

Review

Death by Design: Where Curcumin Sensitizes Drug-resistant Tumours

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Abstract. Chemotherapy remains the core of anticancer treatment. However, despite the tremendous strides made in the development of targeted anticancer therapies, emergence of resistance to chemotherapeutic drugs is still a major obstacle in the successful management of resistant tumours. Therefore, profound investigation into the in-depth molecular mechanisms of drug resistance is essential and may hopefully translate into effective therapies that can flip the switch from drug resistance to susceptibility. Mechanistically, resistance phenomena may be explained by (i) overexpression of drug efflux pumps, (ii) enhanced drug detoxification, (iii) rapid DNA repair efficiency, (iv) defects in apoptosis regulation, and (v) active cell survival signals. Several adverse effects associated with multidrug resistance and the need for safe multi-targeted anticancer drugs instigated the use of the phytochemical, curcumin, the yellow pigment of the spice turmeric, which has pleotropic activities. We performed a structured literature review using PubMed and Medline searches with secondary review of cited publications, identifying studies on the role of curcumin in conquering drug resistance in cancer. This review describes how curcumin sensitizes cancer cells through regulation of multiple multidrug resistance pathways, thus employing one drug for multiple targets. Curcumin helps the cancer cells to regain their 'forgotten' apoptosis, modulates drug-target interaction at different levels, restrains survival pathways when their proteins are overexpressed, and finds an alternate way to carry forward the process of sensitization of different resistant tumours. Additionally, the review dissects the role of

curcumin, if any, in targeting the major culprit of drug resistance, cancer stem cells (CSC), thereby circumventing resistance. Taken together, this review strongly suggests that curcumin is a promising chemosensitizing agent and that the unique properties of curcumin may be exploited for successful management of resistant tumours.

Complete eradication of cancer is the utmost challenge of medicine. The mainstream treatment modalities, chemotherapy and radiotherapy, have experienced setbacks in the hard-fought battle against cancer, with multidrug resistance (MDR) being the greatest hurdle. MDR is defined as resistance of tumour cells to the cytostatic or cytotoxic actions of multiple, structurally dissimilar and functionally divergent chemotherapeutic drugs. MDR is termed 'intrinsic' when the disease is refractory to chemotherapy from the outset, or 'acquired' when the disease becomes insensitive to treatment upon relapse, or arises during the course of treatment. Intrinsic/inherent drug resistance is the rule for melanoma, and lung and pancreatic cancer (1). On the other hand, acquired/adaptive drug resistance is common in relapsing leukemia, and ovarian and breast carcinoma, and seems to involve mechanisms similar to those of intrinsic resistance (2).

MDR has been correlated to the presence of a myriad of defence mechanisms in cancer cells that elude toxic therapy-induced damage. MDR mechanisms can be classified into two general classes: those that impair delivery of anticancer drugs to tumour cells and nullify their cytotoxic effect, and those that arise in the cancer cell itself due to genetic and epigenetic alterations that affect drug sensitivity. Impaired drug delivery can result from the overexpression of drug efflux transporters, rapid drug inactivation/metabolism, increased DNA damage repair or increased excretion resulting in lower levels of drug in the blood and reduced diffusion of drugs from the blood into the tumour mass (3). The other mechanisms of resistance include insensitivity to drug-induced apoptosis due to blockade of death pathways and overexpression of survival pathways. The relative ratio of pro- to anti-apoptotic members or death to survival signals, determines the threshold for

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apoptosis induction in cancer cells post chemotherapy. Deregulation in one or both in pathways is deeply related to drug sensitivity and resistance (4).

MDR mechanisms act simultaneously or in concert, which makes drug resistance in cancer a formidable task to tackle. It is thus evident that intensive laboratory and clinical studies aimed at overcoming drug resistance in cancer have, so far, produced only limited success (5). Still, in many cases, no remedy has been found to overcome these drug resistance mechanisms and improve clinical outcome in resistant cancer, as most current anticancer therapies involve the modulation of a single target (6). In addition, these chemotherapeutic drugs exert concurrent toxic manifestations including oxidative stress (7-8), liver damage (7-10) and immunosuppression (11-13), in the tumour-bearer. The limitations in efficacy, lack of safety, and high cost associated with available mono-targeted therapies underscore the need for novel agents with multitargeted profiles, improved efficacy and safety. Many plant-based products, however, accomplish multitargeting naturally and, in addition, are inexpensive and safe compared to synthetic agents. Among various naturally occurring phytochemicals, curcumin is capturing the attention of cancer investigators worldwide because of its chemopreventive properties against different malignancies (11, 14).

In the present review we discuss the diverse mechanisms employed by tumour cells to evade the cytotoxic damage manifested by chemotherapeutic drugs and we provide an overview of the multitargeted action of curcumin to assert chemosensitivity in tumours with inherent or acquired resistance. The purpose of the current article is to present an appraisal of the current level of knowledge regarding the potential of curcumin as a promising chemosensitizing agent.

MDR: Major Hurdle in Designing Death

Inherent or innate drug resistance. Some types of cancer are naturally resistant to chemotherapy due to genetic and epigenetic alterations and therefore resist therapies from the first exposure to chemotherapy. This is explained by the Goldie-Coldman hypothesis which states that one in one million cancer cells is inherently resistant to a given chemotherapeutic drug or class of drugs (15). This hypothesis is based on an understanding of tumour cell population kinetics and rates of mutations inherent in mammalian cells. Damage to DNA is a constantly occurring fact of life for all cells and even DNA repair is not faultless. Furthermore, although DNA polymerase is a highly efficient enzyme, base mis-incorporation occurs at a predictable rate of 10^6 bases. Since these sources of mutation are inherent in living cells, there is an inherent mutation rate (16). One consequence of this mutation rate is the development of resistance to drugs before any exposure to the drugs takes place. This resistance is thus a form of inherent drug resistance.

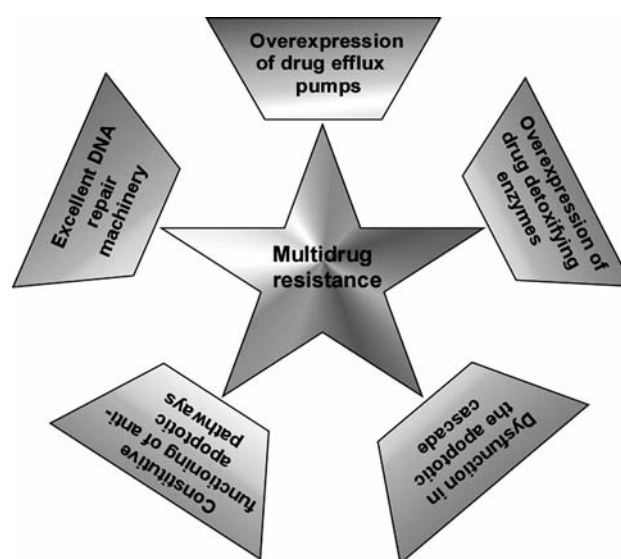


Figure 1. Schematic representation of mechanisms involved in multidrug resistance.

Acquired or adaptive drug resistance. In addition to this inherent drug resistance, drug resistance can be acquired by cellular or organ contact with sub lethal concentrations of certain chemicals, pollutants, and drugs. Some types of cancer respond differentially under the selective pressure of toxic therapy. Chemotherapy kills drug-sensitive cells, but leaves behind a higher proportion of drug-resistant cells. As the tumour begins to grow again, drug-resistant variants of the tumour or its descendants arise, which may result in cross-resistance to the toxic drugs we administer as therapy for cancer (17). These chemicals in fact result in an induced or acquired drug resistance in the organism.

The Canvas of Drug Resistance

It is clear that intrinsic and acquired resistance to chemotherapy critically limit the outcome of cancer treatments. In any population of cancer cells that is exposed to chemotherapy, more than one mechanism of MDR can be present. This phenomenon has been called multifactorial MDR (Figure 1). Clear understanding of the molecular basis of MDR and development of clinical agents or strategies to prevent the occurrence of resistance, or treat resistant tumours are, therefore, of high priority.

We discuss several drug resistance mechanisms, under the following headings: (i) drug efflux and redistribution, (ii) drug detoxification, (iii) DNA repair efficiency, (iv) defects in apoptosis regulation and (v) cellular survival signals.

Drug efflux and redistribution. Multifactorial in aetiology, classic MDR is associated with the overexpression of a

Table I. Drug efflux pumps.

Drug efflux pump		Substrate	Chemotherapeutic drugs
Official HUGO name	Alias/previous name (s)		
ABC B1	P-gp, MDR1	Phospholipids, neutral and cation organic compounds	Vinca alkaloids, taxanes, anthracyclines
ABC C1	MRP1	Organic anions, glutathione, leukotrienes	Vinca alkaloids, anthracyclines, methotrexate
ABC G2	BCRP, ABCP	Prazosin	Anthracyclines, mitoxantrone

specific group of broad substrate spectrum membrane efflux pumps, known as ATP-binding cassette (ABC) transporters, capable of actively transporting diverse chemotherapeutic agents out of the cells against the concentration gradient (18). Resistance arises because increased drug efflux lowers intracellular drug concentrations. So far, 48 human ABC genes have been identified and divided into seven distinct subfamilies (*ABCA–ABCG*) on the basis of their sequence homology and domain organization, a handful of them are likely to be involved in drug disposition and in MDR (19). Among them, we focus on the three major ABC pumps (i) members of ABCB (*ABCB1/P-glycoprotein*), (ii) ABCC (*ABCC1/MRP1*) and ABCG (*ABCG2/MXR/BCRP*) and their role in exerting the resistance phenotype in detail.

P-glycoprotein (P-gp; subfamily B, member 1). *ABCB1*, a protein of 170 kDa is the first discovered multidrug transporter, encoded by the *ABCB1* gene (previously *MDR1*), located on chromosome 7 in humans (20). Overexpression due to gene amplification, gene polymorphism or transcriptional modulation of P-gp in tumour cells manifests acquisition of the MDR phenotype (21). Tumours originating from tissues with naturally high levels of P-gp expression may be intrinsically drug-resistant (*e.g.* colon, kidney, pancreas, and liver carcinoma) (22, 23). On the other hand, tumours with low basic levels of P-gp expression (such as hematological malignancies) sometimes display a marked increase after chemotherapy, and this phenomenon is associated with acquired resistance (24). P-gp efficiently removes cytotoxic drugs and many commonly used pharmaceuticals from the lipid bilayer (Table I). By extruding cytotoxic drugs out of the cells before they reach their cellular target, P-gp expression leads to failure of cancer chemotherapy. P-gp expression is correlated with a reduced complete remission rate, and a higher incidence of refractory disease in patients suffering from acute myelogenous leukaemia (AML) (25).

Multidrug resistance-associated protein (ABCC1): This is a 190-kDa protein, product of the *ABCC1* (previously *MRP1*) gene, located on chromosome 16 (26). Tumours of lung, testis, kidney, and peripheral blood mononuclear cells demonstrate overexpression of *ABCC1* (27). *ABCC1* functions as a multi specific organic anion transporter, with (oxidized) glutathione (GSSG), cysteinyl leukotrienes,

glucuronides, sulfate conjugates of steroid hormones, bile salts, and activated aflatoxin B1 as substrates. It pumps out cytotoxic drugs (such as vincristine) and other hydrophobic compounds in the presence of glutathione (Table I) (28).

Breast cancer resistance protein. *ABCG2/BCRP* is a 95 kDa protein encoded by the *ABCG2* (previously *BCRP*) gene, located on chromosome 4 in humans. Unlike P-gp and *ABCC1* it contains only one transmembrane domain and is thus also called a half transporter (29). Its substrate spectrum and mechanism of transport are quite similar to those of P-gp. *ABCG2* was discovered in MDR cancer cells, with the identification of chemotherapeutic agents, such as mitoxantrone, flavopiridol, methotrexate and irinotecan as substrates (Table I). Later, drugs from other therapeutic groups were also described as substrates, including antibiotics, antivirals, 3-hydroxy-3-methyl glutaryl Coenzyme A (HMG-CoA) reductase inhibitors and flavonoids. Single nucleotide polymorphisms of the gene were shown to alter either the plasma concentrations of substrate drugs or the levels of resistance to chemotherapeutic agents in cell lines (30).

Redistribution of drug. In addition to enhanced drug efflux by virtue of different drug transporters associated with tumour cells, a second mechanism employed by cancer cells to reduce intracellular drug load, can be the redistribution of a drug away from the target. In this case, the total concentration of drug might not be reduced, but the intracellular distribution may be altered, thereby reducing the drug concentration at the site of action (31). These changes in drug distribution are most notable for DNA-interacting drugs, where in drug-resistant cells; the drug is redistributed from the nucleus to the cytoplasm. This redistribution of drug is controlled by an 110-kDa non-ABC drug transporter vault protein, referred to as lung resistance-related protein (LRP) (32). This transporter is not associated with the cytoplasmic membrane, but operates by controlling drug transport from the nucleus to the cytoplasm *via* vaults and its drug substrate spectrum is similar to that of P-gp (Table I) (33). There are reports demonstrating that vaults are overexpressed in MDR cancer cell lines and that LRP expression predicts for drug resistance and poor outcome in patients with acute myelogenous leukemia, ovarian cancer, and possibly other types of cancer (34, 35).

