Review

The Roles of ZFAT in Thymocyte Differentiation and Homeostasis of Peripheral Naive T-Cells

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Abstract. ZFAT (zinc-finger gene in AITD susceptibility region), originally identified as a candidate susceptibility gene for autoimmune thyroid disease, has been reported to be involved in various cellular processes and several common diseases including multiple sclerosis. Recent studies revealed that mouse Zfat is a novel critical regulator for both thymocyte differentiation and peripheral T-cell homeostasis. Zfat deficiency at early thymocyte developmental stages results in the inhibition of the development of CD4⁺CD8⁺ thymocytes with an impaired positive selection. Zfat deficiency in peripheral T-cells results in a reduction in the number of Tcells with decreased expression of the interleukin-7 receptor- α (IL-7R α) that is critical for T-cell homeostasis. In addition, T-cell antigen receptor stimulation-induced responses of Zfatdeficient T-cells are also impaired, with reduced IL-2Ra expression. This review highlights and discusses the roles of Zfat in thymocyte differentiation of T-cells and in the homeostasis of naive T-cells with recent work.

We previously identified zinc-finger gene with AT-hook/zinc-finger gene in autoimmune thyroid disease susceptibility region (*ZFAT*) as a candidate susceptibility gene for autoimmune thyroid disease (1). *ZFAT* encodes an evolutionally-conserved protein with 18 zinc-finger motifs and one AT-hook domain (2). The ZFAT protein is strongly expressed in T- and B-cells in immune-related tissues including the thymus, the spleen, lymph nodes and peripheral blood (2). We reported that *Zfat* deficiency in the mouse (*Zfat*^{-/-}) is embryonically lethal by embryonic day 8.5 and that *Zfat* is a critical transcriptional regulator for *Tal1*

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Key Words: ZFAT, transcription factor, thymocyte differentiation, homeostasis of naive T-cells, review.

(T-cell acute lymphocytic leukemia protein 1), Gata1 (GATAbinding factor 1) and *Lmo2* (LIM domain only 2) expression in primitive hematopoiesis (3, 4). We also found that ZFAT is involved in the regulation of apoptosis in a human leukemia cell line (MOLT-4) (5) and mouse embryonic fibroblasts (6), and in the differentiation of human umbilical vein endothelial cells (7). Furthermore, genetic variants of ZFAT have been reported to be associated with the severity of Hashimoto's disease (8), with adult height in Japanese and Korean populations (9, 10), and with several common diseases including hypertension and cancer (11, 12). Of interest is that a genetic variant of ZFAT is reported to be most strongly associated with interferon-β responsiveness in multiple sclerosis (MS) (13), in which interleukin-7 receptor-α (IL7RA) and IL2RA are susceptibility genes (14-16). Recent studies have shown that ZFAT is a novel imprinted gene expressed in the human placenta (17), and was also reported to be overexpressed in unruptured aneurysms, suggesting that ZFAT could be a possible molecular marker in peripheral blood to predict a high risk of aneurysm rupture (18). The functional roles of ZFAT in biology and genetics have expanded dramatically since its identification and cloning. In our recent studies, we established and analyzed Zfat^{f/f}-LckCre mice and Zfatf/f-Cd4Cre mice, and we found that Zfat is essential for both thymocyte development and peripheral T-cell homeostasis through Erk activation and the expression of Il7ra and Il2ra, respectively (19, 20). Herein, we review the function of Zfat and discuss recent research regarding the signaling factors that control positive selection in thymocyte development, and the cytokine receptors and autoimmune diseases involved in the homeostasis of naive T-cells.

