

## Histone Deacetylase Inhibitors as a Novel Targeted Therapy Against Non-small Cell Lung Cancer: Where Are We Now and What Should We Expect?

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**Abstract.** Non-small cell lung cancer constitutes the most common type of lung cancer, accounting for 85-90% of lung cancer, and is a leading cause of cancer-related death. Despite the progress during the past years, poor prognosis remains a challenge and requires further research and development of novel antitumor treatment. Recently, the role

of histone deacetylases in gene expression has emerged showing their regulation of the acetylation of histone proteins and other non-histone protein targets and their role in chromatin organization, while their inhibitors, the histone deacetylase inhibitors, have been proposed to have a potential therapeutic role in diverse malignancies, including non-small cell lung cancer. This review article focuses on the role of histone deacetylase inhibitors in the treatment of non-small cell lung cancer and the major molecular mechanisms underlying their antitumor activity recognized so far.

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Primary lung cancer remains the most common malignancy after non-melanocytic skin cancer and is a leading cause of cancer-related death globally (1-3). Non-small cell lung cancer (NSCLC) accounts for 85-90% of lung cancer (1). The order of frequency of different histological subtypes has changed; incidence of adenocarcinoma has increased and nowadays represents the most frequent pulmonary malignancy, followed by squamous cell carcinoma (1, 3). Usually NSCLC is diagnosed in advanced stages, with poor

prognosis and limited therapeutic decisions. Despite the recent progress noted in the treatment of NSCLC with the introduction of immunotherapy (3-7) and the approval of tyrosine kinase inhibitors (TKIs) as first-line therapy (3, 8), there is an urgent need for further development of novel and more efficient antitumor treatment.

The role of histones and modifications in their *N*-termini, including acetylation, methylation, phosphorylation and ubiquitination, have been well recognized in chromatin organization and consequently in regulation of gene expression (9-12). More specifically, in eukaryotic cell nuclei, DNA is wrapped around proteins called histones. In this way, the structure of the nucleosome is formed, containing the wrapped DNA around a central histone octamer. Histones often present various modifications that permit the loosening of the nucleosome structure thereby allowing access of RNA polymerase and other transcription factors to their target genes in order to promote the transcriptional process (9-12).

Histone acetylation is the first among histone modifications identified and plays a principal role in the aforementioned regulation of transcription (12). It concerns a dynamic process, involving two groups of enzymes, histone acetyltransferases (HATs) and histone deacetylases (HDACs) (13-15). It is well known that histone acetylation modulates transcription in multiple ways. The result of histone acetylation is reduced interaction between histone and DNA. Increased histone acetylation has been associated with transcriptionally active genes, whilst low levels of acetylation correlate with decreased transcriptional action (13-16).

### HDACs and Their Mechanisms of Action

HDACs are enzymes that remove acetyl groups from histone lysine residues of proteins, such as the core nucleosome histones, in this way not permitting DNA to loosen from the histone octamer and consequently preventing its transcription (Figure 1) (13, 16). Thus, histone deacetylation contributes to transcriptional repression favoring chromatin compaction (17). According to their homology, four classes of HDACs have been recognized: class I, which includes the nuclear localized HDAC-1, -2, -3 and -8; class II, which includes the exonuclear as well as the nuclear localized HDAC-4, -5, -6, -7, -9 and -10; class III, the sirtuins (SIRT-1-7); and class IV, HDAC-11, with features of both class I and II; in total there are 18 HDACs (18). However, they can further divided into  $Zn^{2+}$ -dependent classes (class I, II and IV) and NAD-dependent classes (class III) (Table I). The role of HDACs as transcriptional repressors has been examined in studies including biochemical analyses *in vitro*, cultured cells and HDAC knockdown models (19). Furthermore, recent studies have shown the role of HDACs in the progression of a

variety of malignancies (20-23). Moreover, HDACs have been shown to contribute to cellular homeostasis and regulation of fundamental functions, such as cell-cycle progression, differentiation and apoptosis through deacetylation of non-histone proteins (24).