Drug detoxification/inactivation. The toxic effect of cytotoxic drugs that gain entry inside the cells is successfully reduced with drug-metabolizing enzymes, generally overexpressed in resistant cancer. Both oxidation (cytochrome P-450, phase I enzyme) and conjugation (glutathione-S-hydrolases (GSH)/glutathione-S-transferases (GST), aldehyde dehydrogenases-related, phase II) enzymes play critical roles in protecting cells against many drugs (36). The cytochrome P450 enzymes are a superfamily of haemoproteins, known to be involved in the metabolic activation and detoxification of a number of anticancer drugs (37). In particular, enzymes of the CYP3A subfamily play a role in the metabolism of many anticancer drugs, including epipodophyllotoxins, ifosfamide, tamoxifen, taxol and vinca alkaloids (38). CYP3A4 has been shown to catalyse the activation of the pro-drug ifosfamide, raising the possibility that ifosfamide could be activated in tumour tissues containing this enzyme (39). Whereas in some cancer cell lines, resistance to anticancer drugs, such as mitomycin C, doxorubicin, tamoxifen, cyclophosphamide and their derivatives, is indicated by a high activity of GST and a low activity of P450 in general (40). However, the mechanism of change of these enzyme activities is complicated and different for each drug. On the other hand, reduced glutathione produces species that are usually less toxic and more hydrophilic than the original electrophilic compounds that can be partially metabolized and excreted (41). In cells with acquired resistance to antineoplastic agents, both GSH content and GST activity are frequently elevated, which results in protection of the cells from such agents (42). Moreover, drug-metabolizing enzymes also play an important role in reducing the intracellular concentration of drugs.

DNA damage repair efficiency. An important mechanism that underlies the development of chemotherapeutic resistance is that of cancer cells recognizing DNA lesions, induced by DNA-damaging agents and by ionizing radiation, and repairing these lesions by activating either homology-directed (HR) or non-homologous DNA repair pathways (43). Chemotherapeutic agents in common use, including alkylating agents (cisplatin, carboplatin, and nitrogen mustards, such as melphalan), inhibitors of DNA topoisomerase II (including the anthracyclines, etoposide, and teniposide), and inhibitors of topoisomerase I and antimetabolites are known to be or likely to induce DNA double-strand breaks (DSBs). HR repairs DSBs, removing damage in an error-free process by the radiation sensitive 52 (RAD52) epistasis group of proteins, including replication protein A (RPA), RAD52, RAD54, several RAD51-related accessory proteins, breast cancer 1, early onset (BRCA1), and breast cancer 2, early onset (BRCA2) (44). On the other hand the Ku heterodimer (Ku70 and Ku86), DNA-dependent protein kinase (DNA-PKs) and DNA ligase IV and X-ray repair complementing defective repair in Chinese hamster cells 4 (XRCC4) are involved in the

non-homologous DNA repair pathway (45). Two kinases from the phosphatidylinositol 3-kinase (PI3K)-related protein kinase (PIKKs) family, Ataxia telangiectasia mutated (ATM) and Ataxia telangiectasia and Rad3 related (ATR), are central to cellular responses to DSBs. When activated, ATM and ATR phosphorylate a multitude of proteins, which initiate a cascade that induces cell-cycle arrest and facilitates DNA repair (46). Thus these DNA repair pathways in combination with different cell cycle check point regulators [ATM, ATR, Csk homologous kinase (CHK1) and (Chk2)] provide extra time for cancer cells to repair 'blueprint' damage induced by toxic therapy (47). Thus, therapeutic strategies aimed at inhibiting either one or both DSB repair pathways or at abrogating cell-cycle checkpoints is an appealing strategy to sensitize chemoresistant tumours.

Influence of apoptosis-related genes. Every cell in a multicellular organism has an intrinsic mechanism of self-destruction called programmed cell death or apoptosis, but tumour cells often have faulty apoptotic pathways. These defects not only increase tumour mass, but also render the tumour resistant to therapy. Programmed cell death is executed by a family of cysteine aspartyl-specific proteases (48). In principle, there are two major apoptotic pathways; (i) the death receptor (extrinsic) pathway, and (ii) the mitochondria/cytochrome c-mediated (intrinsic) pathway (Figure 2).

Transmembrane 'extrinsic' pathway. This is activated by the ligation of death receptors [CD95, Tumour necrosis factor (TNF) and TNF-related apoptosis-inducing ligand (TRAIL)] to activate caspase-8 and -10, which in turn cleave and activate executioner caspases such as caspase-3 and -7 (49). In many cell types, death receptor-mediated apoptotic signalling induces a mitochondrial death amplification loop *via* proteolytic activation of the BH3-only protein (BID). The extrinsic pathway is regulated by FADD-like interleukin-1-converting enzyme-like protease (FLICE/caspase-8)-inhibitory proteins (c-FLIP) (50) and inhibitor of apoptosis proteins (IAPs) which can hinder the functions of both activator and effector caspases (51). Overexpression of FLIP in some types of cancer prevents apoptosis induced by some chemotherapeutic drugs (52). Furthermore, the master regulator, p53 can also regulate both CD95 and TRAIL receptor 2 (TRAIL-R2/DR5) (53). In fact, loss of CD95L or TRAIL function can promote drug resistance and metastasis in tumour (54, 55)

Mitochondrial 'intrinsic' pathway. This is triggered in response to DNA damage, hypoxia and survival factor deprivation, it is associated with mitochondrial depolarization and release of cytochrome c from the mitochondrial inter-membrane space into the cytoplasm. Cytochrome c, apoptotic protease-activating factor 1 (APAF-1) and procaspase-9 then form a complex termed the apoptosome, which activates

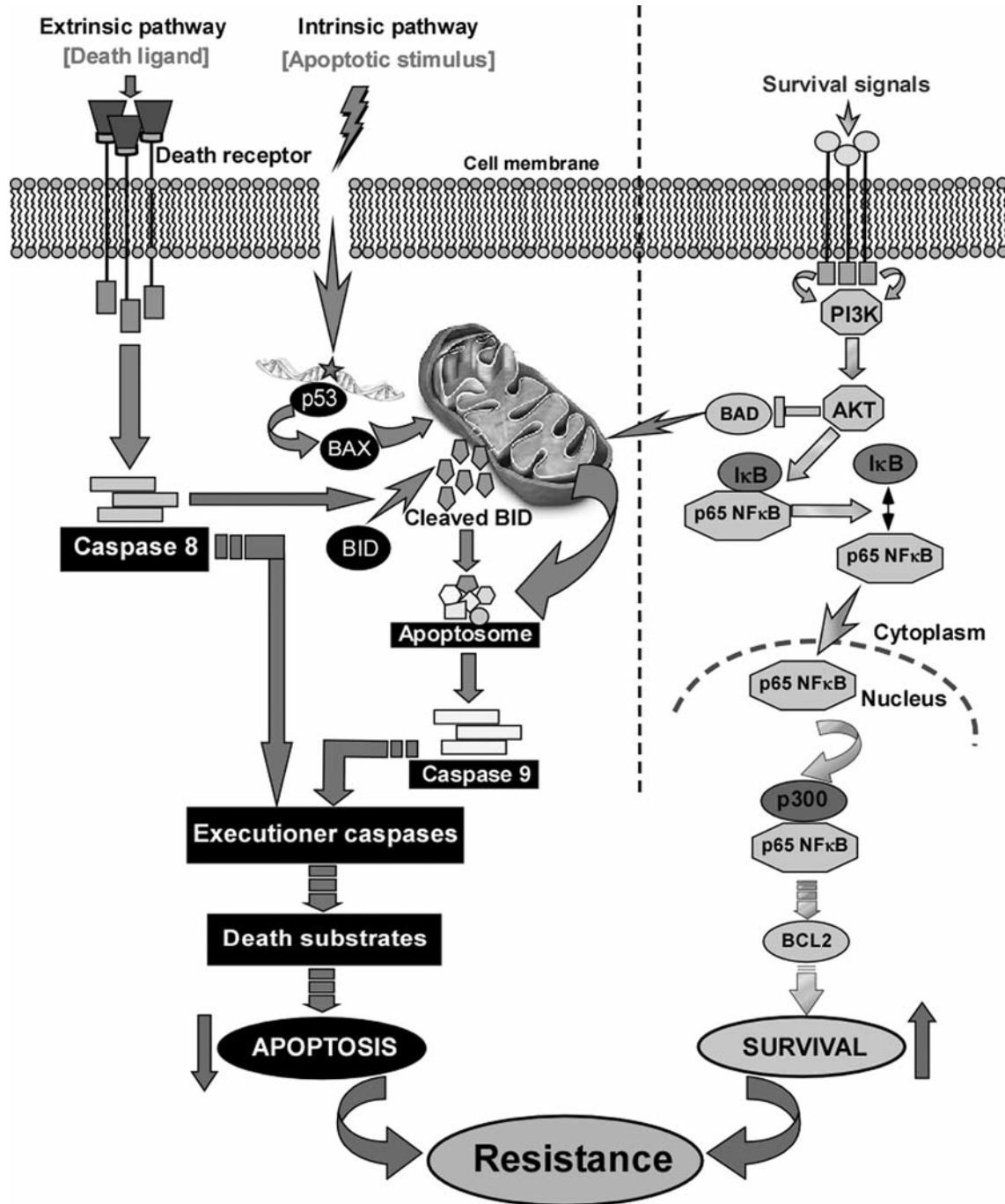


Figure 2. Deregulation of apoptotic survival signals circumvents drug resistance. Schematic illustration of two major apoptotic signalling pathways, transmembrane and mitochondrial down-regulated and protein kinase B (AKT/PKB)-+nuclear factor kappa B (NF-κB) survival signalling pathways, generally overexpressed in most resistant tumours.

caspase-9 and promotes activation of effector caspases (56). The mitochondrial pathway is a critical death pathway common to many different types of chemotherapies. Aberrations in the regulation of this pathway can result in resistance to chemotherapy. As a sensor of cellular stress, p53

is the critical initiator of this pathway (57). p53 can initiate apoptosis by transcriptionally activating pro-apoptotic B-cell lymphoma 2 (BCL2) family members [e.g. BCL2-associated X (BAX), BCL2 homologous antagonist (BAK), p53 up-regulated modulator of apoptosis (PUMA), and phorbol

12-myristate 13-acetate induced protein 1 (NOXA)] and by repressing anti-apoptotic BCL2 proteins (BCL2, BCL-XL) and IAPs (survivin) (58, 59). However, p53 can also transactivate other genes that may contribute to apoptosis including Phosphatase and tensin homolog (*PTEN*), apoptotic protease-activating factor 1 (*APAF1*), p53 apoptosis effector related to PMP-22 (*PERP*), p53-regulated apoptosis-inducing protein 1 (*p53AIP1*), and genes that lead to increases in reactive oxygen species (*ROS*) (60-62). Indeed the gene encoding this master regulator, *TP53*, is frequently mutated and silenced in many tumours which lack apoptosis (62). Reports state that specific mutations in *TP53* have been linked to primary resistance to doxorubicin treatment and early relapse in patients with breast cancer (63). Lymphomas from *TP53*-knockout mice were highly invasive, displayed apoptotic defects and were markedly resistant to chemotherapy *in vitro* and *in vivo* (64). Furthermore, in about 70% of breast cancer cases, wild-type *TP53* is expressed but fails to suppress tumour growth (65). This might be explained by functional mutations or altered expression of *p53* downstream effectors (*PTEN*, *BAX*, *BAK*, and *APAF1*), or upstream regulators [*ATM*, *CHK2*, murine double minute 2 (*MDM2*), and *p19^{ARF}*] (64). The members of the BCL2 family are an important class of regulatory proteins. Over-expression of anti-apoptotic members of the family, *e.g.* BCL2 and BCL-XL, is associated with resistance to various cytotoxic agents and radiotherapy, thereby making them obvious anticancer drug targets (66). In fact, cancer cell resistance to chemotherapeutic agents strongly correlates with the expression levels of BCL-XL.

The information concerning key apoptotic proteins, their regulation, and the manner in which they are altered in MDR tumours can be used for target selection in designing new anticancer agents aiming to restore apoptotic potential through genetic or pharmacological methods.

Survival pathways involved in resistance. Cell survival in the face of cytotoxic therapy is dictated by the intricate balance between the pro-apoptotic and anti-apoptotic signals. The survival signals can mitigate or abrogate the effectiveness of cancer therapy and protect against other cellular insults. Survival signals include growth factors, cytokines, hormones and other stimuli, such as signals initiated by adhesion molecules (Figure 2) (67). Mutations in these pathways are frequent in human cancer, demonstrating their dual role in sustaining tumour growth (carcinogenic potential) and in producing resistance to chemotherapy. Understanding these alterations offers novel opportunities for developing therapeutic strategies against cancer.

