

## **CYP1A2 164 A→C Polymorphism, Cigarette Smoking, Consumption of Well-done Red Meat and Risk of Developing Colorectal Adenomas and Carcinomas**

MONA SÆBØ<sup>1\*</sup>, CAMILLA F. SKJELBRED<sup>1,2\*</sup>, KARIN BREKKE LI<sup>1</sup>, INGER MARIE BOWITZ LOTHE<sup>3</sup>, PER CHR. HAGEN<sup>1</sup>, EGIL JOHNSEN<sup>4</sup>, KJELL M. TVEIT<sup>5</sup> and ELIN H. KURE<sup>1,3,5</sup>

<sup>1</sup>Telemark University College, Faculty of Arts and Sciences, Bø i Telemark;

<sup>2</sup>Department of Laboratory Medicine, Section of Medical Genetics, Telemark Hospital, Skien;

Departments of <sup>3</sup>Pathology, <sup>4</sup>Surgery and <sup>5</sup>Oncology, Ullevål University Hospital, Oslo, Norway

**Abstract.** *Background: Genetic polymorphisms in metabolizing enzymes may modify the association of environmental exposure on colorectal cancer (CRC) and adenoma risk. Materials and Methods: One hundred and ninety-eight CRC cases, 422 adenomas (206 low-risk and 216 high-risk adenomas) and 222 controls were genotyped for the CYP1A2 164 A→C polymorphism and questionnaires were used to assess environmental exposure. Results: The smoking parameter “current smoking” was significantly associated with CRC risk, and all the smoking parameters related to current smoking, having ever smoked or high numbers of cigarette years were significantly associated with risk of adenomas. No association was detected between red meat consumption or how well red meat was cooked and colorectal carcinogenesis. When stratifying the case groups based on CYP1A2 genotype, all the smoking parameters yielded stronger risk association in carriers of the C allele. Conclusion: These findings may indicate that the association between cigarette smoking and colorectal carcinogenesis can be modified by the CYP1A2 genotype.*

Worldwide, colorectal cancer (CRC) is the fourth most frequent type of cancer, while in developed countries it comes second (1). Most CRCs develop through multiple mutations in the normal colonic mucosa and evolve through the adenoma-carcinoma sequence (2, 3). Development of

sporadic colorectal adenomas and carcinomas has been associated with several lifestyle factors, including cigarette smoking (4, 5) and dietary items such as red meat (6-8).

Cigarette smoke is a major source of a wide variety of carcinogens, including nitrosamines, polycyclic hydrocarbons (PAHs), aromatic amines (AAs) and heterocyclic aromatic amines (HCAs) (9, 10). Carcinogens in cigarettes may reach the colorectal mucosa through the circulatory system (11, 12). Long-term, heavy cigarette smokers have a 2- to 3-fold elevated risk of colorectal adenoma and the vast majority of studies in the past several years show an association between cigarette use and CRC (4, 13). In addition, cigarette smoke has been shown to induce the expression of some metabolic genes (14). The risk-enhancing effect of red meat is not precisely known, but meat prepared at high temperatures may contain significant quantities of a number of HCAs (15).

The influence of exposures on cancer development may be affected by variation in biotransformation of carcinogens. The cytochrome P450 (CYP)-dependent monooxygenase (Phase I enzyme) represents the first line of defense against toxic chemicals (16). CYP1A2 is the major enzyme involved in the metabolism of HCAs and AAs (17, 18) and phenotype studies have detected large interindividual variation of CYP1A2 expression in the liver (19, 20). The distribution of CYP1A2 activity in humans is generally trimodal or bimodal (slow/intermediate/rapid or slow/rapid) (20-23). The expression of CYP1A2 is believed to be regulated by two mechanisms: one that controls constitutive expression and one that regulates inducibility (21). Variation in CYP1A2 activity in humans may be due to various environmental exposures, including cigarette smoke (9, 14, 21, 24-27), genetic differences (28) or gene-gene interaction (29).

In phenotype studies, rapid CYP1A2 in combination with reported preference for well-done red meat and smoking status (ever smoking) has been associated with increased CRC risk (30, 31). The hypothesis is that increased CYP1A2 activity increases CRC susceptibility (26, 31). The CYP1A2

\*Both authors contributed equally to this work.

Correspondence to: Elin H. Kure, Telemark University College, Faculty of Arts and Sciences, Hallvard Eikas plass, 3800 Bø i Telemark, Norway. Tel: +47 23027822, Fax: +47 35952703, e-mail: elin.kure@hit.no

Key Words: CYP1A2 164 A→C polymorphism, smoking, red meat, colorectal adenomas, colorectal carcinomas.

