# Curcumin and Rutin Down-regulate Cyclooxygenase-2 and Reduce Tumor-associated Inflammation in HPV16-Transgenic Mice

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Abstract. Aim: Cyclo-oxygenase-2 (COX2) plays a prominent role in carcinogenesis. This study addresses the effects of two nutraceutical compounds on the expression of COX2 and tumor-associated inflammation in human papillomavirus type 16 (HPV16)-transgenic mice. Materials and Methods: Sixweek-old FVB/n mice were supplemented with rutin or curcumin for 24 weeks:  $HPV16^{-/-}$  no treatment, n=12;  $HPV16^{+/-}$  no treatment, n=13;  $HPV16^{+/-}$  rutin, n=12;  $HPV16^{+/-}$  curcumin, n=13. HPV16-induced skin lesions and their inflammatory infiltrates were studied histologically. COX2 expression was assessed immunohistochemically. Results: Rutin reduced COX2 expression in the dermis (immunostaining score 7.83 versus 11.25 in untreated HPV16-transgenic mice) and epidermis (4.5 versus 10.0). Curcumin led to dermal and epidermal scores of 10.5 and 4.5. Both compounds reduced leukocytic infiltration, but neither prevented epidermal dysplasia. Conclusion: COX2 expression in HPV16-induced

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lesions may be modulated by nutraceuticals, reducing tumorassociated inflammation. However, this was not sufficient to block carcinogenesis, calling for additional studies focused on combination therapies.

Infection by high-risk human papillomavirus (HPV) types, most prominently types 16 and 18 (HPV16 and HPV18) is associated with cancer of the uterine cervix and of other anatomic locations, such as the anus and the oropharynx (1). The development of HPV-associated pre-neoplastic and neoplastic lesions is associated with significant inflammatory phenomena (2), which promote stromal remodeling, angiogenesis and metastasis, and are coordinated by nuclear factor kappa B (NF-kB), as recently reviewed (3). Cyclooxygenase-2 (COX2), a key player in cancer-associated inflammation, is up-regulated by HPV16 E5, E6 and E7 oncoproteins, though NF-kB and the epithelial growth factor receptor (EGFR)-Ras-mitogen-activated protein kinase (MAPK) pathways (4, 5). In cervical lesions, COX2 upregulation modulates the immune response, promoting immunotolerance and lesion persistence (6). In line with these findings, celecoxib, a selective COX2 inhibitor, was recently reported to enhance the degranulation of CD8+ T-cells in skin lesions induced by HPV16 oncogenes in transgenic (K14-HPV16) mice (7). In the same study, celecoxib also reduced the incidence of cutaneous dysplastic lesions. This line of evidence suggests that COX2 may be a useful therapeutic target in HPV-induced cancer. Other anti-inflammatory substances have been tested against HPV-induced lesions, with different results. Among these are dietary polyphenols, most prominently curcumin (8, 9). Curcumin and other polyphenols such as quercetin (10) and resveratrol (11) inhibit NF-κB pathway signaling, down-regulating some of its downstream effectors, such as COX2. Recent observations showed that long-term oral administration of rutin (a quercetin glycoside) and curcumin ameliorated systemic inflammation and cachexia in K14-HPV16 mice (12). Considering these previous findings, and the role of COX2 in cancer induced by HPV, it has become essential to evaluate the expression of COX2 in the viral lesions of those animals. Furthermore, this study also describes how curcumin and rutin modulate COX2 expression, and their impact on tumor-associated inflammation.

#### **Materials and Methods**

Animals. K14-HPV16 mice on a FVB/n background (13), were generously donated by Drs. Jeffrey Arbeit and Douglas Hanahan, from the University of California, through the USA National Cancer Institute Mouse Repository. Animals were bred in the University of Trás-os-Montes and Alto Douro animal facility. Genotyping was performed as previously described (14, 15). Hemizygotic (HPV16+/-) and wild-type (HPV16-/-) 6-week-old females were selected for use in the study. The study was authorized by the University de Trás-os-Montes e Alto Douro Ethics Committee (approval no. 10/2013) and the Portuguese Veterinary Directorate (approval no. 0421/000/000/2014).

Chemicals. Curcumin and rutin (Sigma-Aldrich, Saint-Louis, MA, USA) were dispersed in sunflower oil and coated onto diet pellets using an industrial mixer, to a final concentration of 2.0 mg/g. The average daily doses of rutin and curcumin were 9.42±0.62 mg/animal, or 413.0 and 422.4 mg/kg, respectively. The final sunflower oil concentration was 3.0% (w/w).

