# Sorafenib Modulates the Gene Expression of Multi-drug Resistance Mediating ATP-Binding Cassette Proteins in Experimental Hepatocellular Carcinoma

KATRIN HOFFMANN\*, CLEMENS FRANZ\*, ZHI XIAO, ELVIRA MOHR, SUSANNE SERBA, MARKUS W. BÜCHLER and PETER SCHEMMER

Department of General and Transplantation Surgery, Ruprecht-Karls-University, Im Neuenheimer Feld 110, 69120 Heidelberg, Germany

**Abstract.** Background: High ATP-binding cassette (ABC) protein expression leads to intrinsic drug resistance of hepatocellular carcinoma (HCC). The aim of this study was to investigate the potential chemosensitizing effects of sorafenib on the multi-drug resistance (MDR) phenotype. Material and Methods: The ABC-protein gene expression and the cellular survival were determined by RT-PCR analysis and MTT assay in HUH7 cells. Results: Sorafenib inhibits MDR. The ABC-protein mRNA expression decreased by up to 51%  $(p \le 0.01)$ . Addition of sorafenib to conventional chemotherapy restored the chemosensitivity. Combination of gemcitabine plus sorafenib decreased the ABC-protein mRNA levels by up to 77%, compared to gemcitabine monotherapy ( $p \le 0.001$ ). Doxorubicin plus sorafenib decreased the ABC-protein mRNA levels up to 74% compared to doxorubicin monotherapy  $(p \le 0.001)$ . Conclusion: This study provides evidence that the MDR phenotype of HCC cells can be modulated by the multikinase inhibitor sorafenib and consequentially may lead towards personalized therapies in patients with highly resistant tumors.

Hepatocellular carcinoma (HCC) is the third most common cause of cancer-related deaths globally and its incidence is increasing (1). Around 30-40% of HCC patients are diagnosed at an early stage of the disease and are amenable to curative treatment options such as hepatic resection and liver transplantation (2). However, the majority of patients present

\*Both authors contributed equally to this work.

*Correspondence to:* Peter Schemmer, MD, MBA, Department of General and Transplantation Surgery, Ruprecht-Karls-University, Im Neuenheimer Feld 110 D-69120 Heidelberg, Germany. Tel: +49 06221566110, Fax: +49 06221564215, e-mail: peter.schemmer@ med.uni-heidelberg.de

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with advanced-stage tumors and have a median survival of approximately six months (2-4). The results of conventional systemic chemotherapy for HCC are disappointing. None of the available clinical trials reported benefits in terms of survival, and tumor response rates were well below 10% (5, 6). High intrinsic resistance against structurally and functionally unrelated cytostatic drugs has been previously demonstrated in HCC (7, 8). Multi-drug resistance (MDR) in HCC is mediated by an increased cellular efflux of cytostatic compounds via transmembrane ATP-binding cassette proteins (MDR proteins) (9, 10). Physiologically, these proteins modulate the absorption, distribution and excretion of endo- and xenobiotics as well as of various pharmacological compounds in the liver (11-13). However, the overexpression of MDR proteins is associated with impaired overall survival and an aggressive tumor phenotype in primary liver cancer (14-16). The interaction of intracellular signaling pathways and MDR has been discussed with regard to various tumor entities (16, 17). Previously, the involvement of the epidermal growth factor-activated tyrosine kinase pathway has been demonstrated in the regulation of multi-drug resistance in HCC. Selective inhibition of the epidermal growth factor receptor (EGFR) restored the chemosensitivity of resistant HCC cells and increased the efficacy of conventional chemotherapy (18, unpublished data). However, there is further evidence that the downstream kinases of the RAF/MEK/ERK pathway might have a direct impact on the drug resistance phenotype (15-17). The tyrosine kinase inhibitor sorafenib is the current standard of care in patients with advanced HCC and has shown significant survival benefits (19). Combination treatment strategies of sorafenib and conventional chemotherapy showed promising results and acceptable toxicity in phase I clinical trials (20, 21).