164 A→C (*CYP1A2\*1F*) polymorphism (20), located in intron 1 is common in Caucasians. The functional significance of this polymorphism is not yet clear but it appears to affect enzyme inducibility. Some studies have reported that the *CYP1A2* 164 A/A genotype may represent a high inducibility genotype (26, 28, 32), but a recent Korean study found higher *CYP1A2* activity in individuals carrying the *CYP1A2* C allele (33).

In this study, the association between cigarette smoking, intake of well-done red meat, the *CYP1A2* 164 A→C polymorphism and adenoma and CRC risk in a Norwegian population was investigated. We also investigated whether the association between environmental exposure and adenoma and CRC risk could be modified by the *CYP1A2* genotype.

## Materials and Methods

The cohort in the KAM (Kolorektal Cancer, Arv og Miljø/CRC Inheritance and Environment) molecular epidemiological study is based on the screening group of the Norwegian Colorectal Cancer Prevention study (The NORCCAP study) in the county of Telemark (34). In addition, patients diagnosed with CRC operated on at Telemark Hospital (Skien) and Ullevål University Hospital (Oslo) were included. The KAM biobank has been described elsewhere (35). All of the participants completed a questionnaire on demographics, health status, dietary and smoking habits, alcohol consumption, physical exercise and occupation. The Regional Ethics Committee and the Data Inspectorate approved the KAM study.

The KAM cohort is based on an ethnic homogeneous group of Norwegian origin. The ID number for the NORCCAP study at Clinicaltrials.gov is -I NCT00119912 (36). The 424 adenoma cases (206 with low-risk and 216 with high-risk adenomas) and a control group of 222 individuals were drawn from the NORCCAP study. The high-risk adenoma, defined by high-grade dysplasia, size >1 cm or villous features, has a greater potential of developing into a cancerous tumor than a low-risk adenoma (37). All the controls were screen negative participants (negative flexible sigmoidoscopy). For CRC cases, all patients diagnosed with colorectal cancer who were mentally competent to complete the questionnaire were asked to participate in the KAM study. The questionnaire contained information on a family history of cancer and the selected CRC cases had no known personal history of cancer. In this study, 198 CRC cases were drawn from the KAM cohort.

Genomic DNA was isolated from blood samples according to standard procedures (38) with minor modifications as described elsewhere (35). The *CYP1A2* 164 A→C polymorphism (*CYP1A2\*1F*) was genotyped according to the method described by Christiansen *et al.* (39). In short, the fragment was amplified using the sense primer 5'-GGAAGGTATCAGCAGAAAGCC-3' and the anti-sense primer 5'-GGCTCATCCTTGACAGTGCC-3' to produce a 626 bp product. The presence of the A/C polymorphism was detected using *Apa I*, which cleaves the C alleles into two products of 181 bp and 445 bp. A positive control was used in all runs and, in addition, 10% of the samples were retyped with identical results.

Differences in characteristics between groups were assessed using the  $\chi^2$  test for categorical variables and the Mann-Whitney test for continuous variables; *P*-values <0.05 were considered significant. Logistic regression was used to examine the association between cigarette smoking, red meat consumption, doneness level of red meat and the *CYP1A2* 164 A→C polymorphism, and colorectal carcinogenesis measured separately as odds ratio (OR) with 95% confidence interval (CI). The *CYP1A2* A allele was used as wild-type because this allele is more common in both Caucasian and Asian populations (33, 40). The international *CYP* allele nomenclature (<http://imm.ki.se/CYPalleles/cyp1a2.htm>) uses the C allele as wild-type. The genotypes were analysed based on the A/A genotype, or carriers of the C allele. Total intake of red meat was created from the dietary items in the questionnaire and the frequency and portion size were multiplied to obtain the amount in grams per day. Intake of red meat was divided into tertiles based on consumption among controls ( $T_1 \leq 22.5g$  (reference category),  $22.5g > T_2 < 45.0g$  and  $T_3 \geq 45.0g$ ). The doneness level of red meat was divided into two categories rare/medium (reference category) and well-done. In addition to an overall case-control comparison, individuals were further stratified based on *CYP1A2* genotype (A/A genotype or carriers of the C allele) and separately examined for smoking and diet parameters. MiniTab Statistical Software, Release 13.1 Xtra (Minitab Inc. USA) was used for statistical calculations. The data were adjusted for age, sex and ever/never smoking status.

## Results

Selected characteristics of cases and controls are presented in Table I. There were significant differences in the numbers of males and females among the control group and the low- and high-risk adenoma groups,  $p < 10^{-4}$ , and between the control and CRC group,  $p = 0.014$ , respectively. There were also significant differences in age between the control group and all of the case groups,  $p < 10^{-4}$ . There were significantly more ever smokers in the low- and high-risk adenoma groups compared to the control group,  $p < 10^{-4}$ , and between the control and CRC group,  $p = 0.019$ , respectively. There was a significant difference in median red meat intake (grams, per day) between the control group and the CRC case group,  $p = 0.04$ . The response rate regarding cigarette smoking (78% vs. 87%) and consumption of red meat (65% vs. 91%) were significantly different between CRC patients and the controls,  $p = 0.018$  and  $< 0.001$ , respectively.

