

Review

Cancer Immunotherapy Using NKG2D and DNAM-1 Systems

TAKASHI MORISAKI^{1,2}, HIDEYA ONISHI² and MITSUO KATANO²

¹Fukuoka General Cancer Clinic, Fukuoka, Japan;

²Department of Cancer Therapy and Research, Graduate School of Medical Sciences, Kyushu University, Fukuoka, Japan

Abstract. Although tumor antigen-specific immunotherapy, such as dendritic cell vaccine, has recently emerged as a promising clinical approach, one limitation of tumor antigen- and T-cell receptor (TcR)- specific immunotherapy is antigen-specific inhibition by antigen-specific regulatory T-cell and myeloid suppressor cells. Therefore, immunotherapy using a TcR-independent mechanism may be an alternative immunotherapeutic strategy. NKG2D (natural killer, group 2, member D) and DNAX accessory molecule-1 (DNAM-1) are both activated receptors that are strongly expressed on T-cells, $\gamma\delta$ T-cells, and NK cells. Therefore, the expression of ligands for NKG2D and DNAM-1 on tumor cells plays an important role in tumor opsonization by immune effector cell targeting. Various modulatory methods for up-regulating NKG2D and DNAM-1-ligands have been reported, and included chemotherapeutic agents and hyperthermia. Although there are many obstacles to the utilization of NKG2D and DNAM-1 for cancer therapy, combined treatments using immune cell therapy and chemotherapy that take advantage of NKG2D and DNAM-1 may be an ideal approach.

Cancer Immunotherapy Using Tumor Opsonization

Cancer immunotherapy aims to activate the immune system for cancer eradication. After a disappointingly long time, the tide has at last changed due to an understanding of the mechanisms of tumor immunity and success of recent clinical trials (1, 2). Cancer immunotherapy consists of T-cell receptor (TcR)-dependent and -independent mechanisms. Although

tumor antigen-specific and TcR-dependent immunotherapy is an ideal immunotherapy, there are many limitations, such as tumor antigen-specific inhibitory mechanisms exerted by regulatory T-cell and myeloid-derived suppressor cells (3-5). Moreover, it is unknown which antigens can be effectively targeted by immune cells for many types of cancer. Alternatively, a biologically therapeutic strategy is to sensitize or opsonize tumor cells and trigger their death using immune cytotoxic effector cells such as activated natural killer (NK) cells and T cells. Recently there have been many studies reporting the clinical effects of nonspecific cytokine-activated T-cells and NK cells. However, cell therapy alone has not achieved sufficiently effective clinical results (6, 7). In this review, we examine the possibility of combining tumor opsonization methods and immune cell therapy focusing on the natural killer, group 2, member D (NKG2D) and DNAX accessory molecule-1 (DNAM-1) systems, and discuss future perspectives on the combination of chemotherapy and immune cell therapy.

NKG2D and its Ligands

NKG2D is a potent activating receptor expressed on virtually all NK cells, $\gamma\delta$ T-cells, and CD8 T-cells, and the interaction of NKG2D with its ligands in the tumors plays an important role in the immune response to tumors (8-12). NKG2D is a C-type lectin-like receptor expressed on cell surface and has been classified as a killer cell lectin receptor of superfamily K, member 1 (KLRK1), which is encoded by the *Nkg2d* (*Klrk1*) gene that is located within the NK gene complex (NKC) situated on chromosome 12 in humans (10-12). NKG2D is a homodimer and recognizes a number of stress-induced MHC class I-like ligands. The ligands are summarized and shown in Figure 1. In humans, NKG2D binds to the MHC class I-related proteins MICA and MICB (MHC class I chain-related protein A and B), UL-16 binding proteins (ULBPs). There are six members of the ULBP family of proteins, which are closely related to Rae1 molecules in mice. ULBP1, -2, and -3 and -6 are GPI-

Correspondence to: Dr. Takashi Morisaki, Fukuoka General Cancer Clinic, 3-1-1 Sumiyoshi, Hakata-ku, Fukuoka 812-0018, Japan. Tel: +81 922827696, Fax: +81 924056376, e-mail: tmorisaki@cancer-clinic.jp

Key Words: DNAM-1, NKG2D, MICA/B ULBP, CAK, TNK, natural killer, cytokine activated killer cells, immunotherapy, review.

