

## PKC $\eta$ Is a Novel Prognostic Marker in Non-small Cell Lung Cancer

ELLA KRASNITSKY<sup>1</sup>, Yael BAUMFELD<sup>2</sup>, JANNA FREEDMAN<sup>3</sup>, NETTA SION-VARDY<sup>4</sup>,  
SAMUEL ARIAD<sup>3</sup>, VICTOR NOVACK<sup>2</sup> and ETTA LIVNEH<sup>1\*</sup>

<sup>1</sup>The Shrager Segal Department of Microbiology and Immunology, Faculty of Health Sciences,  
Ben Gurion University of the Negev, and the Departments of <sup>2</sup>Clinical Research,  
<sup>3</sup>Oncology and <sup>4</sup>Pathology, Soroka Medical Center, Beer Sheva, Israel

**Abstract.** *Background:* Novel biomarkers which may serve as therapeutic targets are essential for lung cancer treatment. Here we investigated the prognostic significance of protein kinase C $\eta$  (PKC $\eta$ ), a cell cycle regulator involved in tumorigenesis and chemotherapy resistance, in patients diagnosed with non-small cell lung cancer (NSCLC). *Patients and Methods:* Sixty-three chemotherapy-naïve patients were examined for PKC $\eta$  by immunohistochemistry and divided into PKC $\eta$  H-Score tertiles (low, intermediate and high). Time until event (relapse or mortality) within one year was determined using Cochran-Armitage test and Cox proportional hazards regression model. *Results:* The distribution of patients according to clinical stage 1-4 was: 27%, 5%, 26% and 42%, respectively. PKC $\eta$  overexpression was associated with advanced stage ( $p=0.03$ ) and the risk for an event ( $p=0.045$ ). Patients of the lowest tertile were less likely to experience an event. *Conclusion:* PKC $\eta$  is a novel prognostic marker in NSCLC that may predict poor prognosis. The use of PKC $\eta$ -specific inhibitors in NSCLC may prove valuable.

Lung cancer is the world's most common and deadliest type of cancer. Non-small cell lung cancer (NSCLC) accounts for approximately 85% of total lung cancer malignancies. Treatment is often ineffective because of late diagnosis and the emergence of radio- or chemotherapy resistance. Tumor node metastasis (TNM) staging and histological grading are useful, but not satisfactory classification systems to predict prognosis.

Lung tumorigenesis is a multistep process that involves genetic and epigenetic alterations of oncogenes and tumor-suppressor genes. It affects different signal transduction pathways and alters regulation of major cellular functions, such as cell proliferation, differentiation, cell cycle control, survival and apoptosis (1, 2). The protein kinase C (PKC) family of serine/threonine kinases plays an important role in regulation of key cellular processes (3, 4). Alterations in PKC isoform activation and expression have been found in human lung carcinomas (5-7). Moreover, PKC-dependent mechanisms were shown to be involved in lung cell transformation and resistance to chemotherapy (8, 9). The PKC family is divided into three major groups according to structure, activation requirements, tissue distribution and subcellular localization: classical PKCs ( $\alpha$ ,  $\beta$ 1,  $\beta$ 2 and  $\gamma$ ), novel PKCs ( $\delta$ ,  $\epsilon$ ,  $\eta$  and  $\theta$ ) and atypical PKCs ( $\zeta$  and  $\iota$ ) (10). Several PKC isozymes have been identified as being therapeutic targets, and a number of isozyme-selective PKC inhibitors have been developed for clinical use (11, 12). Two PKC isoforms were indicated as prognostic biomarkers in NSCLC: Patients with high PKC $\iota$  expression were found to be at a higher risk of dying from cancer, independently of stage (13). In addition, PKC $\epsilon$  overexpression was detected in more than 90% of NSCLC specimens and its inhibition in NSCLC cells diminished the aggressive phenotype *in vitro*: cells treated with dominant-negative PKC $\epsilon$  exhibited lower cell proliferation and anchorage-independent growth (14, 15).

PKC $\eta$ , a member of the novel PKCs, is primarily expressed in epithelial cells (16). It is implicated in diverse cellular functions, including a role in terminal differentiation (17-21) and in tumor proliferation of transformed cells (22, 23). Recent studies suggested a special role for PKC $\eta$  in drug resistance and regulation of apoptosis. In several types of tumor, its expression was reported to correlate with drug resistance-associated genes, such as multidrug resistance receptors. It was associated with drug-resistance in patients with breast cancer, ovarian cancer and acute myeloid

This article is freely accessible online.