The Role of Zfat in Thymocyte Differentiation

In the thymus, signals transduced by the T-cell antigen receptor (Tcr) promote the transition of CD4⁺CD8⁺ double-positive thymocytes to CD4⁺CD8⁻ single-positive and CD4⁻CD8⁺ single-positive thymocytes, and also regulate CD4

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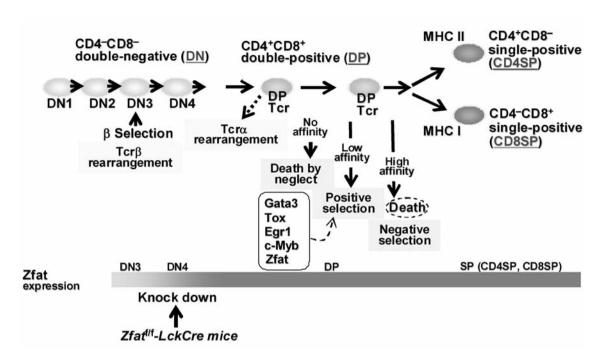


Figure 1. Model of Zfat function in thymocyte differentiation. Thymocyte differentiation is described based on the pattern of cell surface markers expressed at each stage. Double-negative (DN) cells are divided into DN1, DN2, DN3 and DN4 stages. DN4 cells differentiate into double-positive (DP) cells expressing both CD4 and CD8. The expression of mature α/β Tcr by the re-arrangement of Tcra gene leads to the process of positive and negative selection, ensuring a diverse repertoire of T-cells. DP cells that have undergone positive selection differentiate into CD4 single-positive and CD8 single-positive cells. Zfat expression during the thymocyte development begins from the DN3 stage. Zfat deficiency in thymocyte development results in the inhibition of DP thymocyte development with impaired positive selection, indicating that Zfat is essential for the development of DP thymocytes.

versus CD8 lineage commitment (21, 22). In the transition of DP to SP thymocytes, transcriptional factors, including Gata3 (23), Tox(Thymocyte selection-associated high mobility group box) (24), Egr1 (Early growth response protein 1) (25) and c-Myb (V-myb avian myeloblastosis viral oncogene homolog) (26), are critical for positive selection, but not for CD4 versus CD8 lineage commitment. However, the precise molecular mechanisms and the orchestrated gene expression programs in T-cell development are not yet fully-understood. In our study, we generated Zfatf/f-LckCre mice and observed that they exhibited a loss of CD3ζ phosphorylation with dysregulation of Erk(extracellular signal-regulated kinase) and Egr activities, leading to impaired positive selection. We demonstrated that Zfat is required for the proper regulation of Tcr proximal signaling, and that Zfat is a crucial molecule for positive selection in the thymus (Figure 1).

The T-cell repertoire is whittled-down by negative selection, which abolishes T-cells with Tcrs that recognize self-peptides and major histocompatibility complex proteins (self-pMHC) with high affinity, and by positive selection, which requires T-cells with Tcrs that recognize self-pMHC with low affinity. The Tcr signaling pathway can be divided into proximal signaling, Ras(Rat sarcoma viral oncogene

homolog)-mitogen activated protein kinase (Mapk) signaling and calcium-mediated signaling (Figure 2). phosphorylation of Erk1/2 induced by TCR-stimulation through Ras-Mapk signaling was markedly decreased in Zfat^{f/f}-LckCre thymocytes. In agreement with the defects in Erk1/2 activation, the phosphorylation of both Mek1/2 and c-Raf(V-Raf-1 murine leukemia viral oncogene homolog 1), which are located upstream of the Erk signaling pathway, was also reduced in the Zfatf/f-LckCre thymocytes. The phosphorylation of Zap70 (Zeta-chain-associated protein kinase) and Plcy1 (Phospholipase C, gamma-1) was also diminished in Zfatf/f-LckCre thymocytes. Finally, the Tcr stimulation-induced phosphorylation of CD3ζ, which is proximal signaling, was virtually ablated in the Zfatff-LckCre thymocytes, and phosphorylated CD3ζ at nonstimulated status was also apparently diminished due to the Zfat deficiency (Figure 2). By responding to changes of pMHC, the Tcr signal strength provides the effects on signaling mechanisms for positive selection and negative selection during development. The protein schnurri-2 (Shn2; also known as Hivep2) has been reported to be a critical regulator of T-cell development controling the balance between death and differentiation by modulating the Tcr