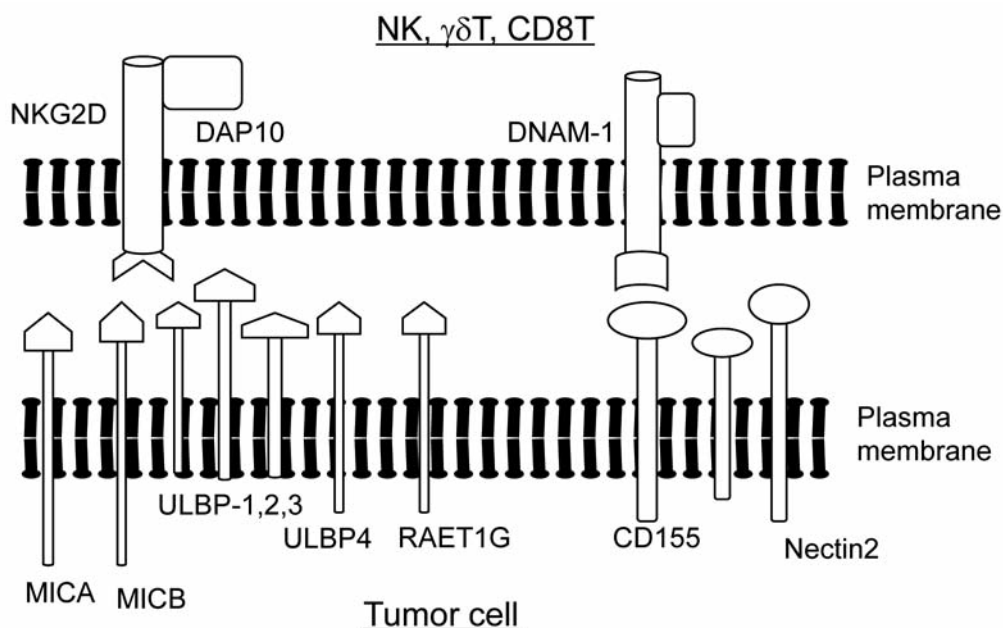


Figure 1. Multiple ligands for the human NKG2D and DNAM-1 receptors. MICA and MICB have extracellular domains and transmembrane domains. Within the UL16-binding protein (ULBP) family, retinoic acid early transcript 1E (RAET1E) and RAET1G also have transmembrane domains.

anchored, while ULBP4 and ULBP5 (also known as RAET1E and -G) are transmembrane proteins. These NKG2D ligands are induced in tumor cells by various stresses such as genotoxic stress, DNA-replication inhibitors, heat stress, photodynamic therapy, and chemotherapeutic agents (13-15). There is no inhibitory counterpart known for NKG2D and it is capable of overriding signals provided by inhibitory receptors on NK cells (9, 10, 12). In human, NKG2D associates with the adaptor protein DAP10 that transduces the activation signal of a specific signaling cascade (16). Since DAP10 deficiency results in complete loss of NKG2D signaling in T-cells, it appears that DAP10 is the most important adaptor for NKG2D signaling in these cells (9, 16). Guerra *et al.* (17) reported that NKG2D-deficient mice are defective in tumor surveillance in spontaneous tumor models and this gives rise to aggressive tumors. This indicates that NKG2D plays a critical role in the immunosurveillance of malignancies.

DNAM-1 and its Ligands

Another activating receptor involved in NK- and T cell-mediated tumor cell killing is DNAM-1, a transmembrane glycoprotein constitutively expressed on the majority of T-cells, NK cells, and macrophages. Its ligands are nectin-2 (CD112) and the poliovirus receptor (PVR, CD155), which belong to the nectin/nectin-like family (18) (Figure 1). CD155 is also expressed on epithelial cells, endothelial cells,

and antigen presenting cells. CD112 is expressed on epithelial cells. *In vitro* studies have shown that DNAM-1 triggers NK cell-mediated killing of tumor cells expressing CD155 and CD112 (19). DNAM-1 also promotes co-stimulation of CD4 and CD8 T-cells, and mediates adhesion of monocytes to endothelial cells facilitating transendothelial migration (20). It was recently demonstrated that DNAM-1 serves to extend the range of target cells that can activate CD8 T-cell and NK cells and so, may be essential for immune surveillance against tumors and may promote activation of cytotoxic lymphocytes by nonprofessional antigen-presenting cells (21). It was shown that DNAM-1 mediated NK cell recognition of freshly isolated ovarian carcinoma and neuroblastoma cells (22, 23). In addition, Iguchi-Manaka *et al.* (24) reported that DNAM-1-deficient mice developed significantly more DNAM-1 ligand-expressing tumors than did wild-type mice, which indicates that DNAM-1 plays an important role in immunosurveillance during tumor development.

