Inhibin A Is Down-regulated During Chemotherapy in Patients with Breast Cancer

N. BURKHARDT, J. JÜCKSTOCK, C. KUHN, B. RACK, W. JANNI, C. SCHINDLBECK, H. SOMMER, K. FRIESE and I. MYLONAS

First Department of Obstetrics and Gynaecology, Ludwig Maximilians University Munich, Munich, Germany

Abstract. Background: Inhibins are dimeric glycoproteins, composed of an alpha-subunit (INH- α) and one of two possible beta-subunits (βA or βB), with substantial roles in human reproduction and in endocrine-responsive tumours. Aims of this study were to determine the serological measurement of inhibin A (α - β A) in breast cancer patients during chemotherapy. Patients and Methods: A series of 30 breast cancer patients who underwent standardised chemotherapy were prospectively evaluated before chemotherapeutic treatment as well as four weeks after chemotherapy and two years after chemotherapy for the serological expression of inhibin A. For statistical analysis the Wilcoxon rank sum test was used for paired samples. Statistical significance was assumed at p<0.05. Results: The concentration of inhibin A showed a significant decrease between data obtained before chemotherapy and after chemotherapy (p < 0.005) and two-year follow-up (p < 0.001). Interestingly, there were no differences in inhibin A concentrations between the four-week and two-year follow-up (p=0.744). Discussion: Chemotherapy significantly decreases inhibin A concentration during chemotherapy. This might reflect a suppression of ovarian function, being also a marker for chemotherapy-induced amenorrhoea. Moreover, it has been suggested that inhibin A might be a tumour marker for breast cancer, and therefore a sudden increase in its concentration might be indicative of breast cancer recurrence.

Inhibins and activins are secreted polypeptides, representing a subgroup of the TGF- β superfamily of growth and differentiation factors (1, 2). Inhibins are heterodimeric glycoproteins, composed of an alpha-subunit (α) and one of two possible beta-subunits, named βA or βB , in the

Key Words: Inhibin A, proliferation, chemotherapy, breast cancer.

formation of either inhibin A (α - β A) or B (α - β B), respectively (1, 2). Two additional β -subunits (named β C and β E) have been cloned, although their precise function and dimerisation potential remains yet unclear (1). Activins are, in contrast to inhibins, homodimers of the β -subunits linked by a disulphide bridge (1-3).

Inhibin subunits have been detected in endocrine tumours (4) and their differential expression in malignant tissue has suggested an important role in malignant cell transformation (4-6). Inhibin subunits have been observed in healthy breast epithelia as well as in benign and malignant breast tissue (7, 8). Moreover, activin A is synthesized in malignant mammary tissue as well as in the serum of postmenopausal women with breast cancer, suggesting it as a possible tumour marker in breast cancer patients, since average activin A levels have been observed to be higher in cancer patients (9).

It has been demonstrated that premenopausal breast cancer survivors have diminished ovarian reserve compared with a control group (10), as assessed by biochemical markers including inhibin B (10). Moreover, pre-chemotherapy inhibin B and anti-müllerian hormone (AMH) have been observed to be lower among women experiencing chemotherapyassociated amenorrhoea, suggesting a predictive value for the occurrence of amenorrhoea (11).

In order to investigate the potential role of inhibin A in breast cancer patients within the context of chemotherapy, serum samples of breast cancer patients obtained from the prospective, randomised German therapeutic SUCCESS trial receiving adjuvant chemotherapy were analysed for inhibin A.

Patients and Methods

Samples. This study used serum samples derived from 30 patients diagnosed with breast cancer from the prospective, randomised German therapeutic SUCCESS trial (12). All of the patients underwent surgery leading to R0 resection of the tumour and received adjuvant chemotherapy. For each patient, three serum samples at three different time-points were taken: before the beginning of chemotherapy, four weeks after termination of chemotherapy and two years after chemotherapy. Therefore, a total of 90 serum samples were tested for their inhibin concentration.