Protein Kinase B (AKT/PKB)-mediated resistance to chemotherapy: AKT is a serine/threonine kinase that plays an important role in survival when cells are exposed to different apoptotic stimuli. Recent studies show that aberrant activation of AKT in cancer cells is associated with a poor prognosis and

resistance to chemotherapy and radiotherapy (67). AKT is activated by phosphatidylinositol 3-phosphate (PIP3), which is produced by PI3Ks. Deregulation of AKT-mediated survival signalling occurs either due to aberrant amplification of AKT, which has been reported in breast, ovarian and pancreatic cancers (68), or due to genetic inactivation, amplification or mutations in epidermal growth factor receptor (69) or serine-threonine kinases or loss of the tumour suppressor *PTEN*, which negatively regulates PI3K signalling. AKT protects cells from apoptosis by phosphorylating and inactivating several key apoptotic molecules: BAD, procaspase-9 and Forkhead protein (FKHR1) (70). AKT also inactivates p53 by phosphorylating MDM2 on Ser166 and Ser186, which promotes p53 degradation (71). Finally, AKT activates nuclear factor kappa B (NFkB) by phosphorylating I kappa B kinase (IKK) on Thr23 (2). Moreover, fibroblasts overexpressing AKT are resistant to staurosporine and etoposide-induced apoptosis (72, 73). *PTEN*-deficient tumour cell lines and tumours derived from *Pten*-knockout mice displaying elevated AKT activity are resistant to apoptotic-inducing stimuli (74). Inactivation of AKT enhances the activity of doxorubicin (75) and etoposide (76). AKT and PI3K inhibitors are rational therapeutic strategies for cancers. Moreover, combination of specific PI3K or AKT inhibitors with existing chemotherapeutic drugs may act synergistically. There are presently no specific inhibitors of the PI3K/AKT survival pathways used clinically (2).

Nuclear factor kappa B (NFkB) and chemoresistance. An important factor influencing apoptosis of tumour cells is the transcription factor NF-κB. Normally, NF-κB remains sequestered in an inactive state by the cytoplasmic inhibitor of NF-κB (IκB) proteins. However, a variety of external stimuli including cytokines, pathogens, stress and chemotherapeutic agents can lead to activation of NF-κB by phosphorylation, ubiquitylation, and the subsequent degradation of IκB (77). Depending on the stimulus and the cellular context, NF-κB can activate pro-apoptotic genes, such as those encoding CD95, CD95L and TRAIL receptors, anti-apoptotic genes, for example, those encoding IAPs and BCL-XL (65) and enhanced expression of the MDR gene product (78, 79). NF-κB can also prevent programmed necrosis by inducing genes that encode antioxidant proteins. As tumour cells often use NF-κB to achieve resistance to anticancer drugs, activation of the NF-κB pathway renders many types of tumour cell more resistant to chemotherapy, presumably *via* induction of anti-apoptotic proteins. Inhibition of NF-κB activation seems to be a promising option to improve the efficacy of conventional anticancer therapies (80). Constitutive nuclear NF-κB activity has been described in many human multiple myeloma cell lines and primary myeloma cells (81). Furthermore, multiple myeloma cells have been shown to be sensitive to growth inhibition and induction of apoptosis upon treatment with various inhibitors

of NF- κ B signalling (82). Constitutive overexpression of NF- κ B signalling pathway is also observed in a number of solid tumours, such as breast, cervical, prostate, renal, lung, colon, liver, pancreatic, oesophageal, gastric, laryngeal, thyroid, parathyroid, bladder and ovarian cancer (83). NF- κ B activation also plays an anti-apoptotic role in human leukemia K562 cells exposed to ionizing radiation (84). Transient inhibition of NF κ B through adenovirus-mediated expression of a degradation-resistant mutant of I κ B can overcome chemoresistance mediated by camptothecin (85) and gemcitabine (86). Disulfiram-mediated inhibition of NF κ B activity enhanced the activity of 5-fluorouracil in human colorectal cancer cell lines (87). The anti-inflammatory drug sulfasalazine sensitizes pancreatic carcinoma cells to etoposide by inhibition of NF- κ B (88). These observations indicate that NF- κ B plays an important role in chemoresistance (85) and establishes the inhibition of NF- κ B as a new adjuvant approach in chemotherapy.

Phytochemicals in Redesigning the Landscape of Drug Resistance

With most traditional anticancer therapies at stake, use of non-toxic natural or synthetic chemicals to intervene in multistage carcinogenesis has emerged as a promising and pragmatic medical approach to reduce the risk of cancer. Phytochemicals are components in the plants ('phyto' is from the Greek word meaning plant) that possess substantial anticarcinogenic and antimutagenic properties (89). There is growing evidence that populations with greater reliance on fruits and vegetables in their diet experience a reduced risk for the major types of cancer (90). The anticancer properties of phytochemicals can be attributed to their strong antioxidant properties. The National Cancer Institute (US) (NCI) has identified about 35 plant-based foods that possess cancer-preventive properties. These include curcumin from turmeric, β -carotene from carrot, epigallocatechin gallate (ECGC) from green tea, theaflavins from black tea, genistein from soybeans, resveratrol from grapes, gingerol from ginger, and capsaicin from chilli (91). ECGC and theaflavins are reported to induce cancer cell apoptosis (92-99). Capsaicin is known to induce apoptosis in several tumour models (100, 101). In addition, a number of natural dietary phytochemicals, including curcumin, quercetin, xanthorrhizol, ginger and genistein, are candidates for inducing chemo/radiosensitization of cancer cells (102-103).

Among these phytochemicals, curcumin has been identified as one of the major natural anticancer agents exerting antineoplastic activity in various types of cancer cells (104-108). Curcumin exerts a minimal effect on normal cells of the body and also protects the immune system from cancer-induced immunosuppression (99, 104, 109-110). Our laboratory has shown that curcumin reversibly arrests non-malignant cells in the G₀ phase but does not induce apoptosis

in them (107). Curcumin and turmeric products have been characterized as safe by health authorities such as the Food and Drug Administration (FDA) in the United States of America, and the Food and Agriculture Organization/World Health Organization (FAO/WHO) (111).

Curcumin Architects the Chemosensitization Programme

Curcumin is a polyphenol derived from the rhizomes of tumeric, *Curcuma longa*, and has been used through the ages as a 'herbal aspirin' and 'herbal cortisone' in Ayurvedic medicine, an ancient Indian healing system that dates back over 5,000 years. *C. longa* is a short-stemmed perennial which grows to about 100 cm in height. It has curved leaves and oblong, ovate or cylindrical rhizomes (Figure 3). Curcumin is the major biologically active compound of turmeric; chemically it is known as diferuloylmethane (C₂₁H₂₀O₆). It has received considerable attention due to its beneficial chemopreventive and chemotherapeutic activity *via* influencing multiple signalling pathways, including those involved in survival, growth, metastasis, angiogenesis and immunopotentiating effects in various types of cancer (11, 14, 112-116) (Figure 3). Furthermore, Ramachandran *et al.* (117) demonstrated by a microarray study that out of the 214 apoptosis-associated genes in the array, the expression of 104 genes was altered by curcumin treatment. These results show that curcumin induces apoptosis by regulating multiple signalling pathways in cancer cells.

In pilot clinical studies in India, Taiwan, USA and UK, curcumin has been associated with regression of pre-malignant lesions of the bladder, soft palate, gastrointestinal tract, cervix, and skin, and with treatment responses in established malignancy (118-120). Doses up to 8-10 g can be administered daily to patients with pre-malignant lesions for three months without overt toxicity. Anecdotal reports suggest that dietary consumption of curcumin of up to 150 mg/day is not associated with any adverse effects in humans (105). All this information not only suggests that curcumin has enormous potential in the prevention and therapy of cancer, but also well-justifies the utility of using curcumin as an antitumour agent.

Signaling pathways implicated in curcumin-induced MDR reversal. In this part of the review, we present curcumin as a significant chemosensitizer in cancer chemotherapy and highlight the signalling networks modulated by curcumin in inducing MDR reversal.

Targeting ATP-driven ABC drug-efflux pumps. As described in the first section of this review, overexpression of drug efflux pumps plays a major role in the development of MDR. Notably, several functional inhibitors of MDR proteins (verapamil, cyclosporine A, tamoxifen, dexverapamil, valspodar and

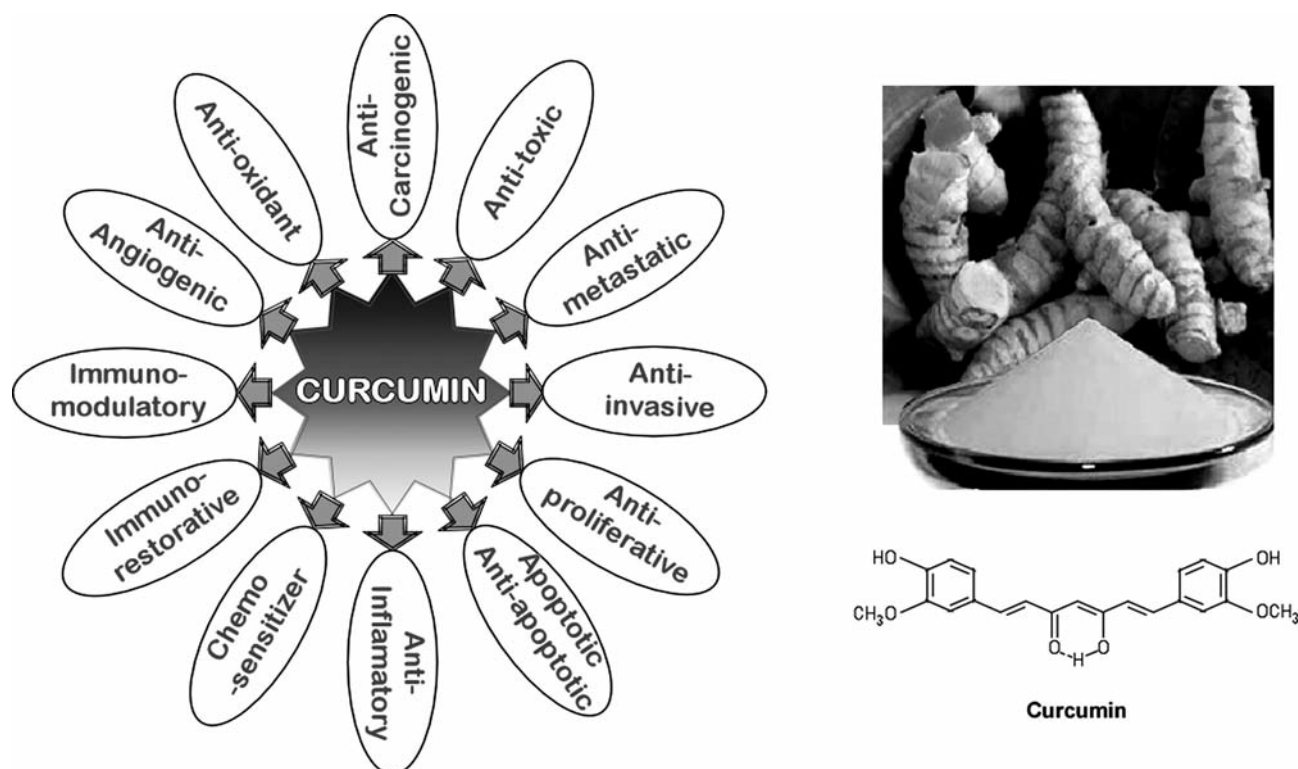


Figure 3. Pictorial depiction of *Curcuma longa* and schematic representation of chemopreventive properties of curcumin in curtailing tumour proliferation and progression via influencing multiple signalling pathways, including those involved in survival, growth, metastasis, angiogenesis and immunopotentiating effects.

biricodar) have been tested, but thus far, none has been clinically successful due to the dose-limiting toxic effect of the modulators (121). On the other hand, curcumin has been reported to reverse the drug resistance phenotype in cancer cells overexpressing ABC transporters, namely ABCB1, ABCG2, and ABCC1 (122-124), without inducing systemic toxicity. Since wild-type p53 represses the expression of different drug transporters (ABCC1), curcumin, by virtue of activating p53, might also contribute indirectly in decreasing the MDR of cells. Curcumin blocked the efflux of fluorescent substrates calcein AM, rhodamine 123, and bodipy-FL-vinblastine in MDR cervical carcinoma cell lines overexpressing ABCB1 (125) and the efflux of mitoxantrone and pheophorbide A mediated by ABCG2 in HEK293 cells (123, 126). Another report stated that sensitivity to vinblastine of cells treated with non-toxic doses of curcumin increased only in the P-gp- overexpressing drug-resistant cell line, KB-V1 (127).

