Prostate-specific antigen stimulates osteoprotegerin production and inhibits receptor activator of nuclear factor-KB ligand expression by human osteoblasts. Prostate 67: 840-848, 2007.
- 69 Atkins GJ, Bouralexis S, Haynes DR, Graves SE, Geary SM, Evdokiou A, Zannettino AC, Hay S and Findlay DM: Osteoprotegerin inhibits osteoclast formation and bone resorbing activity in giant cell tumors of bone. Bone 28: 370-377, 2001.
- 70 Coxon JP, Oades GM, Kirby RS and Colston KW: Zoledronic acid induces apoptosis and inhibits adhesion to mineralized matrix in prostate cancer cells via inhibition of protein prenylation. BJU Int 94: 164-170, 2004.

- 71 Goffinet M, Thoulouzan M, Pradines A, Lajoie-Mazenc I, Weinbaum C, Faye JC and Seronie-Vivien S: Zoledronic acid treatment impairs protein geranyl-geranylation for biological effects in prostatic cells. BMC Cancer 6: 60, 2006.
- 72 Oades GM, Senaratne SG, Clarke IA, Kirby RS and Colston KW: Nitrogen containing bisphosphonates induce apoptosis and inhibit the mevalonate pathway, impairing Ras membrane localization in prostate cancer cells. J Urol 170: 246-252, 2003.
- 73 Lu S, Zhang J, Zhou Z, Liao ML, He WZ, Zhou XY, Li ZM, Xiang JQ, Wang JJ and Chen HQ: Synergistic inhibitory activity of zoledronate and paclitaxel on bone metastasis in nude mice. Oncol Rep 20: 581-587, 2008.
- 74 Kim SJ, Uehara H, Yazici S, He J, Langley RR, Mathew P, Fan D and Fidler IJ: Modulation of bone microenvironment with zoledronate enhances the therapeutic effects of STI571 and paclitaxel against experimental bone metastasis of human prostate cancer. Cancer Res *65*: 3707-3715, 2005.
- 75 Neville-Webbe HL, Rostami-Hodjegan A, Evans CA, Coleman RE and Holen I: Sequence- and schedule-dependent enhancement of zoledronic acid-induced apoptosis by doxorubicin in breast and prostate cancer cells. Int J Cancer *113*: 364-371, 2005.
- 76 Ottewell PD, Lefley DV, Cross SS, Evans CA, Coleman RE and Holen I: Sustained inhibition of tumor growth and prolonged survival following sequential administration of doxorubicin and zoledronic acid in a breast cancer model. Int J Cancer *126*: 522-532, 2010.
- 77 Daubine F, Le Gall C, Gasser J, Green J and Clezardin P: Antitumor effects of clinical dosing regimens of *bisphosphonates* in experimental breast cancer bone metastasis. J Natl Cancer Inst 99: 322-330, 2007.
- 78 Martin CK, Werbeck JL, Thudi NK, Lanigan LG, Wolfe TD, Toribio RE and Rosol TJ: Zoledronic acid reduces bone loss and tumor growth in an orthotopic xenograft model of osteolytic oral squamous cell carcinoma. Cancer Res 70: 8607-8616, 2010.
- 79 Tannehill-Gregg SH, Levine AL, Nadella MV, Iguchi H and Rosol TJ: The effect of zoledronic acid and osteoprotegerin on growth of human lung cancer in the tibias of nude mice. Clin Exp Metastasis 23: 19-31, 2006.
- 80 Buijs JT, Que I, Lowik CW, Papapoulos SE and van der Pluijm G: Inhibition of bone resorption and growth of breast cancer in the bone microenvironment. Bone 44: 380-386, 2009.
- 81 Walker K, Medhurst SJ, Kidd BL, Glatt M, Bowes M, Patel S, McNair K, Kesingland A, Green J, Chan O, Fox AJ and Urban LA: Disease modifying and anti-nociceptive effects of the *bis*phosphonate, zoledronic acid in a model of bone cancer pain. Pain 100: 219-229, 2002.
- 82 Li YY, Chang JW, Liu YC, Wang CH, Chang HJ, Tsai MC, Su SP and Yeh KY: Zoledronic acid induces cell-cycle prolongation in murine lung cancer cells by perturbing cyclin and Ras expression. Anticancer Drugs 22: 89-98, 2011.
- 83 Hoff BA, Chughtai K, Jeon YH, Kozloff K, Galban S, Rehemtulla A, Ross BD and Galban CJ: Multimodality imaging of tumor and bone response in a mouse model of bony metastasis. Transl Oncol 5: 415-421, 2012.
- 84 Hiraga T, Williams PJ, Ueda A, Tamura D and Yoneda T: Zoledronic acid inhibits visceral metastases in the 4T1/luc mouse breast cancer model. Clin Cancer Res 10: 4559-4567, 2004.
- 85 Corey E, Brown LG, Quinn JE, Poot M, Roudier MP, Higano CS and Vessella RL: Zoledronic acid exhibits inhibitory effects on

osteoblastic and osteolytic metastases of prostate cancer. Clin Cancer Res 9: 295-306, 2003.