Correspondence to: Ioannis Mylonas, MD, Ph.D., First Department of Obstetrics and Gynaecology, Ludwig-Maximilians-University Munich Maistrasse 11, 80337 Munich, Germany. Tel: +49 8951604111, Fax: +49 8951604916, email: ioannis.mylonas@med.uni-muenchen.de

The SUCCESS trial is a multicentric, prospective randomised trial comparing the effectiveness and compatibility of two different chemotherapy regimens in breast cancer patients, followed by two- or five-year bisphosphonate therapy (12). The aim of the trial is the comparison of recurrence-free survival after randomisation of patients who received three cycles of epirubicin–5-fluorouracil–cyclo-phosphamide (FEC) chemotherapy, followed by three cycles docetaxel (D) chemotherapy *versus* three cycles of FEC chemotherapy followed by three cycles gemcitabine–docetaxel (DG) chemotherapy.

Immunoassay (ELISA). For serological measurement of inhibin A, Ultrasensitive Inhibin A ELISA (DSL-10-18200 ACTIVE® Ultrasensitive Inhibin A ELISA, DSL, U.S.A) was used. The principle of the test is an enzymatically amplified 'two-step' sandwich type immunoassay. It involves two antibodies which absorb the antigen (inhibin) at two different sites. The primary antibody, anti-inhibin BA subunit antibody, was located in special microtitration wells. In the assay, standards, controls as well as serum samples were incubated in these coated microtitration wells. A protein buffer was also added. After several hours of incubation at room temperature the inhibin βA antibody was able to absorb any inhibin that was present. After washing with wash solution, the wells were treated with biotinylated anti-inhibin alpha subunit detection antibody, the secondary antibody, which also was able to bind inhibin during incubation. The immunoreaction was continued by subsequent addition of streptavidin labelled with enzyme horseradish peroxidase (HRP) to link the enzyme to the secondary antibody. After a third incubation and washing step, the wells were incubated with the substrate tetramethylbenzidine with hydrogen peroxide. This substrate generated a change of colour which was catalysed by the already present HRP. An acidic stopping solution was added afterwards and the degree of enzymatic turnover of the substrate was determined by dual wavelength absorbance measurement at 450 and 620 nm.

The absorbance measured was directly proportional to the concentration of inhibin A present in the samples. A set of inhibin A standards was used to plot a standard curve of absorbance *versus* inhibin A concentration from which the inhibin A concentrations in the unknowns could be calculated.

Statistical analysis. The concentration of inhibin in the serum samples of patients receiving chemotherapy was measured at three time points: before the beginning of chemotherapy, four weeks after the last cycle of chemotherapy and two years after chemotherapy.

For statistical analysis, the statistics programme SPSS (SPSS version 15.0; SPSS Inc., Chicago, IL, USA) for Windows was used. The established data of inhibin concentration were compared by Friedman test and the Wilcoxon rank sum test for paired samples. Statistical significance was assumed at p < 0.05.

Results

In chronological sequence, a significant difference between the three points of time was observed. The concentration of inhibin A showed a significant decrease between data obtained before chemotherapy and after chemotherapy (p<0.005) and between before chemotherapy and two-year follow-up (p<0.001). Interestingly, there were no differences between the four-week and two-year follow-up data (p=0.744) (Figure 1).

Discussion

The inhibin/activin-subunits have been detected in normal female reproductive tissue and endocrine tumours (4), including ovarian cancer (4, 13), endometrial tissue (14-19) and breast cancer (7, 20), suggesting possible roles in cancer proliferation and growth (4, 21).

This widespread expression of these subunits might reflect several autocrine and paracrine roles. In women with ovarian cancer, elevated serological levels of inhibin have been demonstrated, suggesting it as a useful tumour marker in this type of cancer (22). Additionally, similar observations and suggestions have been observed for activin A in breast cancer tissue (9). However, activin might not be a clear-cut marker of breast cancer recurrence, but it still constitutes a potential candidate for recurrence surveillance (20).