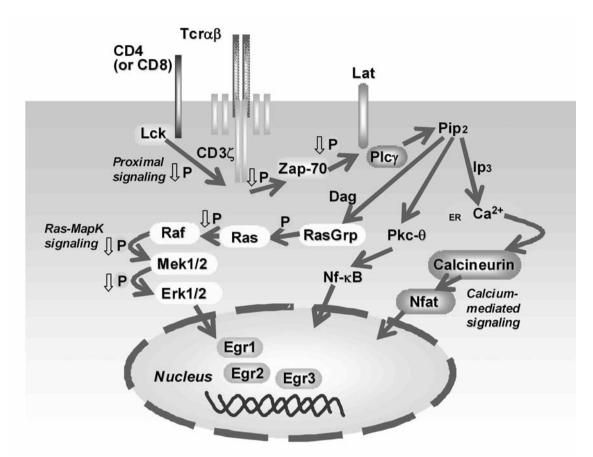


Figure 2. Impaired Tcr signaling pathways coursed by Zfat deficiency. The Tcr signaling pathway can be divided into three pathways: proximal signaling, Ras-mitogen activated protein kinase (Mapk) signaling and calcium-mediated signaling. The recognition of pMHC ligands by Tcr leads to signaling cascades. Zfat deficiency in thymocyte development results in a loss of $CD3\zeta$ phosphorylation with dysregulation of Erk and Egr activities, leading to impaired positive selection. Deregulated phosphorylation by Zfat deficiency is indicated by open arrows.

signal strength (27). In addition, thymocyte-expressed molecule involved in selection (Themis) was reported to set the signal threshold for positive and negative selection in T-cell development (28). Further studies are required to establish the roles of Zfat in the modulation of the Tcr signal strength for both positive and negative selection.

We further explored the relevance of *Zfat* in positive selection, but not in the lineage commitment processes during T-cell development. Gata3, Tox, Egr1 and c-Myb are critical for positive selection, and the positive selection signals are thought to result in up-regulation of *Egr1* (21). The induction of mRNA of *Egr1*, *Egr2* (29, 30) and *Egr3* (31) by Tcr stimulation was impaired in the DP thymocytes of *Zfat*²/₂ LckCre mice. Egr transcriptional factors including Egr1, Egr2 and Egr3, contain highly conserved zinc-finger DNA-binding domains that can bind a number of common target gene promoters (32). Egr proteins are central players in the development of thymocytes, and have both redundant and distinct roles in positive selection (25, 29-31). Thus, our

results collectively suggest that Zfat is involved in positive selection, in part, through the regulation of the expression of Egr1, Egr2 and Egr3. However, the possibility that the decreased *Egr* expression in *Zfat*^{f/f}-LckCre double-positive thymocytes simply reflects the reduced number of positive-selected thymocytes cannot be excluded. A full understanding of the precise mechanisms of Zfat function in positive selection and thymocyte development awaits future studies, which will lead to a better understanding of the orchestrated gene expression programs in T-cell development.

The Role of Zfat in the Homeostasis of Naive T-Cells

In peripheral lymphoid tissues, proper regulation of T-cell homeostasis is highly controlled by both cell-extrinsic and cell-intrinsic factors (33-36). Accumulating evidence demonstrates that peripheral T-cell homeostasis is controlled by cytokine receptor-mediated signals, especially II7r, as well as by interaction between Tcr and MHC (37, 38) (Figure 3).

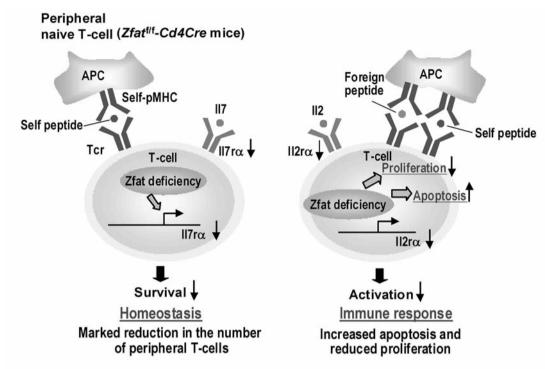


Figure 3. Models of Zfat functions in peripheral T-cell homeostasis and immune response. Naive T-cells receive continuous Tcr signals by interaction with low-affinity self-peptide MHC ligands on antigen-presenting cells (APCs). Continuous Tcr signals together with Il7 signaling in naive T-cells leads to cell survival. In contrast, Tcr signaling induced by high-affinity foreign-peptide MHC ligands results in the activation of T-cells during immune responses. Cd4-Cre-mediated Zfat deficiency results in a reduction in the number of peripheral T-cells with decreased expression of Il7ra. In addition, the Tcr stimulation-induced responses of Zfat-deficient T-cells are also impaired, with reduced Il2ra expression. These results collectively indicate that Zfat plays important roles in peripheral T-cell homeostasis and immune response through the regulation of Il7ra and Il2ra.