Immune Evasion by Downregulation of NKG2D Ligands and NKG2D/DNAM-1 Receptors

Tumor cells use multiple mechanisms to bypass NKG2D-mediated killing. Such a mechanism was observed in tumors which often shed soluble NKG2D ligands from their cell surfaces, which can be detected in the blood of cancer patients (25, 26). These soluble ligands can bind to NKG2D

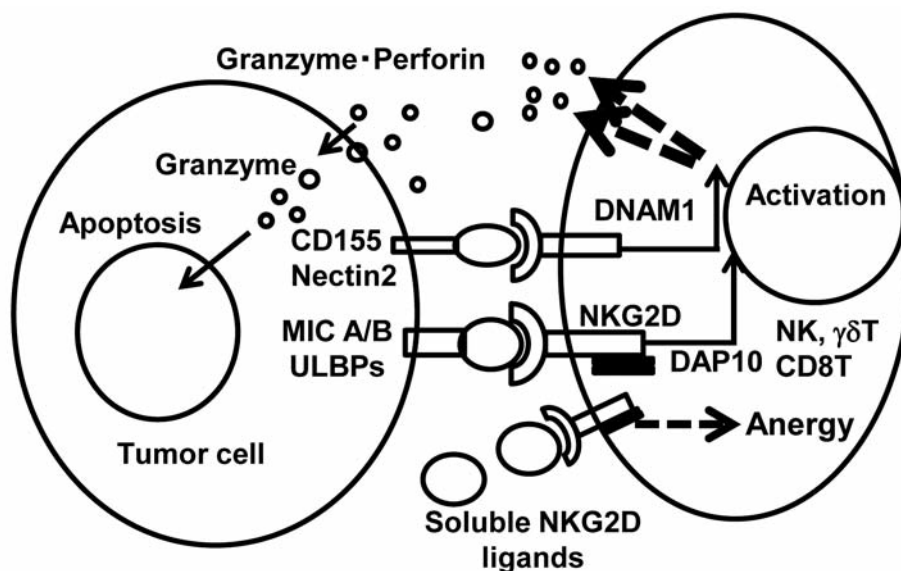


Figure 2. Interaction between activated T/NK cells and tumor cells through the NKG2D and DNAM-1 systems. After recognition and activation by the NKG2D/DNAM-1 receptor on effector cells for the ligands on tumor cell, cytotoxic molecules such as perforin and granzyme are released and induce apoptosis of tumor cell. The soluble form of NKG2D ligands may occupy the NKG2D receptor and inhibit NKG2D binding to the ligands on tumor cells.

and downregulate its expression on T-cells and NK-cells, thus effectively anergizing NKG2D-mediated immune recognition (Figure 2). Tumors reduce surface NKG2D ligand expression by shedding the extracellular domain using metalloproteinases or with the assistance of the disulphide-isomerase ERp5 (26). Another important mechanism for NKG2D ligand down-regulation appears to be the production of immunosuppressive cytokines such as transforming growth factor- β , which can be directly excreted by the tumor cells themselves, or by regulatory immune cells that expanded during tumor progression (27, 28). Many studies reported reduced expression of NKG2D or DNAM-1 on NK cells from cancer patients (29-35). For example, Mamessier *et al.* demonstrated that activating receptors such as DNAM-1 and NKG2D on NK cells infiltrating tumor tissue decreased in correlation with NK cell dysfunction throughout breast cancer progression (33).

Contribution of NKG2D Expression on Activated CD8 T-Cells to TcR-independent Cytotoxicity Toward Tumor Cells

Adoptive T-cell transfer therapy, the infusion of *ex vivo* expanded tumor-reactive T cells, resulted in objective tumor responses in patients with melanoma (6). Indeed, expansion of activating CD8⁺ T-cells with interleukin-2 (IL-2) and the agonistic anti-CD3 antibody OKT3 can be easily achieved for adoptive cell therapy. TNK cells, recently designated by Maccalli *et al.* (36), are NKG2D⁺ CD8⁺ T-cells, and are

relevant T-cell subtypes for immunosurveillance. Negrin's group demonstrated that NKG2D activation can overcome TcR-Class I-restricted cytotoxicity by CD8 T-cells and KIR-inhibition in NK cells (37, 38). Cytokine-activated CD8 T-cells can acquire dual cytotoxic functions: TcR-mediated antigen specific cytotoxicity and TcR-independent NKG2D and DNAM-1-dependent NK-like cytotoxicity against tumor cells (39). It was also shown that cytokine-activated CD8 T-cells induced NK-like cytotoxicity toward tumor cells mainly by NKG2D-ligand interaction (39).

Effects of Chemotherapeutic Agents on Ligands for NKG2D and DNAM-1 (summarized in Table I)

Expression of ligands for NKG2D and DNAM-1 on tumor cells is important for the recognition and killing of tumor cells by effector cells, while shedding of the ligands may inhibit tumor killing by the effector cells (Figure 2). Induced expression of NKG2D ligands on tumors appears to be a promising therapeutic strategy in cancer (9). Heat shock treatment up-regulates MICA on epithelial cells (14, 40). Indeed, the MICA/B promoter contains heat-shock transcriptional elements similar to those found in the promoters of heat shock proteins, such as heat shock protein 70 (41). Moreover, a wide variety of stimuli that causes genotoxic stress and results in DNA replication arrest (summarized in Table I), included histone deacetylase inhibitors, cytarabine, sodium butyrate, sunitinib, retinoic