Recent investigations have focused in assessing ovarian reserve by using several surrogate markers, including inhibin B, in breast cancer survival (23). Meanwhile, it has been demonstrated that premenopausal breast cancer survivors have diminished ovarian reserve compared with a control group (10) as assessed by biochemical markers, including inhibin B (10). Moreover, pre-chemotherapy inhibin B and AMH have been observed to be lower among women experiencing chemotherapy-associated amenorrhea, suggesting a predictive value for the occurrence of amenorrhoea (11). However, the most evaluated serological glycoproteins of the TGF- β group are inhibin B and AMH. In this study it was demonstrated that inhibin A concentration can be detected in serum of breast cancer patients with a significant concentration decrease during the course of chemotherapy. An interesting fact is that there were no differences between the values at end of chemotherapy and the two-year follow-up. Since no patient in the analysed study group has presented with a breast cancer recurrence to date, no definite statement on the value of inhibin A as a marker for tumour recurrence can be made at the time of writing.

The effects and action mechanisms of inhibin in cancer cells are not yet completely elucidated. However, the most important function is the tumour suppressor activity of the α -subunit, first identified after functional deletion of the inhibin- α gene in mice (24, 25). Whether the inhibin- βA and $-\beta B$ subunits have similar tumour-suppressive properties as observed for the inhibin- α subunit remains controversial (4), although recently obtained evidence suggests that activin A, the homodimer of the βA subunit, can inhibit cancer cell proliferation in various experimental models *in vitro* and *in vivo* (26, 27). Whether these assumptions are also true for breast cancer cells is not yet known. Activin A can inhibit breast cancer cell proliferation of cell lines *in vitro* (28-31). Moreover, activin resistance

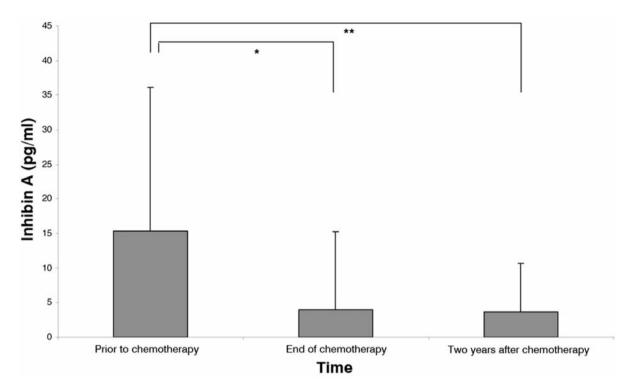


Figure 1. Serological concentrations of inhibin A in patients with mammary carcinomas. The concentration of inhibin A showed a significant decrease between data obtained before and after chemotherapy (*p<0.005), and at two-year follow-up (**p<0.001).

in oestrogen receptor-negative breast tumour cell lines might be involved in breast cancer carcinogenesis and in increased malignancy compared with oestrogen receptorpositive cells (28).

In conclusion, chemotherapy significantly decreased the inhibin A concentration in this study. This might reflect a suppression of ovarian function, being also a marker for chemotherapy-induced amenorrhoea. Whether inhibin A also constitutes a prognostic serological tumour marker in breast cancer patients is still under investigation.

Acknowledgements

We would like to thank Mrs. S. Schulze, Mrs. S. Kunze, Mrs. S. Hoffmann, Mrs. B. Zill and Professor Dr. U. Jeschke for their help in this study.

The Authors declare that they do not have any competing interests regarding this analysis nor do they have any financial, personal, political, intellectual or religious interests in publishing this article.

I. Mylonas is supported by the FöFoLe program of the Ludwig-Maximilians-University, Munich (297/03), the Friedrich-Baur-Institute, Munich and the Weigland Stipendium Program of the Ludwig Maximilians University and the German Research Foundation (Deutsche Forschungsgemeinschaft, DFG BR 3641/3-1).