The purpose of this *in vitro* study was to investigate the effects of tyrosine kinase pathway inhibition on the MDR phenotype in experimental HCC. Sorafenib was evaluated for its potential as part of a chemosensitizing combination treatment in tailored therapies for patients with highly resistant tumors.

## Materials and Methods

*HCC cell line*. The human HCC cell line HuH-7 (gift from Ingrid Herr, Division of Molecular Onco-Surgery, Ruprecht-Karls-University, Heidelberg, Germany) was used for *in vitro* experiments and cultured in DMEM (PAA Laboratories GmbH, Pasching, Austria) containing 10% fetal bovine serum (FBS), 100 U/ml penicillin and 100  $\mu$ g/ml streptomycin (Biochrom AG, Berlin, Germany) in 5% CO<sub>2</sub> at 37°C (22, 23).

*Chemotherapeutic treatment.* Gemcitabine (Lilly, Indianapolis, USA), doxorubicin (Sandoz Pharmaceuticals GmbH, Holzkirchen, Germany) and sorafenib (Bayer Healthcare, Leverkusen, Germany) were prepared according to the manufacturer's instructions. Cells were treated as follows: untreated controls, gemcitabine (11.4 µg/ml, 114 µg/ml or 1140 µg/ml) twice weekly, doxorubicin (0.015 µg/ml, 0.15 µg/ml or 1.5 µg/ml) twice weekly, or sorafenib 5 µM twice weekly. For the evaluation of potential combinative effects, gemcitabine and doxorubicin were combined with sorafenib 5 µM twice weekly at the above-mentioned doses.

*MTT assay.* Cells were treated as mentioned above and MTT assay was performed by adding 10 µl of 5 mg/ml 3-(4,5-dimethylthiazolyl-2)-2,5-diphenyl-2*H*-tetrazoliumbromide. The plates were incubated for 4 hours at 37°C, 5% CO<sub>2</sub> and the resulting formazan crystals dissolved in 100 µl 2-propanol (Carl Roth GmbH, Karlsruhe, Germany). The extinction was measured at 570 nm with a reference wavelength of 620 nm with the Anthos Reader 2010 (Anthos Mikrosysteme GmbH, Krefeld, Germany). A minimum of three independent experiments were performed. The survival of cells was compared to the survival of the untreated controls, which was defined as 100%.

RT-PCR. Total RNA was isolated with the RNeasy mini kit (Qiagen, Hilden, Germany) and cDNA generated with the transcriptor first strand cDNA synthesis kit (Roche Diagnostics GmbH, Mannheim, Germany) according to the manufacturer's instructions. The StepOne Real-Time PCR System (Applied Biosystems, Carlsbad, USA) was used for semi-quantitative analysis with Power SYBR Green PCR Master Mix (Applied Biosystems, Carlsbad, USA) and QuantiTect Primer Assay (Qiagen, Hilden, Germany) for GAPDH (QT01192646) as endogenous control, PGP (QT00081928), MRP1 (QT00061159), MRP2 (QT00056294) and MRP3 (QT00070602). The PCR was performed after 10 min of denaturation at 95°C for 40 cycles with 15 s of denaturation at 95°C and 1 min annealing and polymerization at 60°C. The melting curve was analyzed for unspecific products or primer dimers and a 1:10 dilution series over at least five orders of magnitude revealed PCR efficiency between 89% and 109%. Data were analyzed using the comparative Ct method in the StepOne Software 2.1 (Applied Biosystems, Carlsbad, USA). All samples were analyzed in triplicate and a minimum of three independent experiments were performed.

Statistics. The one-way ANOVA test followed by a Tukey test was used to reveal significant differences between the control and treatment groups. The level of statistical significance was defined as  $p \leq 0.05$ . All statistical computations were performed in SigmaStat 2.03 (Jandel Scientific, San Rafael, CA, USA). The MTT assay results are presented as mean values and standard error of mean (SE), the results of PCR as mean values and 95% confidence interval.