### The Role of HDACs in Cancer

The levels of many HDAC isoenzymes have been found increased in diverse malignancies (25). In particular, overexpression of HDAC-1 has been described in gastric cancer, while HDAC-2 and -3 in colorectal cancer. Similarly, activity of HDAC-6 and HDAC-7 have also been found increased in cutaneous T-cell lymphoma and pancreatic adenocarcinoma, respectively (26, 27). The complete mechanisms of action of HDACs have not been fully elucidated yet, as the involved pathways are considerably complex. However, an example of the exact meaning of the increased expression of HDACs and the subsequent transcriptional suppression of genes of hematopoietic differentiation has been well described in certain types of leukemia. For example in acute promyelocytic leukemia, chromosomal rearrangements of the retinoic acid receptor (RAR) transcription factor and the modified interactions with other transcriptional co-regulators, including nuclear receptor co-repressor (NCoR) and silencing mediator for retinoid or thyroid-hormone receptors (SMRT), result in recruitment of HDACs and contribute to transcriptional repression of specific target genes (28).

### HDAC Inhibitors and Their Role as Anticancer Agents in NSCLC

Recent data has shown that HDAC inhibitors contribute to tumor cell growth arrest, enhancing cell apoptosis and promoting cell-cycle arrest (29-33). Their capacity to enhance the acetylation of cellular proteins by blocking HDAC activity reflects the importance of their potential anticancer action. Diverse studies have highlighted their role in diverse malignancies, including breast cancer (34, 35), and medullary thyroid (36) and pancreatic cancer (37, 38). The potential actions of HDAC inhibitors as novel anticancer agents have been supported based on their capacity to induce apoptosis and autophagy in cancer cells, by activating intrinsic mitochondrial pathways (39). In addition, they have been shown to up-regulate tumor suppressor genes, such as Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) and Death Receptor 5 (*DR5*), involved in the apoptotic pathways and, on the contrary, to inhibit the expression of pro-survival genes, including B-cell lymphoma 2 (*BCL2*) (39, 40). Moreover, HDAC inhibitors also exhibit indirect actions, by enhancing immune responses and up-regulating major histocompatibility complex class I and II

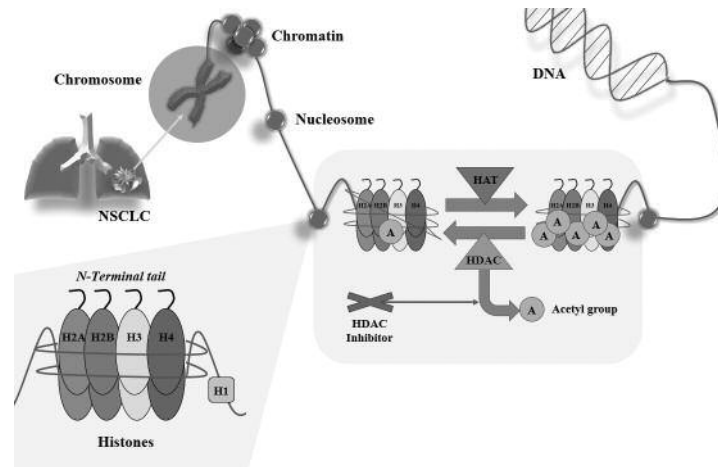


Figure 1. Mechanism of histone deacetylase (HDAC) activity. NSCLC: Non-small cell lung cancer; HAT: histone acetyltransferase.

Table I. Classification of histone deacetylases (HDAC).

Class	HDAC	Dependence
I	HDAC-1, -2, -3 and -8	Zn <sup>2+</sup>
II	HDAC-4, -5, -6, -7, -9 and -10	
III	SIRT-1-7	NAD
IV	HDAC-11	Zn <sup>2+</sup>

SIRT: Sirtuins.

proteins and cytokine secretion, as well as other co-stimulatory molecules, including CD80 and CD86. There exist some evidence that they also have anti-angiogenic effects (39, 40). Furthermore, treatment of cells with HDAC inhibitors such as butyrate, trichostatin A (TSA), and trapoxin A, has been shown to contribute to the transcription of target genes (41). To date three HDAC inhibitors, namely vorinostat, romidepsin and panobinostat (LBH-589, PS), have received US Food and Drug Administration approval and belong to the therapeutic armamentarium against cutaneous and peripheral T-cell lymphoma and multiple myeloma (42).

Lung cancer still remains the leading cause of cancer-related deaths worldwide with approximately 221,200 estimated new cases (39). NSCLC accounts for 85%-90% of lung cancer. Usually the diagnosis is established in advanced stages, with poor prognosis and limited therapeutic decisions (39). Current research focuses on the potential utility of HDAC inhibitors as monotherapy or in combination with other chemotherapeutic regimens in NSCLC in an effort to increase their efficacy or reduce tumor resistance.