Il2/Il2r has a broad array of actions, including the ability to drive T-cell proliferation, mediate activation-induced cell death, promote the development of regulatory T-cells and modulate the expression of cytokine receptors (39, 40). Il2r has three chains: α , β and the common cytokine receptor γ . Resting T-cells express a receptor form composed of β- and γ-chains that bind II2 with moderate affinity, whereas the activation of T-cells induces the α -chain (a high-affinity subunit; CD25) and the formation of the high-affinity heterotrimeric receptor, which plays a critical role in immune response of T-cells (41). In addition, the II7r complex is composed of Il7rα and the γ-chain, and Il7 signaling is mainly regulated by Il7rα expression in T-cells (42). Genetic variants and several haplotypes in the human IL2RA gene have been reported to be associated with type-1 diabetes and MS (14). Furthermore, the IL7RA gene is also a susceptibility gene for MS (15, 16, 43), and IL7 was recently reported to be involved in the generation of T-helper 17 (T_H17) cells, a subset of IL17-producing CD4⁺ T-cells required for the initiation of autoimmune disorders (44, 45).

Several molecules, including forkhead box-O class transcriptional factors (Foxo) (46), Runx (47) and the Klf

family (48), are reported to play critical roles in peripheral T-cell homeostasis as cell-intrinsic factors. Foxo plays important roles in various cellular processes and a wide variety of diseases (36, 49-52). In the immune system, Foxo1 and Foxo3 are predominantly expressed (49, 53), and Foxo1 critically regulates the expression of Il7ra (46, 54). However, the cell-intrinsic factors responsible for the integration of extrinsic signals have not been fully elucidated. In our study, Cd4-Cre-mediated Zfat-deficiency resulted in a remarkable reduction in the number of peripheral T-cells with a decreased expression of II7ra, and in an impaired Tcr stimulation-induced response of T-cells with a reduced Il2ra expression (Figure 3). These findings suggest a functional association of ZFAT with the MS susceptibility genes IL7RA and *IL2RA*. From the viewpoint of autoimmune disorders, an IL7R-mediated signal has been reported to be involved in the generation and survival of T_H17 cells (45), a subset of IL17producing CD4+ T-cells required for the initiation of autoimmune disorders (44), together suggesting a crucial role for ZFAT in autoimmune diseases.

What is the primary event causing the reduced number of peripheral T-cells in *Zfat*^{f/f}-Cd4Cre mice? One of the

remarkable phenotypes observed in the $Zfat^{f/f}$ -Cd4Cre mice is a decrease in the expression of Il7r α on peripheral CD4+T-cells. The responsiveness of the $Zfat^{f/f}$ -Cd4Cre CD44loCD4+-naive T-cells to Il7 is indeed attenuated, which will culminate in reduced survival potency. In addition to the Il7r-mediated signal, when considering that both Il7r α - and Tcr-mediated signals play a crucial role in the survival of naive T-cells (33, 34), it seems possible that the impaired induction of Il2r α expression in response to Tcr underlies the altered T-cell homeostasis and T-cell survival in $Zfat^{f/f}$ -Cd4Cre mice (Figure 3).

The expression of the IL2RA gene is tightly regulated at the transcriptional level, and the positive auto-regulatory loop of the IL2/the high-affinity receptor of IL2R system plays a major role in controlling the magnitude and duration of the T-cell immune response (39). Although positive and negative regulatory regions in the IL2RA gene have been characterized, other regulatory regions or elements are strongly suggested to exist based on the findings of several highly-conserved regions through the IL2RA locus between humans and mice, and the findings of the presence of histone acetylation islands distinct from the known regulatory regions and the autoimmune disorders-associated singlenucleotide polymorphism located in the 5' region of the IL2RA gene (39, 55). In addition, IL2/IL2R signaling has an essential non-redundant role in the production of CD4+CD25+ regulatory T-cells, which are critical for maintaining immune tolerance (56). Thus, both the precise mechanism of the defect in the induction of $I12r\alpha$ expression by Tcr stimulation in Zfat-deficient T-cells and the possibility of the involvement of Zfat in the production of regulatory Tcells and in the development of autoimmune disorders should be examined in future studies.