References

- Xia Y and Schneyer AL: The biology of activin: recent advances in structure, regulation and function. J Endocrinol 202: 1-12, 2009.
- 2 Vale W, Wiater E, Gray P, Harrison C, Bilezikjian L and Choe S: Activins and inhibins and their signaling. Ann NY Acad Sci 1038: 142-147, 2004.
- 3 Ling N, Ying SY, Ueno N, Shimasaki S, Esch F, Hotta M and Guillemin R: Pituitary FSH is released by a heterodimer of the beta-subunits from the two forms of inhibin. Nature *321*: 779-782, 1986.
- 4 Risbridger GP, Schmitt JF and Robertson DM: Activins and inhibins in endocrine and other tumors. Endocr Rev 22: 836-858, 2001.
- 5 Otani T, Minami S, Yamoto M and Umesaki N: Production of activin A in hyperplasia and adenocarcinoma of the human endometrium. Gynecol Oncol 83: 31-38, 2001.
- 6 Worbs S, Shabani N, Mayr D, Gingelmaier A, Makrigiannakis A, Kuhn C, Jeschke U, Kupka MS, Friese K and Mylonas I: Expression of the inhibin/activin subunits (-alpha, -betaA and -betaB) in normal and carcinogenic endometrial tissue: possible immunohistochemical differentiation markers. Oncol Rep 17: 97-104, 2007.
- 7 Mylonas I, Jeschke U, Shabani N, Kuhn C, Friese K and Gerber B: Inhibin/activin subunits (inhibin-alpha, -betaA and -betaB) are differentially expressed in human breast cancer and their metastases. Oncol Rep 13: 81-88, 2005.

- 8 Di Loreto C, Reis FM, Cataldi P, Zuiani C, Luisi S, Beltrami CA and Petraglia F: Human mammary gland and breast carcinoma contain immunoreactive inhibin/activin subunits: evidence for a secretion into cystic fluid. Eur J Endocrinol 141: 190-194, 1999.
- 9 Reis FM, Cobellis L, Tameirao LC, Anania G, Luisi S, Silva IS, Gioffre W, Di Blasio AM and Petraglia F: Serum and tissue expression of activin A in postmenopausal women with breast cancer. J Clin Endocrinol Metab 87: 2277-2282, 2002.
- 10 Partridge AH, Ruddy KJ, Gelber S, Schapira L, Abusief M, Meyer M and Ginsburg E: Ovarian reserve in women who remain premenopausal after chemotherapy for early stage breast cancer. Fertil Steril *94*: 638-644, 2010.
- 11 Anders C, Marcom PK, Peterson B, Gu L, Unruhe S, Welch R, Lyons P, Behera M, Copland S, Kimmick G, Shaw H, Snyder S, Antenos M, Woodruff T and Blackwell K: A pilot study of predictive markers of chemotherapy-related amenorrhea among premenopausal women with early stage breast cancer. Cancer Invest 26: 286-295, 2008.
- 12 Rack B, Schindlbeck C, Juckstock J, Genss EM, Hepp P, Lorenz R, Tesch H, Schneeweiss A, Beckmann MW, Lichtenegger W, Sommer H, Friese K and Janni W: Prevalence of CA 27.29 in primary breast cancer patients before the start of systemic treatment. Anticancer Res 30: 1837-1841, 2010.
- 13 Zheng W, Luo MP, Welt C, Lambert-Messerlian G, Sung CJ, Zhang Z, Ying SY, Schneyer AL, Lauchlan SC and Felix JC: Imbalanced expression of inhibin and activin subunits in primary epithelial ovarian cancer. Gynecol Oncol 69: 23-31, 1998.
- 14 Kimmich T, Bruning A, Kaufl SD, Makovitzky J, Kuhn C, Jeschke U, Friese K and Mylonas I: Inhibin/activin-betaC and -betaE subunits in the Ishikawa human endometrial adenocarcinoma cell line. Arch Gynecol Obstet 282: 185-191, 2010.
- 15 Käufl SD, Makovitzky J, Kuhn C, Kunze S, Jeschke U and Mylonas I: Inhibin/activin-betaC subunit in human endometrial adenocarcinomas and HEC-1a adenocarcinoma cell line. In Vivo 20: 1177-1125, 2010.
- 16 Petraglia F, Florio P, Luisi S, Gallo R, Gadducci A, Vigano P, Di Blasio AM, Genazzani AR and Vale W: Expression and secretion of inhibin and activin in normal and neoplastic uterine tissues. High levels of serum activin A in women with endometrial and cervical carcinoma. J Clin Endocrinol Metab 83: 1194-1200, 1998.
- 17 Mylonas I, Jeschke U, Wiest I, Hoeing A, Vogl J, Shabani N, Kuhn C, Schulze S, Kupka MS and Friese K: Inhibin/activin subunits alpha, beta-A and beta-B are differentially expressed in normal human endometrium throughout the menstrual cycle. Histochem Cell Biol *122*: 461-471, 2004.
- 18 Mylonas I, Worbs S, Shabani N, Kuhn C, Kunze S, Schulze S, Dian D, Gingelmaier A, Schindlbeck C, Bruning A, Sommer H, Jeschke U and Friese K: Inhibin-alpha subunit is an independent prognostic parameter in human endometrial carcinomas: Analysis of inhibin/activin-alpha, -betaA and -betaB subunits in 302 cases. Eur J Cancer 45: 1304-1314, 2009.
- 19 Mylonas I, Makovitzky J, Richter DU, Jeschke U, Briese V and Friese K: Expression of the inhibin-alpha subunit in normal, hyperplastic and malignant endometrial tissue: an immunohistochemical analysis. Gynecol Oncol *93*: 92-97, 2004.