More specifically, TSA is an antifungal antibiotic and has reversible HDAC inhibitory activity, regulating apoptosis, angiogenesis and cell differentiation. Studies based on NSCLC cell lines have reported the antitumor action of vorinostat and TSA by inhibiting tumor cell growth. Mukhopadhyay *et al.*, evaluated the role of TSA in NSCLC lines compared to normal lung fibroblasts (41). Their results confirmed the apoptotic action of TSA. Moreover, the authors concluded that TSA increased histone H4 acetylation and the expression of p21, whilst no significant effect on p16, p27, cyclin-dependent kinase 2 (CDK2), and cyclin D1 were observed (41). All the above findings of the TSA-treated NSCLC cells were consistent with the apoptotic effects that TSA has on tumor cells. The exact mechanism of the induced apoptosis is not clear. However, the authors proposed that TSA may induce expression of p21, which is associated with G<sub>1</sub> phase arrest (41). In addition, it has been demonstrated that lung cancer cells exhibit diverse histone H4 modifications, characterized by either hyperacetylation of H4K5/H4K8 or hypoacetylation of H4K12/H4K16. Seligson *et al.* demonstrated that decreased levels of histone modifications are associated with more aggressive cancer phenotype in lung adenocarcinoma (43). In parallel to the above results, diverse studies have found that HDAC-1 gene expression is associated with lung cancer progression (44, 45). Kim *et al.* found that treatment with TSA resulted in a dose-dependent reduction of H157 lung cancer cells (46). The proposed mechanism of TSA action was the induction of apoptosis by activating the intrinsic mitochondrial and extrinsic/Fas/FasL system death pathways. TSA seems to act synergistically with other HDAC inhibitors, including vorinostat (47) (Table II). Furthermore, Piao *et al.* demonstrated that combination of TSA with a mechanistic

Table II. Proposed actions and involved molecular pathways of histone deacetylase (HDAC) inhibitors in non-small cell lung cancer (NSCLC).

HDAC inhibitor	Action
Trichostatin A (TSA)	Inhibits tumor cell growth Promotes apoptosis in cancer cell lines by activating the intrinsic mitochondrial and extrinsic/Fas/FasL system death pathways Increases histone H4 acetylation and the expression of p21 Sensitizes cancer cells to chemotherapy <i>in vitro</i>
Vorinostat	Exhibits antiproliferative activity Up-regulates the cyclin-dependent kinase inhibitor p21 Synergistic action with carboplatin and paclitaxel and epidermal growth factor receptor tyrosine kinase inhibitors Sensitizes NSCLC cell lines to ionizing radiation through activation of caspase-8
Valproic acid	Synergistic action with cisplatin –vinorelbine Enhances the efficacy of ionizing radiation
Belinostat	Synergistic action with cyclin-dependent kinase inhibitor CYC202 Antiproliferative action
Panobinostat	Increases apoptosis in NSCLC cell lines Synergistic action with erlotinib Induction of apoptosis

Table III. Studies of histone deacetylase (HDAC) inhibitors in non-small cell lung cancer (NSCLC).

Molecule	Phase	Author	Line of treatment	Sample size	Limitations
Vorinostat	Phase II study	Ramalingam <i>et al.</i> (48)	In patients with NSCLC in combination with carboplatin and paclitaxel	94	Increased toxicity
Valproic acid	Phase II study	Traynor <i>et al.</i> (51)	In patients with relapsed NSCLC	16	No objective antitumor activity was seen
	Pre-clinical	Shirsath <i>et al.</i> (57)	In NSCLC cell lines	-	<i>In vitro</i> study, further studies needed
	Pre-clinical	Chen <i>et al.</i> (55)	In NSCLC cell lines	-	<i>In vitro</i> study, further studies needed
	Pre-clinical	Gavrilov <i>et al.</i> (56)	In NSCLC cell lines	-	<i>In vitro</i> study, further studies needed
Belinostat	Pre-clinical	Ong <i>et al.</i> (58)	In NSCLC cell lines	-	<i>In vitro</i> study, further studies needed
Panobinostat	Pre-clinical	Greve <i>et al.</i> (42)	In EGFR-mutated and wild-type NSCLC cells.	-	<i>In vitro</i> study, further studies needed
	Phase I trial	Gray <i>et al.</i> (59)	In patients with advanced NSCLC and head-and-neck cancer	42	Small number of participants

EGFR: Epidermal growth factor receptor.

target of rapamycin inhibitor had an increased synergistic therapeutic antitumor effect on NSCLC cell migration and invasion *in vitro*, suggesting their additional role in sensitizing cancer cells to chemotherapy (48).