Conclusion

The functional role of ZFAT in biology has been dramatically expanded by accumulating evidence regarding the functional consequences of Zfat deficiency in mouse models or cell lines. Our findings for the immune system show that Zfat is critical for thymocyte development and Tcell homeostasis in the periphery and that Zfat is crucial for the proper expression of $I17r\alpha$ and $I12r\alpha$ in peripheral T-cells. However, the detailed molecular mechanisms of action of Zfat remain to be elucidated in the thymocyte differentiation of T-cells and in the homeostasis of naive T-cells. The identification of the Zfat target genes essential for thymic development of T-cells and their homeostasis is expected to uncover novel approaches to manipulating the positive selection process in the thymus and T-cell homeostasis. The elucidation of precise molecular mechanisms of Zfat functions will provide new insights into immune regulation and a wide variety of diseases.

Disclosures

The Authors have no financial conflicts of interest.

Acknowledgements

This work was supported in part by Grants-in-Aid for Scientific Research (B) and Scientific Research (C) from the Japan Society for the Promotion of the Science, and a Grant-in-Aid for the FCAM from the Ministry of Education, Culture, Sports, Science and Technology (MEXT)-Supported Program for the Strategic Research Foundation at Private Universities.

References

- 1 Shirasawa S, Harada H, Furugaki K, Akamizu T, Ishikawa N, Ito K, Tamai H, Kuma K, Kubota S, Hiratani H, Tsuchiya T, Baba I, Ishikawa M, Tanaka M, Sakai K, Aoki M, Yamamoto K and Sasazuki T: SNPs in the promoter of a B cell-specific antisense transcript, SAS-ZFAT, determine susceptibility to autoimmune thyroid disease. Hum Mol Genet 13: 2221-2231, 2004.
- 2 Koyanagi M, Nakabayashi K, Fujimoto T, Gu N, Baba I, Takashima Y, Doi K, Harada H, Kato N, Sasazuki T and Shirasawa S: ZFAT expression in B- and T-lymphocytes and identification of ZFAT-regulated genes. Genomics 91: 451-457, 2008.
- 3 Tsunoda T, Takashima Y, Tanaka Y, Fujimoto T, Doi K, Hirose Y, Koyanagi M, Yoshida Y, Okamura T, Kuroki M, Sasazuki T and Shirasawa S: Immune-related zinc finger gene ZFAT is an essential transcriptional regulator for hematopoietic differentiation in blood islands. Proc Natl Acad Sci USA 107: 14199-14204, 2010.
- 4 Tsunoda T and Shirasawa S: Roles of ZFAT in haematopoiesis, angiogenesis and cancer development. Anticancer Res 33: 2833-2837, 2013.
- 5 Fujimoto T, Doi K, Koyanagi M, Tsunoda T, Takashima Y, Yoshida Y, Sasazuki T and Shirasawa S: ZFAT is an antiapoptotic molecule and critical for cell survival in MOLT-4 cells. FEBS Lett 583: 568-572, 2009.
- 6 Doi K, Fujimoto T, Koyanagi M, Tsunoda T, Tanaka Y, Yoshida Y, Takashima Y, Kuroki M, Sasazuki T and Shirasawa S: ZFAT is a critical molecule for cell survival in mouse embryonic fibroblasts. Cell Mol Biol Lett 16: 89-100, 2011.
- 7 Yoshida Y, Tsunoda T, Takashima Y, Fujimoto T, Doi K, Sasazuki T, Kuroki M, Iwasaki A and Shirasawa S: ZFAT is essential for endothelial cell assembly and the branch point formation of capillary-like structures in an angiogenesis model. Cell Mol Biol Lett 15: 541-550, 2010.
- 8 Inoue N, Watanabe M, Yamada H, Takemura K, Hayashi F, Yamakawa N, Akahane M, Shimizuishi Y, Hidaka Y and Iwatani Y: Associations between autoimmune thyroid disease prognosis and functional polymorphisms of susceptibility genes, CTLA4, PTPN22, CD40, FCRL3, and ZFAT, previously revealed in genome-wide association studies. J Clin Immunol 32: 1243-1252, 2012.
- 9 Cho YS, Go MJ, Kim YJ, Heo JY, Oh JH, Ban HJ, Yoon D, Lee MH, Kim DJ, Park M, Cha SH, Kim JW, Han BG, Min H, Ahn Y, Park MS, Han HR, Jang HY, Cho EY, Lee JE, Cho NH, Shin C, Park T, Park JW, Lee JK, Cardon L, Clarke G, McCarthy MI, Lee JY, Oh B and Kim HL: A large-scale genome-wide association study of Asian populations uncovers genetic factors influencing eight quantitative traits. Nat Genet 41: 527-534, 2009.