Curcumin 'Handcuffs' drug-detoxification machinery. We earlier elaborated the role of endogenous GSH, GST and P450 in cancer drug resistance. Khar *et al.* demonstrated that curcumin induced apoptosis in cancer cells by depleting the levels of glutathione, which increased the generation of ROS (128). Interestingly, curcumin had no effect on normal rat

hepatocytes, which showed no superoxide generation and therefore no cell death. Curcumin has also been shown to directly quench ROS and scavenge superoxide anion radicals and hydroxyl radicals, creating an environment favourable for toxic therapy (129). In K-562 cells curcumin also reduced the levels of GST, which is implicated in the resistance of cancer cells to conventional chemotherapy (116). Curcumin inhibits the phase I enzyme system consisting of cytochrome P450 isoforms, the P450 reductase, the cytochrome b5 and the epoxide hydrolase and which protects from the toxic effects of chemicals and carcinogens (127). On the other hand, curcumin induces phase II enzymes (GST and epoxide hydrolase), which play a protective role by eliminating toxic substances and oxidants and conferring benefit in the prevention of the early stages of carcinogenesis (130).

Curcumin 'Halts' drug-induced DNA repair. As discoursed in detail earlier, abrogation of DNA repair pathways or cell cycle check points widens the therapeutic index of conventional therapies. But small-molecule chemical inhibitor of CHK1/CHK2 XI844 (131), and ATM inhibitor KU55933 (82), in pre-clinical studies manifested potential toxicity towards normal cells, whereas curcumin preferentially induced DNA damage in triple-negative breast cancer cells, sparing

normal cells. A relatively recent article by Rowe *et al.* described that curcumin induced DNA damage in triple-negative breast cancer cells and regulated BRCA1 protein expression and modification (132). Curcumin-induced DNA damage was associated with phosphorylation, increased expression, and cytoplasmic retention of the BRCA1 protein. Interestingly, apoptosis and BRCA1 modulation were not observed in non-transformed mammary epithelial cells, suggesting some breast cancer cells have intrinsic defects that make them more sensitive to curcumin. Lu *et al.* discovered that curcumin induced DNA damage in a mouse-rat hybrid retina ganglion cell line (133). Real-time PCR analysis showed that curcumin reduced expression of DNA damage-response genes, including *ATM*, *ATR*, *BRCA1*, *14-3-3 σ* , *DNA-PK* and O-6 Methyl guanine-DNA methyl transferase (MGMT). Therefore, reduction of DNA damage response may be the reason for curcumin-induced growth inhibition (134). Another study focussed on the combination therapy, where curcumin or cyclophosphamide (CTX) alone failed to induce apoptosis in HT/CTX cells, whereas curcumin with CTX increased apoptosis and reversed MDR of HT/CTX cells, effectively by targeting the BRCA1-DNA repair pathway (135). Curcumin exposure of resistant glioma cells sensitized them to cytotoxic drugs, effects associated with reduced activity of DNA repair enzymes, MGMT, DNA-PK, Ku70, Ku80, and excision repair cross-complementation group 1 (ERCC-1) (136).

These findings suggest that curcumin, by regulating different aspects of anticancer drug delivery to tumour cells and by nullifying their cytotoxic effects, may be beneficial in the chemoprevention of different types of cancer (Figure 4).

Curcumin as apoptosis inducer

Modulating multiple-signals to create a pro-apoptotic milieu. In this section we highlight the potential role of curcumin as an inducer of pro-apoptotic signalling network in combating drug resistance.

Extrinsic apoptotic pathway—stimulating TNFR death signals. Loss of TRAIL function is associated with drug resistance in tumour cells. Several groups have shown that curcumin is able to sensitize cancer cells to TRAIL-induced apoptosis. Wahl *et al.* pointed out that curcumin enhances TRAIL-induced apoptosis in chemoresistant ovarian cancer cells by activating both the extrinsic and the intrinsic apoptotic pathways (137). Similar results were found in LNCaP prostate cancer cells (138). Deeb *et al.* showed that in the prostate cancer cells LNCaP, DU145 and PC3, curcumin is able to block the phosphorylation of I κ B α which leads to the inhibition of the constitutively active NF- κ B and the subsequent enhancement of the sensitivity of prostate cancer cells to TRAIL (138-140). Consistently in a study conducted by Gao *et al.* curcumin and TRAIL cooperatively interacted to promote death of human U87 glioma cells (141).

At low concentrations (curcumin and TRAIL), neither of the two agents alone produced significant cytotoxicity in U87 cells, as measured by 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) dye reduction assay. On the other hand, cell death was markedly enhanced if tumour cells were treated with curcumin and TRAIL together (141). Shankar *et al.* demonstrated that curcumin sensitizes TRAIL-resistant LNCaP xenografts *in vivo* to undergo apoptosis by TRAIL (142). Curcumin can also down-regulate the expression of various pro-inflammatory cytokines including TNF, Interleukin (IL-1), IL-2, IL-6, IL-8, IL-12, and chemokines, most likely through inactivation of the NF κ B (115). Studies from our laboratory showed that curcumin neutralized tumour-induced oxidative stress, restored NF- κ B activity, and inhibited TNF- α production, thereby minimizing tumour-induced T-cell apoptosis (109). As curcumin can activate the extrinsic apoptotic pathways and protect T cells from tumour induced effects, it may circumvent chemoresistance to conventional chemotherapeutic agents.

Intrinsic apoptotic pathway—awakening the p53 death network. Aberrations in the regulation of the intrinsic pathway, governed mainly by the p53 network, result in resistance to chemotherapy. Curcumin by perturbing expressions of p53-regulated BCL2 and IAP family members, sensitized glioma cells to several clinically utilized chemotherapeutic agents (cisplatin, etoposide, camptothecin, and doxorubicin) and radiation (136). Curcumin pre-treatment considerably reduced the dose of cisplatin and of radiation required to inhibit the growth of cisplatin-resistant ovarian cancer cells. During the 6-h pre-treatment, curcumin down-regulated the expression of BCL-XL and myeloid cell leukemia-1 (MCL-1) pro-survival proteins. Another study exhibited that curcumin pre-treatment followed by exposure to low doses of cisplatin increased apoptosis by increasing BAX expression while reducing the expression of BCL and BCL-XL, followed by activation of caspase-9 and caspase-3 (143). A report from our laboratory established the relationship between p53 status, p21 induction, BCL2/BAX ratio, cell cycle deregulation and apoptosis in curcumin-treated tumour cells (105). Findings from our laboratory have also revealed that curcumin can induce breast cancer cell apoptosis in a p53-mediated BAX transactivation-dependent manner through loss of mitochondrial transmembrane potential, release of cytochrome *c* and activation of the death cascade (104), whereas in p53-null chemoresistant leukemia cells, curcumin remarkably induces apoptosis by inducing p73, a member of the p53 family (106).

This information highlights the fact that curcumin targets multiple effectors, modulators and inducers of death, augmenting transcription factors, to work as tumour suppressors in cancer cells, thereby finally helping the cells to regain what they were lacking – the program of apoptosis (Figure 4).

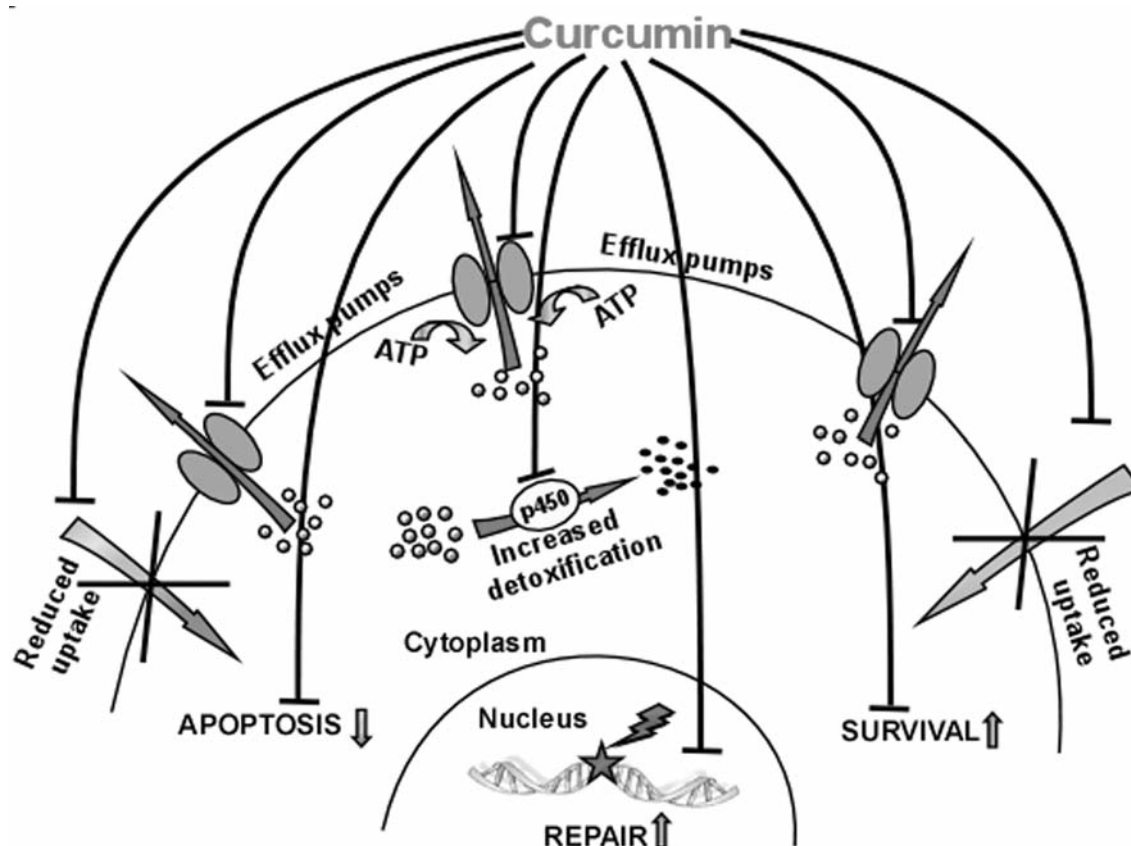


Figure 4. Curcumin, as a potential 'chemosensitizing agent', targets multiple signals for combating drug resistance. Pictorial depiction of modulatory effects of curcumin on drug efflux transporters, drug detoxification machinery, DNA damage repair, apoptotic and survival signals to finally sensitize drug resistant cancer to toxic therapy.

Curcumin as a chemosensitizer

Perturbing survival pathways. In this section we reveal the pivotal role of curcumin in inhibiting multiple survival signals to enhance the efficacy of conventional therapy.

PI3K/AKT pathway. Overexpression of PI3K/AKT-mediated survival pathways via epidermal growth factor receptor (EGFR) kinase activation manifests resistance (144). Reports state that curcumin is a potent inhibitor of EGFR tyrosine kinase. Curcumin inhibits EGF-stimulated phosphorylation of EGFR in breast cells, as well as basal phosphorylation of AKT that may facilitate apoptosis (145). Patel *et al.* revealed that the combination of curcumin with FOLFOX served as a better strategy for inducing apoptosis in folinic acid, fluorouracil, oxaliplatin (FOLFOX)-resistant colorectal cancer (HCT-116 and HT-29) cells (146). These changes were associated with the attenuation of EGFR and insulin-like growth factor 1 receptor (IGF-1R) survival signalling pathways. Furthermore, curcumin inhibits EGFR activation, steroid receptor coactivators (SRC) activity and

inhibits the activity of some nuclear receptors (11). Curcumin reduced cell survival in a p53- and caspase-independent manner, an effect correlated with the inhibition of activator protein 1 (AP-1) and NF- κ B signalling pathways via prevention of constitutive janus kinase (JNK) and AKT activation in chemoresistant human (T98G, U87MG, and T67) and rat (C6) glioma cell lines (136).