Vorinostat represents another HDAC inhibitors with potential therapeutic action in NSCLC. Vorinostat exhibited antiproliferative activity by repressing telomerase activity *via* up-regulation of the cyclin-dependent kinase inhibitor *p21*. The induction of *p21* resulted in G<sub>0</sub>-G<sub>1</sub> cell-cycle arrest when human lung cancer lines were treated with vorinostat (46, 49). The potential therapeutic benefit of the addition of vorinostat to the first-line carboplatin and paclitaxel was

evaluated in pre-clinical and clinical studies, which showed improved response rates (50, 51). Lately, research interest has been focused on the benefit of the use of HDAC inhibitors in combination with epidermal growth factor receptor (EGFR) TKIs (52). The Wisconsin Oncology Network phase II study by Traynor *et al.* showed that monotherapy with vorinostat in patients with relapsed NSCLC provided significant benefit regarding time to progression, however, no objective antitumor activity was observed (53). Vorinostat has been also shown to sensitize NSCLC cell lines to ionizing radiation through activation of caspase-8. These results could imply a potential clinical

benefit of HDAC inhibitors in improving response to radiotherapy in NSCLC (54) (Table II).

Valproic acid (VPA) represents another HDAC inhibitor. Recently, Chen *et al.* demonstrated that despite the fact that VPA did not affect NSCLC cell proliferation, it did induce increased sensitivity to cisplatin (55). VPA has also been also proposed to enhance the efficacy of the anticancer activity of combination therapy with cisplatin-vinorelbine and ionizing radiation and also reduce their side-effects (56) (Table II). Recently, it was shown that VPA had a potential synergistic therapeutic effect on NSCLC cell lines when combined with a CDK inhibitor, supporting more evidence of a potential novel antitumor activity (57).

Belinostat is a composite class I and II HDAC inhibitor. Recently, the novel combination of belinostat and CDK inhibitor CYC202 was evaluated as a potential anticancer strategy in NSCLC in an *in vitro* study of *p53* wild-type A549 cells. Concurrent treatment led to significant reduction in cell proliferation and an increase in apoptosis (58) (Table II).

Panobinostat is a novel HDAC inhibitor that was recently demonstrated to sensitize *EGFR*-mutated and wild-type NSCLC cells to the antiproliferative activity of erlotinib. Furthermore, this combination enhanced the induced acetylation of histone H3 (42). A small phase I trial of combination therapy with panobinostat and TKI in patients with advanced NSCLC and head-and-neck cancer showed that is a well-tolerated therapeutic regimen. However, larger randomized controlled studies are needed to elucidate its clear benefits in erlotinib-resistant NSCLC (59) (Table II). Table III summarizes current studies of HDAC inhibitors in NSCLC.

## Conclusion

Despite much progress, poor prognosis remains a crucial issue in the therapeutic management of NSCLC. Alterations in histone acetylation result in crucial changes in gene expression and HDACs play a vital role in this process, underlying the potential importance of HDAC inhibitors as alternative or additional targets of antitumor therapy. However, further pre-clinical and clinical studies of HDAC inhibitors in NSCLC are required for evaluating their antitumor activity. Clinical trials should also assess the potential benefit of combining HDAC inhibitors with other antitumor drugs. The approach of utilizing HDAC inhibitors in combination with standard chemotherapy deserves further assessment since it could potentially enhance efficacy and reduce the side-effects of current anticancer regimens.