- 20 Reis FM, Luisi S, Carneiro MM, Cobellis L, Federico M, Camargos AF and Petraglia F: Activin, inhibin and the human breast. Mol Cell Endocrinol 225: 77-82, 2004.
- 21 Otani T, Minami S, Kokawa K, Shikone T, Yamoto M and Nakano R: Immunohistochemical localization of activin A in human endometrial tissues during the menstrual cycle and in early pregnancy. Obstet Gynecol 91: 685-692, 1998.
- 22 Robertson DM, Pruysers E, Burger HG, Jobling T, McNeilage J and Healy D: Inhibins and ovarian cancer. Mol Cell Endocrinol 225: 65-71, 2004.
- 23 Johnston RJ and Wallace WH: Normal ovarian function and assessment of ovarian reserve in the survivors of childhood cancer. Pediatr Blood Cancer 53: 296-302, 2009.
- 24 Matzuk MM, Finegold MJ, Su JG, Hsueh AJ and Bradley A: Alpha-inhibin is a tumour-suppressor gene with gonadal specificity in mice. Nature *360*: 313-319, 1992.
- 25 Matzuk MM, Finegold MJ, Mather JP, Krummen L, Lu H and Bradley A: Development of cancer cachexia-like syndrome and adrenal tumors in inhibin-deficient mice. Proc Natl Acad Sci USA 91: 8817-8821, 1994.
- 26 Adkins HB, Bianco C, Schiffer SG, Rayhorn P, Zafari M, Cheung AE, Orozco O, Olson D, De Luca A, Chen LL, Miatkowski K, Benjamin C, Normanno N, Williams KP, Jarpe M, LePage D, Salomon D and Sanicola M: Antibody blockade of the Cripto CFC domain suppresses tumor cell growth *in vivo*. J Clin Invest *112*: 575-587, 2003.
- 27 Razanajaona D, Joguet S, Ay AS, Treilleux I, Goddard-Leon S, Bartholin L and Rimokh R: Silencing of FLRG, an antagonist of activin, inhibits human breast tumor cell growth. Cancer Res 67: 7223-7229, 2007.
- 28 Kalkhoven E, Roelen BA, de Winter JP, Mummery CL, van den Eijnden-van Raaij AJ, van der Saag PT and van der Burg B: Resistance to transforming growth factor beta and activin due to reduced receptor expression in human breast tumor cell lines. Cell Growth Differ 6: 1151-1161, 1995.
- 29 Liu QY, Niranjan B, Gomes P, Gomm JJ, Davies D, Coombes RC and Buluwela L: Inhibitory effects of activin on the growth and morpholgenesis of primary and transformed mammary epithelial cells. Cancer Res *56*: 1155-1163, 1996.
- 30 Cocolakis E, Lemay S, Ali S and Lebrun JJ: The p38 MAPK pathway is required for cell growth inhibition of human breast cancer cells in response to activin. J Biol Chem 276: 18430-18436, 2001.
- 31 Valderrama-Carvajal H, Cocolakis E, Lacerte A, Lee EH, Krystal G, Ali S and Lebrun JJ: Activin/TGF-beta induce apoptosis through Smad-dependent expression of the lipid phosphatase SHIP. Nat Cell Biol 4: 963-969, 2002.

Received June 1, 2010 Revised October 17, 2010 Accepted October 19, 2010