

Identification of The Distinctive Type *i/XhoI+* Strain of Epstein-Barr Virus in Gastric Carcinoma in Peru

PAULA ORDONEZ^{1†}, CHIHAYA KORIYAMA¹, SHAN DING^{1‡}, ELENA YOSHIWARA²,
ALEJANDRO H. CORVALAN³, JUAN TAKANO², JESUS L. CHIRINOS⁴, JOSE WATANABE²,
JUAN MIYAGUI², HERIBERTO HIDALGO⁵, PEDRO CHACON⁶,
VICTOR LINARES⁷, YOSHITO EIZURU⁸ and SUMINORI AKIBA¹

¹Department of Epidemiology and Preventive Medicine, Kagoshima University
Graduate School of Medical and Dental Sciences, Kagoshima, Japan;

²Department of Pathology, Peruano Japonés Polyclinic, Lima, Peru;

³Department of Hematology-Oncology, Pontificia Universidad Católica de Chile, Santiago, Chile;

⁴Academic Department of Public Health and Epidemiology, Faculty of Public Health,
Universidad Peruana Cayetano Heredia, Lima, Peru;

⁵Hermilio Valdizan Medrano Regional Hospital, Huanuco, Peru;

⁶Edgardo Rebagliati Martins Hospital, Lima, Peru;

⁷Chiclayo Regional Hospital, Peru;

⁸Division of Oncogenic and Persistent Viruses, Center for Chronic Viral Diseases,
Kagoshima University Graduate School of Medical and Dental Sciences, Kagoshima, Japan

Abstract. Aim: To clarify the reason for the low frequency of Epstein-Barr virus-associated gastric carcinoma (EBVaGC) in Peru, despite the high frequency reported in neighboring countries, the distribution of the distinctive EBV (type *i/XhoI+*) strain in EBVaGC and a healthy population was examined. Materials and Methods: EBV polymorphisms in *BamHI* W1/I1 and *XhoI* restriction site of the latent membrane protein 1 gene (*LMP1*) were examined among 11 EBVaGCs and 172 healthy controls from Peru, and these frequencies were compared with those in a previous study of Chile and Colombia (n=303). Results: The frequency of the distinctive EBV strain in EBVaGCs (55%) was significantly higher than that in controls (7%). Furthermore, the frequency of this EBV type in Peruvian controls was

significantly lower than that in controls from Chile and Colombia (27%, $p < 0.001$). Conclusion: The low frequency of the distinctive EBV strain among the Peruvian population might be a reason for the lower incidence of EBVaGC in Peru, as compared with neighboring countries.

Epstein-Barr virus (EBV) is associated with lymphoid malignancies and epithelial malignancies, including some gastric carcinomas (1, 2). Previous studies have revealed that the proportion of EBV-associated gastric carcinoma (EBVaGC) was different from country to country, ranging from 2 to 17% (1, 3-10), and the EBVaGC frequency was relatively higher in the American continent than in European and Asian countries. In Latin America, a low frequency of EBVaGC was reported in Lima, Peru (4%) (9), although relatively high frequencies of EBVaGC were observed in the neighboring countries, Chile (17%) and Colombia (16%) (3, 8).

Genetic polymorphisms of prevailing EBV differ among countries. Recently, a significantly high frequency of the distinctive EBV strain type *i/XhoI+* in EBVaGC cases from Chile and Colombia was reported in comparison with healthy populations (11). This strain is determined by polymorphisms of *BamHI* W1/I1 boundary regions and exon 1 of the latent membrane protein 1 gene (*LMP1*). There are two polymorphisms at *BamHI* W1/I1 boundary regions: types I and i. Type I, without a *BamHI* restriction site, predominated among healthy carriers and EBV-associated disease in Japan and China (12, 13). On the other hand, type i, with a *BamHI*

[†]Present address: Division of Antiviral Chemotherapy, Center for Chronic Viral Diseases, Kagoshima University, Japan; [‡]Present address: Drug Safety Research Laboratories, Shin Nippon Biomedical Laboratories Ltd., Kagoshima, Japan.

Correspondence to: Chihaya Koriyama, Department of Epidemiology and Preventive Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, Sakuragaoka 8-35-1, Kagoshima 890-8544, Japan. Tel: +81 992755296, Fax: +81 992755299, e-mail: fiy@m.kufm.kagoshima-u.ac.jp

Key Words: EBV polymorphism, *BamHI* W1/I1, *XhoI* restriction site, EBV-associated gastric cancer, Latin America.

Table I. List of primers used in the present study.

		Sequence 5'-3'	Reference
<i>Bam</i> HI-W1/I1	Sense	ACCTGCTACTCTTCGGAAAC	Lung <i>et al.</i> (12, 25)
	Antisense	TCTGTCACAACCTCACTGTC	
<i>Xho</i> I site in <i>LMP1</i>	Sense	GGAACACGACCTTGAGAGG	Sandvej <i>et al.</i> (26)
	Antisense	AACAGCAGCGCCAAGAGGAG	

restriction site, prevailed in healthy donors and EBV-associated disease in Western countries (14). The polymorphism in the *Xho*I restriction site at exon 1 of the *LMP1* gene is indicated by a G→T mutation resulting in loss of an *Xho*I restriction site. These genotypes are denoted as *Xho*I+ and *Xho*I- (also known as *Xho*I kept and *Xho*I loss), respectively. The *Xho*I+ variant is frequently observed in Western countries (14), while the *Xho*I- variant is common in Asia (15).