- 10 Takeuchi F, Nabika T, Isono M, Katsuya T, Sugiyama T, Yamaguchi S, Kobayashi S, Yamori Y, Ogihara T and Kato N: Evaluation of genetic loci influencing adult height in the Japanese population. J Hum Genet 54: 749-752, 2009.
- 11 Ramakrishna M, Williams LH, Boyle SE, Bearfoot JL, Sridhar A, Speed TP, Gorringe KL and Campbell IG: Identification of candidate growth promoting genes in ovarian cancer through integrated copy number and expression analysis. PLoS One 5: e9983, 2010.
- 12 Slavin TP, Feng T, Schnell A, Zhu X and Elston RC: Two-marker association tests yield new disease associations for coronary artery disease and hypertension. Hum Genet 130: 725-733, 2011.
- 13 Comabella M, Craig DW, Morcillo-Suarez C, Rio J, Navarro A, Fernandez M, Martin R and Montalban X: Genome-wide scan of 500,000 single-nucleotide polymorphisms among responders and nonresponders to interferon beta therapy in multiple sclerosis. Arch Neurol 66: 972-978, 2009.
- 14 Dendrou CA, Plagnol V, Fung E, Yang JH, Downes K, Cooper JD, Nutland S, Coleman G, Himsworth M, Hardy M, Burren O, Healy B, Walker NM, Koch K, Ouwehand WH, Bradley JR, Wareham NJ, Todd JA and Wicker LS: Cell-specific protein phenotypes for the autoimmune locus *IL2RA* using a genotype-selectable human bioresource. Nat Genet 41: 1011-1015, 2009.
- 15 Gregory SG, Schmidt S, Seth P, Oksenberg JR, Hart J, Prokop A, Caillier SJ, Ban M, Goris A, Barcellos LF, Lincoln R, McCauley JL, Sawcer SJ, Compston DA, Dubois B, Hauser SL, Garcia-Blanco MA, Pericak-Vance MA and Haines JL: Interleukin 7 receptor alpha chain (*ILTR*) shows allelic and functional association with multiple sclerosis. Nat Genet 39: 1083-1091, 2007.
- 16 Hoffjan S and Akkad DA: The genetics of multiple sclerosis: an update 2010. Mol Cell Probes 24: 237-243, 2010.
- 17 Barbaux S, Gascoin-Lachambre G, Buffat C, Monnier P, Mondon F, Tonanny MB, Pinard A, Auer J, Bessieres B, Barlier A, Jacques S, Simeoni U, Dandolo L, Letourneur F, Jammes H and Vaiman D: A genome-wide approach reveals novel imprinted genes expressed in the human placenta. Epigenetics 7: 1079-1090, 2012.
- 18 Sabatino G, Rigante L, Minella D, Novelli G, Della Pepa GM, Esposito G, Albanese A, Maira G and Marchese E: Transcriptional profile characterization for the identification of peripheral blood biomarkers in patients with cerebral aneurysms. J Biol Regul Homeost Agents 27: 729-738, 2013.
- 19 Doi K, Fujimoto T, Okamura T, Ogawa M, Tanaka Y, Mototani Y, Goto M, Ota T, Matsuzaki H, Kuroki M, Tsunoda T, Sasazuki T and Shirasawa S: ZFAT plays critical roles in peripheral T-cell homeostasis and its T-cell receptor-mediated response. Biochem Biophys Res Commun 425: 107-112, 2012.
- 20 Ogawa M, Okamura T, Ishikura S, Doi K, Matsuzaki H, Tanaka Y, Ota T, Hayakawa K, Suzuki H, Tsunoda T, Sasazuki T and Shirasawa S: Zfat-deficiency results in a loss of CD3ζ phosphorylation with dysregulation of Erk and Egr activities leading to impaired positive selection. PLoS One 8: e76254, 2013.
- 21 Carpenter AC and Bosselut R: Decision checkpoints in the thymus. Nat Immunol 11: 666-673, 2010.
- 22 Rothenberg EV and Taghon T: Molecular genetics of T-cell development. Annu Rev Immunol 23: 601-649, 2005.
- 23 Hernandez-Hoyos G, Anderson MK, Wang C, Rothenberg EV and Alberola-Ila J: GATA-3 expression is controlled by TCR signals and regulates CD4/CD8 differentiation. Immunity 19: 83-94, 2003.