NF- κ B pathway: Constitutive activation of NF- κ B in different types of cancer creates an environment conducive for chemotherapeutic resistance. Clinical NF- κ B inhibitors, such as bortezomib (Velcade, formerly known as PS-341), generally involve suppression of the proteasome, leading to severe toxicities (146). Studies have shown that curcumin is able to inhibit NF- κ B activation, which manifested chemosensitivity to drug-resistant cancer cells (148). Several teams confirmed these results and reported that curcumin inhibits IL1 α -, TNF α -, 12-tetradecanoyl-13-phorbol acetate (TPA)-, lipopolysaccharide (LPS)- and thrombin-induced NF- κ B activation. This inhibiting effect should be considered for the improvement chemotherapeutic treatment as most

anticancer drugs induce NF- κ B, leading to the development of drug resistance (119). Curcumin abolishes the induction of NF- κ B binding to the DNA, blocks IKK activation, I κ B α phosphorylation and degradation, as well as NF κ Bp65 translocation (149). Furthermore, curcumin and tamoxifen co-treatment has been shown to synergistically sensitize tamoxifen-resistant breast cancer cells and may be a viable strategy to either prevent tamoxifen-resistant disease or to re-sensitize refractory disease to tamoxifen treatment (150). Curcumin promotes the down-regulation of NF- κ B in multiple myeloma cells (11), which results in activation of caspase-7 and caspase-9, and induces polyadenosine-5'-diphosphate-ribose polymerase cleavage, culminating in apoptosis. Curcumin also potentiated the apoptotic effects of thalidomide and bortezomib in multiple myeloma by down-regulating the constitutive activation of NF κ B and AKT, and this correlated with the suppression of NF κ B-regulated gene products, including cyclin D1, BCL-xL, BCL2, TNF receptor-associated factor (TRAF1), cIAP-1, X-linked inhibitor of apoptosis (XIAP), survivin, and vascular endothelial growth factor (VEGF) (151). Studies by Murali *et al.* shows that a 6-h pre-treatment with curcumin effectively sensitized cisplatin, resistant ovarian cancer cells to the cytotoxic effects of cisplatin, at doses at least ten times lower compared to cisplatin treatment alone (152). The inhibitory effect of curcumin upon cyclooxygenase 2 (COX2) and cyclin D1, mediated through NF- κ B, also restrict tumour cell growth. Induction of G₂/M arrest and inhibition of COX2 activity by curcumin in human bladder cancer cells has also been reported (11). Radiation stimulated NF- κ B activity, whereas curcumin suppressed this radiation-induced NF- κ B activation *via* inhibition of radiation-induced phosphorylation and degradation of I κ B α , and inhibition of AKT phosphorylation. Curcumin also suppressed NF- κ B-regulated gene products (BCL2, BCL-xL, IAP2, COX2, and cyclin D1). In parallel curcumin has been reported to reduce cisplatin-induced testicular toxicity in rat testis (153). This was in agreement with our finding where curcumin ameliorated cisplatin-induced toxicity in peripheral blood mononuclear cells and normal lung epithelial cells (110). NF- κ B inhibition by curcumin is certainly an interesting strategy to overcome drug resistance in cancers with constitutive NF- κ B activation.

All the above reports clearly demonstrate how curcumin modulates survival pathways when overexpressed and finds an alternate way to carry forward the process of sensitization in different resistant tumours (Figure 4).

Curcumin chemosensitizes by tailoring p65NF κ B-p300 cross-talk in favor of p53-p300. Considering the deregulation of NF- κ B and p53 pathways in numerous types of cancer, it is not surprising that an extensive cross-talk between these pathways exists at various levels. In fact, after chemotherapy-induced DNA damage, NF- κ B was shown to play a role in neoplastic transformation by inhibiting p53 gene expression

(154). Moreover, NF- κ B and p53 compete for co-activators, for example, the histone acetyltransferases p300 and CREB-binding protein (CBP) (155). An ideal therapeutic approach should, therefore, involve tailoring this cross-talk in favour of p53 to chemosensitize drug-resistant tumours. A combinatorial therapy that not only shifts the cancer cells from resistance to apoptosis, but also prevents systemic toxicity in the cancer patient, will, therefore, be the ideal candidate for regressing drug-resistant cancer. Recent investigations from our laboratory revealed that curcumin sensitizes drug-resistant breast tumours to doxorubicin by inhibiting the NF- κ B-mediated defence pathway and by activating p53 apoptotic signalling. Inhibition of p65NF κ B by curcumin was found to be both scaffold/matrix associated region 1 (SMAR1)-dependent and -independent. In fact, inactivation of the NF κ B pathway by curcumin rescued p300 from p65NF κ B and launched p53-p300 collaboration to induce p53-dependent BAX, PUMA, and NOXA transactivation, and instigation of downstream mitochondria-dependent death cascade in drug-resistant breast cancer cells. Interestingly for induction of p53-dependent apoptosis, curcumin-mediated execution of promyelocytic leukemia (PML)-SMAR1 cross-talk was indispensable. A simultaneous decrease in drug-induced systemic toxicity might also have enhanced the efficacy of doxorubicin by improving the intrinsic defense machineries of the tumour bearer. Therefore, curcumin in combination with standard chemotherapeutics may serve as a double-edged sword in culminating both resistance and toxicity after chemotherapy (156) (Figure 5).

Cancer Stem Cells: New Colour in Drug-Resistance

Over the last decade, there has been a growing body of evidence supporting the concept that tumour is driven by a minor sub-population of self-renewing cancer stem cells (CSCs). CSCs were first identified in the haematopoietic system and subsequently in a variety of solid tumours including brain, breast, colon, prostate, and others (157, 158). Most of the conventional treatment regimens target the non-CSC population of the tumour and fail to eliminate the inherently resistant CSCs (159). The remaining chemotherapy-resistant CSCs lead to chemotherapy refractory tumour, and may explain the difficulty in complete eradication of cancer and recurrence. Therefore, development of therapeutic strategies that specifically target CSCs is warranted in reducing the risk of cancer relapse and recurrence.

Can curcumin uproot the 'root of all evils'? A hypothesis. Normal, non-cancerous stem cells exhibit well-fortified DNA mutation defence systems and overexpression of drug efflux pumps that typically serve to prevent mutation into carcinogenic CSCs. Unfortunately, when mutations that create CSCs do occur, the inherent defence systems of stem cells

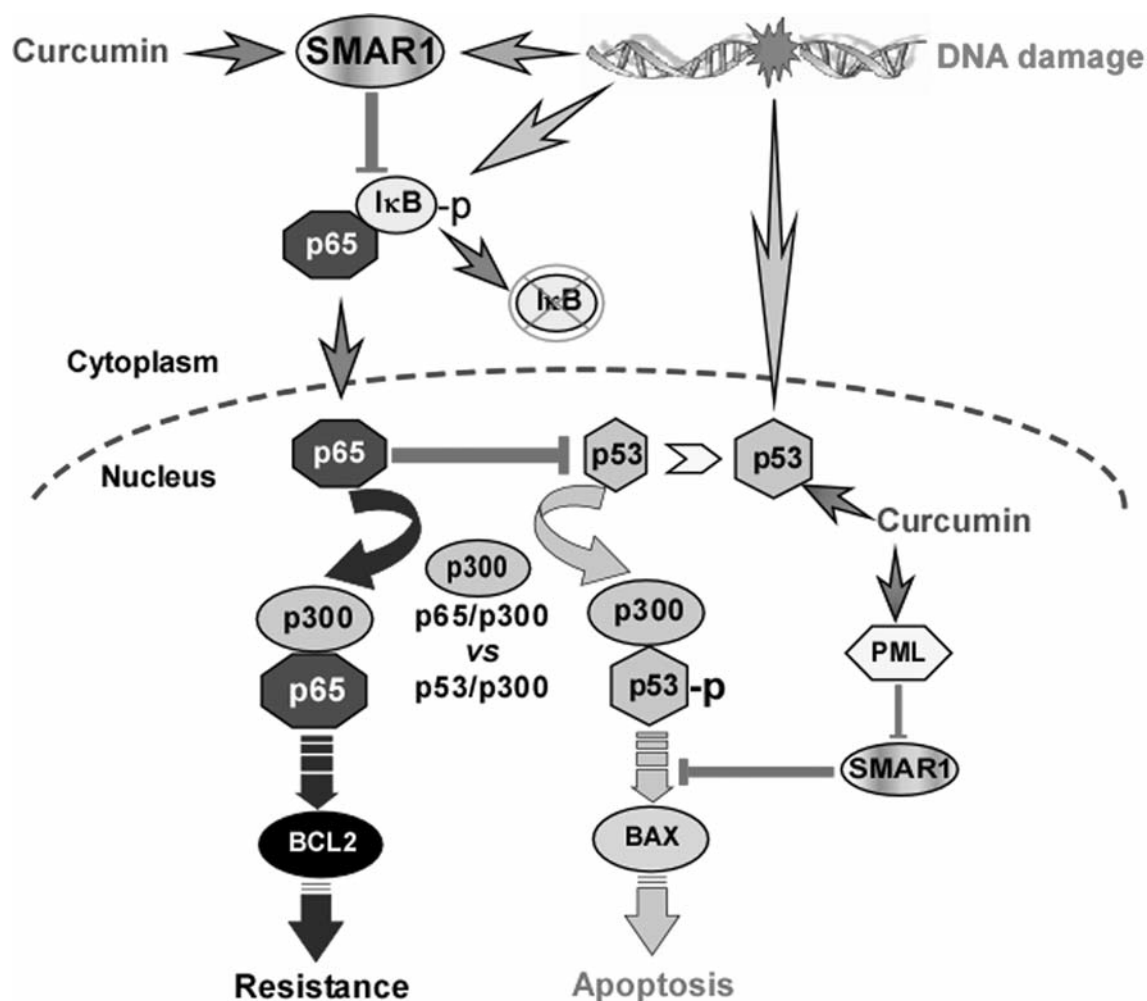


Figure 5. Schematic illustration depicting differential regulation of anti- and pro-apoptotic networks affected by curcumin in drug-resistant cells. Curcumin, by inhibiting p65 nuclear factor kappa B (NF-κB) and inducing promyelocytic leukemia (PML)-scaffold/matrix associated region 1 (SMAR1) cross-talk, censored the resistance pathway, thereby making p300 available for p53 interaction upon doxorubicin treatment, resulting in transcription of pro-apoptotic protein BAX, that effectively sensitizes drug-resistant cancer cells.

serve to protect them from DNA-targeting chemo- and radiation-therapy. In addition, relative dormancy/slow cell-cycle kinetics, sustained telomerase function and resistance to apoptosis promote oncogenic resistance to cytotoxic chemotherapeutic agents. Through the revolutionized concept of CSCs, cancer research has been reinvigorated to study the role of these unique cells in cancer relapse and, more importantly, as targets in innovative therapies. On the basis of the above discussions on the potential role of curcumin in conquering chemotherapy-induced resistance in cancer of different origin, by modulating various MDR mechanisms, we hypothesize that curcumin may uproot the ‘root of all evils’ *i.e.* CSCs. Our discussion further elaborating that to some extent similar MDR mechanisms regulate drug-resistance in CSCs, supports our hypothesis. Further support to our

hypothesis arises from the information that the combinatorial therapy of dasatinib and curcumin inhibited cellular growth, invasion and colonosphere formation and also reduced CSC population as evidenced by the decreased expression of CSC-specific markers (160). Curcumin inhibited signal transducer and activator of transcription-3 (STAT3) phosphorylation, cell viability and tumour sphere formation in colon cancer stem cells (161). Curcumin also inhibited the side population (SP) of rat C6 glioma cells, characteristic of CSCs (162). Another report highlighted that curcumin in combination with piperine targets breast CSCs (163). Additionally, our recent findings indicate that curcumin pre-treatment of breast cancer cells remarkably regressed the CSC repertoire (unpublished data). However, further work is required to quell the as yet unresolved debate on curcumin in the inhibition of CSCs.

Conclusion

As illustrated in this review, MDR is the major impediment in the cure of cancer. Being multifactorial in nature, the conventional one drug-one target theory fails to overcome chemotherapy resistance; moreover, such drugs exert toxicity towards normal cells. As reviewed above, curcumin single-handedly, or as an adjuvant with cytotoxic drugs (combinatorial therapy), alters intricate networks of signals and molecular interactions *i.e.* 'one drug, multiple targets' which regulate several aspects of MDR manifesting chemosensitivity of otherwise resistant tumours. Moreover curcumin confers a high therapeutic index in tumours by simultaneously protecting the host from adverse side-effects, thus widening the index from both sides. For these compounds to exert maximum potency *in vivo*, an understanding of the absorption and metabolism of curcumin is crucial. Hence it is of utmost importance to synthesize compounds retaining the molecular framework of curcumin, while exhibiting enhanced bioavailability. However, further in-depth mechanistic studies, *in vivo* animal experiments, and clinical trials are needed to bring this concept into practice to fully appreciate the value of curcumin in combinatorial therapy of human cancer.