## References

- Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D: Global cancer statistics. *CA Cancer J Clin* 61: 69-90, 2011.
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A: Global cancer statistics, 2012. *CA Cancer J Clin* 65: 87-108, 2015.
- Novello S, Barlesi F, Califano R, Cufer T, Ekman S, Levra MG, Kerr K, Popat S, Reck M, Senan S, Simo GV, Vansteenkiste J and Peters S: ESMO Guidelines Committee. Metastatic non-small-cell lung cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 27(Suppl 5): v1-v27, 2016.
- Herbst RS, Baas P, Kim DW, Felip E, Pérez-Gracia JL, Han JY, Molina J, Kim JH, Arvis CD, Ahn MJ, Majem M, Fidler MJ, de Castro G Jr., Garrido M, Lubiniecki GM, Shentu Y, Im E, Dolled-Filhart M and Garon EB: Pembrolizumab *versus* docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. *Lancet* 387: 1540-1550, 2016.
- Rizvi NA, Mazières J, Planchard D, Stinchcombe TE, Dy GK, Antonia SJ, Horn L, Lena H, Minenza E, Mennecier B, Otterson GA, Campos LT, Gandara DR, Levy BP, Nair SG, Zalcman G, Wolf J, Souquet PJ, Baldini E, Cappuzzo F, Chouaid C, Dowlati A, Sanborn R, Lopez-Chavez A, Grohe C, Huber RM, Harbison CT, Baudalet C, Lestini BJ and Ramalingam SS: Activity and safety of nivolumab, an anti- PD-1 immune checkpoint inhibitor, for patients with advanced, refractory squamous non-small-cell lung cancer (CheckMate 063): a phase 2, single-arm trial. *Lancet Oncol* 16: 257-265, 2015.
- Brahmer J, Reckamp KL, Baas P, Crinò L, Eberhardt WE, Poddubskaya E, Antonia S, Pluzanski A, Vokes EE, Holgado E, Waterhouse D, Ready N, Gainor J, Arén Frontera O, Havel L, Steins M, Garassino MC, Aerts JG, Domine M, Paz-Ares L, Reck M, Baudalet C, Harbison CT, Lestini B and Spigel DR: Nivolumab *versus* docetaxel in advanced squamous-cell non-small-cell lung cancer. *N Engl J Med* 373: 123-135, 2015.
- Reckamp K, Brahmer JR, Spigel DR, Rizvi NA, Poddubskaya E, West H, Eberhardt WEE, Baas P, Antonia SJ, Pluzanski A, Vokes E, Holgado E, Waterhouse D, Ready N, Gainor JF, Frontera OA, Horn L, Paz-Ares L, Li A and Lynch M: 1 Phase 3, randomized trial (CheckMate 017) of nivolumab (NIVO) *vs.* docetaxel in advanced squamous (SQ) cell non-small cell lung cancer (NSCLC). *J Thorac Oncol* 10(Suppl 2): S174, 2015.
- Lindeman NI, Cagle PT, Beasley MB, Chitale DA, Dacic S, Giaccone G, Jenkins RB, Kwiatkowski DJ, Saldivar JS, Squire J, Thunnissen E and Ladanyi M: Molecular testing guideline for selection of lung cancer patients for *EGFR* and *ALK* tyrosine kinase inhibitors: guideline from the College of American Pathologists, International Association for the Study of Lung Cancer, and Association for Molecular Pathology. *J Thorac Oncol* 8: 823-859, 2013.
- Glozak MA and Seto E: Histone deacetylases and cancer. *Oncogene* 26: 5420-5432, 2007.
- Körner M and Tibes U: Histone deacetylase inhibitors: a novel class of anti-cancer agents on its way to the market. *Prog Med Chem* 46: 205-280, 2008.
- Kothapalli N, Sarath G and Zemleni J: Biotinylation of K12 in histone H4 decreases in response to DNA double-strand breaks in human JAr choriocarcinoma cells. *J Nutr* 135: 2337-2342, 2005.
- Li Z and Zhu WG: Targeting histone deacetylases for cancer therapy: from molecular mechanisms to clinical implications. *Int J Biol Sci* 10: 757-770, 2014.
- Kouraklis G and Theocharis S: Histone deacetylase inhibitors: a novel target of anticancer therapy (review). *Oncol Rep* 15: 489-494, 2006.