Experimental procedures. The animals were maintained in accordance with Portuguese (Decree-Law 113/2013, dated 7th August) and European (EU Directive 2010/63/EU) legislation, under controlled conditions of temperature (23±2°C), light-dark cycle (12 h light/12 h dark) and relative humidity (50±10%), using hardwood bedding. Health checks were performed daily. As previously described (12), 50 mice were divided into four groups: 1: HPV16-/-, n=12; 2: HPV16+/-, n=13; 3: HPV16+/-, n=12; and 4: HPV16+/-, n=13. Groups 3 and 4 were administered rutin, and curcumin, respectively, for 24 consecutive weeks. Groups 1 and 2 received a control diet (3.0% sunflower oil). At 30 weeks old, the animals were sacrificed by intraperitoneal pentobarbital overdose followed by cardiac puncture and exsanguination.

Histological analysis. Chest skin samples were collected and fixed in 10% neutral-buffered formalin. Following histological processing, each skin sample was stained with hematoxylin and eosin and classified histologically as normal skin, epidermal hyperplasia epidermal dysplasia. Total infiltrating leukocytes and specific leukocytic populations (neutrophils, macrophages,

lymphocytes, plasma cells, mast cells) were counted in high-power (×400) fields and leukocyte counts are expressed as mean values±standard error to the mean.

Immunohistochemical analysis. Immunohistochemistry for COX2 (1:100; SP21; Transduction Laboratories, Lexington, KY, USA) was performed as previously described (16). Briefly, heat-induced epitope retrieval was performed in a microwave oven (700 W) for 20 minutes in a citrate buffer solution. Endogenous peroxidase activity was blocked with 3% hydrogen peroxide. Nonspecific staining was minimized by a 30-minute incubation with normal rabbit serum (X 0902; DakoCytomation, Glostrup, Denmark). The primary antibody was incubated overnight (4°C) in a humid chamber. Immunoreactivity was detected using a streptavidin-biotinperoxidase complex (TS-125-HR; Labvision Corporation, Freemont, CA, USA). Color development was performed using 3,3'diaminobenzidine tetrahydrochloride and the preparations were counterstained with Mayer's hematoxylin. For negative controls, sections were incubated with normal rabbit serum instead of the primary antibody. Macrophages on submandibular lymph nodes were used as internal positive controls. Immunostaining for COX2 was assessed on the dermis and epidermis according to its distribution and intensity. Concerning the distribution of positive cells, four staining patterns were observed: pattern 1, comprising cases where immunostaining was present in fewer than 25% of cells, pattern 2 with 26-50% of stained cells, pattern 3 with 51-75% of stained cells and pattern 4 with 76-100% of stained cells. The staining intensity was scored as light (1 point), moderate (2 points) or intense (3 points). The COX2 score represents the product of the pattern of positive tumor cells (1 to 4 points) and staining intensity (1 to 3 points) and ranged between 0 and 12.

Statistics. The Shapiro–Wilk test rejected the hypothesis that group data were samples from a normal distribution. Scale transformations did not change the previous hypothesis rejection. The non-parametric Kruskal–Wallis test was applied, the significance level was set to 0.05 and the significance values were adjusted by the Bonferroni correction for multiple tests. All statistical analyses were carried out using SPSS 24 (IBM Corp., Armonk, NY, USA). Results were expressed as mean±standard deviation and mean rank, when appropriate.

## Results

Cutaneous lesions and leukocytic infiltrates. No histological alterations were observed in control (group 1) animals. All treated and untreated transgenic mice (groups 2-4) showed varying degrees of epidermal hyperplasia and dysplasia (Figure 1), and there were no significant differences between groups, as previously described (12). HPV-transgenic mice showed increased numbers of tumor-associated leukocytes compared with wild-type animals (Table I). Rutin and, to a lesser extent, curcumin, reduced the leukocytic infiltration in HPV-induced lesions (Table I).

*COX2 immunoexpression*. The normal skin of wild-type mice did not express COX2, neither at the dermal nor epidermal level (Figure 2a). Untreated transgenic animals (group 2) showed

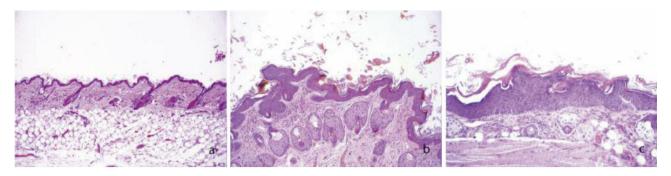


Figure 1. Histopathological changes induced by HPV16 in FVB/n mice, hematoxylin-eosin staining. a: Normal skin histology (group 1 animal), ×40. b: Epidermal hyperplasia and papillomatosis (group 2 animal), ×100. c: Epidermal dysplasia (group 2 animal), ×100.