- 86 Montague R, Hart CA, George NJ, Ramani VA, Brown MD and Clarke NW: Differential inhibition of invasion and proliferation by *bis*phosphonates: Anti-metastatic potential of zoledronic acid in prostate cancer. Eur Urol 46: 389-401, 2004.
- 87 Jones DH, Nakashima T, Sanchez OH, Kozieradzki I, Komarova SV, Sarosi I, Morony S, Rubin E, Sarao R, Hojilla CV, Komnenovic V, Kong YY, Schreiber M, Dixon SJ, Sims SM, Khokha R, Wada T and Penninger JM: Regulation of cancer cell migration and bone metastasis by RANKL. Nature 440: 692-696, 2006.
- 88 Kupas V, Weishaupt C, Siepmann D, Kaserer ML, Eickelmann M, Metze D, Luger TA, Beissert S and Loser K: RANK is expressed in metastatic melanoma and highly upregulated on melanoma-initiating cells. J Invest Dermatol 131: 944-955, 2011.
- 89 Zhang J, Dai J, Qi Y, Lin DL, Smith P, Strayhorn C, Mizokami A, Fu Z, Westman J and Keller ET: Osteoprotegerin inhibits prostate cancer-induced osteoclastogenesis and prevents prostate tumor growth in the bone. J Clin Invest 107: 1235-1244, 2001.
- 90 Lipton A, Fizazi K, Stopeck AT, Henry DH, Brown JE, Yardley DA, Richardson GE, Siena S, Maroto P, Clemens M, Bilynskyy B, Charu V, Beuzeboc P, Rader M, Viniegra M, Saad F, Ke C, Braun A and Jun S: Superiority of denosumab to zoledronic acid for prevention of skeletal-related events: A combined analysis of three pivotal, randomised, phase III trials. Eur J Cancer 48: 3082-3092, 2012.
- 91 Branstetter DG, Nelson SD., Manivel JC, Blay JY, Chawla S, Thomas DM, Jun S and Jacobs I: Denosumab induces tumor reduction and bone formation in patients with giant-cell tumor of bone. Clin Cancer Res 18: 4415-4424, 2012.
- 92 Fizazi K, Carducci M, Smith M, Damiao R, Brown J, Karsh L, Milecki P, Shore N, Rader M, Wang H, Jiang Q, Tadros S, Dansey R and Goessl C: Denosumab *versus* zoledronic acid for treatment of bone metastases in men with castration-resistant prostate cancer: A randomised, double-blind study. Lancet *377*: 813-822, 2011.
- 93 Stopeck AT, Lipton A, Body JJ, Steger GG, Tonkin K, de Boer RH, Lichinitser M, Fujiwara Y, Yardley DA, Viniegra M, Fan M, Jiang Q, Dansey R, Jun S and Braun A: Denosumab compared with zoledronic acid for the treatment of bone metastases in patients with advanced breast cancer: A randomized, double-blind study. J Clin Oncol 28: 5132-5139, 2010.
- 94 Henry DH, Costa L, Goldwasser F, Hirsh V, Hungria V, Prausova J, Scagliotti GV, Sleeboom H, Spencer A, Vadhan-Raj S, von Moos R, Willenbacher W, Woll PJ, Wang J, Jiang Q, Jun S, Dansey R and Yeh H: Randomized, double-blind study of denosumab *versus* zoledronic acid in the treatment of bone metastases in patients with advanced cancer (excluding breast and prostate cancer) or multiple myeloma. J Clin Oncol 29: 1125-1132, 2011.
- 95 Cleeland CS, Body JJ, Stopeck A, von Moos R, Fallowfield L, Mathias SD, Patrick DL, Clemons M, Tonkin K, Masuda N, Lipton A, de Boer R, Salvagni S, Oliveira CT, Qian Y, Jiang Q, Dansey R, Braun A and Chung K: Pain outcomes in patients with advanced breast cancer and bone metastases: Results from a randomized, double-blind study of denosumab and zoledronic acid. Cancer 119: 832-838, 2013.

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