*Correspondence to:* Etta Livneh, The Shrager Segal Department of Microbiology and Immunology, Faculty of Health Sciences, Ben Gurion University of the Negev, Beer Sheva 84105, Israel. Tel: +972 86477294, Fax: +972 86477626, e-mail: etta@bgu.ac.il

**Key Words:** Non-small cell lung cancer, protein kinase C, PKC $\eta$ , biomarker, prognosis.

Table I. Patients' demographical, clinical and histological data according to event within one year of diagnosis.

Variable	All patients n=63	Event n=38	No event n=25	p-Value
Male, n (%)	50 (79.4)	33 (86.8)	17 (68.0)	0.07
Age at diagnosis (mean±SD), years	64.94±9.87	64.65±10.84	65.36±8.38	0.79
Ethnicity, n (%)				
Ashkenazi	23 (36.5)	11 (28.9)	12 (48.0)	0.29
Sephardic	26 (41.3)	17 (44.7)	9 (36.0)	
Other	14 (22.2%)	10 (26.3)	4 (16.0)	
Smoking, n (%)	49 (77.8)	31 (81.6)	18 (72.0)	0.37
Histological type, n (%)				
Adenocarcinoma	26 (41.3)	14 (36.8)	12 (48.0)	0.23
Squamous cell	22 (34.9)	15 (39.5)	7 (28.0)	
Large cell	12 (19.0)	9 (23.7)	3 (12.0)	
Bronchoalveolar	3 (4.8)	0 (0.0)	3 (4.8)	
Adenocarcinoma histological grade, n (%)				
Well-differentiated	15 (60)	6 (46.2)	9 (75.0)	0.23
Poorly differentiated	10 (40)	7 (53.8)	3 (25.0)	
Stage, n (%)				
1	17 (27.4)	2 (5.3)	15 (62.5)	<0.001
2	3 (4.8)	0 (0.0)	3 (12.5)	
3	16 (25.8)	16 (42.1)	0 (0.0)	
4	26 (41.9)	10 (52.6)	6 (25.0)	
Surgery, n (%)	23 (36.5)	4 (10.5)	19 (76.0)	0.44
Chemotherapy, n (%)	29 (46.0)	19 (50.0)	10 (40.0)	0.01
Radiotherapy, n (%)	31 (49.2)	24 (63.2)	7 (28.0)	
PKC $\eta$ tertiles, n (%)				
≤90	21 (33.3)	10 (26.3)	11 (44.0)	0.12
91-150	23 (36.5)	13 (34.2)	10 (40.0)	
≥151	19 (30.2)	15 (39.5)	4 (16.0)	

leukemia blasts (24-26). In A549 lung cancer cells, down-regulation of PKC $\eta$  resulted in a significant increase in caspase-3 activity and sensitization of these cells to vincristine and paclitaxel (27). In addition, PKC $\eta$  was implicated in antagonizing the apoptotic response in a number of cancer lines; however, the molecular mechanisms are largely unknown (22, 28-34). We have recently shown that PKC $\eta$  contributes to the resistance of breast adenocarcinoma MCF-7 cells by inhibiting c-Jun N-terminal kinase (JNK) activity (33).

In the present study, we examined PKC $\eta$  expression in biopsies from NSCLC patients.

## Materials and Methods

**Patients.** Sixty-three patients with histological diagnosis of NSCLC were included in this study. All patients were diagnosed and treated at the Soroka University Medical Center (SUMC) during 1998-2008. Demographical data, including gender, age and ethnicity was obtained for all patients. Medical records were also reviewed for clinical data, including patient smoking status, types of treatment, and outcome. Pathological data included stage of disease and histological type of cancer. The study was conducted with the approval of the Ethical Review Board of the SUMC.

**Histological examination.** Hematoxylin and eosin stained slides of our patient's biopsies were reviewed for confirmation of histopathological diagnosis and for selection of adequate specimens for analysis as described previously (35).

**Immunohistochemistry.** Paraffin-embedded human lung tissue specimens of pre-treatment core biopsies from 63 patients of our study group were stained with PKC $\eta$ -specific antibody (Santa Cruz Biotechnology, Inc., CA, USA) was performed using the avidin-biotin peroxidase complex method with the Vectastain kit of Vector Laboratories (Burlingame, CA, USA), as described previously (35).