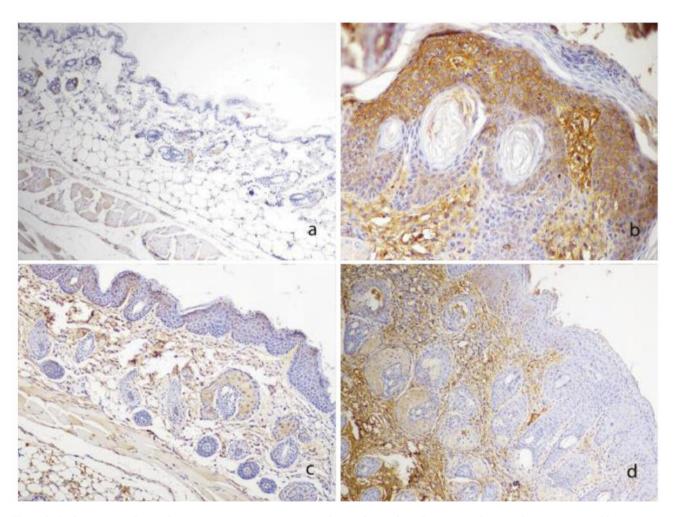


Figure 2. Cycle-oxygenase-2 (COX2) immunoexpression in experimental mice, diaminobenzidine-Mayer's hematoxylin staining. a: Wild-type mouse (group 1). COX2 expression limited to scattered dermal leukocytes, ×40. b: Untreated HPV16-transgenic mouse (group 2). Intense diffuse dermal and epidermal COX2 immunoexpression, ×100. c: Rutin-treated HPV16-transgenic mouse (group 3). Light to moderate, multifocal COX2 immunoexpression in the dermis and epidermis, ×40. d: Curcumin-treated HPV16-transgenic mouse (group 4). Light, multifocal COX2 immunoexpression epidermis, but intense diffuse immunoexpression in the dermis, ×40.

Table I. Leukocyte counts on normal skin and lesions from K14HPV16 mice. Data are the mean±SD (mean rank).

| Group                                   | Macrophages                 | Neutrophils                 | Lymphocytes                 | Mast cells     | Plasma cells   | Total leukocytes            |
|-----------------------------------------|-----------------------------|-----------------------------|-----------------------------|----------------|----------------|-----------------------------|
| 1: HPV <sup>-/-</sup> (n=12)            | 0.7±0.5 (6.5 <sup>a</sup> ) | 0.2±0.4 (6.5 <sup>a</sup> ) | 0.5±0.7 (7.1 <sup>a</sup> ) | 0.9±0.7 (6.5a) | 0.2±0.4 (15.1) | 2.3±0.8 (6.5 <sup>a</sup> ) |
| 2: HPV <sup>+/-</sup> (n=8)             | 6.6±1.2 (35.9)              | 5.8±2.3 (31.6)              | 2.9±1.1 (24.25)             | 6.4±1.2 (35.6) | 1.0±1.1 (26.4) | 21.6±3.6 (34.75)            |
| 3: HPV <sup>+/-</sup> + rutin (n=12)    | 4.8±1.3 (24.75)             | 4.3±1.6 (25.5)              | 3.7±1.4 (29.6)              | 5.1±1.3 (27.2) | 0.8±0.9 (24.7) | 17.8±3.4 (24.8)             |
| 4: HPV <sup>+/-</sup> + curcumin (n=12) | 5.5±1.7 (28.7)              | 5.2±1.9 (30.6)              | 3.9±1.6 (30.8)              | 5.0±1.4 (26.7) | 1.0±1.0 (26.6) | 19.5±3.3 (29.4)             |

<sup>&</sup>lt;sup>a</sup>Mean rank statistically different from groups 2, 3 and 4 (p<0.05).

Table II. Immunostaining scores for cyclo-oxygenase-2 on normal skin and skin lesions from K14HPV16 mice for each experimental group. Data are the mean±SD (mean rank).

| Group                                   | Extension       | Dermis intensity | Score            | Extension      | Epidermis intensity | Score            |
|-----------------------------------------|-----------------|------------------|------------------|----------------|---------------------|------------------|
| 1: HPV <sup>-/-</sup> (n=12)            | 0.0±0.0 (6.5a)  | 0.0±0.0 (6.5a)   | 0.0±0.0 (6.5a)   | 0.0±0.0 (6.5a) | 0.0±0.0 (6.5a)      | 0.0±0.0 (6.5a)   |
| 2: HPV+/- (n=8)                         | 3.8±0.5 (31.25) | 3.0±0.0 (32.5)   | 11.3±1.4 (34.75) | 3.5±0.8 (36.3) | 2.9±0.4 (37.25)     | 10.0±2.3 (39.9b) |
| 3: $HPV^{+/-} + rutin (n=12)$           | 3.1±0.7 (22.7)  | 2.6±0.5 (25.8)   | 7.8±1.9 (21.5)   | 2.7±0.9 (28.5) | 1.8±0.7 (23.8)      | 4.5±1.2 (24.5)   |
| 4: HPV <sup>+/-</sup> + curcumin (n=12) | 3.8±0.4 (32.5)  | 2.8±0.5 (28.5)   | 10.5±1.9 (31.3)  | 2.1±0.7 (23.3) | 2.2±0.4 (27.3)      | 4.5±1.5 (24.8)   |

aStatistically different from groups 2, 3 and 4 (p<0.05); bstatistically different from groups 1, 3 and 4 (p<0.05).

moderate to intense, diffuse COX2 immunoexpression in the epidermis (Table II), where it assumed a cytoplasmic or membrane-associated pattern. There was also intense, diffuse dermal COX2 immunoexpression (Figure 2b). The expression of COX2 was reduced by rutin administration in group 3 (Figure 2c) in the dermis (immunostaining score 7.83 *versus* 11.25) and epidermis (4.5 *versus* 10.0, p<0.05) of transgenic mice. Curcumin (Figure 2d) also reduced the immunoexpression of COX2 in the epidermis (4.5 score, p<0.05) but not on the dermis (10.5 score) of group 4 mice.

#### Discussion

Curcumin, isolated from the rhizomes of *Curcuma longa*, has long been used for nutritional and pharmaceutical purposes due to its anti-inflammatory and anti-neoplastic properties (17). Rutin, another polyphenolic compound, is a glycoside of the flavonoid quercetin, found in numerous edible plants and fruits. Rutin is one of quercetin's most common presentations and its digestion releases the sugar moiety, allowing the absorption of the quercetin aglycone (18). Curcumin inhibits the I kappa B kinase (IkK) complex, blocking I kappa B (IkB) phosphorylation and the nuclear translocation of NF-kB dimers, down-regulating COX2 (19), but not the expression of constitutive COX1 (20). Other polyphenols, such as quercetin, are also reported to down-regulate COX2 by inhibiting NF-kB signallng (19, 21). In many HPV-induced lesions, multistep carcinogenesis is accompanied by increasing

inflammation, underpinned by NF-kB, which is considered essential for tumor progression and metastasis (3).

Accordingly, in K14-HPV16 mice, the dysplastic stage coincides with enhanced sub-epidermal inflammation and angiogenesis (22, 23). In line with these findings, we observed significantly enhanced COX2 expression in skin samples from HPV16-transgenic animals compared with samples from wildtype ones (Figure 2). In transgenic mice, COX2 was overexpressed in the dermis and epidermis, accompanying the strong dermal inflammatory infiltrates observed in these animals. This agrees with the idea that HPV16 oncoproteins drive the expression of pro-inflammatory mediators to modulate the tumor microenvironment (2). Prolonged oral rutin administration effectively reduced dermal and epidermal expression of COX2, resulting in reduced leukocyte (mostly macrophages and neutrophils) infiltration (Table I), although this effect did not reach statistical significance. Curcumin was less effective, reducing epidermal, but not dermal COX2 expression and led to a more intense leukocyte infiltration, especially concerning macrophages and neutrophils. Although neither compound was able to reduce the incidence of dysplastic lesions, both showed interesting immunomodulatory properties in this model. This agrees with our previously published observations on this model (12), showing reduced systemic inflammation in mice treated with either curcumin or rutin. It is also in line with results by other research groups, which have shown quercetin to exert interesting antiinflammatory and anti-neoplastic properties (24).

In the K14-HPV16 mouse model, curcumin seemed to exert a less potent anti-inflammatory effect, despite its known efficacy against some HPV-induced lesions (25), a discrepancy that may be explained by its poor pharmocokinetic properties (26). Taken together, the present results describe the expression pattern of COX2 in the K14HPV16 mouse model and help validate it for testing anti-inflammatory and immunoregulatory compounds. Both rutin and curcumin reduced COX2 expression in HPV16-transgenic mice, but this was not sufficient to stop cancer progression. Future studies may take advantage of the anti-inflammatory properties of these compounds by including them in combination therapies.

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