In EBVaGC, at least five EBV genes are expressed in carcinoma cells: EBV-encoded RNAs (EBERs), Epstein-Barr nuclear antigen 1 (*EBNA1*), latent membrane protein 2 (*LMP2*), *Bam*HI A rightward transcript 0 (*BARF0*), and *BARF1* (16). Although EBV was able to immortalize human gastric primary epithelial cells (17), *LMP1*, a well-known viral oncogene in other EBV-related malignancies, was only rarely expressed in EBVaGC (18, 19). Recent studies have proposed that EBER1 (20), *LMP2A* (21), and *BARF1* (22) play important role(s) in the carcinogenesis of the stomach. Interestingly, these potentially oncogenic viral products are encoded by neighboring regions of the *Bam*HI W1/I1 boundary region and exon 1 of the *LMP1* gene.

These findings suggest that the geographical variance of EBVaGC frequency may be explained by different distributions of EBV variants around the world. In the present study, to clarify the reason for the low frequency of EBVaGC in Peru, the prevalence of the distinct EBV strain, type i/*Xho*I+, in patients with EBVaGC and non-cancer controls in Peru was examined. In addition, frequencies of this distinctive EBV strain were compared with controls from Peru and the neighboring countries, Colombia and Chile.

Materials and Methods

Patients and samples. Paraffin-embedded tumor specimens obtained from 11 EBVaGC cases (6 males and 5 females) were used. The EBV presence was examined previously (9) using an *in situ* hybridization assay with oligonucleotide probes specific to detect EBER1 in gastric tissue, as described before (23). In 2006, throat-washing specimens were obtained from 196 non-cancer individuals, with informed consent, in Lima, Peru. Thirteen were excluded since demographic information was missing. DNA specimens of the other 183 participants were subjected to viral genotype analysis. However, 11 were excluded because of low quality and/or an insufficient amount of DNA for viral typing. Thus, the number of controls was

172, aged between 20 and more than 80 years old (64 males and 108 females). Throat-washing samples were collected by gargling with 15 ml of phosphate-buffered saline (PBS), which was then stored at -20°C until use. The Institutional Review Boards of Kagoshima University Graduate School of Medical and Dental Sciences, the Peruano Japonese Polyclinic, Hermilio Valdizan Medrano Regional Hospital, Edgardo Rebagliati Martins Hospital and Chiclayo Regional Hospital previously approved this study.

DNA extraction. Paraffin-embedded specimens were cut into 10-µm-thick slices, treated with xylene and ethanol and centrifuged at 22,000× *g* for 20 min. The resulting pellets were re-suspended in 100 µl of extraction buffer following a method described before (24). Throat-washing specimens were centrifuged at 16,000× *g* for 50 min, and the pellets re-suspended in 100 µl of extraction buffer (1 M Tris, pH 8.0, 50 mM EDTA, 0.5% Tween 20). The suspended samples were treated with 200 µg ml⁻¹ of Proteinase K at 37°C overnight and boiled at 100°C for 10 min for Proteinase K inactivation. Samples were then subjected to phenol/chloroform extraction and ethanol precipitation. Finally, the extracted DNA was dissolved in 50 µl of TE buffer [10 mM Tris/HCl (pH 8.0) and 1 mM EDTA] and kept at -20°C until amplification.

Polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP) analysis. The polymorphisms of *Bam*HI W1/I1 boundary region and *Xho*I restriction site at exon 1 of the *LMP1* gene were investigated as reported previously (11). In brief, a 205-bp fragment of the *Bam*HI W1/I1 boundary region was amplified using the primers reported by Lung *et al.* (25). Analysis of *Xho*I restriction site polymorphism in exon 1 of *LMP1* gene was performed with a primer set to amplify a 113-bp fragment (26). The sequences of primers are shown in Table I. The *Bam*HI W1/I1 boundary region and *Xho*I restriction site of *LMP1* were amplified by PCR using 2-5 µl of DNA template in a 25 µl reaction mixture of PCR buffer, Ampdirect Plus (Shimazu, Japan) (10 mM Tris-HCl pH 8.0, 35 mM KCl, 1.5 mM MgCl₂), containing 200 µM dNTPs, 1 µM of each primer and 1.25 U NovaTaq Polymerase (Shimazu, Japan). Amplification cycles for these two regions were performed as described before (11). Restriction enzyme digestion was performed in 20 µl volumes with 10 U of restriction enzyme for 1 h at 37°C. After *Bam*HI restriction enzyme digestion, type i was determined by the presence of *Bam*HI restriction site, producing fragments of 130-bp and 75-bp length. Digestion with *Xho*I restriction enzyme resulted in 67- and 46-bp fragments for the *Xho*I+ type, and the undigested 113-bp PCR product indicated the *Xho*I- type. Whole and digested PCR products were confirmed by electrophoresis in a 3% agarose gel and by staining with 0.5 µg ml⁻¹ of ethidium bromide. The B95-8 cell line, initiated by exposing marmoset blood leukocytes to EBV extracted from a human leukocyte line, served as positive control for type I and *Xho*I+ strain. A DNA

Table II. Age, gender and Epstein-Barr virus genotype distribution in EBVaGC cases and non-cancer controls.