**PKC $\eta$  expression score.** The immunohistochemical expression of PKC $\eta$  was determined by applying a semiquantitative method, incorporating both the intensity and the distribution of specific staining (35, 36). All tumor cell areas in each slide were evaluated for their intensity of staining with PKC $\eta$ -specific antibody, according to the following four category scale: 0=no staining, 1=weak staining, 2=moderate staining and 3=intense staining. The percentage of tumor cells stained within each category of intensity was then determined. For each tissue, a value designating the H-Score was derived by summing the percentages of cells stained at each intensity (Pi) multiplied by the weighted intensity of staining; H-Score= $\sum Pi(i+1)$ , where: i=1, 2, 3 and Pi ranges from 0 to 100%. For statistical purposes, PKC $\eta$  H-Score values were divided into tertiles: the first tertile included patients with low PKC $\eta$  staining

Table II. *Patients' demographic, clinical and histological data according to PKC $\eta$  expression within one year of diagnosis.*

Variable	Tertiles			<i>p</i> -Value
	1st	2nd	3rd	
Male, n (%)	16 (76.2)	20 (87.0)	14 (73.7)	0.52
Age at diagnosis (mean $\pm$ SD), years	63.62 $\pm$ 10.88	66.87 $\pm$ 9.44	64.06 $\pm$ 9.34	0.50
Ethnicity, n (%)				
Ashkenazi	8 (38.1)	11 (47.8)	4 (21.1)	0.21
Sephardic	7 (33.3)	9 (39.1)	10 (52.6)	
Other	6 (28.6)	3 (13.0)	5 (26.3)	
Smoking, n (%)	14 (66.7)	19 (82.6)	16 (84.2)	0.33
Histological type, n (%)				
Adenocarcinoma	8 (38.1)	7 (30.4)	11 (57.9)	0.22
Squamous cell	6 (28.6)	11 (47.8)	5 (26.3)	
Large cell	5 (23.8)	4 (17.4)	3 (15.8)	
Bronchoalveolar	2 (9.5)	1 (4.3)	0 (0.0)	
Adenocarcinoma histological grade, n (%)				
Well-differentiated	6 (75.0)	3 (42.9)	6 (60.0)	0.46
Poorly differentiated	2 (25.0)	4 (57.1)	4 (40.0)	
Stage, n (%)				
1	7 (33.3)	8 (34.8)	2 (11.1)	0.03
2	0 (0.0)	3 (13.0)	0 (0.0)	
3	4 (19.0)	7 (30.4)	5 (27.8)	
4	10 (47.6)	5 (21.7)	11 (61.1)	
Surgery, n (%)	7 (33.3)	11 (47.8)	5 (26.3)	0.34
Chemotherapy, n (%)	12 (57.1)	9 (39.1)	8 (42.1)	0.45
Radiotherapy, n (%)	10 (47.6)	12 (52.2)	9 (47.4)	0.94
Event in one year, n (%)	10 (47.6)	13 (56.5)	15 (78.9)	0.12
Death in one year, n (%)	10 (47.6)	10 (43.5)	12 (66.7)	0.31

with H-Score values in the range of 0-90; the second tertile included patients with intermediate PKC $\eta$  staining, with H-Score values in the range of 91-150; the third tertile included patients with high PKC $\eta$  staining, with H-Scores above 150.

**Statistical analysis.** An event was defined as death of a patient or lung cancer relapse within twelve months of diagnosis; all patients with stage 4 disease were regarded as relapsed disease to begin with, so that an event in this group of patients was defined as death. Patients alive at the end of the follow-up period or those who died throughout it of any cause other than lung cancer were censored from the cumulative survival calculation. Quantitative and qualitative variables are reported with the use of descriptive statistics. The SPSS statistical program, version 16.01 (SPSS Inc, Chicago, IL, USA) was used for data extraction and analysis. Student's *t*-test and the Wilcoxon rank sum test were used to formally test normally and non-normally distributed continuous variables, respectively. The Cochran-Armitage test for trend was used to compare risk ratios for primary outcome. The Kruskal-Wallis test was used to compare groups of non-parametrical variables. The Cox-proportional-regression model was used to compare groups of different PKC $\eta$  H-Score values (in tertiles) and events. The model included age, stage, histology and PKC $\eta$  H-Score values. Stepwise forward regression models with a stay criterion of 0.10 were used. Kaplan-Meier analysis with the log-rank test was applied to compare survival between the groups. A *p*-value of  $\leq 0.05$  (two-sided) was considered significant.

## Results

**Patients' characteristics.** Table I describes the characteristics of the 63 patients included in the present study, with regard to relapse or death within one year of diagnosis. The study group consisted of 20 (32%) patients diagnosed at early clinical stages 1+2, 16 (26%) diagnosed at clinical stage 3 and 26 (42%) diagnosed at clinical stage 4. Primary outcome of relapse or mortality within one year of diagnosis was observed in 38 (60%). Table II describes demographic and clinical data of patients according to PKC $\eta$  H-Score tertiles. The distribution of patients according to tertiles was: 21 (33%) in the first tertile, 23 (37%) in the second tertile and 19 (30%) in the third tertile.

**Immunohistochemistry of patient's biopsies.** The staining intensity of PKC $\eta$  was assessed using the H-Score method. Most of the staining was in the cytoplasm, while the plasma membrane and nuclear envelope exhibited almost no staining. Figure 1 shows tumors with different degrees of staining for PKC $\eta$ , demonstrating tumors with negative, low, intermediate and high staining.