The smoking parameter "current smoker or stopped smoking  $\leq 10$  years ago" was significantly associated with CRC risk OR=2.37 (CI 1.29-4.38) (see Table II). All the smoking parameters, except cigarette years <260 for high-risk adenomas, yielded significant associations for adenomas (see Table II). None of the diet parameters were associated with increased risk of adenoma or carcinoma (see Table II).

The genotype distribution for the *CYP1A2* 164 A→C polymorphism was in Hardy-Weinberg equilibrium and comparable to frequencies reported in other studies of Caucasian populations (28, 41). The *CYP1A2* 164 A→C

Table I. Distribution of selected characteristics by case-control status.

Characteristic	Cases			
	Controls n=222	CRC n=198	High-risk adenomas n=216	Low-risk adenomas n=206
Men	91 (41%)	105 (53%)	141 (65%)	126 (61%)
Women	131 (59%)	93 (47%)	75 (35%)	80 (39%)
Mean age at cohort entry (years)	54.8±3.7	67.7±11.9	57.2±3.5	59.0±3.1
Never smoked*	95 (49%)	59 (38%)	48 (26%)	53 (28%)
Ever smoked*	99 (51%)	97 (62%)	135 (74%)	140 (72%)
Median smoking dose (smoking years)*	260	296	350	350
Never smoked or stopped >10 years ago	145 (73%)	103 (66%)	80 (44%)	92 (48%)
Current smoker or stopped ≤10 years ago	49 (27%)	53 (34%)	103 (56%)	99 (52%)
Median red meat intake (g)*	28.5	24	27	27
Cooking preference well-done <sup>1</sup> *	121 (62%)	73 (62%)	109 (58%)	104 (54%)
<i>CYP1A2</i> 164 A→C Distribution in %, AA/AC/CC	55/38/7	49/44/7	47/44/9	52/40/8

<sup>1</sup>The cooking preference of red meat was divided into two categories rare/medium and well-done. \*Missing values for smoking parameters, red meat intake and doneness level of red meat gave rise to diminished numbers of cases.

Table II. Result of logistic regression analysis for *CYP1A2*-164 A→C polymorphism and smoking and diet parameters.

	Controls	CRC	OR (95% CI) <sup>a</sup>	High-risk adenomas	OR (95% CI) <sup>a</sup>	Low-risk adenomas	OR (95% CI) <sup>a</sup>
<i>CYP1A2</i> -164 A→C*							
A/A	106	73	1(ref)	87	1(ref)	97	1(ref)
Any C	88	83	0.91 (0.52-1.60)	96	1.01 (0.64-1.59)	96	0.99 (0.60-1.62)
Smoking parameters*							
Never smoked	95	59	1(ref)	48	1(ref)	53	1(ref)
Ever smoked	99	97	1.61 (0.90-2.89)	135	2.57 (1.58-4.18)	140	2.48 (1.48-4.17)
Cigarette years <260	49	41	1.41 (0.69-2.89)	42	1.65 (0.91-2.99)	56	2.25 (1.21-4.19)
Cigarette years ≥260	50	54	1.66 (0.83-3.31)	93	3.50 (2.03-6.04)	84	2.70 (1.49-4.89)
Never smoked or stopped >10 years ago	145	103	1(ref)	80	1(ref)	92	1(ref)
Current smoker or stopped ≤10 years ago	49	53	2.37 (1.29-4.38)	103	4.51 (2.75-7.38)	99	4.83 (2.79-8.35)
Red meat (g/day) *							
1 (≤22.5)	74	58	1(ref)	74	1(ref)	78	1(ref)
2 (>22.5 and ≤ 45.0)	77	48	1.07 (0.54-2.14)	78	1.25 (0.74-2.11)	66	1.14 (0.64-2.04)
3 (>45.0)	50	23	1.58 (0.71-3.47)	45	1.05 (0.57-1.92)	50	1.47 (0.75-2.85)
Doneness level*							
Rare/medium	75	45	1(ref)	85	1(ref)	89	1(ref)
Well-done	121	73	0.69 (0.36-1.32)	111	0.69 (0.43-1.10)	104	0.70 (0.42-1.17)

\*Missing values for smoking parameters, red meat intake and doneness level of red meat gave rise to diminished numbers of cases. <sup>a</sup>All data are adjusted for age and gender, and in addition genotyping and diet parameters are adjusted for ever smoking.

polymorphism was not associated with increased adenoma or CRC risk (see Table II). When stratifying the CRC case groups based on *CYP1A2* genotype, all the smoking parameters yielded stronger risk association in carriers of the

C allele than in cases with the A/A genotype (see Table III). None of the case groups showed any significant association with consumption of red, or well-done red meat, even after stratification by the *CYP1A2* genotype (results not shown).

Table III. Effects of smoking parameters on adenoma and CRC case groups stratified by CYP1A2 genotype.

Smoking parameter	CYP1A2 – 164 A→C polymorphism					
	Colorectal cancer <sup>a</sup>		High-risk adenomas <sup>a</sup>		Low-risk adenomas <sup>a</sup>	
	A/A	Any C	A/A	Any C	A/A	Any C
Never smoked	1(ref)	1(ref)	1(ref)	1(ref)	1(ref)	1(ref)
Case/control	33/52	26/43	30/52	18/43	29/52	24/43
Ever smoked						
Case/control	40/54	57/45	57/54	78/45	68/54	72/45
OR (95% CI)	1.02 (0.47-2.21)	2.92 (1.15-7.41)	1.68 (0.87-3.24)	4.43 (2.09-9.38)	2.15 (1.04-4.43)	2.98 (1.40-6.35)
Cigarette years <260*						
Case /control	16/28	25/21	21/28	21/21	24/28	32/21
OR (95% CI)	0.59 (0.21-1.64)	3.84 (1.25-11.78)	1.23 (0.55 -2.78)	2.55 (1.03-6.29)	1.57 (0.63-3.89)	3.13 (1.31-7.48)
Cigarette years ≥260*						
Case /control	24/26	30/24	36/26	57/24	44/26	40/24
OR (95% CI)	1.56 (0.63-3.86)	1.89 (0.61-5.83)	2.15 (1.01 - 4.54)	6.15 (3.71-13.97)	2.67 (1.17-6.06)	2.85 (1.19-6.83)
Never smoked or stopped						
>10 years ago*	1(ref)	1(ref)	1(ref)	1(ref)	1(ref)	1(ref)
Case/control	52/77	51/68	39/77	47/68	48/77	44/68
Current smoker or stopped						
≤10 years ago*						
Case/control	21/29	32/20	48/29	59/20	49/29	50/20
OR (95% CI)	1.42 (0.61-3.27)	4.61 (1.79-11.87)	3.36 (1.71-6.60)	6.29 (3.00-13.16)	3.72 (1.75-7.91)	6.53 (2.91-14.64)

\*Missing values for smoking parameters, gave rise to diminished numbers of cases. <sup>a</sup>All data are adjusted for age and gender.

## Discussion

Sporadic CRC is a complex disease and several environmental and genetic factors may have an impact on its development. We detected significant associations for cigarette smoking and adenoma and carcinoma risk, which is in agreement with the majority of studies in the past several years (4, 13). We detected no association between consumption of red meat or the doneness level of red meat and risk of colorectal carcinogenesis. A number of studies have focused on the relationship between red meat consumption and CRC risk, but the results remain controversial (6-8, 42-45). The most convincing evidence of a positive association between meat intake and risk of colorectal carcinogenesis comes from case-control studies (46, 47), and the risk is more consistent for red and processed meat than for white meat. Production of HCAs during preparation of meat at high temperature has been suggested as a potential mechanism, but epidemiological evidence is sparse (48, 49). Sinha *et al.* (50) showed that the level of some HCAs in cooked white meat exceeds the levels in cooked red meat, but studies have shown that diets high in white meat (poultry, fish) are not associated with an increased risk (8).

Information regarding exposure may be influenced by recall bias and there is also the chance that CRC patients have changed their diet (more vegetables). In our study group, the median daily intake of red meat was significantly lower among CRC (24.0 g) cases compared to controls (28.5 g) and there was no difference between the groups in the doneness level of consumed red meat.

Case-control comparisons of the CYP1A2 164 A→C polymorphism yielded no significant association for any of the case groups. This finding is in agreement with previous studies on colorectal cancer risk in Caucasians (41, 51). A recent Korean study detected a stronger association in CRC patients with the CYP1A2 164 C allele in individuals ≥55 years of age after adjusting for smoking habits (33). Moonen *et al.* (52) suggested that individuals carrying the CYP1A2 A/A genotype may be at higher risk for developing high-risk adenomas compared to those with the any C allele, but this study was small with only 19 controls and 74 adenomas (37 adenomas <1 cm and 38 adenomas ≥1 cm and carcinoma *in situ*) (52).

In our study, we stratified the case groups based on the CYP1A2 genotype (A/A or any C allele). All the smoking parameters yielded stronger risk association in carriers of the C allele than in cases with the A/A genotype. This may

indicate that the risk associated with cigarette smoking may be modified by the *CYP1A2* genotype. The smoking parameter “ever smoking” yielded significant associations only in carriers of the C allele for CRC and high-risk adenomas, but for low-risk adenomas both *CYP1A2* genotypes yielded significant results. This may imply that smoking can initiate carcinogenesis regardless of genotype, but that progression is more dependent on genotype.

The prevailing hypothesis is that increased *CYP1A2* activity increases the likelihood of CRC development (26, 31). In which way the C allele affects inducibility and enzyme activity is not clear. Studies of the *CYP1A2* 164 A→C polymorphism and protein activity in humans have reported conflicting evidence. Both the A/A and any C allele had either no effect (53-55), or increased, or decreased activity (26, 28, 32, 33). Various markers have been used to assess protein activity (urinary caffeine metabolites, plasma metabolic ratio, urinary PhIP metabolites, clozapine serum concentrations) which makes it difficult to compare results from different studies. A Korean study used the urinary caffeine challenge test to analyze the genotype phenotype association and found that the *CYP1A2* activity in healthy smokers with the C allele was significantly higher than that in individuals with the A/A genotype (33). The genotype frequencies of the *CYP1A2* 164 A→C polymorphism in the Korean study (33) were comparable to the result in this study and other Caucasian studies (28, 41). To clarify the effect of *CYP1A2* 164 A→C polymorphism on activity, identical methods for measuring activity should be used in additional studies to enhance our understanding of the genotype–phenotype associations.

Most CRCs evolve through the adenoma–carcinoma sequence (56) and high-risk adenomas have a greater potential of developing into a cancerous tumor (37). The fact that we detected a stronger effect modification for smoking parameters and the *CYP1A2* genotype in high-risk adenomas and the CRC case group compared to low-risk adenomas strengthens our findings. This may imply that the modifying effect of the *CYP1A2* genotype in combination with smoking is more strongly associated with progression than initiation.

The *CYP1A2* 164 A→C polymorphism is located in intron 1 and variation in activity may be due to both environmental exposures and gene–gene interactions (29). A shortcoming of this study is the lack of genotyping in other genes that may influence the biotransformation of carcinogens in cigarette smoke, such as *CYP1A1*, *N*-acetyltransferase (*NAT*) and glutathione *S*-transferases (*GSTs*). The analysis was also limited by a relatively small sample size. This is a pilot study, so we did not commit all our samples to the genotype analysis. The response rate for both cigarette smoking and consumption of red meat were significantly different between CRC patients and the controls. This may affect the

result for overall association for cigarette smoking and red meat consumption. However, in this study we wanted to see if the *CYP1A2* genotype could modify the association of environmental exposure. There is no risk of bias in relation to genotype and the difference in response rate should therefore not affect the outcome.

Our results show an association between cigarette smoking and colorectal carcinogenesis. This association may be modified by the *CYP1A2* genotype since all the smoking parameters yielded stronger risk association in carriers of the C allele, than among cases with the A/A genotype. These results highlight the importance of considering genetic susceptibility when evaluating external exposure. A larger study would be required to clarify this issue.

### Acknowledgements

We thank Dr. Steinar Aase for contributing to the pathology of the cancer cases and Dr. Gunter Bock, Hans-Olaf Johannessen and Gro Wiedswang for collecting tumor tissues.

This study was supported by the Norwegian Cancer Society (Grant numbers 51024/001 and E01-0851001), Telemark University College (Grant number 22069) and the Norwegian Colorectal Cancer Prevention (NORCCAP) study (Grants from the Norwegian Cancer Society and the Department of Health and Social Affairs), Eastern Norway Regional Health Authority and with the aid of EXTRA funds from the Norwegian Foundation for Health and Rehabilitation (2001/2/0110).

### References

- 1 Parkin DM, Bray FI, Devesa SS: Cancer burden in the year 2000. The global picture. *Eur J Cancer* 37(Suppl 8): S4-66, 2001.
- 2 Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM and Bos JL: Genetic alterations during colorectal-tumor development. *N Engl J Med* 319: 525-532, 1988.
- 3 Bond JH: Clinical evidence for the adenoma carcinoma sequence and the management of patients with colorectal adenomas. *Semin Gastrointest Dis* 11: 176-184, 2000.
- 4 Giovannucci E: An updated review of the epidemiological evidence that cigarette smoking increases risk of colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 10: 725-731, 2001.
- 5 Almendingen K, Hofstad B, Trygg K, Hoff G, Hussain A and Vatn MH: Smoking and colorectal adenomas: a case-control study. *Eur J Cancer Prev* 9: 193-203, 2000.
- 6 Heavey PM, McKenna D and Rowland IR: Colorectal cancer and the relationship between genes and the environment. *Nutr Cancer* 48: 124-141, 2004.
- 7 Potter JD: Colorectal cancer: molecules and populations. *J Natl Cancer Inst* 91: 916-932, 1999.
- 8 Giovannucci E, Rimm EB, Stampfer MJ, Colditz GA, Ascherio A and Willett WC: Intake of fat meat and fiber in relation to risk of colon cancer in men. *Cancer Res* 54: 2390-2397, 1994.
- 9 Manabe S, Tohyama K, Wada O and Aramaki T: Detection of a carcinogen 2-amino-1-methyl-6-phenylimidazo[45-b]pyridine (PhIP) in cigarette smoke condensate. *Carcinogenesis* 12: 1945-1947, 1991.

- 10 Wogan GN, Hecht SS and Felton JS: Environmental and chemical carcinogenesis. *Semin Cancer Biol* 14: 473-486, 2004.
- 11 Yamasaki E and Ames BN: Concentration of mutagens from urine by absorption with the nonpolar resin XAD-2: cigarette smokers have mutagenic urine. *Proc Natl Acad Sci USA* 74: 3555-3559, 1977.
- 12 Kune GA, Kune S, Vitetta L and Watson LF: Smoking and colorectal cancer risk: data from the Melbourne Colorectal Cancer Study and brief review of literature. *Int J Cancer* 50: 369-372, 1992.
- 13 International Agency for Research on Cancer: Tobacco smoke and involuntary smoking. IARC Monogr Eval Carcinog Risks Hum Vol 83, IARC, Lyon, France, pp. 1-1438, 2004.
- 14 Koide A, Fuwa K, Furukawa F, Hirose M, Nishikawa A and Mori Y: Effect of cigarette smoke on the mutagenic activation of environmental carcinogens by rodent liver. *Mutat Res* 428: 165-176, 1999.
- 15 Ishibe N, Sinha R, Hein D, Kulldorff M, Strickland P, Fretland AJ, Chow WH, Kadlubar FF, Lang NP and Rothman N: Genetic polymorphisms in heterocyclic amine metabolism and risk of colorectal adenomas. *Pharmacogenetics* 12: 145-150, 2002.
- 16 Guengerich FP: Catalytic selectivity of human cytochrome P450 enzymes: relevance to drug metabolism and toxicity. *Toxicol Lett* 70: 133-138, 1994.
- 17 Eaton DL, Gallagher EP, Bammler TK and Kunze KL: Role of cytochrome P4501A2 in chemical carcinogenesis: implications for human variability in expression and enzyme activity. *Pharmacogenetics* 5: 259-274, 1995.
- 18 Boobis AR, Lynch AM, Murray S, de la Torre R, Solans A, Farre M, Segura J, Gooderham NJ and Davies DS: CYP1A2-catalyzed conversion of dietary heterocyclic amines to their proximate carcinogens is their major route of metabolism in humans. *Cancer Res* 54: 89-94, 1994.
- 19 Kalow W and Tang BK: Use of caffeine metabolite ratios to explore CYP1A2 and xanthine oxidase activities. *Clin Pharmacol Ther* 50: 508-519, 1991.
- 20 Aitchison KJ, Gonzalez FJ, Quattrochi LC, Sapone A, Zhao JH, Zaher H, Elizondo G, Bryant C, Munro J, Collier DA, Makoffa and Kerwin RW: Identification of novel polymorphisms in the 5' flanking region of CYP1A2 characterization of interethnic variability and investigation of their functional significance. *Pharmacogenetics* 10: 695-704, 2000.
- 21 Landi MT, Sinha R, Lang NP and Kadlubar FF: Human cytochrome P4501A2. *IARC Sci Publ* 148: 173-195, 1999.
- 22 Ou-Yang DS, Huang SL, Wang W, Xie HG, Xu ZH, Shu Y and Zhou HH: Phenotypic polymorphism and gender-related differences of CYP1A2 activity in a Chinese population. *Br J Clin Pharmacol* 49: 145-151, 2000.
- 23 Butler MA, Lang NP, Young JF, Caporaso NE, Vineis P, Hayes RB, Teitel CH, Massengill JP, Lawsen MF and Kadlubar FF: Determination of CYP1A2 and NAT2 phenotypes in human populations by analysis of caffeine urinary metabolites. *Pharmacogenetics* 2: 116-127, 1992.
- 24 Schrenk D, Brockmeier D, Morike K, Bock KW and Eichelbaum M: A distribution study of CYP1A2 phenotypes among smokers and non-smokers in a cohort of healthy Caucasian volunteers. *Eur J Clin Pharmacol* 53: 361-367, 1998.
- 25 Kitada M, Taneda M, Ohta K, Nagashima K, Itahashi K and Kamataki T: Metabolic activation of aflatoxin B1 and 2-amino-3-methylimidazo[45-f]-quinoline by human adult and fetal livers. *Cancer Res* 50: 2641-2645, 1990.
- 26 Sachse C, Bhambra U, Smith G, Lightfoot TJ, Barrett JH, Scollay J, Garner RC, Boobis AR, Wolf CR and Gooderham NJ: Polymorphisms in the cytochrome P450 CYP1A2 gene (CYP1A2) in colorectal cancer patients and controls: allele frequencies linkage disequilibrium and influence on caffeine metabolism. *Br J Clin Pharmacol* 55: 68-76, 2003.
- 27 Nakajima M, Yokoi T, Mizutani M, Kinoshita M, Funayama M and Kamataki T: Genetic polymorphism in the 5'-flanking region of human CYP1A2 gene: effect on the CYP1A2 inducibility in humans. *J Biochem (Tokyo)* 125: 803-808, 1999.
- 28 Sachse C, Brockmoller J, Bauer S and Roots I: Functional significance of a C->A polymorphism in intron 1 of the cytochrome P450 CYP1A2 gene tested with caffeine. *Br J Clin Pharmacol* 47: 445-449, 1999.
- 29 MacLeod S, Sinha R, Kadlubar FF and Lang NP: Polymorphisms of CYP1A1 and GSTM1 influence the *in vivo* function of CYP1A2. *Mutat Res* 376: 135-142, 1997.
- 30 Le Marchand L, Hankin JH, Wilkens LR, Pierce LM, Franke A, Kolonel LN, Seifried A, Custer LJ, Chang W, Lum-Jones A and Donlon T: Combined effects of well-done red meat, smoking and rapid N-acetyltransferase 2 and CYP1A2 phenotypes in increasing colorectal cancer risk. *Cancer Epidemiol Biomarkers Prev* 10: 1259-1266, 2001.
- 31 Lang NP, Butler MA, Massengill J, Lawson M, Stotts RC, Hauer-Jensen M and Kadlubar FF: Rapid metabolic phenotypes for acetyltransferase and cytochrome P4501A2 and putative exposure to food-borne heterocyclic amines increase the risk for colorectal cancer or polyps. *Cancer Epidemiol Biomarkers Prev* 3: 675-682, 1994.
- 32 Moonen HJ, Moonen EJ, Maas L Dallinga JW, Kleinjans JC and de Kok TM: CYP1A2 and NAT2 genotype/phenotype relations and urinary excretion of 2-amino-1-methyl-6-phenylimidazo[45-b]pyridine (PhIP) in a human dietary intervention study. *Food Chem Toxicol* 42: 869-878, 2004.
- 33 Bae SY, Choi SK, Kim KR, Park CS, Lee SK, Roh HK, Shin DW, Pie JE, Woo ZH and Kang JH: Effects of genetic polymorphisms of MDR1 FMO3 and CYP1A2 on susceptibility to colorectal cancer in Koreans. *Cancer Sci* 97: 774-779, 2006.
- 34 Gondal G, Grotmol T, Hofstad B, Bretthauer M, Eide TJ and Hoff G: The Norwegian Colorectal Cancer Prevention (NORCCAP) screening study: baseline findings and implementations for clinical work-up in age groups 50-64 years. *Scand J Gastroenterol* 38: 635-642, 2003.
- 35 Hansen R, Saebø M, Skjelbred CF, Nexø BA, Hagen PC, Bock G, Bowitz Lothe IM, Johnson E, Aase S, Hansteen IL, Vogel U and Kure EH: GPX Pro198Leu and OGG1 Ser326Cys polymorphisms and risk of development of colorectal adenomas and colorectal cancer. *Cancer Lett* 229: 85-91, 2005.
- 36 <http://www.clinicaltrials.gov>
- 37 Winawer SJ and Zauber AG: The advanced adenoma as the primary target of screening. *Gastrointest Endosc Clin N Am* 12: 1-9 v, 2002.
- 38 Miller SA, Dykes DD and Polesky HF: A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res* 16: 1215, 1988.

- 39 Christiansen L, Bygum A, Jensen A, Thomsen K, Brandrup F, Horder M and Petersen NE: Association between *CYP1A2* polymorphism and susceptibility to porphyria cutanea tarda. *Hum Genet* 107: 612-614, 2000.
- 40 Todesco L, Torok M, Krahenbuhl S and Wenk M: Determination of -3858G→A and -164C→A genetic polymorphisms of *CYP1A2* in blood and saliva by rapid allelic discrimination: large difference in the prevalence of the -3858G→A mutation between Caucasians and Asians. *Eur J Clin Pharmacol* 59: 343-346, 2003.
- 41 Sachse C, Smith G, Wilkie MJ, Barrett JH, Waxman R, Sullivan F, Forman D, Bishop DT and Wolf CR: A pharmacogenetic study to investigate the role of dietary carcinogens in the etiology of colorectal cancer. *Carcinogenesis* 23: 1839-1849, 2002.
- 42 Gaard M, Tretli S and Loken EB: Dietary factors and risk of colon cancer: a prospective study of 50535 young Norwegian men and women. *Eur J Cancer Prev* 5: 445-454, 1996.
- 43 Yoon H, Benamouzig R, Little J, Francois-Collange M and Tome D: Systematic review of epidemiological studies on meat dairy products and egg consumption and risk of colorectal adenomas. *Eur J Cancer Prev* 9: 151-164, 2000.
- 44 Kampman E, Slattery ML, Bigler J, Leppert M, Samowitz W, Caan BJ and Potter JD: Meat consumption genetic susceptibility and colon cancer risk: a United States multicenter case-control study. *Cancer Epidemiol Biomarkers Prev* 8: 15-24, 1999.
- 45 Sandhu MS, White IR and McPherson K: Systematic review of the prospective cohort studies on meat consumption and colorectal cancer risk: a meta-analytical approach. *Cancer Epidemiol Biomarkers Prev* 10: 439-446, 2001.
- 46 Breuer-Katschinski B, Nemes K, Marr A, Rump B, Leiendecker B, Breuer N and Goebell H: Colorectal adenomas and diet: a case-control study. Colorectal Adenoma Study Group. *Dig Dis Sci* 46: 86-95, 2001.
- 47 Norat T and Riboli E: Meat consumption and colorectal cancer: a review of epidemiologic evidence. *Nutr Rev* 59: 37-47, 2001.
- 48 Augustsson K, Skog K, Jagerstad M, Dickman PW and Steineck G: Dietary heterocyclic amines and cancer of the colon rectum bladder and kidney: a population-based study. *Lancet* 353: 703-707, 1999.
- 49 Butler LM, Sinha R and Millikan RC: Heterocyclic amines meat intake and association with colon cancer in a population-based study. *Am J Epidemiol* 157: 434-445, 2003.
- 50 Sinha R, Rothman N, Brown ED, Salmon CP, Knize MG, Swanson CA, Rossi SC, Mark SD, Levander OA and Felton JS: High concentrations of the carcinogen 2-amino-1-methyl-6-phenylimidazo-[4,5-b]pyridine (PhIP) occur in chicken but are dependent on the cooking method. *Cancer Res* 55: 4516-4519, 1995.
- 51 Kiss I, Orsos Z, Gombos K, Bogner B, Csejtei A, Tibold A, Varga Z, Pazsit E, Magda I, Zolyomi A and Ember I: Association between allelic polymorphisms of metabolizing enzymes (*CYP1A1*, *CYP1A2*, *CYP2E1*, *mEH*) and occurrence of colorectal cancer in Hungary. *Anticancer Res* 27: 2931-2937, 2007.
- 52 Moonen H, Engels L, Kleinjans J and Kok T: The *CYP1A2*-164A→C polymorphism (*CYP1A2\*1F*) is associated with the risk for colorectal adenomas in humans. *Cancer Lett* 229: 25-31, 2005.
- 53 Kootstra-Ros JE, Smallegoor W and van der Weide J: The cytochrome P450 *CYP1A2* genetic polymorphisms \*1F and \*1D do not affect clozapine clearance in a group of schizophrenic patients. *Ann Clin Biochem* 42: 216-219, 2005.
- 54 van der Weide J, Steijns LS and van Weelden MJ: The effect of smoking and cytochrome P450 *CYP1A2* genetic polymorphism on clozapine clearance and dose requirement. *Pharmacogenetics* 13: 169-172, 2003.
- 55 Nordmark A, Lundgren S, Ask B, Granath F and Rane A: The effect of the *CYP1A2\*1F* mutation on *CYP1A2* inducibility in pregnant women. *Br J Clin Pharmacol* 54: 504-510, 2002.
- 56 Bond JH: Polyp guideline: diagnosis treatment and surveillance for patients with colorectal polyps. Practice Parameters Committee of the American College of Gastroenterology. *Am J Gastroenterol* 95: 3053-3063, 2000.
- 57 Lieberman DA, Weiss DG, Bond JH, Ahnen DJ, Garewal H and Chejfec G: Use of colonoscopy to screen asymptomatic adults for colorectal cancer. Veterans Affairs Cooperative Study Group 380. *N Engl J Med* 343: 162-168, 2000.

Received February 21, 2008

Revised April 23, 2008

Accepted May 5, 2008