	EBVaGC (N=11)		TW (N=135)		p-Value*
	N	%	N	%	
Gender					
Female	5	45	82	61	0.320
Male	6	55	53	39	
Age (years)					
40-49	1	9	15	11	0.114
50-59	3	27	25	19	
60-69	4	36	15	11	
70-79	2	18	65	48	
80+	1	9	15	11	
BamHI W1/I1					
I	5	45	101	75	0.001
i	6	55	17	13	
ND	0	0	17	13	
XhoI site					
-	2	18	105	78	<0.001
+	9	82	17	13	
ND	0	0	13	10	
Combination of BamHI W1/I1 & XhoI site					<0.001
I/-	2	18	83	61	
I/+	3	27	5	4	
i/-	0	0	8	6	
i/+	6	55	9	7	
ND	0	0	30	22	

EBVaGC: Epstein-Barr virus-associated gastric carcinoma; TW: throat washing of non-cancer controls; ND: not determined. *Obtained by χ^2 test.

sample of EBVaGC, whose genotype was determined in the previous study, was used as control for type i and XhoI-.

Statistical analysis. For a case-control comparison in Peru, non-cancer controls were limited to those aged 40 or over, since all cases with EBVaGC were over 40. Furthermore, distributions of EBV polymorphisms in all Peruvian controls were compared with the healthy controls from Chile and Colombia reported in the previous study (11). The associations between EBV polymorphisms and the risk of EBVaGC were analyzed using the chi-square test and logistic regression models. Maximum likelihood estimates of odds ratios (ORs) and corresponding 95% confidence intervals (CIs) were obtained using a logistic regression model. A p-value less than 0.05 was considered to be statistically significant.

Results

There was no statistically significant differences in gender and age distributions between EBVaGC cases and non-cancer controls (Table II). Polymorphisms of BamHI W1/I1 region were determined in all EBVaGCs and 118 controls (87%), and the frequency of type i was significantly higher in

Table III. Co-segregation analysis of BamHI-I and XhoI restriction sites in LMP1 gene in EBVaGC cases and non-cancer controls.

BamHI-W1/I1	XhoI site	OR (95% CI)*	Adjusted OR (95% CI)†
I	-	1.0 (reference)	1.0 (reference)
	+	25 (3.4-185)	29 (3.3-263)
i	-	-	-
	+	28 (4.8-158)	39 (4.7-320)

OR: Odds ratio; CI: confidence interval. *OR and 95% CI calculated by logistic regression model. †OR and 95% CI obtained by exact method after adjusting for the effects of age and sex.

EBVaGC (55%) than in controls (13%). Regarding the presence of XhoI restriction site in LMP1, the polymorphism in all EBVaGC cases and 122 controls (90%) was determined. The frequency of EBV with XhoI+ was significantly higher in EBVaGC (82%) than in controls (13%). The sequence of the XhoI variants was further analyzed in all EBVaGC cases by direct sequencing of the 113-bp amplified fragment of the XhoI region in exon 1 of LMP1. All XhoI+ variants showed the XhoI restriction site by sequencing (data not shown).

In a case-control comparison of the combination of these polymorphisms, the frequency of type i/XhoI+ strain in EBVaGC (55%) was significantly higher than that of controls (7%). This frequency was not changed when controls under 40 were included (7%). After adjusting for the effects of age and gender distributions, the EBVaGC risk for carrying this distinct strain was statistically significant (Table III). The OR of type I/XhoI+ was also significantly high in reference to the EBVaGC risk of type I/XhoI-, which was the most frequent strain among controls.

The frequency of EBV polymorphisms in all Peruvian non-cancer controls (n=172) was compared with those from Chile and Colombia (Table IV). In this comparison, 26 controls were excluded because the quality and quantity of DNA was not guaranteed for viral typing. Thus, 303 controls from Chile and Colombia were used in the present analysis. Among Peruvian controls, the most prevalent type was type I/XhoI- (52%). The frequency of EBV type i/XhoI+ was the lowest (7%). On the other hand, the distinctive type i/XhoI+ (27%) was as common as other types among Chilean and Colombian controls. Note that the distribution pattern of EBV type varies across age groups, and type i/XhoI+ tends to prevail among younger individuals in both populations (Table V). The difference in the frequency of this distinctive type between controls from Peru (15%) and Chile/Colombia (29%) still existed when the data were limited to those younger than 50 (p=0.043 by Chi-square test). In contrast, EBV type I/XhoI- was the most frequent type among older individuals, over 50 years of age, in Peru. Although the

Table IV. Age, gender and Epstein-Barr virus genotype distribution in non-cancer subjects from Peru, Chile and Colombia.