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References

- Sosin MD and Handa SI: Spontaneous remission of large granular lymphocytic leukaemia. *Int J Clin Pract* 57: 551-552, 2003.
- Pommier Y, Sordet O, Antony S, Hayward RL and Kohn KW: Apoptosis defects and chemotherapy resistance: molecular interaction maps and networks. *Oncogene* 23: 2934-2949, 2004.
- Pluen A, Boucher Y, Ramanujan S, McKee TD, Gohongi T, di Tomaso E, Brown EB, Izumi Y, Campbell RB, Berk DA and Jain RK: Role of tumor-host interactions in interstitial diffusion of macromolecules: cranial *vs.* subcutaneous tumors. *Proc Natl Acad Sci USA* 98: 4628-4633, 2001.
- Diaconu CC, Szathmari M, Keri G and Venetianer A: Apoptosis is induced in both drug-sensitive and multidrug-resistant hepatoma cells by somatostatin analogue TT-232. *Br J Cancer* 80: 1197-1203, 1999.
- Dalton WS and Jove R: Drug resistance in multiple myeloma: approaches to circumvention. *Semin Oncol* 26: 23-27, 1999.
- Mohanty S, Adhikary A, Chakrabarty S, Sa G and Das T: Operation 'p53 Hunt' to combat cancer: theaflavins in action. *Front Biosci (Schol Ed)* 4: 300-320, 2012.
- Gopalakrishna R and Jaken S: Protein kinase C signaling and oxidative stress. *Free Radic Biol Med* 28: 1349-1361, 2000.
- Bhimani RS, Troll W, Grunberger D and Frenkel K: Inhibition of oxidative stress in HeLa cells by chemopreventive agents. *Cancer Res* 53: 4528-4533, 1993.
- Bhattacharyya A, Mandal D, Lahiry L, Bhattacharyya S, Chattopadhyay S, Ghosh UK, Sa G and Das T: Black tea-induced amelioration of hepatic oxidative stress through antioxidative activity in EAC-bearing mice. *J Environ Pathol Toxicol Oncol* 26: 245-254, 2007.
- Ray PK, Das T, Sa G, Ghosh AK and Chattopadhyay S: Protection of apoptotic cell death by protein A. *Apoptosis* 5: 509-514, 2000.
- Sa G and Das T: Anticancer effects of curcumin: cycle of life and death. *Cell Div* 3: 14, 2008.
- Sosin M and Handa S: Low dose methotrexate and bone marrow suppression. *Bmj* 326: 266-267, 2003.
- Rayman P, Wesa AK, Richmond AL, Das T, Biswas K, Raval G, Storkus WJ, Tannenbaum C, Novick A, Bukowski R and Finke J: Effect of renal cell carcinomas on the development of type 1 T-cell responses. *Clin Cancer Res* 10: 6360S-6366S, 2004.
- Hossain DM, Bhattacharyya S, Das T and Sa G: Curcumin: the multi-targeted therapy for cancer regression. *Front Biosci (Schol Ed)* 4: 335-355, 2012.
- Koty PP, Zhang H and Levitt ML: Antisense BCL-2 treatment increases programmed cell death in non-small cell lung cancer cell lines. *Lung Cancer* 23: 115-127, 1999.
- Nilsen H and Krokan HE: Base excision repair in a network of defence and tolerance. *Carcinogenesis* 22: 987-998, 2001.
- Persidis A: Cancer multidrug resistance. *Nat Biotechnol* 17: 94-95, 1999.
- Nooter K and Stoter G: Molecular mechanisms of multidrug resistance in cancer chemotherapy. *Pathol Res Pract* 192: 768-780, 1996.
- Dean M, Hamon Y and Chimini G: The human ATP-binding cassette (ABC) transporter superfamily. *J Lipid Res* 42: 1007-1017, 2001.
- Juliano RL and Ling V: A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochim Biophys Acta* 455: 152-162, 1976.
- Shain KH and Dalton WS: Cell adhesion is a key determinant in *de novo* multidrug resistance (MDR): new targets for the prevention of acquired MDR. *Mol Cancer Ther* 1: 69-78, 2001.
- Huang CC, Wu MC, Xu GW, Li DZ, Cheng H, Tu ZX, Jiang HQ and Gu JR: Overexpression of the MDR1 gene and P-glycoprotein in human hepatocellular carcinoma. *J Natl Cancer Inst* 84: 262-264, 1992.
- Kramer R, Weber TK, Morse B, Arcenci R, Staniunas R, Steele G Jr. and Summerhayes IC: Constitutive expression of multidrug resistance in human colorectal tumours and cell lines. *Br J Cancer* 67: 959-968, 1993.
- Petrini M, Di Simone D, Favati A, Mattii L, Valentini P and Grassi B: GST-pi and P-170 co-expression in multiple myeloma. *Br J Haematol* 90: 393-397, 1995.
- Grundy M, Seedhouse C, Russell NH and Pallis M: P-Glycoprotein and breast cancer resistance protein in acute myeloid leukaemia cells treated with the aurora-B kinase inhibitor barasertib-hQPA. *BMC Cancer* 11: 254, 2011.
- Szakacs G, Varadi A, Ozvegy-Laczka C and Sarkadi B: The role of ABC transporters in drug absorption, distribution, metabolism, excretion and toxicity (ADME-Tox). *Drug Discov Today* 13: 379-393, 2008.

- 27 Laberge RM, Karwatsky J, Lincoln MC, Leimanis ML and Georges E: Modulation of GSH levels in ABCC1 expressing tumor cells triggers apoptosis through oxidative stress. *Biochem Pharmacol* 73: 1727-1737, 2007.
- 28 Sharom FJ: ABC multidrug transporters: structure, function and role in chemoresistance. *Pharmacogenomics* 9: 105-127, 2008.
- 29 Ozvegy C, Litman T, Szakacs G, Nagy Z, Bates S, Varadi A and Sarkadi B: Functional characterization of the human multidrug transporter, ABCG2, expressed in insect cells. *Biochem Biophys Res Commun* 285: 111-117, 2001.
- 30 Polgar O, Robey RW and Bates SE: ABCG2: structure, function and role in drug response. *Expert Opin Drug Metab Toxicol* 4: 1-15, 2008.
- 31 Dalton WS and Scheper RJ: Lung resistance-related protein: determining its role in multidrug resistance. *J Natl Cancer Inst* 91: 1604-1605, 1999.
- 32 Scheffer GL, Wijngaard PL, Flens MJ, Izquierdo MA, Slovak ML, Pinedo HM, Meijer CJ, Clevers HC and Scheper RJ: The drug resistance-related protein LRP is the human major vault protein. *Nat Med* 1: 578-582, 1995.
- 33 Scheffer GL, Schroeijers AB, Izquierdo MA, Wiemer EA and Scheper RJ: Lung resistance-related protein/major vault protein and vaults in multidrug-resistant cancer. *Curr Opin Oncol* 12: 550-556, 2000.
- 34 Izquierdo MA, van der Zee AG, Vermorken JB, van der Valk P, Belien JA, Giaccone G, Scheffer GL, Flens MJ, Pinedo HM, Kenemans P, Chris JLM Meijer, Elisabeth G. E.de Vriesand and Rik J. Scheper: Drug resistance-associated marker LRP for prediction of response to chemotherapy and prognoses in advanced ovarian carcinoma. *J Natl Cancer Inst* 87: 1230-1237, 1995.
- 35 List AF, Spier CS, Grogan TM, Johnson C, Roe DJ, Greer JP, Wolff SN, Broxterman HJ, Scheffer GL, Scheper RJ and Dalton WS: Overexpression of the major vault transporter protein lung-resistance protein predicts treatment outcome in acute myeloid leukemia. *Blood* 87: 2464-2469, 1996.
- 36 Talalay P: Chemoprotection against cancer by induction of phase 2 enzymes. *Biofactors* 12: 5-11, 2000.
- 37 McFadyen MC, McLeod HL, Jackson FC, Melvin WT, Doehmer J and Murray GI: Cytochrome P450 CYP1B1 protein expression: a novel mechanism of anticancer drug resistance. *Biochem Pharmacol* 62: 207-212, 2001.
- 38 Kivisto KT, Kroemer HK and Eichelbaum M: The role of human cytochrome P450 enzymes in the metabolism of anticancer agents: implications for drug interactions. *Br J Clin Pharmacol* 40: 523-530, 1995.
- 39 Kivisto KT, Bookjans G, Fromm MF, Griese EU, Munzel P and Kroemer HK: Expression of CYP3A4, CYP3A5 and CYP3A7 in human duodenal tissue. *Br J Clin Pharmacol* 42: 387-389, 1996.
- 40 Harris AL and Hochhauser D: Mechanisms of multidrug resistance in cancer treatment. *Acta Oncol* 31: 205-213, 1992.
- 41 Hayes JD and Pulford DJ: The glutathione S-transferase supergene family: regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. *Crit Rev Biochem Mol Biol* 30: 445-600, 1995.
- 42 Schecter RL, Alaoui-Jamali MA and Batist G: Glutathione S-transferase in chemotherapy resistance and in carcinogenesis. *Biochem Cell Biol* 70: 349-353, 1992.
- 43 Rassool FV and Tomkinson AE: Targeting abnormal DNA double-strand break repair in cancer. *Cell Mol Life Sci* 67: 3699-3710.
- 44 Fojo T: Cancer, DNA repair mechanisms, and resistance to chemotherapy. *J Natl Cancer Inst* 93: 1434-1436, 2001.
- 45 Braastad CD, Leguia M and Hendrickson EA: Ku86 autoantigen related protein-1 transcription initiates from a CpG island and is induced by p53 through a nearby p53 response element. *Nucleic Acids Res* 30: 1713-1724, 2002.
- 46 Lavin MF: Ataxia-telangiectasia: from a rare disorder to a paradigm for cell signalling and cancer. *Nat Rev Mol Cell Biol* 9: 759-769, 2008.
- 47 Rajewsky N, Socci ND, Zapotocky M and Siggia ED: The evolution of DNA regulatory regions for proteo-gamma bacteria by interspecies comparisons. *Genome Res* 12: 298-308, 2002.
- 48 Sakamaki K and Satou Y: Caspases: evolutionary aspects of their functions in vertebrates. *J Fish Biol* 74: 727-753, 2009.
- 49 Bratton SB and Cohen GM: Death receptors leave a caspase footprint that Smacs of XIAP. *Cell Death Differ* 10: 4-6, 2003.
- 50 Safa AR and Pollok KE: Targeting the anti-apoptotic protein c-FLIP for cancer therapy. *Cancers (Basel)* 3: 1639-1671, 2011.
- 51 Fulda S and Debatin KM: Targeting inhibitor of apoptosis proteins (IAPs) for diagnosis and treatment of human diseases. *Recent Pat Anticancer Drug Discov* 1: 81-89, 2006.
- 52 Tepper CG and Seldin MF: Modulation of caspase-8 and FLICE-inhibitory protein expression as a potential mechanism of Epstein-Barr virus tumorigenesis in Burkitt's lymphoma. *Blood* 94: 1727-1737, 1999.
- 53 Herr I and Debatin KM: Cellular stress response and apoptosis in cancer therapy. *Blood* 98: 2603-2614, 2001.
- 54 Rosen D, Li JH, Keidar S, Markon I, Orda R and Berke G: Tumor immunity in perforin-deficient mice: a role for CD95 (Fas/Apo-1). *J Immunol* 164: 3229-3235, 2000.
- 55 Takeda K, Smyth MJ, Cretney E, Hayakawa Y, Yamaguchi N, Yagita H and Okumura K: Involvement of tumor necrosis factor-related apoptosis-inducing ligand in NK cell-mediated and IFN-gamma-dependent suppression of subcutaneous tumor growth. *Cell Immunol* 214: 194-200, 2001.
- 56 Nicholson DW and Thornberry NA: Apoptosis. Life and death decisions. *Science* 299: 214-215, 2003.
- 57 Lowe SW and Lin AW: Apoptosis in cancer. *Carcinogenesis* 21: 485-495, 2000.
- 58 Bartke T, Siegmund D, Peters N, Reichwein M, Henkler F, Scheurich P and Wajant H: p53 up-regulates cFLIP, inhibits transcription of NF-kappaB-regulated genes and induces caspase-8-independent cell death in DLD-1 cells. *Oncogene* 20: 571-580, 2001.
- 59 Hoffman WH, Biade S, Zilfou JT, Chen J and Murphy M: Transcriptional repression of the anti-apoptotic survivin gene by wild type p53. *J Biol Chem* 277: 3247-3257, 2002.
- 60 Hwang PH, Yi HK, Kim DS, Nam SY, Kim JS and Lee DY: Suppression of tumorigenicity and metastasis in B16F10 cells by *PTEN/MMAC1/TEP1* gene. *Cancer Lett* 172: 83-91, 2001.
- 61 Moroni MC, Hickman ES, Lazzerini Denchi E, Caprara G, Colli E, Cecconi F, Muller H and Helin K: APAF-1 is a transcriptional target for E2F and p53. *Nat Cell Biol* 3: 552-558, 2001.
- 62 Ryan KM, Phillips AC and Vousden KH: Regulation and function of the p53 tumor suppressor protein. *Curr Opin Cell Biol* 13: 332-337, 2001.

- 63 Gasco M, Shami S and Crook T: The p53 pathway in breast cancer. *Breast Cancer Res* 4: 70-76, 2002.
- 64 Schmitt CA, McCurrach ME, de Stanchina E, Wallace-Brodeur RR and Lowe SW: *INK4a/ARF* mutations accelerate lymphomagenesis and promote chemoresistance by disabling p53. *Genes Dev* 13: 2670-2677, 1999.
- 65 Igney FH and Krammer PH: Death and anti-death: tumour resistance to apoptosis. *Nat Rev Cancer* 2: 277-288, 2002.
- 66 Youle RJ and Strasser A: The BCL-2 protein family: opposing activities that mediate cell death. *Nat Rev Mol Cell Biol* 9: 47-59, 2008.
- 67 Huang WC and Hung MC: Induction of AKT activity by chemotherapy confers acquired resistance. *J Formos Med Assoc* 108: 180-194, 2009.
- 68 Steelman LS, Navolanic P, Chappell WH, Abrams SL, Wong EW, Martelli AM, Cocco L, Stivala F, Libra M, Nicoletti F, Drobot LB, Franklin RA and McCubrey JA: Involvement of AKT and mTOR in chemotherapeutic- and hormonal-based drug resistance and response to radiation in breast cancer cells. *Cell Cycle* 10: 3003-3015.
- 69 Iida K, Nakayama K, Rahman MT, Rahman M, Ishikawa M, Katagiri A, Yeasmin S, Otsuki Y, Kobayashi H, Nakayama S and Miyazaki K: *EGFR* gene amplification is related to adverse clinical outcomes in cervical squamous cell carcinoma, making the *EGFR* pathway a novel therapeutic target. *Br J Cancer* 105: 420-427.
- 70 Choy G, Liu JW, Chandra D and Tang DG: Cell survival signaling during apoptosis: implications in drug resistance and anticancer therapeutic development. *Prog Drug Res* 63: 115-145, 2005.
- 71 Sordet O, Khan QA, Kohn KW and Pommier Y: Apoptosis induced by topoisomerase inhibitors. *Curr Med Chem Anticancer Agents* 3: 271-290, 2003.
- 72 Tang D, Okada H, Ruland J, Liu L, Stambolic V, Mak TW and Ingram AJ: Akt is activated in response to an apoptotic signal. *J Biol Chem* 276: 30461-30466, 2001.
- 73 Karpinich NO, Tafani M, Rothman RJ, Russo MA and Farber JL: The course of etoposide-induced apoptosis from damage to DNA and p53 activation to mitochondrial release of cytochrome c. *J Biol Chem* 277: 16547-16552, 2002.
- 74 Wu H, Goel V and Haluska FG: PTEN signaling pathways in melanoma. *Oncogene* 22: 3113-3122, 2003.
- 75 Huang H, Chevillat JC, Pan Y, Roche PC, Schmidt LJ and Tindall DJ: PTEN induces chemosensitivity in PTEN-mutated prostate cancer cells by suppression of BCL-2 expression. *J Biol Chem* 276: 38830-38836, 2001.
- 76 Yuan XJ and Whang YE: PTEN sensitizes prostate cancer cells to death receptor-mediated and drug-induced apoptosis through a FADD-dependent pathway. *Oncogene* 21: 319-327, 2002.
- 77 Karin M and Ben-Neriah Y: Phosphorylation meets ubiquitination: the control of NF-[kappa]B activity. *Annu Rev Immunol* 18: 621-663, 2000.
- 78 Zhou G and Kuo MT: NF-kappaB-mediated induction of mdrlb expression by insulin in rat hepatoma cells. *J Biol Chem* 272: 15174-15183, 1997.
- 79 Antonsson B: Bax and other pro-apoptotic Bcl-2 family killer proteins and their victim the mitochondrion. *Cell Tissue Res* 306: 347-361, 2001.
- 80 Nakashio A, Fujita N, Rokudai S, Sato S and Tsuruo T: Prevention of phosphatidylinositol 3'-kinase-Akt survival signaling pathway during topotecan-induced apoptosis. *Cancer Res* 60: 5303-5309, 2000.
- 81 Markovina S, Callander NS, O'Connor SL, Kim J, Werndli JE, Raschko M, Leith CP, Kahl BS, Kim K and Miyamoto S: Bortezomib-resistant nuclear factor-kappaB activity in multiple myeloma cells. *Mol Cancer Res* 6: 1356-1364, 2008.
- 82 Peng H, Wen J, Li H, Chang J and Zhou X: Drug inhibition profile prediction for NFkB pathway in multiple myeloma. *PLoS One* 6: e14750.
- 83 Cortes Sempere M, Rodriguez Fanjul V, Sanchez Perez I and Perona R: The role of the NFkappaB signalling pathway in cancer. *Clin Transl Oncol* 10: 143-147, 2008.
- 84 Cataldi A, Rapino M, Centurione L, Sabatini N, Grifone G, Garaci F and Rana R: NFkB activation plays an antiapoptotic role in human leukemic K562 cells exposed to ionizing radiation. *J Cell Biochem* 89: 956-963, 2003.
- 85 Wang CY, Cusack JC Jr., Liu R and Baldwin AS Jr.: Control of inducible chemoresistance: enhanced antitumor therapy through increased apoptosis by inhibition of NF-kappaB. *Nat Med* 5: 412-417, 1999.
- 86 Jones DR, Broad RM, Madrid LV, Baldwin AS Jr. and Mayo MW: Inhibition of NF-kappaB sensitizes non-small cell lung cancer cells to chemotherapy-induced apoptosis. *Ann Thorac Surg* 70: 930-936; discussion 936-937, 2000.
- 87 Ling MT, Wang X, Ouyang XS, Xu K, Tsao SW and Wong YC: Id-1 expression promotes cell survival through activation of NF-kappaB signalling pathway in prostate cancer cells. *Oncogene* 22: 4498-4508, 2003.
- 88 Muerkoster S, Arlt A, Witt M, Gehrz A, Haye S, March C, Grohmann F, Wegehenkel K, Kalthoff H, Folsch UR and Schafer H: Usage of the NF-kappaB inhibitor sulfasalazine as sensitizing agent in combined chemotherapy of pancreatic cancer. *Int J Cancer* 104: 469-476, 2003.
- 89 Surh YJ: Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 3: 768-780, 2003.
- 90 Wargovich MJ: Nutrition and cancer: the herbal revolution. *Curr Opin Gastroenterol* 15: 177-180, 1999.
- 91 Aggarwal BB and Shishodia S: Molecular targets of dietary agents for prevention and therapy of cancer. *Biochem Pharmacol* 71: 1397-1421, 2006.
- 92 Shim JH, Su ZY, Chae JI, Kim DJ, Zhu F, Ma WY, Bode AM, Yang CS and Dong Z: Epigallocatechin gallate suppresses lung cancer cell growth through RAS-GTPase-activating protein SH3 domain-binding protein 1. *Cancer Prev Res (Phila)* 3: 670-679, 2010.
- 93 Luo T, Wang J, Yin Y, Hua H, Jing J, Sun X, Li M, Zhang Y and Jiang Y: (-)-Epigallocatechin gallate sensitizes breast cancer cells to paclitaxel in a murine model of breast carcinoma. *Breast Cancer Res* 12: R8, 2010.
- 94 Lahiry L, Saha B, Chakraborty J, Adhikary A, Mohanty S, Hossain DM, Banerjee S, Das K, Sa G and Das T: Theaflavins target FAS/caspase-8 and AKT/pBAD pathways to induce apoptosis in p53-mutated human breast cancer cells. *Carcinogenesis* 31: 259-268 2010.
- 95 Lahiry L, Saha B, Chakraborty J, Bhattacharyya S, Chattopadhyay S, Banerjee S, Choudhuri T, Mandal D, Bhattacharyya A, Sa G and Das T: Contribution of p53-mediated Bax transactivation in theaflavin-induced mammary epithelial carcinoma cell apoptosis. *Apoptosis* 13: 771-781, 2008.
- 96 Adhikary A, Mohanty S, Lahiry L, Hossain DM, Chakraborty S and Das T: Theaflavins retard human breast cancer cell migration by inhibiting NF-kappaB via p53-ROS cross-talk. *FEBS Lett* 584: 7-14, 2010.

- 97 Chattopadhyay S, Bhattacharyya S, Saha B, Chakraborty J, Mohanty S, Sakib Hossain DM, Banerjee S, Das K, Sa G and Das T: Tumor-shed PGE(2) impairs IL2R γ -signaling to inhibit CD4 T cell survival: regulation by theaflavins. *PLoS One* 4: e7382, 2009.
- 98 Bhattacharyya A, Choudhuri T, Pal S, Chattopadhyay S, G KD, Sa G and Das T: Apoptogenic effects of black tea on Ehrlich's ascites carcinoma cell. *Carcinogenesis* 24: 75-80, 2003.
- 99 Bhattacharyya S, Mandal D, Saha B, Sen GS, Das T and Sa G: Curcumin prevents tumor-induced T cell apoptosis through Stat-5a-mediated Bcl-2 induction. *J Biol Chem* 282: 15954-15964, 2007.
- 100 Brown KC, Witte TR, Hardman WE, Luo H, Chen YC, Carpenter AB, Lau JK and Dasgupta P: Capsaicin displays anti-proliferative activity against human small cell lung cancer in cell culture and nude mice models *via* the E2F pathway. *PLoS One* 5: e10243.
- 101 Huang SP, Chen JC, Wu CC, Chen CT, Tang NY, Ho YT, Lo C, Lin JP, Chung JG and Lin JG: Capsaicin-induced apoptosis in human hepatoma HepG2 cells. *Anticancer Res* 29: 165-174, 2009.
- 102 Cheah YH, Nordin FJ, Sarip R, Tee TT, Azimahtol HL, Sirat HM, Rashid BA, Abdullah NR and Ismail Z: Combined xanthorrhizol-curcumin exhibits synergistic growth inhibitory activity *via* apoptosis induction in human breast cancer cells MDA-MB-231. *Cancer Cell Int* 9: 1, 2009.
- 103 Siddiqui IA, Malik A, Adhami VM, Asim M, Hafeez BB, Sarfaraz S and Mukhtar H: Green tea polyphenol EGCG sensitizes human prostate carcinoma LNCaP cells to TRAIL-mediated apoptosis and synergistically inhibits biomarkers associated with angiogenesis and metastasis. *Oncogene* 27: 2055-2063, 2008.
- 104 Pal S, Bhattacharyya S, Choudhuri T, Datta GK, Das T and Sa G: Amelioration of immune cell number depletion and potentiation of depressed detoxification system of tumor-bearing mice by curcumin. *Cancer Detect Prev* 29: 470-478, 2005.
- 105 Pal S, Choudhuri T, Chattopadhyay S, Bhattacharya A, Datta GK, Das T and Sa G: Mechanisms of curcumin-induced apoptosis of Ehrlich's ascites carcinoma cells. *Biochem Biophys Res Commun* 288: 658-665, 2001.
- 106 Chakraborty J, Banerjee S, Ray P, Hossain DM, Bhattacharyya S, Adhikary A, Chattopadhyay S, Das T and Sa G: Gain of cellular adaptation due to prolonged p53 impairment leads to functional switchover from p53 to p73 during DNA damage in acute myeloid leukemia cells. *J Biol Chem* 285: 33104-33112, 2010.
- 107 Choudhuri T, Pal S, Das T and Sa G: Curcumin selectively induces apoptosis in deregulated cyclin D1-expressed cells at G2 phase of cell cycle in a p53-dependent manner. *J Biol Chem* 280: 20059-20068, 2005.
- 108 Choudhuri T, Pal S, Aggarwal ML, Das T and Sa G: Curcumin induces apoptosis in human breast cancer cells through p53-dependent BAX induction. *FEBS Lett* 512: 334-340, 2002.
- 109 Bhattacharyya S, Mandal D, Sen GS, Pal S, Banerjee S, Lahiry L, Finke JH, Tannenbaum CS, Das T and Sa G: Tumor-induced oxidative stress perturbs nuclear factor-kappaB activity-augmenting tumor necrosis factor-alpha-mediated T-cell death: protection by curcumin. *Cancer Res* 67: 362-370, 2007.
- 110 Bhattacharyya S, Md Sakib Hossain D, Mohanty S, Sankar Sen G, Chattopadhyay S, Banerjee S, Chakraborty J, Das K, Sarkar D, Das T and Sa G: Curcumin reverses T-cell-mediated adaptive immune dysfunctions in tumor-bearing hosts. *Cell Mol Immunol* 7: 306-315, 2010.
- 111 Strimpakos AS and Sharma RA: Curcumin: preventive and therapeutic properties in laboratory studies and clinical trials. *Antioxid Redox Signal* 10: 511-545, 2008.
- 112 Das T, Sa G, Saha B and Das K: Multifocal signal modulation therapy of cancer: ancient weapon, modern targets. *Mol Cell Biochem* 336: 85-95, 2010.
- 113 Aggarwal BB, Sundaram C, Malani N and Ichikawa H: Curcumin: the Indian solid gold. *Adv Exp Med Biol* 595: 1-75, 2007.
- 114 Anand P, Sundaram C, Jhurani S, Kunnumakkara AB and Aggarwal BB: Curcumin and cancer: an old-age disease with an age-old solution. *Cancer Lett* 267: 133-164, 2008.
- 115 Jagetia GC and Aggarwal BB: Spicing up of the immune system by curcumin. *J Clin Immunol* 27: 19-35, 2007.
- 116 Kuttan G, Kumar KB, Guruvayoorappan C and Kuttan R: Antitumor, anti-invasion, and antimetastatic effects of curcumin. *Adv Exp Med Biol* 595: 173-184, 2007.
- 117 Ramachandran C, Rodriguez S, Ramachandran R, Raveendran Nair PK, Fonseca H, Khatib Z, Escalon E and Melnick SJ: Expression profiles of apoptotic genes induced by curcumin in human breast cancer and mammary epithelial cell lines. *Anticancer Res* 25: 3293-3302, 2005.
- 118 Dhillon N, Aggarwal BB, Newman RA, Wolff RA, Kunnumakkara AB, Abbruzzese JL, Ng CS, Badmaev V and Kurzrock R: Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clin Cancer Res* 14: 4491-4499, 2008.
- 119 Aggarwal BB, Kumar A and Bharti AC: Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 23: 363-398, 2003.
- 120 Cheng AL, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, Lin JT, Lin BR, Ming-Shiang W, Yu HS, Jee SH, Chen GS, Chen TM, Chen CA, Lai MK, Pu YS, Pan MH, Wang YJ, Tsai CC and Hsieh CY: Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res* 21: 2895-2900, 2001.
- 121 Sharma SK, George N, Kadhiraan T, Saha PK, Mishra HK and Hanif M: Prevalence of extensively drug-resistant tuberculosis among patients with multidrug-resistant tuberculosis: a retrospective hospital-based study. *Indian J Med Res* 130: 392-395, 2009.
- 122 Anuchapreeda S, Leechanachai P, Smith MM, Ambudkar SV and Limtrakul PN: Modulation of P-glycoprotein expression and function by curcumin in multidrug-resistant human KB cells. *Biochem Pharmacol* 64: 573-582, 2002.
- 123 Chearwae W, Shukla S, Limtrakul P and Ambudkar SV: Modulation of the function of the multidrug resistance-linked ATP-binding cassette transporter ABCG2 by the cancer chemopreventive agent curcumin. *Mol Cancer Ther* 5: 1995-2006, 2006.
- 124 Chearwae W, Wu CP, Chu HY, Lee TR, Ambudkar SV and Limtrakul P: Curcuminoids purified from turmeric powder modulate the function of human multidrug-resistance protein 1 (ABCC1). *Cancer Chemother Pharmacol* 57: 376-388, 2006.
- 125 Sharma M, Manoharlal R, Shukla S, Puri N, Prasad T, Ambudkar SV and Prasad R: Curcumin modulates efflux mediated by yeast ABC multidrug transporters and is synergistic with antifungals. *Antimicrob Agents Chemother* 53: 3256-3265, 2009.

- 126 Chearwae W, Anuchapreeda S, Nandigama K, Ambudkar SV and Limtrakul P: Biochemical mechanism of modulation of human P-glycoprotein (ABCB1) by curcumin I, II, and III purified from Turmeric powder. *Biochem Pharmacol* 68: 2043-2052, 2004.
- 127 Limtrakul P, Anuchapreeda S and Buddhasukh D: Modulation of human multidrug-resistance MDR-1 gene by natural curcuminoids. *BMC Cancer* 4: 13, 2004.
- 128 Khar A, Ali AM, Pardhasaradhi BV, Varalakshmi CH, Anjum R and Kumari AL: Induction of stress response renders human tumor cell lines resistant to curcumin-mediated apoptosis: role of reactive oxygen intermediates. *Cell Stress Chaperones* 6: 368-376, 2001.
- 129 M Khopde S, Priyadarsini KI, Venkatesan P and Rao MN: Free radical scavenging ability and antioxidant efficiency of curcumin and its substituted analogue. *Biophys Chem* 80: 85-91, 1999.
- 130 Mori Y, Tatematsu K, Koide A, Sugie S, Tanaka T and Mori H: Modification by curcumin of mutagenic activation of carcinogenic *N*-nitrosamines by extrahepatic cytochromes P-450 2B1 and 2E1 in rats. *Cancer Sci* 97: 896-904, 2006.
- 131 Liu A, Yoshioka K, Salerno V and Hsieh P: The mismatch repair-mediated cell cycle checkpoint response to fluorodeoxyuridine. *J Cell Biochem* 105: 245-254, 2008.
- 132 Rowe DL, Ozbay T, O'Regan RM and Nahta R: Modulation of the BRCA1 protein and induction of apoptosis in triple-negative breast cancer cell lines by the polyphenolic compound curcumin. *Breast Cancer (Auckl)* 3: 61-75, 2009.
- 133 Lu HF, Yang JS, Lai KC, Hsu SC, Hsueh SC, Chen YL, Chiang JH, Lu CC, Lo C, Yang MD and Chung JG: Curcumin-induced DNA damage and inhibited DNA repair genes expressions in mouse-rat hybrid retina ganglion cells (N18). *Neurochem Res* 34: 1491-1497, 2009.
- 134 Lu HF, Lai KC, Hsu SC, Lin HJ, Yang MD, Chen YL, Fan MJ, Yang JS, Cheng PY, Kuo CL and Chung JG: Curcumin induces apoptosis through FAS and FADD, in caspase-3-dependent and -independent pathways in the N18 mouse-rat hybrid retina ganglion cells. *Oncol Rep* 22: 97-104, 2009.
- 135 Xiao H and Zhang KJ: Antiproliferative effect of curcumin combined with cyclophosphamide on the growth of human lymphoma cell line HT/CTX with drug resistance and its relation with FA/BRCA pathway. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 16: 804-808, 2008 (in Chinese).
- 136 Dhandapani KM, Mahesh VB and Brann DW: Curcumin suppresses growth and chemoresistance of human glioblastoma cells *via* AP-1 and NF κ B transcription factors. *J Neurochem* 102: 522-538, 2007.
- 137 Wahl H, Tan L, Griffith K, Choi M and Liu JR: Curcumin enhances APO2L/TRAIL-induced apoptosis in chemoresistant ovarian cancer cells. *Gynecol Oncol* 105: 104-112, 2007.
- 138 Deeb D, Xu YX, Jiang H, Gao X, Janakiraman N, Chapman RA and Gautam SC: Curcumin (diferuloyl-methane) enhances tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in LNCaP prostate cancer cells. *Mol Cancer Ther* 2: 95-103, 2003.
- 139 Deeb DD, Jiang H, Gao X, Divine G, Dulchavsky SA and Gautam SC: Chemosensitization of hormone-refractory prostate cancer cells by curcumin to TRAIL-induced apoptosis. *J Exp Ther Oncol* 5: 81-91, 2005.
- 140 Deeb D, Jiang H, Gao X, Al-Holou S, Danyluk AL, Dulchavsky SA and Gautam SC: Curcumin [1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadine-3,5-dione; C₂₁H₂₀O₆] sensitizes human prostate cancer cells to tumor necrosis factor-related apoptosis-inducing ligand/APO2L-induced apoptosis by suppressing nuclear factor-kappaB *via* inhibition of the prosurvival AKT signaling pathway. *J Pharmacol Exp Ther* 321: 616-625, 2007.
- 141 Gao X, Deeb D, Jiang H, Liu YB, Dulchavsky SA and Gautam SC: Curcumin differentially sensitizes malignant glioma cells to TRAIL/Apo2L-mediated apoptosis through activation of procaspases and release of cytochrome *c* from mitochondria. *J Exp Ther Oncol* 5: 39-48, 2005.
- 142 Shankar S, Ganapathy S, Chen Q and Srivastava RK: Curcumin sensitizes TRAIL-resistant xenografts: molecular mechanisms of apoptosis, metastasis and angiogenesis. *Mol Cancer* 7: 16, 2008.
- 143 Woo JH, Kim YH, Choi YJ, Kim DG, Lee KS, Bae JH, Min DS, Chang JS, Jeong YJ, Lee YH, Park JW and Kwon TK: Molecular mechanisms of curcumin-induced cytotoxicity: induction of apoptosis through generation of reactive oxygen species, down-regulation of BCL-XL and IAP, the release of cytochrome *c* and inhibition of Akt. *Carcinogenesis* 24: 1199-1208, 2003.
- 144 Kulik GI: Comparative *in vitro* study of the effects of the new antitumor drug Ukrain and several cytostatic agents on the thiol groups in the tissue of Guerin carcinoma and its resistance to cisplatin variant. *Drugs Exp Clin Res* 24: 277-280, 1998.
- 145 Aggarwal S, Ichikawa H, Takada Y, Sandur SK, Shishodia S and Aggarwal BB: Curcumin (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and metastatic gene products through suppression of I κ B α kinase and AKT activation. *Mol Pharmacol* 69: 195-206, 2006.
- 146 Patel BB, Gupta D, Elliott AA, Sengupta V, Yu Y and Majumdar AP: Curcumin targets FOLFOX-surviving colon cancer cells *via* inhibition of EGFRs and IGF-1R. *Anticancer Res* 30: 319-325, 2010.
- 147 Madonna G, Dansky Ullman C, Gentilecore G, Palmieri G and Ascierto PA: NF κ B as potential target in the treatment of melanoma. *J Transl Med* 10: 53, 2012.
- 148 Divya CS and Pillai MR: Antitumor action of curcumin in human papillomavirus associated cells involves down-regulation of viral oncogenes, prevention of NF κ B and AP-1 translocation, and modulation of apoptosis. *Mol Carcinog* 45: 320-332, 2006.
- 149 Duvoix A, Blasius R, Delhalle S, Schnekenburger M, Morceau F, Henry E, Dicato M and Diederich M: Chemopreventive and therapeutic effects of curcumin. *Cancer Lett* 223: 181-190, 2005.
- 150 Mimeault M and Batra SK: Potential applications of curcumin and its novel synthetic analogs and nanotechnology-based formulations in cancer prevention and therapy. *Chin Med* 6: 31, 2011.
- 151 Sung B, Kunnumakkara AB, Sethi G, Anand P, Guha S and Aggarwal BB: Curcumin circumvents chemoresistance *in vitro* and potentiates the effect of thalidomide and bortezomib against human multiple myeloma in nude mouse model. *Mol Cancer Ther* 8: 959-970, 2009.
- 152 Yallapu MM, Maher DM, Sundram V, Bell MC, Jaggi M and Chauhan SC: Curcumin induces chemo/radio-sensitization in ovarian cancer cells and curcumin nanoparticles inhibit ovarian cancer cell growth. *J Ovarian Res* 3: 11, 2010.
- 153 Ilbey YO, Ozbek E, Cekmen M, Simsek A, Otunctemur A and Somay A: Protective effect of curcumin in cisplatin-induced oxidative injury in rat testis: mitogen-activated protein kinase and nuclear factor-kappa B signaling pathways. *Hum Reprod* 24: 1717-1725, 2009.

- 154 Kim DS, Park SS, Nam BH, Kim IH and Kim SY: Reversal of drug resistance in breast cancer cells by transglutaminase 2 inhibition and nuclear factor-kappaB inactivation. *Cancer Res* 66: 10936-10943, 2006.
- 155 Schneider G, Henrich A, Greiner G, Wolf V, Lovas A, Wiczorek M, Wagner T, Reichardt S, von Werder A, Schmid RM, Weih F, Heinzel T, Saur D and Kramer OH: Cross talk between stimulated NF-kappaB and the tumor suppressor p53. *Oncogene* 29: 2795-806, 2010.
- 156 Sen GS, Mohanty S, Hossain DM, Bhattacharyya S, Banerjee S, Chakraborty J, Saha S, Ray P, Bhattacharjee P, Mandal D, Bhattacharya A, Chattopadhyay S, Das T and Sa G: Curcumin enhances the efficacy of chemotherapy by tailoring p65NFkappaB-p300 cross-talk in favor of p53-p300 in breast cancer. *J Biol Chem* 286: 42232-42247, 2011.
- 157 Wang JC and Dick JE: Cancer stem cells: lessons from leukemia. *Trends Cell Biol* 15: 494-501, 2005.
- 158 Ailles LE and Weissman IL: Cancer stem cells in solid tumors. *Curr Opin Biotechnol* 18: 460-466, 2007.
- 159 Dean M, Fojo T and Bates S: Tumour stem cells and drug resistance. *Nat Rev Cancer* 5: 275-284, 2005.
- 160 Nautiyal J, Kanwar SS, Yu Y and Majumdar AP: Combination of dasatinib and curcumin eliminates chemoresistant colon cancer cells. *J Mol Signal* 6: 7, 2011.
- 161 Lin L, Fuchs J, Li C, Olson V, Bekaii-Saab T and Lin J: STAT3 signaling pathway is necessary for cell survival and tumorsphere forming capacity in ALDH/CD133 stem cell-like human colon cancer cells. *Biochem Biophys Res Commun* 416: 246-251, 2011.
- 162 Fong D, Yeh A, Naftalovich R, Choi TH and Chan MM: Curcumin inhibits the side population (SP) phenotype of the rat C6 glioma cell line: towards targeting of cancer stem cells with phytochemicals. *Cancer Lett* 293: 65-72, 2010.
- 163 Kakarala M, Brenner DE, Korkaya H, Cheng C, Tazi K, Ginestier C, Liu S, Dontu G and Wicha MS: Targeting breast stem cells with the cancer preventive compounds curcumin and piperine. *Breast Cancer Res Treat* 122: 777-785, 2010.

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