- 14 Vigushin DM and Coombes RC: Histone deacetylase inhibitors in cancer treatment. *Anticancer Drugs* 13: 1-13, 2002.
- 15 Johnstone RW: Histone-deacetylase inhibitors: novel drugs for the treatment of cancer. *Nature* 1: 287-299, 2002.
- 16 Ausio J and van Holde KE: Histone hyperacetylation: its effects on nucleosome conformation and stability. *Biochemistry* 25: 1421-1428, 1986.
- 17 Ruthenburg AJ, Li H, Patel DJ and Allis CD: Multivalent engagement of chromatin modifications by linked binding modules. *Nature Rev Mol Cell Biol* 8: 983-994, 2007.
- 18 Trapp J and Jung M: The role of NAD<sup>+</sup>-dependent histone deacetylases (sirtuins) in ageing. *Curr Drug Targets* 7: 1553-1560, 2006.
- 19 Haberland M, Montgomery RL and Olson EN: The many roles of histone deacetylases in development and physiology: implications for disease and therapy. *Nat Rev Genet* 10: 32-42, 2009.
- 20 Li A, Liu Z, Li M, Zhou S, Xu Y, Xiao Y and Yang W: HDAC5, a potential therapeutic target and prognostic biomarker, promotes proliferation, invasion and migration in human breast cancer. *Oncotarget* 7: 37966-37978, 2016.
- 21 Krusche CA, Wülfing P, Kersting C, Vloet A, Böcker W, Kiesel L, Beier HM and Alfer J: Histone deacetylase-1 and -3 protein cancer: a tissue microarray analysis. *Breast Cancer Res Treat* 90: 15-23, 2005.
- 22 Stojanovic N, Hassan Z, Wirth M, Wenzel P, Beyer M, Schäfer C, Brand P, Kroemer A, Stauber RH, Schmid RM, Arlt A, Sellmer A, Mahboobi S, Rad R, Reichert M, Saur D, Krämer OH and Schneider G: HDAC1 and HDAC2 integrate the expression of p53 mutants in pancreatic cancer. *Oncogene* 36: 1804-1815, 2017.
- 23 Zhang Z, Yamashita H, Toyama T, Sugiura H, Ando Y, Mita K, Hamaguchi M, Hara Y, Kobayashi S and Iwase H: Quantitation of *HDAC1* mRNA expression in invasive carcinoma of the breast. *Breast Cancer Res Treat* 94: 11-16, 2005.
- 24 Minucci S and Pelicci PG: Histone deacetylase inhibitors and the promise of epigenetic (and more) treatments for cancer. *Nat Rev Cancer* 6: 38-51, 2006.
- 25 Yoo CB and Jones PA: Epigenetic therapy of cancer: Past, present and future *Nat Rev Drug Discov* 5: 37-50, 2006.
- 26 Ellis L and Pili R: Histone deacetylase inhibitors: Advancing therapeutic strategies in hematological and solid malignancies. *Pharmaceuticals* 3: 2441-2469, 2010.
- 27 Ouaiissi M, Giger U, Sielezneff I, Pirrò N, Sastre B and Ouaiissi A: Rationale for possible targeting of histone deacetylase signaling in cancer diseases with a special reference to pancreatic cancer. *J Biomed Biotechnol* 2011: 315939, 2011.
- 28 Minucci S, Nervi C, Lo Coco F and Pelicci PG: Histone deacetylases: a common molecular target for differentiation treatment of acute myeloid leukemias? *Oncogene* 20: 3110-3115, 2001.
- 29 Sambucetti LC, Fischer DD, Zabluoff S, Kwon PO, Chamberlain H, Trogani N, Xu H and Cohen D: Histone deacetylase inhibition selectively alters the activity and expression of cell cycle proteins leading to specific chromatin acetylation and antiproliferative effects. *J Biol Chem* 274: 34940-34947, 1999.
- 30 Kim MS, Kwon HJ, Lee YM, Baek JH, Jang JE, Lee SW, Moon EJ, Kim HS, Lee SK, Chung HY, Kim CW and Kim KW: Histone deacetylases induce angiogenesis by negative regulation of tumor suppressor genes. *Nat Med* 7: 437-443, 2001.