	Chile & Colombia* (N=303)		Peru (N=172)		p-Value†
	N	%	N	%	
Gender					
Female	159	52	108	63	0.029
Male	144	48	64	37	
Age (years)					
20-29	203	67	26	15	<0.001
30-39	50	17	11	6	
40-49	32	11	15	9	
50-59	15	5	25	15	
60-69	3	1	15	9	
70-79	0	0	65	38	
80+	0	0	15	9	
BamHI W1/I1					
I	85	28	116	67	<0.001
i	169	56	24	14	
ND	49	16	32	19	
XhoI site					
-	92	30	118	69	<0.001
+	140	46	38	22	
ND	71	23	16	9	
Combination of BamHI W1/I1 & XhoI site					<0.001
I/-	23	8	89	52	
I/+	46	15	13	8	
i/-	60	20	12	7	
i/+	82	27	12	7	
ND	92	30	46	27	

ND: Not determined. *Reported by Corvalan *et al.* (11). †Obtained by χ^2 test.

number of controls over 50 years old was limited in Chile and Colombia, the EBV type I/XhoI- has the lowest frequency (one out of 18, 6%). After adjusting for age and gender distributions, the heterogeneity of the four combinations of EBV polymorphisms between Peru and Chile/Colombia was statistically significant ($p < 0.001$). When non-type i/XhoI+ was used as a reference group, however, the difference in the frequency of the distinctive strain was not statistically significant ($p = 0.483$).

Discussion

In the present study, genetic polymorphisms of EBV in BamHI W1/I1 boundary regions and the XhoI restriction site at exon 1 of the LMPI gene were examined among 11 EBVaGC tissue specimens and 172 throat-washing samples obtained from non-cancer individuals in Peru, where the prevalence of EBVaGC is lower than in other Latin

American countries (3, 8, 9). As was reported in the previous study (11), the frequency of EBV type i/XhoI+ in EBVaGC cases from Colombia and Chile (55%) was higher than that of non-cancer controls (7%). More recently, a high frequency of this strain was also reported in Iranian EBVaGC (five out of nine EBVaGC), although its prevalence in the general population of Iran is unknown (10). On the other hand, this EBV type was not detected in a recent study of EBVaGC conducted in Guangzhou, Southern China. The EBV type I/XhoI- was the most predominant type in Chinese EBVaGC cases (75.6%) (27). In the present study, the frequency of EBV type I/XhoI+ in Peruvian EBVaGC (27%) was also significantly higher than that of non-cancer controls (4%). However, as the frequency of this strain in EBVaGC was very low (1.6%) in the previous study (11), and the sample size in this study is small, this result should be interpreted cautiously.

Although the findings in the present study support the hypothesis, proposed by Corvalan *et al.* (11), that the EBV type i/XhoI+ is related to the risk of EBVaGC, there are some discrepancies between the results of the two studies. Among Chilean and Colombian EBVaGC, most of the cases (92%; 59 out of 64 determined) harbored the distinctive EBV strain (type i/XhoI+). In the present study, however, only six out of nine EBVaGC cases had this type. The difference was statistically significant, even after adjusting for the effects of age and gender distributions ($p = 0.019$). This was true in a comparison with non-cancer controls ($p < 0.001$). The EBV type I/XhoI- was the most frequent type among older Peruvians, those over 50 years old, but the frequency of this type was the lowest in older Chilean and Colombian controls (Table V). The point to notice here is that there are 1 to 12-year intervals between the study periods of EBVaGC cases and controls. In the case of Peru, the intervals were 5-12 years, since the sampling periods of EBVaGC cases and throat washing specimens were 1994-2001 (9) and 2006, respectively. Since the mean age of Peruvian EBVaGC cases was 62.5 years (95% CI, 53.8-71.1 years), the corresponding background populations are approximately 70 years old. In a similar way, the intervals of sampling in Chile and Colombia were 1-8 years. In these countries, the sampling periods of EBVaGC cases and throat-washing specimens were 1993-1999 (3, 8) and 2000-2001, respectively. The distribution pattern of EBV type in individuals in their early sixties should be regarded as background populations because the mean age of EBVaGC cases in Chile and Colombia was 58.1 years (95% CI, 54.8-61.4 years). Thus, the distribution pattern of EBV type in control populations significantly differed between Peru and Chile/Colombia, even taking account of sampling intervals. This might be one of the reasons for the low EBVaGC frequency in Peru. This hypothesis should be confirmed in other cohorts with different incidence rates of EBVaGC.

Table V. Distribution patterns of the four EBV types, determined in *BamHI-I* and *XhoI* restriction site in *LMP1* genes, by age in non-cancer controls from Peru, and Chile and Colombia.

Country	Age group (years)	N	EBV genotype (%)				
			Type I/ <i>XhoI</i> -	Type I/ <i>XhoI</i> +	Type i/ <i>XhoI</i> -	Type i/ <i>XhoI</i> +	ND
Peru	20-29	26	4 (15)	8 (31)	2 (8)	0	12 (46)
	30-49	26	5 (19)	1 (4)	4 (15)	8 (31)	8 (31)
	50-69	40	28 (70)	1 (3)	4 (10)	3 (8)	4 (10)
	70+	80	52 (65)	3 (4)	2 (3)	1 (1)	22 (28)
Chile & Colombia	20-29	203	11 (5)	31 (15)	34 (17)	73 (36)	54 (27)
	30-49	82	11 (13)	13 (16)	19 (23)	9 (11)	30 (37)
	50-69	18	1 (6)	2 (11)	7 (39)	0	8 (44)
	70+	0	-	-	-	-	-

ND: Not determined.

The observations of the present and previous studies suggest a possibility that particular EBV strains, variants in neighboring regions of *BamHI* W1/I1 and *LMP1*, may be related to EBVaGC development, transformation ability or cytotoxic immune responses. Note that the polymorphisms examined in this study are located in regions near genes encoding potential oncogenic viral products: *LMP1*, *LMP2A*, *BARF1*, and EBERs. Although *LMP1* is an important EBV oncoprotein in epithelial malignancies (28, 29), it is rarely expressed in EBVaGC (18).

An *XhoI* restriction site is also located in the intron between exons 1 and 2 of *LMP2A*. Tanaka *et al.* (30) reported that EBV strains in EBVaGC tended to have *LMP2A* gene with threonine substitution at codon 348, which corresponds to an HLA A-11-restricted cytotoxic T lymphocyte (CTL) epitope, in comparison with EBV strains in healthy individuals. This substitution may confer an advantage for viral persistence in tumor cells. *LMP2A* up-regulates the cellular survivin gene, through the NF- κ B pathway (21), and DNA methyltransferase 1 (31), which leads to anti-apoptotic effects and an increase in methylation of the phosphatase and tensin homolog gene (*PTEN*) promoter, respectively. Recently, a possible sequence of events within the epithelial mucosa was proposed in which EBV infection is followed by expression of viral latent genes, abnormal signal pathway in host cells, DNA methylation-mediated repression of tumor suppressor genes, and the growth of the predominant clone by the interaction with other etiologic factors (32).

BARF1 is frequently expressed in EBVaGC (16, 33), and is able to immortalize primary monkey and human epithelial cells *in vitro* (34, 35). Transfection of *BARF1* into the rodent fibroblast cell line BALB/c 3T3, or into the EBV-negative B-cell line, Louckes, resulted in tumorigenic transformation (36, 37). In EBVaGC, *BARF1* was reported to play an anti-apoptotic role (22), and to induce cyclin D1 overexpression (38).

EBERs are widely expressed in EBVaGC, suggesting an important role in the development and maintenance of this carcinoma. According to the previous studies, EBERs played roles in colony formation and tumorigenicity in immunodeficient mice, had anti-apoptotic effects in Burkitt lymphoma cells (39), and caused induction of an anti-inflammatory cytokine IL-10 through retinoic acid-inducible gene I-mediated IRF-3 (40). Iwakiri *et al.* (41) reported that EBER1 induced the secretion of the insulin-like growth factor 1, which plays a role as an autocrine growth factor in EBVaGC.

The associations between cancer risk and EBV polymorphisms in these regions, *BamHI* W1/I1 and *XhoI* restriction site at exon 1 of the *LMP1* gene, were also reported in other types of EBV-related cancer. The polymorphism in the *XhoI* restriction site of *LMP1* is strongly suspected to relate to the etiology of nasopharyngeal carcinoma (42). EBV with *XhoI* loss has been found in 97-100% of Chinese nasopharyngeal carcinoma, whereas such a subtype was detected in only 30-40% of the throat-washing specimens obtained from healthy Chinese (43, 44). NK/T-cell lymphoma, an aggressive type of non-Hodgkin's lymphoma, is also associated with EBV infection, and striking geographical variations are known. Recently, Cabrera *et al.* (45) suggested an association between a recombinant EBV strain (type i/*XhoI*-) and the risk of nasal NK/T-cell lymphoma in Chile.

These findings suggest a possibility that either variant of *BamHI* W1/I1 or *XhoI* restriction site at exon 1 of *LMP1* can increase the risk of EBV-related malignancies. In fact, an increase of the EBVaGC risk in those with EBV type I/*XhoI*+ was observed in this study (Table III). Interestingly, EBVaGC cases with either EBV variant were younger than those with the prototype of EBV, type I/*XhoI*-. For Peruvian cases, the average/median age of EBVaGC cases with type I/*XhoI*-, type I/*XhoI*+, and type i/*XhoI*+ was 68/68, 62/66, and 61/56 years, respectively. The average/median age of non-EBVaGC

cases was 65/67 years. Although this age difference was not statistically significant ($p=0.350$, non-EBVaGC vs. EBVaGC with either variant type), similar trends were also observed for Chilean and Colombian gastric cancer cases (data not shown). Thus, these findings suggest that EBV types with either or both variants of *BamHI* W1/I1 and *XhoI* restriction site at exon 1 of the *LMP1* may increase the risk of early development of EBVaGC.

In a recent study conducted by Chen *et al.* (27), although the most prevalent EBV type in Chinese EBVaGC cases was type I/*XhoI*- (75.6%) non-variant EBV, they found a new hotspot mutation in the *BamHI* W1/I1 boundary region (148,972 T C) in 86.7% of the tumors. This result suggest the implication of a different EBV strain in EBVaGC, however, it supports the involvement of EBV variants in neighboring regions of *BamHI* W1/I1 and/or *LMP1* in carcinogenesis of EBVaGC. It is worth investigating this mutation among EBVaGC cases with neither variant of *BamHI* W1/I1 (type i) nor *XhoI* restriction site at exon 1 of *LMP1* (*XhoI*+).

Some of the individuals were examined for other EBV polymorphisms in *EBNA3C* and *BamHI* F regions. The two major types of EBV, type 1 and 2, were determined in the sequence of *EBNA3C*. Type 1 was more prevalent in both EBVaGC (9 out of 11, 82%) and controls (57 out of 58, 98%). In the *BamHI* F region, the prototype F and f variants were determined, and the prototype F was the major type in both EBVaGC (11 out of 11, 100%) and non-cancer controls (108 out of 113, 96%).

In conclusion, as in the previous studies of individuals in Chile and Colombia, the distinctive EBV strain, type i/*XhoI*+, was also predominant among EBVaGC cases in Peru. However, the frequency of this type among populations at risk of gastric cancer in Peru was significantly lower than that in neighboring countries, which might explain the low incidence of EBVaGC in Peru.

Acknowledgements

This work was supported by a Grant-in-Aid for Scientific Research on Priority Areas of the Ministry of Education, Science, Sports and Culture of Japan (12218231).

References

- 1 Takada K: Epstein-Barr virus and gastric carcinoma. *Mol Pathol* 53: 255-261, 2000.
- 2 Rickinson AB and Kieff E: Epstein-Barr virus. *In: Fields Virology*, 4th edn. Knipe DM and Howley PM (eds.). Philadelphia, Lippincott Williams & Wilkins, pp. 2597-2612, 2001.
- 3 Corvalan A, Koriyama C, Akiba S, Eizuru Y, Backhouse C, Palma M, Argandona J and Tokunaga M: Epstein-Barr virus in gastric carcinoma is associated with location in the cardia and with a diffuse histology: a study in one area of Chile. *Int J Cancer* 94: 527-530, 2001.
- 4 Koriyama C, Akiba S, Iriya K, Yamaguti T, Hamada GS, Itoh T, Eizuru Y, Aikou T, Watanabe S, Tsugane S and Tokunaga M: Epstein-Barr virus-associated gastric carcinoma in Japanese Brazilians and non-Japanese Brazilians in Sao Paulo. *Jpn J Cancer Res* 92: 911-917, 2001.
- 5 Burgess DE, Woodman CB, Flavell KJ, Rowlands DC, Crocker J, Scott K, Biddulph JP, Young LS and Murray PG: Low prevalence of Epstein-Barr virus in incident gastric adenocarcinomas from the United Kingdom. *Br J Cancer* 86: 702-704, 2002.
- 6 Hao Z, Koriyama C, Akiba S, Li J, Luo X, Itoh T, Eizuru Y and Zou J: The Epstein-Barr virus-associated gastric carcinoma in Southern and Northern China. *Oncol Rep* 9: 1293-1298, 2002.
- 7 Kattoor J, Koriyama C, Akiba S, Itoh T, Ding S, Eizuru Y, Nair RR, Varghese C and Nair K: Epstein-Barr virus-associated gastric carcinoma in southern India – A comparison with a large scale Japanese series. *J Med Virol* 68: 384-389, 2002.
- 8 Carrascal E, Koriyama C, Akiba S, Tamayo O, Itoh T, Eizuru Y, Garcia F, Sera M, Carrasquilla G, Piazuolo MB, Florez L and Bravo JC: Epstein-Barr virus-associated gastric carcinoma in Cali, Colombia. *Oncol Rep* 10: 1059-1062, 2003.
- 9 Yoshiwara E, Koriyama C, Akiba S, Itoh T, Minakami Y, Chirinos JL, Watanabe J, Takano J, Miyagui J, Hidalgo H, Chacon P, Linares V and Eizuru Y: Epstein-Barr virus-associated gastric carcinoma in Lima, Peru. *J Exp Clin Cancer Res* 24: 49-54, 2005.
- 10 Abdirad A, Ghaderi-Sohi S, Shuyama K, Koriyama C, Nadimi-Barforoosh H, Emami S, Mosavi-Jarrahi A, Nahvijou A and Akiba S: Epstein-Barr virus associated gastric carcinoma: a report from Iran in the last four decades. *Diagn Pathol* 2: 25, 2007.
- 11 Corvalan A, Ding S, Koriyama C, Carrascal E, Carrasquilla G, Backhouse C, Urzua L, Argandona J, Palma M, Eizuru Y and Akiba S: Association of a distinctive strain of Epstein-Barr virus with gastric cancer. *Int J Cancer* 118: 1736-1742, 2006.
- 12 Lung ML and Chang GC: Detection of distinct Epstein-Barr virus genotypes in NPC biopsies from southern Chinese and Caucasians. *Int J Cancer* 52: 34-37, 1992.
- 13 Sidagis J, Ueno K, Tokunaga M, Ohyama M and Eizuru Y: Molecular epidemiology of Epstein-Barr virus (EBV) in EBV-related malignancies. *Int J Cancer* 72: 72-76, 1997.
- 14 Khanim F, Yao QY, Niedobitek G, Sihota S, Rickinson AB and Young LS: Analysis of Epstein-Barr virus gene polymorphisms in normal donors and in virus-associated tumors from different geographic locations. *Blood* 88: 3491-3501, 1996.
- 15 Lung ML, Chang RS and Jones JH: Genetic polymorphism of natural Epstein-Barr virus isolates from infectious mononucleosis patients and healthy carriers. *J Virol* 62: 3862-3866, 1988.
- 16 zur Hausen A, Brink AA, Craanen ME, Middeldorp JM, Meijer CJ and van den Brule AJ: Unique transcription pattern of Epstein-Barr virus (EBV) in EBV-carrying adenocarcinomas: expression of the transforming *BARF1* gene. *Cancer Res* 60: 2745-2748, 2000.
- 17 Danve C, Decaussin G, Busson P and Ooka T: Growth transformation of primary epithelial cells with a NPC-derived Epstein-Barr virus strain. *Virology* 288: 223-235, 2001.
- 18 Gully ML, Pulitzer DR, Eagan PA and Schneider BG: Epstein-Barr virus infection is an early event in gastric carcinogenesis and is independent of BCL-2 expression and p53 accumulation. *Hum Pathol* 27: 20-27, 1996.
- 19 Ryan JL, Morgan DR, Dominguez RL, Thorne LB, Elmore SH, Mino-Kenudson M, Lauwers GY, Booker JK and Gully ML: High levels of Epstein-Barr virus DNA in latently infected gastric adenocarcinoma. *Lab Invest* 89: 80-90, 2009.

- 20 Takada K and Nanbo A: The role of EBERs in oncogenesis. *Semin Cancer Biol* 11: 461-467, 2001.
- 21 Hino R, Uozaki H, Inoue Y, Shintani Y, Ushiku T, Sakatani T, Takada K and Fukayama M: Survival advantage of EBV-associated gastric carcinoma: Survivin up-regulation by viral latent membrane protein 2A. *Cancer Res* 68: 1427-1435, 2008.
- 22 Wang Q, Tsao SW, Ooka T, Nicholls JM, Cheung HW, Fu S, Wong YC and Wnag X: Anti-apoptotic role of BARF1 in gastric cancer cells. *Cancer Lett* 238: 90-103, 2006.
- 23 Chang KL, Chen YY, Shibata D and Weiss LM: Description of an *in situ* hybridization methodology for detection of Epstein-Barr virus RNA in paraffin-embedded tissues, with a survey of normal and neoplastic tissues. *Diagn Mol Pathol* 1: 246-255, 1992.
- 24 Greer CE, Wheeler CM and Manos MM: PCR amplification from paraffin-embedded tissues: Sample preparation and the effects of fixation. *In: PCR Primer: A Laboratory Manual*. Carl WD and Gabriela SD (eds.). New York: Cold Spring Harbor Laboratory Press, pp. 99-112, 1995.
- 25 Lung ML, Chang GC, Miller TR, Wara WM and Phillips TL: Genotypic analysis of Epstein-Barr virus isolates associated with nasopharyngeal carcinoma in Chinese immigrants to the United States. *Int J Cancer* 59: 743-746, 1994.
- 26 Sandvej K, Gratama JW, Munch M, Zhou XG, Bolhuis RL, Andresen BS, Gregersen N and Hamilton-Dutoit S: Sequence analysis of the Epstein-Barr virus (EBV) latent membrane protein-1 gene and promoter region: identification of four variants among wild-type EBV isolates. *Blood* 90: 323-330, 1997.
- 27 Chen JN, Ding YG, Feng ZY, Li HG, He D, Du H, Wu B and Shao CK: Association of distinctive Epstein-Barr virus variants with gastric carcinoma in Guangzhou, southern China. *J Med Virol* 82: 658-667, 2010.
- 28 Miller WE, Earp HS and Raab-Traub N: The Epstein-Barr virus latent membrane protein 1 induces expression of the epidermal growth factor receptor. *J Virol* 69: 4390-4398, 1995.
- 29 Thornburg NJ and Raab-Traub N: Induction of epidermal growth factor receptor expression by Epstein-Barr virus latent membrane protein 1 C-terminal-activating region 1 is mediated by NF-kappaB p50 homodimer/BCL-3 complexes. *J Virol* 81: 12954-12961, 2007.
- 30 Tanaka M, Kawaguchi Y, Yokofujita J, Takagi M, Eishi Y and Hirai K: Sequence variations of Epstein-Barr virus *LMP2A* gene in gastric carcinoma in Japan. *Virus Genes* 19: 103-111, 1999.
- 31 Hino R, Uozaki H, Murakami N, Ushiku T, Shinozaki A, Ishikawa S, Morikawa T, Nakaya T, Sakatani T, Takada K and Fukayama M: Activation of DNA methyltransferase 1 by EBV latent membrane protein 2A leads to promoter hypermethylation of *PTEN* gene in gastric carcinoma. *Cancer Res* 69: 2766-2774, 2009.
- 32 Fukayama M, Hino R and Uozaki H: Epstein-Barr virus and gastric carcinoma: virus-host interactions leading to carcinoma. *Cancer Sci* 99: 1726-1733, 2008.