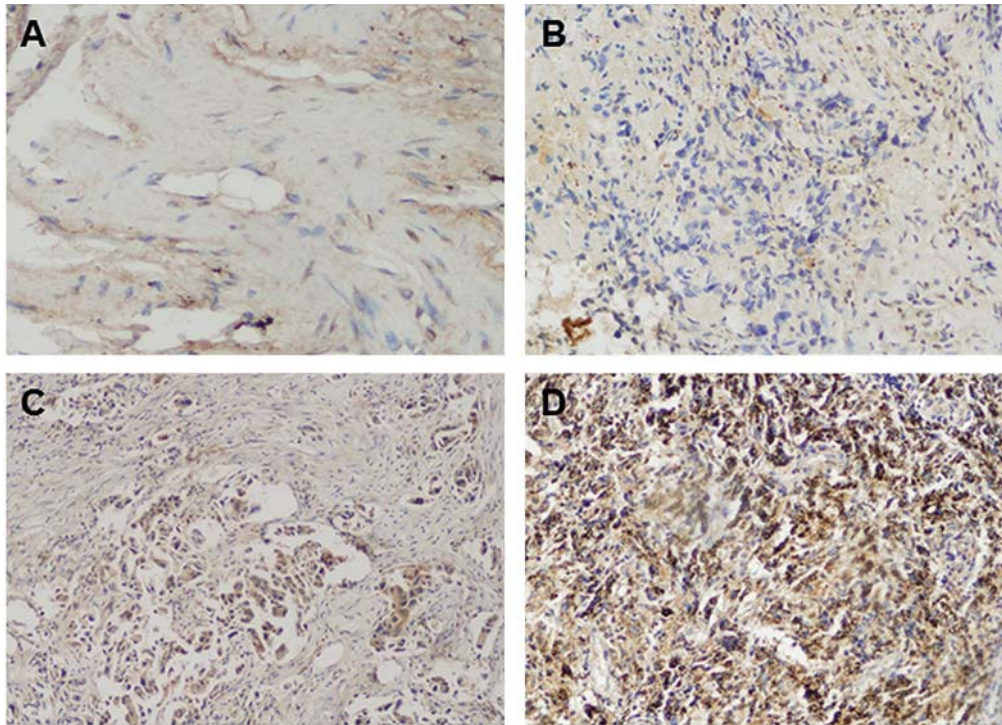


Figure 1. Protein kinase C $\eta$  (PKC $\eta$ ) staining in non-small cell lung cancer (NSCLC) biopsies according to H-Score values. Core biopsies were stained by the labeled avidin-biotin technique using an anti-PKC $\eta$  specific antibody. Cytoplasmic protein variation in tumors was assessed by a semiquantitative method (H-Score) based on stained cell number and intensity. A: H-Score=0 (no staining),  $\times 20$ . B: H-Score=110 (intermediate staining),  $\times 20$ . C: H-Score=190 (high-intermediate staining)  $\times 20$ . D: H-Score=240 (highest staining in the study group)  $\times 40$ .

**PKC $\eta$  distribution according to stage at diagnosis.** As shown in Figure 2, lung cancer patients with clinical stage 4 disease had higher PKC $\eta$  H-Score values compared to patients with disease at earlier stages. The Kruskal-Wallis test showed that PKC $\eta$  expression increased with disease progression ( $p=0.03$ ). Stages 1 and 2 were merged into one group for purposes of statistical convenience.

**Risk for an event within one year of diagnosis according PKC $\eta$  expression.** The risk for an event in one year according to PKC $\eta$  H-Score values was evaluated using the Cochran-Armitage test for trend (Figure 3). Patients exhibiting high PKC $\eta$  expression were found to be at a greater risk for an event compared to patients with low expression ( $p=0.045$ ).

**Multivariate survival analysis of the primary event.** The variables included in the multivariate model for prediction of an event within one year of diagnosis were stage (hazard ratio=2.36,  $p<0.001$ ) and PKC $\eta$  tertiles (hazard ratio=1.45,  $p=0.06$ ). Patients with low H-Scores (first tertile) were less likely to experience an event within one year compared to patients with intermediate and high H-Scores (second and third tertiles) (Figure 4).

## Discussion

In the present study, we show that the expression of PKC $\eta$  in patients with NSCLC correlates with poor prognosis within one year of diagnosis. High H-Score values were in correlation with the stage of disease progression. Patients with high PKC $\eta$  expression exhibited greater risk for an event compared to patients with intermediate and low PKC $\eta$  expression. Accordingly, survival rates were better for patients with low PKC $\eta$  H-Scores. Thus, PKC $\eta$  expression may be of prognostic value and identifies groups of patients with tumors of a more aggressive biology.