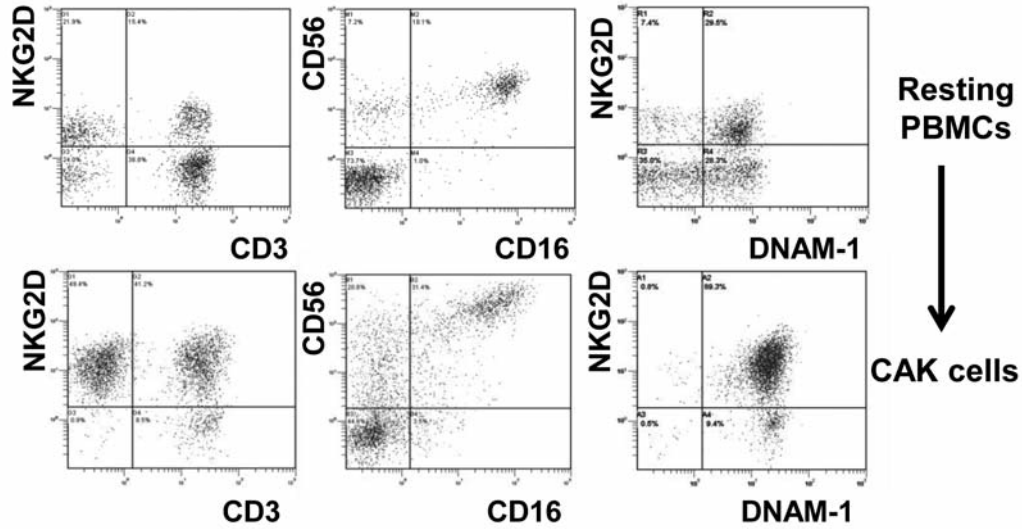


Figure 3. Increased expression of NKG2D and DNAM-1 on cytokine activated killer (CAK) cells. Peripheral blood mononuclear leukocytes (PBMC) were cultured in the presence of high dose interleukin-2 (IL-2) and the agonistic anti-CD3 antibody OKT3. NKG2D/DNAM-1 expression on mononuclear leukocytes was analyzed by fluorescence activated cell sorting (FACS) analysis as previously described in reference 50. The notable increase in expression of NKG2D and DNAM-1 on CAK cells is shown.

Table I. Chemotherapeutic agents.

Reagents	Cancer cell type	Reference
HDAC inhibitors (Tricostatin A)	Various	47
(FR9011228)		47
Retinoic acid	HCC	49
Sodium valproate	HCC, osteosarcoma	45, 46
Sodium butyrate	Hela, HCC, osteosarcoma	41
Cytarabine	Leukemia cell	42
Sunitinib, sorafenib	Nasopharyngeal carcinoma	48
Bortezomib, MG132	HCC, squamous carcinoma	44,53
Gemcitabine	HCC	50
Hydroxyurea	Leukemia	51
5-Aza-2'-deoxycytidine	Leukemia	43
Hydralazine, valproic acid	Various	54
HSP90 inhibitors	Myeloma	52
Melphalan, doxorubicin	Myeloma	55

HCC: Hepatocellular carcinoma.

acid, gemcitabine, and hydroxyurea (41-55). In our previous study, we demonstrated that gemcitabine induced MICA/B expression at both the protein and mRNA levels in hepatocellular carcinoma cell line HepG2 (50). The DNA damage response pathway, initiated by the ataxia telangiectasia mutated (ATM) or ataxia telangiectasia and the Rad3 related (ATR) kinases, was implicated in the regulation of NKG2D ligands expression in response to these insults. For example, Soriani *et al.* demonstrated that doxorubicin and melphalan up-regulated DNAM-1 and NKG2D ligands

on myeloma cells in an ATM-ATR-dependent manner (55). However, in squamous cell carcinoma cells treated with proteasome inhibitors such as bortezomib and M231, the enhanced expression of ULBP-1 was induced by an alternative pathway from ATM/ATR (53).

Modulation of NKG2D and DNAM-1 Systems by Antibody or Inhibitor via Ligand Shedding

The NKG2D and DNAM-1 receptor systems can be used as targets for anticancer therapy. Therapeutic MICA-specific antibodies effectively opsonized cancer cells and induced DC-mediated cross-presentation of tumor antigens (56). Bifunctional proteins consisting of a tumor-antigen directed antibody fused to NKG2D ligands effectively coated tumor cells with activating ligands and increased their killing (57, 58). The prevention of MICA/B shedding may also be an important strategy for enhancing cytokine activated killer cell (CAK) cytotoxicity via the NKG2D system. There are several reports that have shown the effect of MMP inhibitors on preventing MICA/B shedding by tumor cells (59, 60). Kohga *et al.* demonstrated that a disintegrin and metalloproteinase 9 (ADAM9) are involved in MICA shedding in HCC cells, and sorafenib can modulate ADAM9 expression that resulted in increase of MICA expression (59). Huang *et al.* demonstrated that combined treatment using histone deacetylase inhibitors and metalloproteinase inhibitor caused up-regulation of MICA/B and that inhibition of MICA/B resulted in increased susceptibility to killing by cytokine induced killer lymphocytes (60).