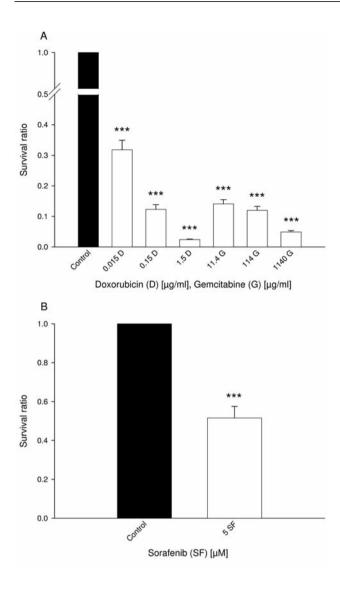
## Results

Impact of conventional chemotherapeutic and molecular targeted treatment on cellular survival. Conventional chemotherapy, as well as targeted-molecular therapy with sorafenib, significantly inhibited the survival of HuH-7 compared to the untreated controls. Gemcitabine monotherapy, at concentrations of 11.4 mg/ml, 114 mg/ml and 1140 mg/ml, reduced the survival significantly by 86%, 88% and 95%, respectively, compared to the control group ( $p \le 0.001$ ). Doxorubicin, at concentrations of 0.015 mg/ml, 0.15 mg/ml and 1.5 mg/ml, revealed a dose-dependent survival decrease by 68%, 88% and 98%, respectively, compared to the untreated controls (p≤0.001) (Figure 1A). Sorafenib monotherapy reduced the cellular survival by 48% compared to the control group ( $p \le 0.05$ ) (Figure 1B). DMSO, which was used as a solvent for sorafenib, did not influence the cellular survival (data not shown).

Combinative treatment of conventional chemotherapy plus sorafenib also inhibited the survival of HuH-7 cells significantly compared to the untreated controls. The survival rate decreased by 83%, 86% and 93%, respectively, after combinative treatment with gemcitabine at 11.4 mg/ml, 114 mg/ml or 1140 mg/ml plus sorafenib compared to the controls. Furthermore, a dose-dependent survival decrease of 71%, 88% and 98%, respectively, was detectable after treatment with a combination of doxorubicin at 0.015 mg/ml, 0.15 mg/ml or 1.5 mg/ml plus sorafenib compared to the untreated controls (Figure 1C). However, the inhibitory effect on the cellular survival of the combinative treatment was not significantly different from the effects of monotherapy with gemcitabine or doxorubicin (data not shown).

Induction of MDR protein gene expression by conventional chemotherapy. As described previously (18, unpublished data), conventional chemotherapy induced MDR in HUH7 cells. After treatment with gemcitabine, a significant dose-dependent increase of MDR protein gene expression was detectable compared to the controls ( $p \le 0.05$ ). The PGP, MRP1, MRP2 and MRP3 mRNA expression increased two-, five-, four- and ten-fold, respectively, compared to the control group (Figure 2A). Doxorubicin led to enhanced mRNA detection compared to controls ( $p \le 0.05$ ). The levels of PGP, MRP1, MRP2 and MRP3 mRNA increased in a dose-dependent manner by up to five-, four- and eight-fold, respectively, compared to the controls ( $p \le 0.05$ ).

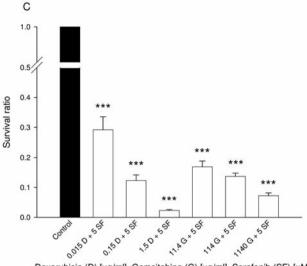
Inhibition of MDR protein gene expression by sorafenib. Sorafenib monotherapy reduced the expression of *PGP* and *MRP2* mRNA significantly. The mRNA expression of *PGP* and *MRP2* was 36% ( $p \le 0.01$ ) and 51% lower compared to the control group ( $p \le 0.0001$ ) (Figure 3).



Effects of combined conventional and molecular targeted therapy on MDR gene expression. Simultaneous treatment with conventional chemotherapy plus sorafenib significantly reduced the MDR protein gene expression. The combination of gemcitabine (11.4 mg/ml) plus sorafenib reduced the PGP, MRP1, MRP2 and MRP3 mRNA levels by 75%, 67%, 59% and 77%, respectively, compared to monotherapy with gemcitabine ( $p\leq0.001$ ) (Figure 4A). Doxorubicin (0.15 mg/dl) plus sorafenib reduced the PGP, MRP1, MRP2 and MRP3 mRNA levels by 47%, 74%, 44% and 53%, respectively, compared to monotherapy with doxorubicin ( $p\leq0.001$ ) (Figure 4B).

#### Discussion

Hepatocellular carcinoma is one of the most intrinsically resistant tumors and conventional chemotherapy has shown only minor effectiveness, with response rates below 10% (5, 24). Individually designed treatment strategies to overcome



Doxorubicin (D) [µg/ml], Gemcitabine (G) [µg/ml], Sorafenib (SF) [µM]

Figure 1. Impact of conventional chemotherapeutic and molecular targeted treatment on cellular survival. A: Conventional chemotherapy reduces the survival of HuH7 cells. Gemcitabine (11.4 µg/ml, 114 µg/ml or 1140 µg/ml) or doxorubicin (0.015 mg/ml, 0.15 µg/ml or 1.5 µg/ml) were added to HuH7 cells twice weekly. B: Sorafenib inhibits the survival of HuH7 cells. Sorafenib 5 µM was added to HuH7 cells twice weekly. C: Combinative treatment of conventional chemotherapy and sorafenib reduces the survival of HuH7 cells. Gemcitabine (11.4 µg/ml, 114 µg/ml or 1.15 µg/ml) or doxorubicin (0.015 mg/ml, 0.15 µg/ml or 1.15 µg/ml) plus Sorafenib 5 µM were added to HuH7 cells twice weekly. \*\*\*p≤0.001 Compared to the untreated control group.

MDR in HCC are being progressively investigated (25, 26). This study provides evidence that the gene expression of MDR proteins can be modulated by the multi-kinase inhibitor sorafenib and implicates the involvement of the RAF/MEK/ERK pathway in MDR regulation in HCC.

As previously described, standard chemotherapy induces MDR in HCC cell lines by increased ATP-binding cassette protein gene expression and, consequently, increased survival of drugresistant cells (18, unpublished data). One new approach towards the abrogation of tumor-cell resistance to treatmentinduced growth inhibition and cell death might be the combination of molecular-targeted and conventional systemic therapies. Several studies have implicated an involvement of tyrosine kinase-mediated signaling in the development of the chemoresistance phenotype in tumor cells (14, 27, 28). However, although reduced mRNA expression of MRP1 has been described after ERK inhibition in HepG2 cells, data for HCC are scarce (16). To date, two potential mechanisms for the modulation of drug resistance by tyrosine kinase inhibitors have been discussed. Firstly, a direct interaction of tyrosine kinase inhibitors with the transporter and secondly, an influence of tyrosine kinase inhibitors on the expression level of MDR proteins, have been hypothesized (29). It has been reported that sorafenib itself is only a weak PGP substrate in vitro and in

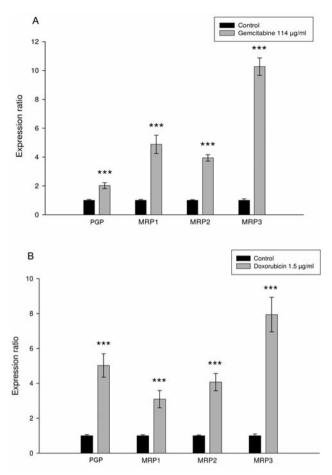


Figure 2. Induction of MDR protein gene expression by conventional chemotherapy. A: Gemcitabine (114  $\mu$ g/ml) was added to HuH7 cells twice weekly. B: Doxorubicin (1.5  $\mu$ g/ml) was added to HuH7 cells twice weekly. \*\*\*p≤0.001 Compared to the untreated control group.

*vivo* (30). However, a reduction of the ATP-dependent efflux function of ATP-binding cassette proteins, as found after treatment with the tyrosine kinase inhibitors gefitinib and vandetanib, has not yet been described (29).

Data presented in this study clearly show that sorafenib monotherapy significantly reduced the expression of ATPbinding cassette protein mRNA in HuH7 cells. The PGP and MRP2 mRNA expression decreased by 36% and 51%, respectively. The multi-kinase inhibitor sorafenib strongly inhibits VEGFR, RAF isoforms and reduces the phosphorylation of ERK and MEK1/2 (21, 31). The potential link between the inhibitory effects of sorafenib on drug resistance can therefore either be the result of receptor inhibition or modulation of the down-stream kinases. There is evidence that RAF-1 plays a role in drug resistance acquisition by prolongation of cellular survival via anti-MEK/ERK-dependent mechanisms apoptotic (32).Furthermore, increased resistance was found in MCF-7

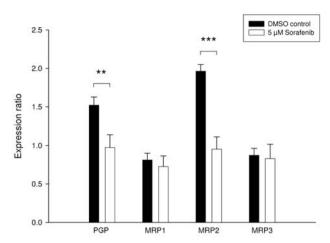


Figure 3. Inhibition of MDR protein gene expression by sorafenib. Sorafenib 5  $\mu$ M was added to HuH7 cells twice weekly. \*\*p≤0.01, \*\*\*p≤0.001 compared to the DMSO control group.

breast cancer cells which overexpress active RAF-1. An effect of activated RAF-1 kinase on the *PGP* gene promoter has been described previously (33, 34). Additionally, VEGF seems to play a role in the drug-resistance phenotype. A simultaneous increase of PGP and VEGF expression was associated with a more aggressive and invasive phenotype of resistant laryngeal carcinoma cells (35).

Interestingly, in the present study, combinative treatment with conventional chemotherapy and sorafenib restored the chemosensitivity and inhibited the survival of HuH-7 cells significantly. The ABC-transport protein gene expression was found to be significantly lower after treatment with sorafenib plus gemcitabine or doxorubicin compared to monotherapy with these cytostatics. This supports the observation of the potentially chemosensitizing effects of sorafenib in phase I trials of patients with advanced refractory solid tumors. Combination treatment strategies of sorafenib and conventional chemotherapy have previously shown promising results and an acceptable toxicity (21, 36). Abou-Alfa et al. demonstrated in their phase II trial a significant prolongation of survival in patients treated with the combination of sorafenib and doxorubicin (37). Furthermore, Zhu et al. investigated the effects of conventional chemotherapy, targeting the vascular endothelial growth factor pathway with bevacizumab in a phase II trial involving patients with advanced HCC. The combination had moderate antitumor activity and lead to a median progression-free survival of 5.3 months (26).

However, despite the significant effects on ATP-binding cassette protein gene expression that were found in the present study, no additive or super-additive effect on the cellular survival was detectable in comparison to chemomonotherapy. One reason for this might be the activation of cellular escape mechanisms and up-regulation of other

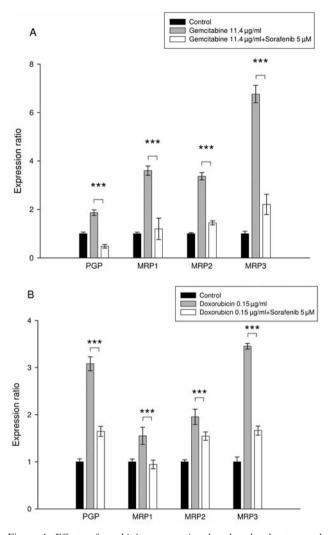


Figure 4. Effects of combining conventional and molecular-targeted therapy on MDR gene expression. A: Gemcitabine (11.4  $\mu$ g/ml) plus sorafenib 5  $\mu$ M were added to HuH7 cells twice weekly and cells cultured for two weeks. \*\*\*p≤0.001 Compared to gemcitabine monotherapy. B: Doxorubicin (0.15  $\mu$ g/ml) plus sorafenib 5  $\mu$ M were added to HuH7 cells twice weekly and cells cultured for two weeks. \*\*\*p≤0.001 Compared to doxorubicin monotherapy.

resistance pathways, such as alterations in the cell cycle and increased repair of DNA damage (38). Previously, it has been shown that the anti-tumoral efficiency of growth factor receptor targeting is counteracted in HCC cells due to the ability of alternative stimulation of signaling pathways (39).

The multi-kinase inhibitor sorafenib is the current gold standard for treating advanced HCC. The present study provides evidence that the gene expression of ATP-binding cassette proteins can be modulated by sorafenib and also suggests the involvement of the RAF/MEK/ERK pathway in regulating MDR in HCC. Further studies are warranted to identify the signaling pathways involved in the enduring restoration of chemosensitivity to maximize the anti-tumoral effects. Nevertheless, the combination of biologically targeted therapies and established chemotherapeutic approaches might have important implications for the development of tailored treatment strategies in patients with highly resistant hepatocellular carcinomas.

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### References

- Parkin DM, Bray F, Ferlay J and Pisani P: Global cancer statistics, 2002. CA Cancer J Clin 55(2): 74-108, 2005.
- 2 Llovet JM and Bruix J: Hepatocellular carcinoma. The Lancet *362*: 1907-1917, 2003.
- 3 Bruix J and Sherman M: Management of hepatocellular carcinoma. Hepatology 42(5): 1208-1236, 2005.
- 4 Bruix J, Sherman M, Llovet JM, Beaugrand M, Lencioni R, Burroughs AK, Christensen E, Pagliaro L, Colombo M and Rodes J: Clinical management of hepatocellular carcinoma. Conclusions of the Barcelona-2000 EASL conference. European Association for the Study of the Liver. J Hepatol 35(3): 421-430, 2001.
- 5 Ganne-Carrie N and Trinchet JC: Systemic treatment of hepatocellular carcinoma. Eur J Gastroent and Hepatol 16(3): 275-281, 2004.
- 6 Lopez PM, Villanueva A and Llovet JM: Systematic review: evidence-based management of hepatocellular carcinoma – an updated analysis of randomized controlled trials. Aliment Pharmacol Ther 23(11): 1535-1547, 2006.
- 7 Nies AT, Konig J, Pfannschmidt M, Klar E, Hofmann WJ and Keppler D: Expression of the multidrug resistance proteins MRP2 and MRP3 in human hepatocellular carcinoma. Int J Cancer 94(4): 492-499, 2001.
- 8 Li B, Ye T, Zhao L, Li DH, Gou XH, Zhao LY, Han L, Chen L, Yan LN and Gong JP: Effects of multidrug resistance, antisense RNA on the chemosensitivity of hepatocellular carcinoma cells. Hepatobiliary Pancreat Dis Int 5(4): 552-559, 2006.
- 9 Borst P and Elferink RO: Mammalian ABC transporters in health and disease. Annu Rev Biochem 71: 537-592, 2002.
- 10 Zollner G, Wagner M, Fickert P, Silbert D, Fuchsbichler A, Zatloukal K, Denk H and Trauner M: Hepatobiliary transporter expression in human hepatocellular carcinoma. Liver Int 25(2): 367-379, 2005.
- 11 Kato A, Miyazaki M, Ambiru S, Yoshitomi H, Ito H, Nakagawa K, Shimizu H, Yokosuka O and Nakajima N: Multidrug resistance gene (*MDR-1*) expression as a useful prognostic factor in patients with human hepatocellular carcinoma after surgical resection. J Surg Oncol 78(2): 110-115, 2001.
- 12 Soini Y, Virkajarvi N, Raunio H and Paako P: Expression of Pglycoprotein in hepatocellular carcinoma: a potential marker of prognosis. J Clin Pathol 49: 470-473, 1996.
- 13 Vander BS, Komuta M, Libbrecht L, Katoonizadeh A, Aerts R, Dymarkowski S, Verslype C, Nevens F and Roskams T: Expression of multidrug resistance-associated protein 1 in hepatocellular carcinoma is associated with a more aggressive tumour phenotype and may reflect a progenitor cell origin. Liver Int 28(10): 1370-1380, 2008.

- 14 Barancik M, Bohacova V, Kvackajova J, Hudecova S, Krizanova O and Breier A: SB203580, a specific inhibitor of p38-MAPK pathway, is a new reversal agent of P-glycoprotein-mediated multidrug resistance. Eur J Pharm Sci 14(1): 29-36, 2001.
- 15 Garcia R, Franklin RA and McCubrey JA: EGF induces cell motility and multi-drug resistance gene expression in breast cancer cells. Cell Cycle *5*(*23*): 2820-2826, 2006.
- 16 Guan J, Chen XP, Zhu H, Luo SF, Cao B and Ding L: Involvement of extracellular signal-regulated kinase/mitogenactivated protein kinase pathway in multidrug resistance induced by HBx in hepatoma cell line. World J Gastroenterol 10(23): 3522-3527, 2004.
- 17 Katayama K, Yoshioka S, Tsukahara S, Mitsuhashi J and Sugimoto Y: Inhibition of the mitogen-activated protein kinase pathway results in the down-regulation of P-glycoprotein. Mol Cancer Ther 6(7): 2092-2102, 2007.
- 18 Hoffmann K, Xiao Z, Franz C, Mohr E, Schultze D, Serba S, Büchler MW and Schemmer P: Involvement of the epidermal growth factor receptor in the modulation of multidrug resistance in experimental hepatocellular carcinoma. Unpublished data.
- 19 Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Raoul JP and Zeuzem S: Sorafenib improves survival in advanced hepatocellular carcinoma (HCC): results of a phase III randomized placebo controlled trial. J Clin Oncol 25: LBA1, 2007.
- 20 Richly H, Kupsch P, Passage K, Grubert M, Hilger RA, Voigtmann R, Schwartz B, Brendel E, Christensen O, Haase CG and Stuart-Harris H: Results of a phase I trial of Bay 43-9006 in combination with doxorubicin in patients with primary hepatic cancer. Int J Clinical Pharmacol *42(11)*: 650-651, 2004.
- 21 Mross K, Steinbild S, Baas F, Reil M, Buss P, Mersmann S, Voliotis D, Schwartz B and Brendel E: Drug–drug interaction pharmacokinetic study with the Raf kinase inhibitor (RKI) BAY 43-9006 administered in combination with irinotecan (CPT-11) in patients with solid tumors. Int J Clin Pharmacol Ther 41(12): 618-619, 2003.
- 22 Nakabayashi H, Taketa K, Miyano K, Yamane T and Sato J: Growth of human hepatoma cells lines with differentiated functions in chemically defined medium. Cancer Res 42(9): 3858-3863, 1982.
- 23 Knowles BB, Howe CC and Aden DP: Human hepatocellular carcinoma cell lines secrete the major plasma proteins and hepatitis B surface antigen. Science 209(4455): 497-499, 1980.
- 24 Herr I, Schemmer P and Buchler MW: On the TRAIL to therapeutic intervention in liver disease. Hepatology *46(1)*: 266-274, 2007.
- 25 Villanueva A, Toffanin S and Llovet JM: Linking molecular classification of hepatocellular carcinoma and personalized medicine: preliminary steps. Curr Opin Oncol 20(4): 444-453, 2008.
- 26 Zhu AX, Blaszkowsky LS, Ryan DP, Clark JW, Muzikansky A, Horgan K, Sheehan S, Hale KE, Enzinger PC, Bhargava P and Stuart K: Phase II study of gemcitabine and oxaliplatin in combination with bevacizumab in patients with advanced hepatocellular carcinoma. J Clin Oncol 24(12): 1898-1903, 2006.
- 27 Yang JM, Sullivan GF and Hait WN: Regulation of the function of P-glycoprotein by epidermal growth factor through phospholipase C. Biochem Pharmacol 53(11): 1597-1604, 1997.
- 28 Yang JM, Vassil AD and Hait WN: Activation of phospholipase C induces the expression of the multidrug resistance (*MDR1*) gene through the Raf-MAPK pathway. Mol Pharmacol 60(4): 674-680, 2001.

- 29 Azzariti A, Porcelli L, Simone GM, Quatrale AE, Colabufo NA, Berardi F, Perrone R, Zucchetti M, D'Incalci M, Xu JM and Paradiso A: Tyrosine kinase inhibitors and multidrug resistance proteins: interactions and biological consequences. Cancer Chemother Pharmacol 65(2): 335-346, DOI: 10.1007/560280-009-1039-0.
- 30 Gnoth MJ, Sandmann S, Engel K and Radtke M: *In vitro* to *in vivo* comparison of the substrate characteristics of sorafenib tosylate toward P-glycoprotein. Drug Metab Dispos 38(8): 1341-1346, 2010.
- 31 Richly H, Kupsch P, Passage K, Grubert M, Hilger RA, Kredtke S, Voliotis D, Scheulen ME, Seeber S and Strumberg D: A phase I clinical and pharmacokinetic study of the Raf kinase inhibitor (RKI) BAY 43-9006 administered in combination with doxorubicin in patients with solid tumors. Int J Clin Pharmacol Ther 41(12): 620-621, 2003.
- 32 Odabaei G, Chatterjee D, Jazirehi AR, Goodglick L, Yeung K and Bonavida B: Raf-1 kinase inhibitor protein: structure, function, regulation of cell signaling, and pivotal role in apoptosis. Adv Cancer Res *91*: 169-200, 2004.
- 33 Davis JM, Navolanic PM, Weinstein-Oppenheimer CR, Steelman LS, Hu W, Konopleva M, Blagosklonny MV and McCubrey JA: Raf-1 and Bcl-2 induce distinct and common pathways that contribute to breast cancer drug resistance. Clin Cancer Res *9*(*3*): 1161-1170, 2003.
- 34 Kim SH, Lee SH, Kwak NH, Kang CD and Chung BS: Effect of the activated Raf protein kinase on the human multidrug resistance 1 (MDR1) gene promoter. Cancer Lett 98(2): 199-205, 1996.
- 35 Li L, Jiang AC, Dong P, Wang H, Xu W and Xu C: MDR1/P-gp and VEGF synergistically enhance the invasion of Hep-2 cells with multidrug resistance induced by taxol. Ann Surg Oncol *16*(5): 1421-1428, 2009.
- 36 Richly H, Kupsch P, Passage K, Grubert M, Hilger RA, Voigtmann R, Schwartz B, Brendel E, Christensen O, Haase CG and Strumberg D: Results of a phase I trial of BAY 43-9006 in combination with doxorubicin in patients with primary hepatic cancer. Int J Clin Pharmacol Ther *42(11)*: 650-651, 2004.
- 37 Abou-Alfa GK, Schwartz L, Ricci S, Amadori D, Santoro A, Figer A, De GJ, Douillard JY, Lathia C, Schwartz B, Taylor I, Moscovici M and Saltz LB: Phase II study of sorafenib in patients with advanced hepatocellular carcinoma. J Clin Oncol 24(26): 4293-4300, 2006.
- 38 Gottesman MM: Mechanisms of cancer drug resistance. Annu Rev Med 53: 615-627, 2002.
- 39 Desbois-Mouthon C, Baron A, Blivet-Van Eggelpoel MJ, Fartoux L, Venot C, Bladt F, Housset C and Rosmorduc O: Insulin-like growth factor-1 receptor inhibition induces a resistance mechanism via the epidermal growth factor receptor/HER3/AKT signaling pathway: rational basis for cotargeting insulin-like growth factor-1 receptor and epidermal growth factor receptor in hepatocellular carcinoma. Clin Cancer Res 15(17): 5445-5456, 2009.

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