- 33 Seto E, Yang L, Middeldorp J, Sheen TS, Chen JY, Fukayama M, Eizuru Y, Ooka T and Takada K: Epstein-Barr Virus (EBV)-Encoded *BARF1* gene is expressed in nasopharyngeal carcinoma and EBV-associated gastric carcinoma tissues in the absence of lytic gene expression. *J Med Virol* 76: 82-88, 2005.
- 34 Griffin BE and Karran L: immortalization of monkey epithelial cells by specific fragments of Epstein-Barr virus DNA. *Nature* 309: 78-82, 1984.
- 35 Karran L, Teo CG, King D, Hitt MM, Gao YN, Wedderburn N and Griffin BE: Establishment of immortalized primate epithelial cells with sub-genomic EBV DNA. *Int J Cancer* 45: 763-772, 1990.
- 36 Wei MX and Ooka T: A transforming function of the *BARF1* gene encoded by Epstein-Barr virus. *EMBO J* 8: 2897-2903, 1989.
- 37 Wei MX, Moulin JC, Decaussin G, Berger F and Ooka T: Expression and tumorigenicity of the Epstein-Barr virus *BARF1* gene in human Louckes B-lymphocyte cell line. *Cancer Res* 54: 1843-1848, 1994.
- 38 Wiech T, Nikolopoulos E, Lassman S, Heidt T, Schöpflin A, Sarbia M, Werner M, Shimizu Y, Sakka E, Ooka T and zur Hausen A: Cyclin D1 expression is induced by viral BARF1 and is overexpressed in EBV-associated gastric cancer. *Virchows Arch* 452: 621-627, 2008.
- 39 Ernberg I, Kallin B, Dillner J, Falk K, Ehlin-Henriksson B, Hammarskjöld ML and Klein G: Lymphoblastoid cell lines and Burkitt-lymphoma-derived cell lines differ in the expression of a second Epstein-Barr virus encoded nuclear antigen. *Int J Cancer* 38: 729-737, 1986.
- 40 Samanta M, Iwakiri D and Takada K: Epstein-Barr virus-encoded small RNA induces IL-10 through RIG-I-mediated IRF-3 signaling. *Oncogene* 27: 4150-4160, 2008.
- 41 Iwakiri D, Eizuru Y, Tokunaga M and Takada K: Autocrine growth of Epstein-Barr virus-positive gastric carcinoma cells mediated by an Epstein-Barr virus-encoded small RNA. *Cancer Res* 63: 7062-7067, 2003.
- 42 Abdel-Hamid M, Chen JJ, Constantine N, Massoud M and Raab-Traub N: EBV strain variation: geographical distribution and relation to disease state. *Virology* 190: 168-175, 1992.
- 43 Hu LF, Zabarovsky ER, Chen F, Cao SL, Erberg I, Klein G and Winberg G: Isolation and sequencing of the Epstein-Barr virus BNLF-1 gene *LMP1* from a Chinese nasopharyngeal carcinoma. *J Gen Virol* 72: 2399-2409, 1991.
- 44 Chen ML, Tsai CN, Liang CL, Shu CH, Huang CR, Sulitzeanu D, Liu ST and Chang YS: Cloning and characterization of the latent membrane protein (LMP) of a specific Epstein-Barr virus variant derived from the nasopharyngeal carcinoma in the Taiwanese population. *Oncogene* 7: 2131-2140, 1992.
- 45 Cabrera ME, Eizuru Y, Itoh T, Koriyama C, Tashiro Y, Ding S, Rey S, Akiba S and Corvalan A: Nasal natural killer/T-cell lymphoma and its association with type *iXhoI* loss strain Epstein-Barr virus in Chile. *J Clin Pathol* 60: 656-660, 2007.

Received June 3, 2011
 Revised July 15, 2011
 Accepted July 16, 2011