Combinatorial Chemotherapy and Activated Lymphocyte Therapy *via* Enhanced NKG2D and DNAM-1-oriented Systems

Although various kinds of cell-based immunotherapy have been reported, the clinical effects of the therapy alone have been modest (6, 7). Combining immunotherapy with chemotherapy is a promising advancement. Chemotherapy that up-regulates cell surface expression of NKG2D and DNAM-1 ligand and down-regulates shedding of these molecules will need to be effectively combined with cell therapy using activated T-cells and NK cells (60). We previously reported that CAK cells, which consist a heterogenous population composed of activated NK cells and activated killer T-cells, can be induced *in vitro* using peripheral blood lymphocytes treated with high doses of IL-2 and OKT3 (50). We further demonstrated that gemcitabine induces MICA/B expression in hepatocellular carcinoma cells and results in the synergistic enhancement of the cytotoxic effects of CAK cells. Upregulation of NKG2D and DNAM-1 receptors on effector cells can be achieved by *ex vivo* activation with a cytokine such as high dose IL-2 (Figure 3). Thus, combining gemcitabine with CAK cell immunotherapy may have clinical significance in the treatment of various types of cancer. In the near future, harnessing the benefits of immunotherapy using conventional chemotherapy that strengthens the NKG2D/DNAM-1 system will be a promising combinatorial therapy approach.

References

- Mellman I, Coukos G and Dranoff G: Cancer immunotherapy comes of age. *Nature* 480: 480-489, 2011.
- Dougan M and Dranoff G: Immune therapy for cancer. *Ann Rev Immunol* 27: 83-117, 2009.
- Gabrilovich DI and Nagaraj S: Myeloid-derived-suppressor cells as regulators of the immune system. *Nat Rev Immunol* 9: 162-174, 2009.
- Rabinovich GA, Gabrilovich D and Sotomayor EM: Immunosuppressive strategies that are mediated by tumor cells. *Annu Rev Immunol* 25: 267-296, 2007.
- Bonertz A, Weitz J, Pietsch D-H K, Rahbari NN, Schlude C, Ge Y, Juenger S, Vlodavsky I, Khazaie K, Jaeger D, Reissfelder C, Antolovic D, Aigner M, Koch M and Beckhove P: Antigen-specific Tregs control T cell responses against a limited repertoire of tumor antigens in patients with colorectal carcinoma. *J Clin Invest* 119: 3311-3321, 2009.
- Dudley ME and Rosenberg SA: Adoptive cell transfer therapy. *Semin Oncol* 34: 524-531, 2007.
- June CH: Adoptive T cell therapy for cancer in the clinic. *J Clin Invest* 117: 1466-1476, 2007.
- Eagle RA and Trowsdale J: Promiscuity and the single receptor: NKG2D. *Nature Rev Immunol* 7: 737-743, 2007.
- Nausch N and Cerwenka A: NKG2D ligands in tumor immunity. *Oncogene* 27: 5944-5958, 2008.
- Ho EL, Heusel JW, Brown MG, Matsumoto K, Scalzo AA and Yokoyama WM: Murine NKG2D and CD94 are clustered within the natural killer complex and are expressed independently in natural killer cells. *Proc Natl Acad Sci USA* 95: 6320-6325, 1998.
- Raulet DH: Roles of the NKG2D immunoreceptor and its ligands. *Nat Rev Immunol* 3: 781-790, 2003.
- Champsaur M and Lanier LL: Effect of NKG2D ligand expression on host immune responses. *Immunol Rev* 235: 267-285, 2010.
- Gasser S, Orsulic S, Brown EJ and Raulet DH: The DNA damage pathway regulates innate immune system ligands for the NKG2D receptor. *Nature* 436: 1186-1190, 2005.
- Ostberg JR, Dayanc BE, Yuan M, Oflazoglu E and Repasky EA: Enhancement of natural killer (NK) cell cytotoxicity by fever-range thermal stress is dependent on NKG2D function and is associated with plasma membrane NKG2D clustering and increased expression of MICA on target cells. *J Leukoc Biol* 82: 1322-1331, 2007.
- Park MJ, Bae JH, Chung JS, Kim SH and Kang CD: Induction of NKG2D ligands and increased sensitivity of tumor cells to NK cell-mediated cytotoxicity by hematoporphyrin-based photodynamic therapy. *Immunol Invest* 40: 367-382, 2011.
- Wu J, Song Y, Bakker AB, Bauer S, Spies T, Lanier LL and Phillips JH: An activating immunoreceptor complex formed by NKG2D and DAP10. *Science* 285: 730-732, 1999.
- Guerra N, Tan YX, Jonker NT, Choy A, Gallardo F, Xiong N, Knoblaugh S, Cado D, Greenberg NR and Raulet DH: NKG2D-deficient mice are defective in tumor surveillance in models of spontaneous malignancy. *Immunity* 28: 571-580, 2008.
- Shibuya A, Campbell D, Hannum C, Yssel H, Franz-Bacon K, McClanahan T, Kitamura T, Nicholl J, Sutherland GR, Lanier LL and Phillips JH: DNAM-1, a novel adhesion molecule involved in the cytolytic function of T lymphocytes. *Immunity* 4: 573-581, 1996.
- Bottino C, Castriconi R, Pende D, Rivera P, Nanni M, Carnemolla B, Cantoni C, Grassi J, Marcenaro S, Reymond N, Vitale M, Moetta L, Lopez M and Moretta A: Identification of PVR (CD155) and nectin-2 (CD112) as cell surface ligands for the human DNAM-1 (CD226) activating molecule. *J Exp Med* 198: 557-567, 2003.
- Shibuya K, Shirakawa J, Kameyama T, Honda S, Tahara-Hanaoka S, Miyamoto A, Onodera M, Sumida T, Nakauchi H, Miyoshi H and Shibuya A: CD226 (DNAM-1) is involved in lymphocyte function-associated-antigen 1 costimulatory signal for naïve T-cell differentiation and proliferation. *J Exp Med* 198: 1829-1839, 2003.
- Gillfillan S, Chan CJ, Cella CM, Haynes NM, Rapaport AS, Boles KS, Andrews DM, Smyth MJ and Colonna M: DNAM-1 promotes activation of cytotoxic lymphocytes by nonprofessional antigen-presenting cells and tumors. *J Exp Med* 205: 2965-2973, 2008.
- Carlsten M, Björkström N, Norell H, Bryceon Y, van Hall T, Baumann BC, Hanson M, Schedvins K, Kiessling R, Ljunggren H-G and Malmberg K-J: DNAX accessory molecule-1-mediated recognition of freshly isolated ovarian carcinoma by resting natural killer cells. *Cancer Res* 67: 1317-1325, 2007.
- Castriconi R, Dondero A, Corrias MV, Lanino E, Pende D, Moretta L, Bottino C and Moretta A: Natural killer cell-mediated killing of freshly isolated neuroblastoma cell: critical role of DNAX accessory molecule-1-poliiovirus receptor interaction. *Cancer Res* 64: 9180-9184, 2004.

- 24 Iguchi-Manaka A, Kai H, Yamashita Y, Shibata K, Tahara-Hanaoka S, Honda S, Yasui T, Kikutani H, Shibuya K and Shibuya A: Accelerated tumor growth in mice deficient in DNAM-1 receptor. *J Exp Med* 205: 2959-2964, 2008.
- 25 Salih HR, Rammensee H-G and Steinle A: Cutting edge: down-regulation of MICA on human tumors by proteolytic shedding. *J Immunol* 169: 4098-4102, 2002.
- 26 Kaiser BK, Yim D, Chow IT, Gonzalez S, Dai Z, Mann HH, Strong RK, Groh V and Spies T: Disulphide-isomerase-enabled shedding of tumour-associated NKG2D ligands. *Nature* 447: 482-486, 2007.
- 27 Li H, Han Y, Guo Q, Zhang M and Cao X: Cancer-expanded myeloid-derived suppressor cells induce anergy NK cells through membrane-bound TGF-beta 1. *J Immunol* 182: 240-249, 2009.
- 28 Lee JC, Lee KM, Kim DW and Heo DS: Elevated TGF-beta 1 secretion and down-modulation of NKG2D underlines impaired NK cytotoxicity in cancer patients. *J Immunol* 172: 7335-7340, 2004.
- 29 Saito H, Osaki T and Ikeguchi M: Decreased NKG2D expression on NK cells correlates with impaired NK cell function in patients with gastric cancer. *Gastric Cancer* 15: 27-33, 2012.
- 30 Mirjačić MK, Konjević G, Babović N and Inić M: The stage dependent changes in NK cell activity and the expression of activating and inhibitory NK cell receptors in melanoma patients. *J Surg Res* 171: 637-649, 2011.
- 31 Caristen M, Baumann BC, Simonsson M, Jadersten M, Forsblom AM, Hammarstedt C, Bryceson YT, Ljunggren HG, Hellstrom-Lidberg E and Malmberg KJ: Reduced DNAM-1 expression on bone marrow NK cells associated with impaired killing of CD34+ blasts in myelodysplastic syndrome. *Leukemia* 24: 1607-1616, 2010.
- 32 Duan X, Deng L, Chen X, Lu Y, Zhang Q, Zhang K, Hu Y, Zeng J and Sun W: Clinical significance of immunostimulatory MHC class I chain-related molecule A and NKG2D receptor on NK cells in pancreatic cancer. *Med Oncol* 28: 466-474, 2011.
- 33 Mamessier E, Sylvain A, Thibault M-L, Houvenaeghel G, Jacquemier J, Castellano R, Goncalves A, André P, Romagné F, Thibaut G, Viens P, Birnbaum D, Bertucci F, Moretta A and Olive D: Human breast cancer cells enhance self tolerance by promoting evasion from NK cell antitumor immunity. *J Clin Invest* 121: 3609-3622, 2011.
- 34 Osaki T, Saito H, Yoshikawa T, Matsumoto S, Tatebe S, Tsujitani S and Ikeguchi M: Decreased NKG2D expression on CD8+T cell is involved in immune evasion in patients with gastric cancer. *Clin Cancer Res* 13: 382-387, 2007.
- 35 Shen Y, Lu C, Tian W, Wang L, Cui B, Jiao Y, Ma C, Ju Y, Zhu L, Shao C, Liu X, Wang J, Zhang B and Lu Z: Possible association of decreased NKG2D expression levels and suppression of the activity of natural killer cells in patients with colorectal cancer. *Int J Oncol* 40: 1285-1290, 2012.
- 36 Maccalli C, Scaramuzza S and Parmiani G: TNK cells (NKG2D+CD8T+ or CD4+ T lymphocytes) in the control of human tumors. *Cancer Immunol Immunother* 58: 801-808, 2009.
- 37 Verneris MR, Karimi M, Baker J, Jayaswal A and Negrin RS: Role of NKG2D signaling in the cytotoxicity of activated and expanded CD8+ T-cells. *Blood* 103: 3065-3072, 2004.
- 38 Karimi M, Cao TM, Baker JA, Verneris MR, Soares L and Negrin RS: Silencing human NKG2D, DAP10, and DAP12 reduces cytotoxicity of activated CD8+T cells and NK cells. *J Immunol* 175: 7819-7828, 2005.