- 24 Wilkinson B, Chen JY, Han P, Rufner KM, Goularte OD and Kaye J: TOX: an HMG box protein implicated in the regulation of thymocyte selection. Nat Immunol 3: 272-280, 2002.
- 25 Bettini M, Xi H, Milbrandt J and Kersh GJ: Thymocyte development in early growth response gene 1-deficient mice. J Immunol 169: 1713-1720, 2002.
- 26 Maurice D, Hooper J, Lang G and Weston K: c-Myb regulates lineage choice in developing thymocytes via its target gene Gata3. EMBO J 26: 3629-3640, 2007.
- 27 Staton TL, Lazarevic V, Jones DC, Lanser AJ, Takagi T, Ishii S and Glimcher LH: Dampening of death pathways by schnurri-2 is essential for T-cell development. Nature 472: 105-109, 2011.
- 28 Fu G, Casas J, Rigaud S, Rybakin V, Lambolez F, Brzostek J, Hoerter JA, Paster W, Acuto O, Cheroutre H, Sauer K and Gascoigne NR: Themis sets the signal threshold for positive and negative selection in T-cell development. Nature 504: 441-445, 2013.
- 29 Lauritsen JP, Kurella S, Lee SY, Lefebvre JM, Rhodes M, Alberola-Ila J and Wiest DL: Egr2 is required for Bcl-2 induction during positive selection. J Immunol 181: 7778-7785, 2008.
- 30 Lawson VJ, Weston K and Maurice D: Early growth response 2 regulates the survival of thymocytes during positive selection. Eur J Immunol 40: 232-241, 2010.
- 31 Xi H and Kersh GJ: Sustained early growth response gene 3 expression inhibits the survival of CD4/CD8 double-positive thymocytes. J Immunol *173*: 340-348, 2004.
- 32 Swirnoff AH and Milbrandt J: DNA-binding specificity of NGFI-A and related zinc finger transcription factors. Mol Cell Biol 15: 2275-2287, 1995.
- 33 Sprent J and Surh CD: Normal T-cell homeostasis: the conversion of naive cells into memory-phenotype cells. Nat Immunol 12: 478-484, 2011.
- 34 Marrack P and Kappler J: Control of T-cell viability. Annu Rev Immunol 22: 765-787, 2004.
- 35 Takada K and Jameson SC: Naive T-cell homeostasis: from awareness of space to a sense of place. Nat Rev Immunol 9: 823-832, 2009.
- 36 Ouyang W and Li MO: Foxo: in command of T-lymphocyte homeostasis and tolerance. Trends Immunol 32: 26-33, 2011.
- 37 Khaled AR and Durum SK: Lymphocide: cytokines and the control of lymphoid homeostasis. Nat Rev Immunol 2: 817-830, 2002.
- 38 Surh CD and Sprent J: Homeostasis of naive and memory T-cells. Immunity 29: 848-862, 2008.
- 39 Kim HP, Imbert J and Leonard WJ: Both integrated and differential regulation of components of the IL-2/IL-2 receptor system. Cytokine Growth Factor Rev 17: 349-366, 2006.
- 40 Liao W, Lin JX, Wang L, Li P and Leonard WJ: Modulation of cytokine receptors by IL-2 broadly regulates differentiation into helper T_cell lineages. Nat Immunol 12: 551-559, 2011.
- 41 Malek TR and Bayer AL: Tolerance, not immunity, crucially depends on IL-2. Nat Rev Immunol 4: 665-674, 2004.
- 42 Mazzucchelli R and Durum SK: Interleukin-7 receptor expression: intelligent design. Nat Rev Immunol 7: 144-154, 2007.
- 43 Hafler DA, Compston A, Sawcer S, Lander ES, Daly MJ, De Jager PL, de Bakker PI, Gabriel SB, Mirel DB, Ivinson AJ, Pericak-Vance MA, Gregory SG, Rioux JD, McCauley JL, Haines JL, Barcellos LF, Cree B, Oksenberg JR and Hauser SL: Risk alleles for multiple sclerosis identified by a genomewide study. N Engl J Med 357: 851-862, 2007.

- 44 Veldhoen M and Seddon B: Empowering T-helper 17 cells in autoimmunity. Nat Med 16: 166-168, 2010.
- 45 Liu X, Leung S, Wang C, Tan Z, Wang J, Guo TB, Fang L, Zhao Y, Wan B, Qin X, Lu L, Li R, Pan H, Song M, Liu A, Hong J, Lu H and Zhang JZ: Crucial role of interleukin-7 in T-helper type 17 survival and expansion in autoimmune disease. Nat Med 16: 191-197, 2010.
- 46 Kerdiles YM, Beisner DR, Tinoco R, Dejean AS, Castrillon DH, DePinho RA and Hedrick SM: Foxo1 links homing and survival of naive T-cells by regulating L-selectin, Ccr7 and interleukin 7 receptor. Nat Immunol 10: 176-184, 2009.
- 47 Egawa T, Tillman RE, Naoe Y, Taniuchi I and Littman DR: The role of the Runx transcription factors in thymocyte differentiation and in homeostasis of naive T-cells. J Exp Med 204: 1945-1957, 2007.
- 48 Carlson CM, Endrizzi BT, Wu J, Ding X, Weinreich MA, Walsh ER, Wani MA, Lingrel JB, Hogquist KA and Jameson SC: Kruppel-like factor 2 regulates thymocyte and T-cell migration. Nature 442: 299-302, 2006.
- 49 Peng SL: Foxo in the immune system. Oncogene 27: 2337-2344, 2008.
- 50 van der Horst A and Burgering BM: Stressing the role of Foxo proteins in lifespan and disease. Nat Rev Mol Cell Biol 8: 440-450, 2007.
- 51 Wijchers PJ, Burbach JP and Smidt MP: In control of biology: of mice, men and Foxes. Biochem J 397: 233-246, 2006.
- 52 Accili D and Arden KC: Foxos at the crossroads of cellular metabolism, differentiation, and transformation. Cell 117: 421-426, 2004.

- 53 Dejean AS, Beisner DR, Ch'en IL, Kerdiles YM, Babour A, Arden KC, Castrillon DH, DePinho RA and Hedrick SM: Transcription factor Foxo3 controls the magnitude of T-cell immune responses by modulating the function of dendritic cells. Nat Immunol 10: 504-513, 2009.
- 54 Ouyang W, Beckett O, Flavell RA and Li MO: An essential role of the Forkhead-box transcription factor Foxo1 in control of Tcell homeostasis and tolerance. Immunity 30: 358-371, 2009.
- 55 Chistiakov DA, Voronova NV and Chistiakov PA: The crucial role of IL-2/IL-2RA-mediated immune regulation in the pathogenesis of type 1 diabetes, an evidence coming from genetic and animal model studies. Immunol Lett 118: 1-5, 2008.
- 56 Yamanouchi J, Rainbow D, Serra P, Howlett S, Hunter K, Garner VE, