Review

A Significant Role of Lipogenic Enzymes in Colorectal Cancer

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Abstract. In this review, we summarize recent progress regarding the study of the main enzymes of lipid metabolism involved in colorectal cancer development, namely of a) farnesyltransferase (Ftase), a cytosolic enzyme that catalyzes the first step in the protein farnesylation; b) farnesyl diphosphate synthase (FPPS, which yields FPP, a substrate for Ftase; c) fatty acid synthase (FAS), an enzyme required for the conversion of acetyl-CoA and malonyl-CoA to palmitate; and d) lipoprotein lipase (LPL), the crucial enzyme for intravascular catabolism of triglyceride-rich lipoproteins. Alterations in the levels of these enzymes may contribute to a cell growth advantage acquired during the carcinogenic process and to the development of malignancy. We have demonstrated an elevated Ftase activity in human colorectal cancer (CRC), with differences in Ftase activity related to histological grading, tumor location and KRAS mutation status. Moreover, the first evidence of FPPS activity in human CRC was demonstrated by our study, where a higher FPPS activity and mRNA expression was present in cancer rather than in normal mucosa. We also detected a hyperactivation of FAS in colon cancer, related to tumor location, sex and, p53 mutation status. Our data reinforce the role of lipid metabolism in the regulation of cellular metabolic processes and in carcinogenesis. Moreover, our findings suggest that biological factors including sex, gene mutation status, as well as the stratification of patients with colorectal cancer into right- and left-sided subsets may be important in patient selection for targeted therapies. Our studies in vitro demonstrated that FAS might also be a

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molecular target for the antiproliferative activity of olive oil polyphenols in a metabolically defined subset of patients with colon cancer. Moreover, we detected that the serum levels of FAS in patients with colorectal cancer are associated with tumor stage. Recently, we found a significant reduction in the levels of FAS and another lipogenic enzyme, LPL, in adipose tissue adjacent to tumor lesions, compared to the levels of FAS detected in paired tissue distant from neoplasia in patients with colorectal cancer. The study of metabolic changes in lipogenic enzyme pathways, as well as the determination of the distribution of individual roles within each biochemical pathway provide a rationale for selecting a particular reaction step suitable for therapeutic intervention.

Colorectal cancer (CRC) may involve different genetic pathways or different combinations of genetic and biochemical alterations during tumorigenesis (1, 2). An important feature of malignant transformation is the loss of the cholesterol feedback inhibition mechanism that regulates cholesterol synthesis (3). Alterations in biosynthetic processes of the cholesterol pathway and in the levels of enzyme products participating in this biochemical system may contribute to the cell growth advantage acquired during carcinogenesis and to the development of malignancy (4). Several Hydroxy-Methyl-Glutaryl-Coenzyme A (HMGCoA) metabolites, such as farnesyl pyrophosphate (FPP) and geranyl pyrophosphate (GPP) are implicated in oncogene activation and tumorigenesis (4) while the production of isoprenoid intermediates has been found intrinsically enhanced in rapidly proliferating cells (5).

Farnesyltransferase

FPP, serving as a precursor for lipid end-products, is a substrate for protein isoprenylation by farnesyltransferase (Ftase), a cytosolic enzyme that catalyzes the first step of the post-translational modification, "farnesylation" of a number of cellular polypeptides, including RAS (6), the nuclear intermediate filament proteins lamin B (7) and

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prelamin A (8), the centromere proteins CENP-E and CENP-F (9), protein tyrosine phosphatases PRL-1, -2 and -3 (10), cyclic GMP phosphodiesterase alpha (11) and others (12-14). Ftase is a heterodimer of α and β subunits. The α subunit appears to be the carrier of its substrate FPP, while the β subunit binds to RAS protein. Ftase has attracted attention because of its role in the processing of RAS proteins, which function as molecular switches linking receptor and non-receptor tyrosine kinase activation to downstream cytoplasmic and nuclear events. Given the role these proteins play in pathways regulating cell survival, proliferation, differentiation and cytoskeletal organization, it seems likely that their altered expression may manifest a markedly abnormal function.

Oncogenic RAS mutations have been identified in ~30% of various human cancers, including CRC, (15). Since the maturation of RAS proteins was originally reported to be dependent on farnesylation (16), Ftase activity inhibition was envisioned as a strategy for interfering with RAS-mediated cell transformation (17-19), although it is now clear that there might also be additional targets (20, 21). The interest in the study of Ftase in tumor growth has been also reinforced by recent experimental observations. Khan *et al.* observed enhanced Ftase activity and mRNA expression in human skin basal carcinoma compared with normal tissue (22); Tanimoto *et al.* found that *Ftase* β -subunit mRNA overexpression in ovarian carcinoma is associated with the *KRAS* mutation (23).

Our study showed an increased Ftase activity in human CRC, showing differences in Ftase activity in relation to histological grading, tumor location and *KRAS* mutation status (24). These findings confirm that Ftase activity may be correlated with more aggressive tumor behavior. Likely, the transformed colonic cells acquire a growth advantage over the non-transformed cells, because the former are capable of utilizing isoprenoid compounds, such as farnesyl, for other important pathways that diverge from the main cholesterogenic route (the *flux diversion hypothesis*) (25).

Therapeutic strategies targeted to the enzymatic activities of the mevalonate pathway are in clinical evaluation. The Ftase inhibitors (FTIs) are currently being investigated in phase I and phase II clinical trials for several solid tumors, including CRC (26-28).

Farnesyl Diphosphate Synthase

FPP is yielded by farnesyl diphosphate synthase (FPPS), which is the pharmacological target of nitrogen-containing biphosphonates, such as alendronate, and pamidronate, potent inhibitors of bone resorption (29). Recent data suggest that, besides inhibiting osteoclast activity, biphosphonate may also have an anticancer action by a pro-apototic and antiproliferative effect on tumor cells (30-32). These effects of biphosphonates have been attributed to different

biochemical mechanisms (33), but the most important seems to be the inhibition of FPPS activity (34). This inhibition blocks the biosynthesis of cholesterol with changes in the signaling events which lead to the induction of the caspase cascade and apoptosis (35).

Our previous study provided the first evidence of FPPS activity in human CRC (36). A higher FPPS activity and higher mRNA expression levels has been found in cancer than in normal mucosa. This observation is in agreement with a recent study of Sung et al. (37) who detected overexpression of the FPPS gene in human hepatocellular carcinoma (HCC), as well as an upregulation of the FPPS protein in about 85% of HCC cases examined. An elevated FPPS expression has been also observed in rat prostate tumor cell lines (38). FPPS seems to be regulated by androgens in rat prostatic cells, suggesting that it likely plays a significant role in androgen action and prostate cancer progression. Furthermore, the demonstration of increased FPPS activity and of an overexpression of its mRNA in CRC, not only clarifies our understanding of colon tumorigenesis, but also contributes to the development of therapeutic targets. In this study, we also investigated the effects of pamidronate on cell growth and apoptosis in a human colon cancer cell line. Cell proliferation inhibition by pamidronate on DLD-1 cells underlines the involvement of FPPS in cell growth and proliferation. The antiproliferative effect of pamidronate observed in this cell line may occur via the apoptotic pathway, as the exposure of DLD-1 cells to pamidronate promoted apoptosis. Apoptosis, or programmed cell death, and cell proliferation act concurrently in cancer cells, and often the balance between the two processes determines the net tumor growth rate. In our experiments, the inhibition of cell growth and the apoptotic effects exerted by pamidronate appear to be due to inhibition of FPPS, reinforcing the role of isoprenoids in the regulation of cellular metabolic processes.

Fatty Acid Synthase

Another enzyme known to play an important role in the growth and pathogenesis of colon carcinoma is fatty acid synthase (FAS), a multi-enzyme protein containing domains for acyl-carrier peptide and the seven different catalytic activities required for the conversion of acetyl-CoA and malonyl-CoA to palmitate (39). The expression of FAS is linked to specific functions, such as *de novo* biosynthesis of fatty acids, the conversion and storage of energy in the liver and in the adipose tissue (40), while FAS expression is possibly involved in the regulation of food intake (39, 41). Rapidly proliferating cells and tissues, including proliferative endometrium and some fetal tissues, have high levels of FAS expression (42, 43), which likely support membrane synthesis (44). In most normal human tissuesa, however, FAS is generally expressed at low levels because cells

preferentially use circulating dietary fatty acids for the synthesis of new structural lipids (40). High levels of FAS expression have been found in many types of human cancers, including cancer of the breast, prostate, colon, ovary, thyroid and endometrium (45, 46). In breast epithelial cells, the malignant transformation leads to the up-regulation of FAS and high levels of FAS are associated with poor clinical outcome (47), suggesting a relationship between FAS expression and tumor aggressiveness (48). Several studies have shown that FAS inhibitors suppress mammary carcinogenesis, suggesting that FAS is a promising molecular target for breast cancer chemoprevention (49, 50). FAS is overexpressed at both protein and mRNA levels in prostate carcinoma and its high expression has also been associated with aggressive biological behaviour of tumors (46). Biochemical studies of FAS activity and fatty acid synthesis in colon cancer cells have shown that its levels of expression correlates with the overall activity of the fatty acid synthetic pathway (45, 51, 52).