- 39 Pievani A, Borleri G, pende D, Moretta L, Rambaldi A, Golay J and Introna M: Dual functional capability of CD3+CD56+ CIK cells, a T-cell subset that acquires NK function and retains TCR-mediated specific cytotoxicity. *Blood* 118: 3301-3310, 2011.
- 40 Groh V, Steinle A, Bauer S and Spies T: Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells. *Science* 279:1737-1740, 1998.
- 41 Zhang C, Wang Y, Zhou Z, Zhang J and Tian Z: Sodium butyrate upregulates expression of NKG2D ligand MICA/B in HeLa and HepG2 cell lines and increases their susceptibility to NK lysis. *Cancer Immunol Immunother* 58: 1275-1285, 2009.
- 42 Ogbomo H, Michaelis M, Klassert D, Doerr HM, and Cinatl Jr. J: Resistance to cytarabine induces the up-regulation of NKG2D ligands and enhances natural killer cell lysis of leukemic cells. *Neoplasia* 10: 1402-1410, 2008.
- 43 Tang K-F, He C-X, Zeng G-Li, Wu J, Song G-B, Shi Y-S, Zhang W-G, Huang A-L, Steinle A and Ren H: Induction of MHC Class I-related chain B (MICB) by 5-aza-2'-deoxycytidine. *Biochem Biophys Res Com* 370: 578-583, 2008.
- 44 Armeanu S, Krusch M, Baltz KM, Weiss TS, Smirnow I, Steinle A, Lauer UM, Bitzer M and Salih HR: Direct and natural killer cell-mediated antitumor effects of low-dose bortezomib in hepatocellular carcinoma. *Clin Cancer Res* 14: 3520-3528, 2008.
- 45 Armeanu S, Bitzer M, Lauer UM, Venturelli S, Pathil A, Krusch M, Kaiser S, Jobst J, Smirnow I, Wagner A, Stenle A and Salih HR: Natural killer cell-mediated lysis of hepatoma cells via specific induction of NKG2D ligands by the histone deacetylase inhibitor sodium valproate. *Cancer Res* 65: 6321-6329, 2005.
- 46 Yamanegi K, Yamane J, Kobayashi K, Kato-Kogoe N, Ohyama H, Nakasho K, Yamada N, Hata M, Nishioka T, Fukunaga S, Futani H, Okamura H and Terada N: Sodium valproate, a histone deacetylase inhibitor, augments the expression of cell-surface NKG2D ligands, MICA/B, without increasing their soluble forms to enhance susceptibility of human osteosarcoma cells to NK cell-mediated cytotoxicity. *Oncol Rep* 24: 1621-1627, 2010.
- 47 Skov S, Pederson MT, Andresen A, Straten PT, Woetmann A and Ødum N: Cancer Cells become susceptible to natural killer cell killing after exposure to histone deacetylase inhibitors due to glycogen synthase kinase-3-dependent expression of MHC Class I-related chain A and B. *Cancer Res* 65: 11136-11145, 2005.
- 48 Huang Y, Wang Y, Li Y, Guo K and He Y: Role of sorafenib and sunitinib in the induction of expressions of NKG2D ligands in nasopharyngeal carcinoma with high expression of ABCG2. *J Cancer Res Clin Oncol* 137: 829-837, 2010.
- 49 Jinushi M, Takehara T, Tatsumi T, Kanto T, Groh V, Spies T, Kimura R, Miyagi T, Mochizuki K, Sasaki Y and Hayashi N: Expression and role of MICA and MICB in human hepatocellular carcinomas and their regulation by retinoic acid. *Int J Cancer* 104: 354-361, 2003.
- 50 Morisaki T, Onishi H, Koya N, Kiyata A, Tanaka H, Umebayashi M, Ogino T, Nagamatsu I and Katano M: Combinatorial cytotoxicity of gemcitabine and cytokine-activated killer cells in hepatocellular carcinoma via NKG2D-MICA/B system. *Anticancer Res* 31: 2505-2510, 2011.
- 51 Lu X, Ohata K, Kondo Y, Espinoza JL, Qi Z and Nakao S: Hydroxyurea upregulates NKG2D ligand expression in myeloid leukemia cells synergistically with valproic acid and potentially enhances susceptibility of leukemic cells to natural killer cell-mediated cytotoxicity. *Cancer Sci* 101: 609-615, 2010.