- 31 Butler LM, Zhou X, Xu WS, Scher HI, Rifkind RA, Marks PA and Richon VM: The histone deacetylase inhibitor SAHA arrests cancer cell growth, up-regulates thioredoxin-binding protein-2, and down-regulates thioredoxin. *Proc Natl Acad Sci USA* 99: 11700-11705, 2002.
- 32 Richon VM, Sandhoff TW, Rifkind RA and Marks PA: Histone deacetylase inhibitors selectively induces *p21<sup>WAF1</sup>* expression and gene-associated histone acetylation. *Proc Natl Acad Sci USA* 97: 10014-10019, 2000.
- 33 Yu X, Guo S, Marcu MG, Necker L, Nguyen DM, Chen GA and Schrupp DS: Modulation of p53, ErbB1, ErbB2 and Raf-1 expression in lung cancer cells by depsipeptide FR901228. *J Natl Cancer Inst* 94: 504-513, 2002.
- 34 Damaskos C, Garmpis N, Valsami S, Kontos M, Spartalis E, Kalampokas T, Kalampokas E, Athanasiou A, Moris D, Daskalopoulou A, Davakis S, Tsourouflis G, Kontzoglou K, Perrea D, Nikiteas N and Dimitroulis D: Histone deacetylase inhibitors: an attractive therapeutic strategy against breast cancer. *Anticancer Res* 37: 35-46, 2017.
- 35 Garmpis N, Damaskos C, Garmpi A, Kalampokas E, Kalampokas T, Spartalis E, Daskalopoulou A, Valsami S, Kontos M, Nonni A, Kontzoglou K, Perrea D, Nikiteas N and Dimitroulis D: Histone deacetylases as new therapeutic targets in triple-negative breast cancer: Progress and promises. *Cancer Genomics Proteomics* 14: 299-313, 2017.
- 36 Damaskos C, Garmpis N, Valsami S, Spartalis E, Antoniou EA, Tomos P, Karamaroudis S, Zoumpou T, Pergialiotis V, Stergios K, Michaelides C, Kontzoglou K, Perrea D, Nikiteas N and Dimitroulis D: Histone deacetylase inhibitors: a novel therapeutic weapon against medullary thyroid cancer? *Anticancer Res* 36: 5019-5024, 2016.
- 37 Damaskos C, Garmpis N, Karatzas T, Nikolidakis L, Kostakis ID, Garmpi A, Karamaroudis S, Boutsikos G, Damaskou Z, Kostakis A and Kouraklis G: Histone deacetylase (HDAC) inhibitors: current evidence for therapeutic activities in pancreatic cancer. *Anticancer Res* 35: 3129-3135, 2015.
- 38 Giaginis C, Damaskos C, Koutsounas I, Zizi-Serbetzoglou A, Tsoukalas N, Patsouris E, Kouraklis G and Theocharis S: Histone deacetylase (HDAC)-1, -2, -4 and -6 expression in human pancreatic adenocarcinoma: associations with clinicopathological parameters, tumor proliferative capacity and patients' survival. *BMC Gastroenterol* 15: 148, 2015.
- 39 Ansari J, Shackelford RE and El-Osta H: Epigenetics in non-small cell lung cancer: from basics to therapeutics. *Transl Lung Cancer Res* 5: 155-171, 2016.
- 40 Miyana A, Gemma A, Noro R, Kataoka K, Matsuda K, Nara M, Okano T, Seike M, Yoshimura A, Kawakami A, Uesaka H, Nakae H and Kudoh S: Antitumor activity of histone deacetylase inhibitors in non-small cell lung cancer cells: development of a molecular predictive model. *Mol Cancer Ther* 7: 1923-1930, 2008.
- 41 Mukhopadhyay NK, Weisberg E, Gilchrist D, Bueno R, Sugarbaker DJ and Jaklitsch MT: Effectiveness of trichostatin A as a potential candidate for anticancer therapy in non-small-cell lung cancer. *Ann Thorac Surg* 81: 1034-1042, 2006.
- 42 Greve G, Schiffmann I, Pfeifer D, Pantic M, Schüler J and Lübbert M: The pan-HDAC inhibitor panobinostat acts as a sensitizer for erlotinib activity in *EGFR*-mutated and -wild-type non-small cell lung cancer cells. *BMC Cancer* 15: 947, 2015.