Our recent study has demonstrated increased FAS activity in colorectal tumors compared with the adjacent normal tissue (53). Our data are in accordance with those of Rhashid *et al*. (39), who demonstrated elevated levels of FAS activity in colorectal neoplasia as compared to adjacent normal mucosa in six surgical specimens of colon. In addition, in our study, we demonstrated that colon tumors on the left side had higher FAS activity than tumors located on the right. A biological difference between proximal and distal colon tumors has also been observed in regard other enzymes, growth factors and multiple genetic alterations (54, 55), underlining the regional biochemical variability of colon neoplastic transformation.

Hyperactivation of FAS seems also to be related to sex in our study. Tumors from male patients had higher levels of FAS activity than tumors from females, suggesting an androgen imprint on FAS activity regulation in cancer tissue from male patients. Sex steroid hormones and growth factors have been demonstrated to up-regulate FAS through SREBPs, which stimulate FAS transcription (46). Several investigations have demonstrated that FAS expression is markedly stimulated by androgens in human prostate cancer cell lines (38) and that an activated androgen signaling pathway in prostate cancer cells contributes to the up-regulation of FAS expression (56, 57). Our findings support the idea that FAS may also be an androgen-regulated enzyme in CRC.

A different behaviour with respect to protein activity was observed for FAS mRNA levels of carcinomas when compared to normal adjacent mucosa: higher FAS mRNA levels were found in normal colonic tissue compared to cancer tissue. It is possible that the mRNA of the FAS gene has an accelerated turnover and degradation in neoplastic compared with normal tissue, or that mRNA modifications in tumor tissue occur at post-transcriptional level.

Recently, important roles for p53 have been recognized in the cellular responses to a variety of metabolic stresses, including hypoxia, acidosis and perturbations of protein synthesis (58). It seems that perturbation of fatty acid synthesis also belongs to the list of metabolic stresses regulated by p53. Changes in FAS activity in colorectal tumors may be modulated by p53, since we have found lower levels of FAS activity in patients carrying the p53 mutation, even if the difference was not statistically significant. Accumulation of the p53 protein, subsequent to the p53 gene mutation has been demonstrated to increase the sensitivity of tumor cells to FAS inhibitors (58). The relatively higher FAS activity in tumors with intact p53 function suggests that the p53 protein may be involved in the regulation of FAS in human CRC. Our data, therefore, suggest that biological factors, including sex and gene mutation status, as well as the stratification of patients with CRC into right- and left-sided subsets may be important in patient selection for targeted therapies and for the subsequent assessment of objective therapeutic responses.

Our recent study shows for the first time, to our knowledge, down-regulation of FAS after hydroxytyrosol treatment in the SW620 human CRC cell line (59). Among phenolic compounds of olive oil, oleuropein and hydroxytyrosol are those which give extra-virgin olive oil its bitter, pungent taste, and they possess powerful antioxidant properties in vitro (60). As antioxidants, polyphenols may protect cell constituents against oxidative damage and may act as highly effective chemopreventive agents (61-63). Moreover, hydroxytyrosol significantly inhibits the proliferation of SW620 cell, through a pro-apoptotic effect. Exposure to increasing concentrations of hydroxytyrosol also led to a remarkable reduction in the proliferation of HT-29 cells. In these cells, the reduction in cancer cell growth detected after such an exposure seems to be independent of FAS gene expression and its enzymatic activity. These findings suggest that FAS might be a molecular target for antiproliferative activity of olive oil polyphenols in a metabolically defined subset of patients with colon cancer. Interestingly our findings demonstrate that the main olive oil polyphenols may induce antiproliferative effects in human CRC cells. These inhibitory effects on cell growth are mediated by FAS in certain human CRC cell types, suggesting that more differentiated and specialized cells are more able to control the expression of genes involved in cell proliferation.

In order to detect the presence of metastasis in CRC, several studies have been carried out in order to improve the prognosis of patients with aggressive disease. Identification of serum protein markers of CRC, together with other markers already known, could help provide such a non-invasive diagnostic screening tool. In a recent study of ours (64) we described for the first time a significant association between circulating levels of FAS and clinical stage CRC. Patients with tumor stage III and IV have significantly higher serum levels of FAS than patients with tumor stage I and II.

The quantitative determination of FAS in the blood might represent a tool to evaluate the aggressiveness during the progression of CRC.

Experiments are currently underway in our laboratory to establish if the measurement of serum FAS might be a rapid and non-invasive test of adequate prognostic/diagnostic accuracy capable of monitoring the effects of antineoplatic treatment. Moreover, it is possible that serum FAS levels could be helpful in identifying and following-up patients with colorectal cancer, when used in combination with other markers, emulating their use in breast and pancreatic cancer (65-68).

Lipoprotein Lipase

Several studies have demonstrated a clear association between obesity and the risk of CRC (69-72). Mature adipocytes seem to influence colon cancer cell proliferation (73). Adipose tissue synthesizes lipoprotein lipase (LPL), the crucial enzyme for intravascular catabolism of triglyceriderich lipoproteins. Patients with resectable non-small cell lung cancer have higher LPL activity in cancer tissue than in adjacent, apparently healthy, non-cancer lung tissue (74). Moreover, increased LPL activity in non-small cell lung cancer tissue predicts shorter patient survival, independent of standard prognostic factors (73).

The involvement of both LPL and FAS in tumor biology has been widely demonstrated in different studies and regional differences in the expression of these enzymes in visceral adipose tissue collected from patients with CRC might be representative of events which sustain tumor growth.

Therefore, the aim of our recent study was to evaluate LPL and FAS activity and the expression of their genes in adipose tissue adjacent to neoplasia as well as distant from it, in patients with CRC (75). In this study, both LPL and FAS were differentially expressed in the two different regions of adipose tissue. Our findings showed a significant reduction in both *LPL* and *FAS* gene expression and activity in adipose tissue adjacent to tumor lesion compared to those detected in paired tissue distant from the neoplasia. Adipose and colon tissue may affect the enzymatic expression of proteins involved in cell proliferation. The increased demand for long-chain fatty acids by the tumor may lead to adipose atrophy, resulting in alterations of lipid metabolism.

These results underline the influence of the tumor microenvironment on lipid metabolism in adipose tissue, demonstrating a tumor-induced impairment in the formation and lipid-storing capacity of adipose tissue in patients with CRC.

Conclusion

The results presented here demonstrate the extent at which lipogenic enzymes are involved in tumor development and progression. Understanding the distribution of roles within individual biochemical pathways is clearly important and it provides a rationale for selecting a particular reaction step suitable for therapeutic intervention. The molecular mechanisms linking the inhibition of the lipogenic pathway with the induction of cell death in tumor cells remain unclear. More work is needed to conclusively show that inhibition of lipogenesis can be efficacious in the long-term in order to control tumor growth *in vivo*.

References

- 1 Fearon ER and Vogelstein A: A genetic model for colorectal tumorigenesis. Cell 61: 759-767, 1990.
- 2 Srivastava S, Verma M and Henson DE: Biomarkers for early detection of colon cancer. Clinical Cancer Res 7: 1118-1126, 2001
- 3 Rao KN: The significance of cholesterol biosynthetic pathway in cell growth and carcinogenesis. Anticancer Res 15: 309-314, 1995.
- 4 Newmark HL: Squalene, olive oil, and cancer risk. Review and hypothesis. Ann NY Acad Sci 889: 193-203, 1999.
- 5 Krisans SK, Ericsson J, Edwards PA and Keller GA: Farnesyldiphosphate synthase is localized in peroxisomes. J Biol Chem 269: 14165-14169, 1994.
- 6 Clarke S: Protein isoprenylation and methylation at carboxylterminal cysteine residues. Annu Rev Biochem 61: 355-386, 1992.
- 7 Farnsworth CC, Wolda SL, Gelb MH and Glomset JA: Human lamin B contains a farnesylated cysteine residue. J Biol Chem *264*: 20422-20429, 1989.
- 8 Sinensky M, Famtle K, Triyillo M, McLain T, Kupfer A and Dalton M: The processing pathway of prelamin A. J Cell Sci 107: 61-67, 1994.
- 9 Ashar HR, James L, Gray K, Carr D, Black S, Armstrong L, Bishop WR and Kirschmeier P: FTIs block the farnesylation of CENP-E and CENP-F and alter the association of CENP-E with the microtubules. J Biol Chem 275: 30451-30457, 2000.
- 10 Zeng Q, Si X, Horstmann H, Xu Y, Hong W and Pallen CJ: Prenylation-dependent association of protein-tyrosine phosphatases PRL-1, -2, and -3 with the plasma membrane and the early endosome. J Biol Chem 275: 21444-21452, 2000.
- 11 Reiss Y, Seabra MC, Armstrong SA, Slaughter CA, Goldstein JL and Brown MS: Non-identical subunits of p21 Ras-farnesyltransferase:peptide binding and farnesylpirophosphate carrier functions. J Biol Chem 266: 10672-10677, 1991.
- 12 Reiss Y, Goldstein JL, Seabra MC, Casey P and Brown MS: Inhibition of purified p21 ras farnesyl protein transferase by cys-AAX tetrapeptides. Cell 62: 81-88, 1990.
- 13 Manne V, Roberts D, Tobin A, O'Rourke E, De Virgilio M, Meyers C, Ahmed N, Kurz B, Resh M, Kung HF and Barbacid M: Identification and preliminary characterization of proteincysteine farnesyltransferase. Proc Natl Acad Sci USA 87: 7541-7545, 1990.
- 14 Agarwal R, Khan SG, Athar M, Zaidi SY, Bichers DR and Mukhtar H: Ras protein p21 processing enzyme farnesyltransferase in chemical carcinogen-induced murine skin tumors. Mol Carcinog 8: 290-298, 1993.
- 15 Downward J: RAS signaling and apoptosis. Curr Opin Genet Dev 8: 49-54, 1998.

- 16 Bos J: RAS oncogenes in human cancer: a review. Cancer Res 49: 4682-4689, 1989.
- 17 Kato K, Cox AD, Hisaka MM, Graham SM, Buss JE and Der CJ: Isoprenoid addition to ras proteins is the critical modification for its membrane association and transforming activity. Proc Natl Acad Sci USA 89: 6403-6407, 1992.
- 18 Rowinsky EK, Windle JJ and Von Hoff DD: RAS protein farnesyltransferase: a strategic target for anticancer therapeutic development. J Clin Oncol 17: 3631-3652, 1999.
- 19 Reuter CW, Morgan MA and Bergmann L: Targeting the RAS signaling pathway: a rational mechanism-based treatment for hematologic malignancies? Blood 96: 1655-1669, 2000.
- 20 Lebowitz PF, Casey PJ, Prendergast GC and Thissen JA: Farnesyl-transferase inhibitors is mediated by gain of geranylgeranylated Rho B. J Biol Chem 272: 15591-15594, 1997.
- 21 Du W, Lebowitz PF and Prendergast GC: Cell growth inhibition by farnesyltransferase inhibitors is mediated by gain of geranylgeranylated Rho B. Mol Cell Biol 19: 1831-1834, 1999.
- 22 Khan SG, Dummer R, Siddiqui J, Bickers DR, Agarwal R and Mukhtar H: Farnesyltransferase activity and mRNA expression in human skin basal cell carcinomas. Biochem Biophys Res Comm 220: 795-801, 1996.
- 23 Tanimoto H, Mehta KD, Parmley TH, Shigemasa K, Parham GP, Clarke J and O'Brien TJ: Expression of the farnesyltransferase beta-subunit gene in human ovarian carcinoma: correlation to K-RAS mutation. Gynecol Oncol 66: 308-312, 1997.
- 24 Caruso MG, Notarnicola M, Bifulco M, Laezza C, Guerra V, Altomare DF, Memeo V, Lorusso D, Demma I and Di Leo A: Increased farnesyltransferase activity in human colorectal cancer: relationship with clinicopathological features and K-RAS mutation. Scand J Gastroenterol 38: 80-85, 2003.
- 25 Faust JR, Brown MS and Goldstein JL: Synthesis of Isopentenyl tRNA from mevalonate in cultured human fibroblasts. J Biol Chem 255: 6546-6548, 1980.
- 26 Eskens F, Awada A, Verweig J, Cutler DL, Hanauske A and Piccart M: Phase I and pharmacocinetic study of the oral Ftase inhibitor SCH66336 given twice daily to patients with advanced solid tumors. J Clin Oncol 19: 1167-1175, 2001.
- 27 Zujewski BJ, Horak ID, Bol CJ, Woestenborghs R, Bowden C, End DW, Piotrovsky VK, Chiao J, Belly RT, Todd A, Kopp WC, Kohler DR, Chow C, Noone M, Hakim FT, Larkin G, Gress RE, Nussenblatt RB, Kremer AB and Cowan KH: Phase I and pharmacokinetic study of farnesyl protein transferase inhibitor R115777 in advanced cancer. J Clin Oncol 18: 927-941, 2000.
- 28 Fitton A and Mc Tavish D: Pamidronate: a review of its pharmacological properties and therapeutic efficacy in resorptive bone disease. Drugs 41: 289-318, 1991.
- 29 Diel IJ, Solomayer EF and Bastert G: Biphosphonates and the prevention of metastasis: first eveidences from preclinical and clinical studies. Cancer 88(12 Suppl): 3080-3088, 2000.
- 30 Senaratne SG, Pirianov G, Mansi JL, Arnett TR and Colston KW: Biphosphonates induce apoptosis in human breast cancer cell lines. Br J Cancer 82: 1459-1468, 2000.
- 31 Tassone P, Forciniti S, Galea E, Morrone G, Turco MC, Martinelli V, Tagliaferri P and Venuta S: Growth inhibition and synergistic induction of apoptosis by zoledronate and dexamethasone in human myeloma cell lines. Leukemia 14: 841-844, 2000.
- 32 Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP and Wilhelm M: Stimulation of gamma/delta T-cells by

- aminobiphosphonates and induction of antiplasma cell activity in multiple myeloma. Blood *96*: 384-392, 2000.
- 33 Green JR: Antitumor potential of biphosphonates. Med Klin 95(2 Suppl): 23-28, 2000.
- 34 van Beek E, Pieterman E, Cohen L, Lowik C and Papapoulos S: Farnesyl pyrophosphate synthase is the molecular target of nitrogen-containing biphosphonates. Biochem Biophys Res Commun 264(1): 108-111, 1999.
- 35 Glomset JA, Gelb MH and Farnsworth CC: Geranylgeranylated proteins. Biochem Soc Trans 20(2): 479-484, 1992.
- 36 Notarnicola M, Messa C, Cavallini A, Bifulco M, Tecce MF, Eletto D, Di Leo A, Montemurro S, Laezza C and Caruso MG: Higher farnesyl diphosphate synthase activity in human colorectal cancer: inhibition of cellular apoptosis. Oncology 67: 351-358, 2004.
- 37 Sung YK, Hwang SY, Park MK, Farooq M, Han IS, Bae HI, Kim JC and Kim M: Glypican-3 is overexpressed in human hepatocellular carcinoma. Cancer Sci 94: 259-262, 2003.
- 38 Jiang F, Yang L, Cai X, Cyriac J, Shechter I and Wang Z: Farnesyl diphosphate synthase is abundantly expressed and regulated by androgen in rat prostatic epithelial cells. J Steroid Biochem Mol Biol 78: 123-130, 2001.
- 39 Rashid A, Pizer ES, Moga M, Milgraum LZ, Zahurak M, Pastermack GR, Kuhajda FP and Hamilton SR: Elevated expression of fatty acid synthase and fatty acid synthetic activity in colorectal neoplasia. American J Pathol 150: 201-208, 1997.
- 40 Kuhajda FP: Fatty acid synthase and human cancer: new perspectives on its role in tumor biology. Nutrition 16: 202-208, 2000.
- 41 Kim EK, Miller I, Landree LE, Borisy-Rudin FF, Brown P, Tihan T, Townsend CA, Witters LA, Moran TH, Kuhajda FP and Ronnett GV: Expression of FAS within hypothalamic neurons: a model for decreased food intake after c75 treatment. Am J Physiol Endocrinol Metab 283: E867-E879, 2002.
- 42 Pizer ES, Kurman RJ, Pastenack GR and Kuhajda FP: Expression of fatty acid synthase is closely linked to proliferation and stromal decidualization in cycling endometrium. Int J Gynecol Pathol 16: 45-51, 1997.
- 43 Wilentz R, Witters LA and Pizer E: Lipogenic enzyme fatty acid synthase and acetyl-CoA carboxylase are co-expressed with sterol regulatory element binding protein and Ki-67 in fetal tissue. Pediatr Dev Pathol 3: 525-531, 2000.
- 44 Jackowski S, Wang J and Baburina I: Activity of the phosphatidylcholine biosynthetic pathway modulates the distribution of fatty acids into glycerolipids in proliferating cells. Biochem Biophys Acta 1483: 301-315, 2000.
- 45 Chalbos D, Chambon M, Aohaud G and Rochefort H: Fatty acid synthetase and its mRNA are induced by progestins in breast cancer cells. J Biol Chem 262: 9923-9926, 1987.
- 46 Swinnen JV, Heemers H, Deboel L, Foufelle F, Heyns W and Verhoeven G: Stimulation of tumor-associated fatty acid synthase expression by growth factor activation of the sterol regulatory element-binding protein pathway. Oncogene 19: 5173-5181, 2000.
- 47 Kuhajda FP, Piantadosl S and Pasternack GR: Hepatoglobinrelated protein (HPR) epitopes in breast cancer as a predictor of recurrence of the disease. N Engl J Med 321: 636-641, 1989.
- 48 Alo P, Visca P, Marci A, Mangoni A, Botti C and Di Tondo U: Expression of fatty acid synthase (FAS) as a predictor of recurrence in stage I breast carcinoma patients. Cancer 77: 474-482, 1996.

- 49 Lu S, Zhang X, Badawi AF, El-Sohemy A and Archer MC: Cyclooxygenase-2 inhibitor celecoxib inhibits promotion of mammary tumorigenesis in rats fed a high fat diet rich in n-6 polyunsaturated fatty acids. Cancer Lett 184: 7-12, 2002.
- 50 Lu S and Archer MC: Fatty acid synthase is a potential molecular target for the chemoprevention of breast cancer. Carcinogenesis 26: 153-157, 2005.
- 51 Visca P, Alo PL, Del Nonno F, Botti C, Trombetta G, Marandino F, Filippi S, Di Tondo U and Donnorso RP: Immunohistochemical expression of fatty acid synthase, apoptotic-regulating genes, proliferating factors, and ras protein product in nonneoplastic mucosa. Clin Cancer Res 5: 4111-4118, 1999.
- 52 Chelbos D, Joyeux C, Galtier F and Rochefort H: Progestininduced fatty acid synthesase in breast cancer: from molecular biology to clinical applications. Ann NY Acad Sci 595: 67-73, 1990.
- 53 Notarnicola M, Altomare DF, Calvani M, Orlando A, Bifulco M, D'Attoma B and Caruso MG: Fatty acid synthase hyperactivation in human colorectal cancer: relationship with tumor side and sex. Oncology 71: 327-332, 2007.
- 54 Messa C, Russo F, Caruso MG and Di Leo A: EGF, TGF-α and EGF-R in human colorectal adenocarcinoma. Acta Oncologica *37*: 285-289, 1998.
- 55 Pascal G, Bouzourene H, Cerottini JP, Chaubert P, Benhattar J, Secic M, Wexner S, Givel JC and Belin B: Dukes B colorectal cancer. Distinct genetic categories and clinical outcome based on proximal or distal tumor location. Dis Colon Rectum 44: 364-373, 2001.
- 56 Swinnen JV, Vanderhoydonc F, Elgamal AA, Eelen M, Vercaeren I, Joniau S, Van Poppel H, Baert L, Goossens K, Heyns W and Verhoeven G: Selective activation of the fatty acid synthesis pathway in human prostate cancer. Int J Cancer 88: 176-179, 2000.
- 57 Rossi S, Graner E, Febbo P, Weinstein L, Bhattacharya N, Onody T, Bubley G, Balk S and Loda M: Fatty acid expression defines distinct molecular signatures in prostate cancer. Mol Cancer Res 1: 707-715, 2003.
- 58 Li JN, Gorospe M, Chrest FJ, Kumaravel TS, Evans MK, Han WF and Pizer ES: Pharmacological inhibition of fatty acid synthase activity produces both cytostatic and cytotoxic effects modulates by p53. Cancer Res *61*: 1493-1499, 2001.
- 59 Notarnicola M, Pisanti S, Tutino V, Bocale D, Rotelli MT, Gentile A, Memeo V, Bifulco M, Perri E and Caruso MG: Effects of olive oil polyphenols on fatty acid synthase gene expression and activity in human colorectal cancer cells. Genes Nutr 6(1): 63-69, 2011.
- 60 Gill CI, Boyd A, McDermott E, McCann M, Servili M, Selvaggini R, Taticchi A, Esposto S, Montedoro G, McGlynn H and Rowland I: Potential anticancer effects of virgin olive phenols on colorectal carcinogenesis models in vitro. Int J Cancer 117: 1-7, 2005.
- 61 Obied HK, Prenzler PD, Konczak I, Rheman A and Robard K: Chemistry and bioactivity of olive biophenols in some antioxidant and antiproliferative "in vitro" bioassays. Chem Res Toxicol 22: 227-234, 2009.
- 62 Fini L, Hotchkiss E, Fogliano V, Graziani G, Romano M, De Vol EB, Qin H, Selgrad M, Boland CR and Ricciardiello L: Chemopreventive properties of pinoresinol-rich olive oil involve a selective activation of the ATM-p53 cascade in colon cancer cell lines. Carcinogenesis 29: 139-146, 2008.

- 63 Owen RW, Giacosa A, Hull WE, Haubner R, Wurtele G, Spiegelhalder and Bartsch H: Olive-oil consumption and health: the possible role of antioxidants. Lancet Oncology 1: 107-112, 2000.
- 64 Notarnicola M, Tutino V, Calvani M, Lorusso D, Guerra V and Caruso MG: Serum levels of Fatty acid synthase in colorectal cancer are associated with tumor stage. J Gastrointestinal Cancer 2011, DOI: 10.1007/s12029-011-9300-2
- 65 Wang Y, Kuhajda FP, Li JN, Pizer ES, Han WF, Sokoll LJ and Chan DW: Fatty acid synthase expression in human breast cancer cell culture supernatants and in breast cancer patients. Cancer Lett 167: 99-104, 2001.
- 66 Walter K, Hong S-M, Nyhan S, Canto M, Fedarko N, Klein A, Griffith M, Omura N, Medghalchi S, Kuhajda F and Goggins M: Serum fatty acid synthase as a marker of pancreatic neoplasia. Cancer Epidemiol Biomarkers Prev 18(9): 2380-2385, 2009.
- 67 Vazquez-Martin A, Fernandez-Real JM, Oliveras-Ferraros C, Navarrete JM, Martin-Castillo B, Del Barco S, Brunet J and Menendez JA: Fatty acid synthase activity regulates HER2 extracellular domain shedding into the circulation of HER2positive metastatic breast cancer patients. Int J Oncology 35: 1369-1376, 2009.
- 68 Wang Y, Kuhajda FP, Li J, Finch TT, Cheng P, Koh C, Li T, Sokoll LJ and Chan DW: Fatty acid synthase as a tumor marker: its extracellular expression in human breast cancer. J Exp Ther Oncol *4*: 101-110, 2004.
- 69 Bianchini F, Kaaks R and Vainio H: Overweight, obesity and cancer risk. Lancet Oncology *3*: 565-574, 2002.
- 70 Calle EE, Rodriguez C, Walker-Thurmond K and Thun MJ: Overweight, obesity, and mortality from cancer in a prospectively studied cohort of US adults. N Engl J Med 348: 1625-1638, 2003.
- 71 Calle EE and Kaaks R: Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. Nat Rev Cancer 4: 579-591, 2004.
- 72 Amemori S, Ootani A, Aoki S, Fujise T, Shimoda R, Kakimoto T, Shiraishi R, Sakata Y, Tsunada S, Iwakiri R and Fujimoto K: Adipocytes and preadipocytes promote the proliferation of colon cancer cells "in vitro". Am J Physiol Gastrointest Liver Physiol 292: G923-G929, 2007.
- 73 Cerne D, Zitnik IP and Sok M: Increased fatty acid synthase activity in non-small cell lung cancer tissue is a weaker predictor of shorter patient survival than increased lipoprotein lipase activity. Arch Med Res 41: 405-409, 2010.
- 74 Trost Z, Sok M, Marc J and Cerne D: Increased lipoprotein lipase activity in non-small cell lung cancer tissue predicts shorter patient survival. Arch Med Res 40: 364-368, 2009.
- 75 Notarnicola M, Miccolis A, Tutino V, Lorusso D and Caruso MG: Low levels of lipogenic enzymes in peritumoral adipose tissue of colorectal cancer patients. Lipids *47(1)*: 59-63, 2012.

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