- 52 Fionda C, Soriani A, Malgarini G, Iannitto ML, Santoni A, and Cippitelli M: Heat shock protein-90 inhibitors increase MHC class I-related chain A and B ligand expression on multiple myeloma cells and their ability to trigger NK cell degranulation. *J Immunol* 183: 4385-4394, 2009.
- 53 Butler JE, Moore MB, Presnell SR, Chan H-W, Chalupny NJ and Lutz CT: Proteasome regulation of ULBP1 transcription. *J Immunol* 182: 6600-6609, 2009.
- 54 Chávez-Blanco A, De la Cruz-Hernández E, Dominguez GI, Rodríguez-Cortez O, Alatorre B, Pérez-Cárdenas E, Chacon-Salinas R, Trjo-Becerril C, Taja-Chayeb L, Trujillo JE, Contreras-Paredes A and Duenas-González A: Upregulation of NKG2D ligands and enhanced natural killer cytotoxicity by hydralazine and valproate. *Int J Oncol* 39: 1491-1499, 2011.
- 55 Soriani A, Zingoni A, Cerboni C, Iannitto ML, Ricciardi MR, Di Gialleonardo V, Cippitelli M, Fionda C, Petrucci MT, Guarini A, Foá R and Santoni A: ATM-ATR-dependent up-regulation of DNAM-1 and NKG2D ligands on multiple myeloma cells by therapeutic agents results in enhanced NK-cell susceptibility and is associated with a senescent phenotype. *Blood* 113: 3503-3511, 2009.
- 56 Jinushi M, Hodi FS and Dranoff G: Therapy-induced antibodies to MHC class I chain-related protein A antagonize immune suppression and stimulate antitumor cytotoxicity. *Proc Natl Acad Sci USA* 103: 9190-9195, 2006.
- 57 Germain C, Campigna E, Salhi I, Morisseau S, Navarro-Teulon I, Mach JP, Pèlegriin and Robert B: Redirecting NK cells mediated tumor cell lysis by a new recombinant bifunctional protein. *Protein Eng Des Sel* 21: 665-672, 2008.
- 58 Von Strandmann EP, Hansen HP, Reiners KS, Schnell R, Borchmann P, Merkert S, Simhadri VR, Draube A, Reiser M, Purr I, Hallek M and Engert A: A novel bispecific protein (ULBP2-BB4) targeting the NKG2D receptor on natural killer (NK) cells and CD138 activates NK cells and has potent antitumor activity against human multiple myeloma. *Blood* 107: 1955-1962, 2006.
- 59 Kohga K, Takehara T, Tatsumi T, Ishida H, Miyagi T, Hosui A and Hayashi N: Sorafenib inhibits the shedding major histocompatibility complex class-I-related chain A on hepatocellular carcinoma cells by down-regulating a disintegrin and metalloproteinase 9. *Hepatology* 51: 1264-1273, 2010.
- 60 Huang B, Sikorski R, Sampath P and Thorne SH: Modulation of NKG2D-ligand cell surface expression enhances immune cell therapy of cancer. *J Immunother* 34: 289-296, 2011.

Received April 4, 2012
Revised May 16, 2012
Accepted May 17, 2012