- 43 Seligson DB, Horvath S, McBrien MA, Mah V, Yu H, Tze S, Wang Q, Chia D, Goodglick L and Kurdastani SK: Global levels of histone modifications predict prognosis in different cancers. *Am J Pathol* 174: 1619-1628, 2009.
- 44 Sasaki H, Moriyama S, Nakashima Y, Kobayashi Y, Kiriya M, Fukai I, Yamakawa Y and Fujii Y: Histone deacetylase 1 mRNA expression in lung cancer. *Lung Cancer* 46: 171-178, 2004.
- 45 Minamiya Y, Ono T, Saito H, Takahashi N, Ito M, Motoyama S and Ogawa J: Strong expression of HDAC3 correlates with a poor prognosis in patients with adenocarcinoma of the lung. *Tumour Biol* 31: 533-539, 2010.
- 46 Kim HR, Kim EJ, Yang SH, Jeong ET, Park C, Lee JH, Youn MJ, So HS and Park R: Trichostatin A induces apoptosis in lung cancer cells *via* simultaneous activation of the death receptor-mediated and mitochondrial pathway? *Exp Mol Med* 38: 616-624, 2006.
- 47 Seo SK, Jin HO, Woo SH, Kim YS, An S, Lee JH, Hong SI, Lee KH, Choe TB and Park IC: Histone deacetylase inhibitors sensitize human non-small cell lung cancer cells to ionizing radiation through acetyl p53-mediated c-myc down-regulation. *J Thorac Oncol* 6: 1313-1319, 2011.
- 48 Piao J, Chen L, Quan T, Li L, Quan C, Piao Y, Jin T and Lin Z: Superior efficacy of co-treatment with the dual PI3K/mTOR inhibitor BEZ235 and histone deacetylase inhibitor Trichostatin A against NSCLC. *Oncotarget* 7: 60169-60180, 2016.
- 49 Komatsu N, Kawamata N, Takeuchi S, Yin D, Chien W, Miller CW and Koeffler HP: SAHA, a HDAC inhibitor, has profound anti-growth activity against non-small cell lung cancer cells. *Oncol Rep* 15: 187-191, 2006.
- 50 Ramalingam SS, Maitland ML, Frankel P, Argiris AE, Koczywas M, Gitlitz B, Thomas S, Espinoza-Delgado I, Vokes EE, Gandara DR and Belani CP: Carboplatin and paclitaxel in combination with either vorinostat or placebo for first-line therapy of advanced non-small-cell lung cancer. *J Clin Oncol* 28: 56-62, 2010.
- 51 Owonikoko TK, Ramalingam SS, Kanterewicz B, Balis TE, Belani CP and Hersherberger PA: Vorinostat increases carboplatin and paclitaxel activity in non-small cell lung cancer cells. *Int J Cancer* 126: 743-755, 2010.
- 52 Neal JW and Sequist LV: Complex role of histone deacetylase inhibitors in the treatment of non-small-cell lung cancer. *J Clin Oncol* 30: 2280-2282, 2012.
- 53 Traynor AM, Dubey S, Eickhoff JC, Kolesar JM, Schell K, Huie MS, Groteluschen DL, Marcotte SM, Hallahan CM, Weeks HR, Wilding G, Espinoza-Delgado I and Schiller JH: Vorinostat (NSC# 701852) in patients with relapsed non-small cell lung cancer: a Wisconsin Oncology Network phase II study. *J Thorac Oncol* 4: 522-526, 2009.
- 54 McLaughlin KA, Nemeth Z, Bradley CA, Humphreys L, Stasik I, Fenning C, Majkut J, Higgins C, Crawford N, Holohan C, Johnston PG, Harrison T, Hanna GG, Butterworth KT, Prise KM and Longley DB: FLIP: A targetable mediator of resistance to radiation in non-small cell lung cancer. *Mol Cancer Ther* 15: 2432-2441, 2016.
- 55 Chen JH, Zheng YL, Xu CQ, Gu LZ, Ding ZL, Qin L, Wang Y, Fu R, Wan YF and Hu CP: Valproic acid (VPA) enhances cisplatin sensitivity of non-small cell lung cancer cells *via* HDAC2 mediated down-regulation of *ABCA1*. *Biol Chem* 398: 785-792, 2016.
- 56 Gavrilov V, Lavrenkov K, Ariad S and Shany S: Sodium valproate, a histone deacetylase inhibitor, enhances the efficacy of vinorelbine-cisplatin-based chemoradiation in non-small cell lung cancer cells. *Anticancer Res* 34: 6565-6572, 2014.
- 57 Shirsath N, Rathos M, Chaudhari U, Sivaramakrishnan H and Joshi K: Potentiation of anticancer effect of valproic acid, an antiepileptic agent with histone deacetylase inhibitory activity, by the cyclin-dependent kinase inhibitor P276-00 in human non-small-cell lung cancer cell lines. *Lung Cancer* 82: 214-221, 2013.
- 58 Ong PS, Wang L, Chia DM, Seah JY, Kong LR, Thuya WL, Chinnathambi A, Lau JY, Wong AL, Yong WP, Yang D, Ho PC, Sethi G and Goh BC: A novel combinatorial strategy using Seliciclib(®) and Belinostat(®) for eradication of non-small cell lung cancer *via* apoptosis induction and BID activation. *Cancer Lett* 381: 49-57, 2016.
- 59 Gray JE, Haura E, Chiappori A, Tanvetyanon T, Williams CC, Pinder-Schenck M, Kish JA, Kreaehling J, Lush R, Neuger A, Tetteh L, Akar A, Zhao X, Schell MJ, Bepler G and Altiock S: A phase I, pharmacokinetic and pharmacodynamic study of panobinostat, an HDAC inhibitor, combined with erlotinib in patients with advanced aerodigestive tract tumors. *Clin Cancer Res* 20: 1644-1655, 2014.

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