PKC $\eta$  is primarily expressed in epithelial tissues including skin, heart, and lung (16). It was also found to be activated by cigarette smoke extracts (37). Here we show that PKC $\eta$  overexpression in lung cancer could be a biomarker for clinical outcome. Its expression was in positive correlation with disease/stage progression and with a tendency for increased relapse or death of NSCLC patients within one year of diagnosis. Moreover, PKC $\eta$  overexpression was of significant predictive value for risk of an event. Although our study group was not large enough, PKC $\eta$  expression was in correlation with poor survival within one year of diagnosis, irrespective of clinical stage; patients with low PKC $\eta$



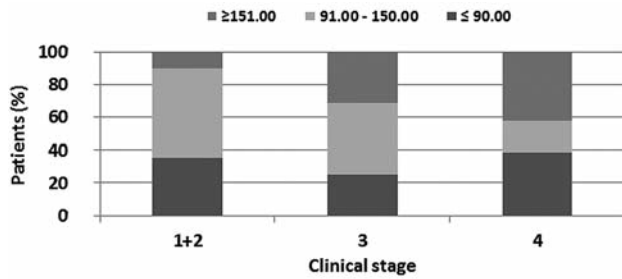


Figure 2. Expression of Protein kinase C $\eta$  (PKC $\eta$ ) increases with disease progression. Patients were divided according to the stage of the disease at diagnosis and the percentage of patients in each tertile was determined as described in the Materials and Methods ( $p=0.03$ ). Clinical stages 1 and 2 were merged due to small study group in stage 2.

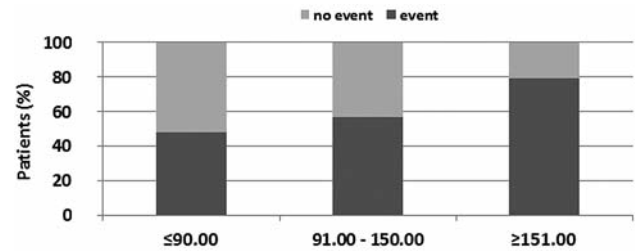


Figure 3. The risk for an event within one year of diagnosis according to Protein kinase C $\eta$  (PKC $\eta$ ) expression. A significant positive correlation was found between PKC $\eta$  expression and the risk for an event within one year of diagnosis using the Cochran-Armitage test ( $p=0.045$ ).

expression had better prognosis compared to patients with intermediate and high PKC $\eta$  expression. The fact that PKC $\eta$  expression is associated with tumor aggressiveness in lung cancer patients is in agreement with its reported role in promoting cell proliferation; in MCF-7 adenocarcinoma and glioblastoma cells, its expression correlated with enhanced growth rates (23, 39). In some of these studies, its mechanism of action appeared to be through modulation of cell-cycle components (21, 40) and association with the complex of cyclin E and the cell cycle-dependent kinase 2 (CDK2) (41). PKC $\eta$  elevated expression was also found to be associated with tumor progression of renal cell carcinoma (42) and with hyperplasia in prostate carcinoma (43). In breast cancer, positive correlation was shown between PKC $\eta$  overexpression and invasion to lymph nodes (38). Our results suggest that PKC $\eta$  plays an important role in tumor and metastasis progression, although the underlying molecular mechanism is currently unknown. Indeed, here we show that PKC $\eta$  expression increases with stage progression and predicts lower survival rates. This suggests its role during late clinical stages. This is unlike PKC $\iota$  which was reported to predict risk of relapse in early stages (13). It is therefore possible that different PKC isoforms affect different stages of NSCLC.

Previous studies have shown that PKC $\eta$  confers protection against DNA damage induced by chemotherapeutic agents (33, 34, 44). Moreover, we have recently shown that increased expression and localization of PKC $\eta$  in cellular membranes correlated with poor clinical outcome in patients with locally advanced breast cancer undergoing neoadjuvant chemotherapy (consisting of cyclophosphamide, doxorubicin and fluorouracil, CAF protocol) (45). It is therefore possible that the increased risk for an event seen in the present study for patients with high H-Scores resulted from the resistance to chemotherapy conferred by PKC $\eta$ . Accordingly, the better prognosis during the first 12 months after diagnosis, seen in

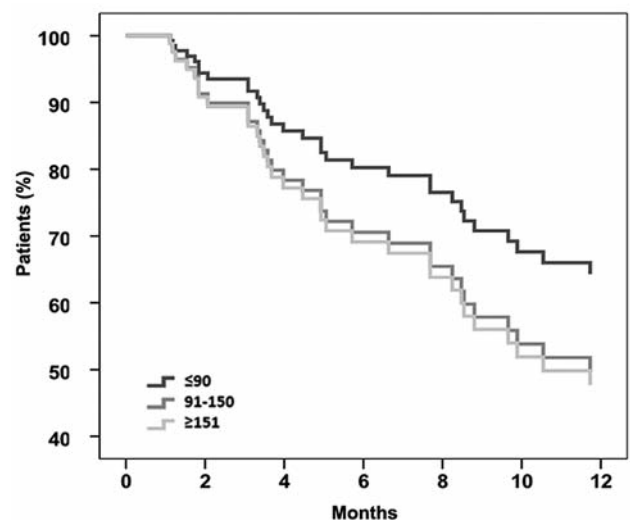


Figure 4. The risk for an event decreases in patients with tumors exhibiting low PKC $\eta$  expression. The variables included in the multivariate model for prediction of an event within one year (relapse or death) were stage (hazard ratio=2.36,  $p<0.001$ ) and PKC $\eta$  expression (hazard ratio=1.45,  $p=0.06$ ), as described in the Materials and Methods.

patients exhibiting low PKC $\eta$  expression, could be related to their greater response to chemotherapy. Thus, our findings suggest that PKC $\eta$  may serve as a predictive factor for patients who will benefit from chemotherapy, especially in the setting of patients with early lung cancer and adjuvant chemotherapy.

Our study suggests that PKC $\eta$  could be a novel prognostic biomarker in patients with NSCLC. Its expression correlates with the stage of the disease. Once diagnosed with lung cancer, PKC $\eta$  may predict outcome; patients of the third tertile were at greater risk for an event within one year of diagnosis. When adjusted for stage, patients with PKC $\eta$

overexpression tended to exhibit an increased risk of death within the first year after diagnosis. A larger study group is needed in order to confirm this observation. Due to the biological importance of PKC $\eta$  for tumor progression and resistance, the use of specific kinase inhibitors may potentially benefit patients with NSCLC.

## Acknowledgements

This study was supported by the Israel Science Foundation (ISF grant No. 1413/10).

## References

- Minna JD, Roth JA and Gazdar AF: Focus on lung cancer. *Cancer Cell* 1: 49-52, 2002.
- Osada H and Takahashi T: Genetic alterations of multiple tumor suppressors and oncogenes in the carcinogenesis and progression of lung cancer. *Oncogene* 21: 7421-7434, 2002.
- Nishizuka Y: Protein kinase C and lipid signaling for sustained cellular responses. *FASEB J* 9: 484-496, 1995.
- Dempsey EC, Newton AC, Mochly-Rosen D, Fields AP, Reyland ME, Insel PA and Messing RO: Protein kinase C isozymes and the regulation of diverse cell responses. *Am J Physiol Lung Cell Mol Physiol* 279: L429-438, 2000.
- Schuller HM, Orloff M and Reznik GK: Inhibition of protein kinase C-dependent cell proliferation of human lung cancer cell lines by the dihydropyridine dextrigulipine. *J Cancer Res Clin Oncol* 120: 354-358, 1994.
- Regala RP, Weems C, Jamieson L, Khor A, Edell ES, Lohse CM and Fields AP: Atypical protein kinase C  $\iota$  is an oncogene in human non-small cell lung cancer. *Cancer Res* 65: 8905-8911, 2005.
- Bae KM, Wang H, Jiang G, Chen MG, Lu L and Xiao L: Protein kinase C epsilon is overexpressed in primary human non-small cell lung cancers and functionally required for proliferation of non-small cell lung cancer cells in a p21/Cip1-dependent manner. *Cancer Res* 67: 6053-6063, 2007.
- Volm M and Pommerenke EW: Associated expression of protein kinase C with resistance to doxorubicin in human lung cancer. *Anticancer Res* 15: 463-466, 1995.
- Jull BA, Plummer HK III and Schuller HM: Nicotinic receptor-mediated activation by the tobacco-specific nitrosamine NNK of a Raf-1/MAP kinase pathway, resulting in phosphorylation of c-myc in human small cell lung carcinoma cells and pulmonary neuroendocrine cells. *J Cancer Res Clin Oncol* 127: 707-717, 2001.
- Mellor H and Parker PJ: The extended protein kinase C superfamily. *Biochem J* 332: 281-292, 1998.
- Liu Y, Wang B, Wang J, Wan W, Sun R, Zhao Y and Zhang N: Down-regulation of PKC $\zeta$  expression inhibits chemotaxis signal transduction in human lung cancer cells. *Lung Cancer* 63: 210-218, 2009.
- Wang Y, Yang H, Liu H, Huang J and Song X: Effect of staurosporine on the mobility and invasiveness of lung adenocarcinoma A549 cells: an *in vitro* study. *BMC cancer* 9: 174, 2009.
- Fields AP, Frederick LA and Regala RP: Targeting the oncogenic protein kinase C $\iota$  signalling pathway for the treatment of cancer. *Biochem Soc Trans* 35: 996-1000, 2007.
- Gorin MA and Pan Q: Protein kinase C epsilon: an oncogene and emerging tumor biomarker. *Mol Cancer* 8: 1-9, 2009.
- Ding L, Wang H, Lang W and Xiao L: Protein kinase C-epsilon promotes survival of lung cancer cells by suppressing apoptosis through dysregulation of the mitochondrial caspase pathway. *J Biol Chem* 277: 35305-35313, 2002.
- Bacher N, Zisman Y, Berent E and Livneh E: Isolation and characterization of PKC-L, a new member of the protein kinase C-related gene family specifically expressed in lung, skin, and heart. *Mol Cell Biol* 11: 126-133, 1991.
- Osada S, Hashimoto Y, Nomura S, Kohono Y, Chida K, Tajima O, Kubo K, Akimoto K, Koizumi H, Kitamura Y, Suzuki K, Ohno S and Kuroki T: Predominant expression of nPKC $\eta$ , a Ca<sup>2+</sup>-independent isoform of protein kinase C in epithelial tissues, in association with epithelial differentiation. *Cell Growth and Diff* 4: 167-175, 1993.
- Gherzi R, Sparatore B, Patrone M, Sciutto A and Briata P: Protein kinase C mRNA levels and activity in reconstituted normal human epidermis: relationship to cell differentiation. *Biochem Biophys Res Commun* 184: 283-291, 1992.
- Denning MF, Dlugosz AA, Williams EK, Szallasi Z, Blumberg PM and Yuspa SH: Specific protein kinase C isozymes mediate the induction of keratinocyte differentiation markers by calcium. *Cell Growth and Diff* 6: 149-157, 1995.
- Ohba M, Ishino K, Kashiwagi M, Kawabe S, Chida K, Huh N and Kuroki T: Induction of differentiation in normal human keratinocytes by adenovirus-mediated induction of  $\eta$  and  $\delta$  isoforms of protein kinase C. *Mol Cell Biol* 18: 5199-5207, 1998.
- Livneh E, Shimon T, Bechor E, Doki Y, Schieren I and Weinstein IB: Linking protein kinase C to the cell cycle: ectopic expression of PKC $\eta$  in NIH-3T3 cells alters the expression of cyclins and CDK inhibitors and induces adipogenesis. *Oncogene* 12: 1545-1555, 1996.
- Hussaini IM, Karns LR, Vinton G, Carpenter JE, Redpath GT, Sando JJ and Vandenberg SR: Phorbol 12-myristate 13-acetate induces protein kinase C $\eta$ -specific proliferative response in astrocytic tumor cells. *J Biol Chem* 275: 22348-22354, 2000.
- Fima E, Shtutman M, Libros P, Missel A, Shahaf G, Kahana G and Livneh E: PKC $\eta$  enhanced cell cycle progression, the expression of G1 cyclins and p21 in MCF-7 cells. *Oncogene* 20: 6794-6804, 2001.
- Beck J, Bohnet B, Brugger D, Bader P, Dietl J, Scheper RJ, Kandolf R, Liu C, Niethammer D and Gekeler V: Multiple gene expression analysis reveals distinct differences between G2 and G3 stage breast cancers, and correlations of PKC $\eta$  with MDR1, MRP and LRP gene expression. *Br J Cancer* 77: 87-91, 1998.
- Beck JF, Bohnet B, Brugger B, Dietl J and Scheper RJ: Expression analysis of protein kinase C isoenzymes and multidrug resistance associated genes in ovarian cancer cells. *Anticancer Res* 18: 701-705, 1998.
- Beck JF, Handgretinger R, Klingebiel T, Dopfer R, Schaich M and Ehninger G: Expression of PKC isozyme and MDR-associated genes in primary and relapsed state AML. *Leukemia* 10: 426-433, 1996.
- Sonnemann J, Gekeler V, Ahlbresht K, Brischwein K, Liu C, Bader P, Muller C, Niethammer D and Beck JF: Down-regulation of protein kinase C $\eta$  by antisense oligonucleotides sensitises A549 lung cancer cells to vincristine and paclitaxel. *Cancer Lett* 209: 177-185, 2004.

- 28 Basu A: The involvement of novel protein kinase C isoenzymes in influencing sensitivity of breast cancer MCF-7 cells to tumor necrosis factor- $\alpha$ . *Mol Pharmacol* 53: 105-111, 1998.
- 29 Akkaraju GR and Basu A: Overexpression of protein kinase C- $\eta$  attenuates caspase activation and tumor necrosis factor  $\alpha$ -induced cell death. *Biochem Biophys Res Commun* 279: 103-107, 2000.
- 30 Hussaini IM, Carpenter JE, Redpath GT, Sando JJ, Shaffrey ME and VandenBerg SR: Protein kinase C- $\eta$  regulates resistance to UV and  $\gamma$ -irradiation-induced apoptosis in glioblastoma cells by preventing caspase-9 activation. *Neuro-Oncol* 4: 9-21, 2002.
- 31 Matsumura M, Tanaka N, Kuroli T, Ichihashi M and Ohba M: The  $\eta$  isoform of protein kinase C inhibits UV-induced activation of caspase-3 in normal human keratinocytes. *Biochem Biophys Res Commun* 303: 350-356, 2003.
- 32 Sonnemann J, Gekeler V, Sagrauske A, Muller C, Hofmann HP and Beck JF: Down-regulation of PKC $\eta$  potentiates the cytotoxic effects of exogenous tumor necrosis factor-related apoptosis-inducing ligand in PC-3 prostate cancer cells. *Mol Cancer Ther* 3: 773-781, 2004.
- 33 Rotem-Dai N, Oberkovitz G, Abu-Ghanem S and Livneh E: PKC $\eta$  confers protection against apoptosis by inhibiting the pro-apoptotic JNK activity in MCF-7 cells. *Exp Cell Res* 315: 2616-2623, 2009.
- 34 Abu-Ghanem S, Oberkovitz G, Benharroch D, Gopas J and Livneh E: PKC $\eta$  expression contributes to the resistance of Hodgkin's lymphoma cell lines to apoptosis. *Cancer Biology & Therapy* 6: 1375-1380, 2007.
- 35 Sion-Vardy N, Freedman J, Lazarov I, Bolotin A and Ariad S: p27<sup>kip1</sup> Expression in non-small cell lung cancer is not an independent prognostic factor. *Anticancer Res* 30: 3699-3704, 2010.
- 36 McClelland RA, Finlay P, Walker KJ, Nicholson D, Robertson JF, Blamey RW and Nicholson RI: Automated quantitation of immunocytochemically localized estrogen receptors in human breast cancer. *Cancer Res* 50: 3545-3550, 1990.
- 37 Ye H, Ma WL, Jin S, Liu SY, Wang DX and Hu QH: Cigarette smoke extract activates PKC isoforms and down-regulates the expressions of potassium channels BK(Ca) and Kv1.5 in rat bronchial smooth muscle cells. *Sheng Li Xue Bao* 60: 709-714, 2008.
- 38 Masso-Welch PA, Winston JS, Edge S, Darcy KM, Asch H, Vaughan MM and Ip MM: Altered expression and localization of PKC $\eta$  in human breast tumors. *Breast Cancer Res Treat* 68: 211-223, 2001.
- 39 Martin PM and Hussaini IM: PKC $\eta$  as a therapeutic target in glioblastoma multiforme. *Expert Opin Ther Targets* 9: 1-15, 2005.
- 40 Livneh E and Fishman DD: Linking protein kinase C to cell cycle control. *Eur J Biochem* 248: 1-9, 1997.
- 41 Shtutman M, Hershko T, Maissel A, Fima E and Livneh E: PKC $\eta$  associates with cyclin E/CDK2 complex in serum-starved MCF-7 and NIH-3T3 cells. *Exp Cell Res* 286: 22-29, 2003.
- 42 Brenner W, Faber G, Herget T, Wiesner C, Hengstler JG and Thuroff JW: Protein kinase C  $\eta$  is associated with progression of renal cell carcinoma. *Anticancer Res* 23: 4001-4006, 2003.
- 43 Koren R, Ben Meir D, Langzam L, Dekel Y, Baniel J, Livne PM, Gal R and Sampson S: Expression of protein kinase C isoenzymes in benign hyperplasia and carcinoma of prostate. *Oncol Rep* 11: 321-326, 2004.
- 44 Tamarkin A, Zurgil U, Braiman A, Hai N, Krasnitsky E, Maissel A, Ben-Ari A, Yankelovich L and Livneh E: DNA damage targets PKC $\eta$  to the nuclear membrane *via* its C1B domain. *Exp Cell Res* 10: 1465-1475, 2011.
- 45 Karp G, Abu-Ghanem S, Novak V, Mermerstain W, Ariad S, Sion-Vardy N and Livneh E: Localization of PKC $\eta$  in cell membranes as a predictor for breast cancer response to treatment. *Onkologie* In Press, 2012.

*Received February 15, 2012*

*Revised March 13, 2012*

*Accepted March 14, 2012*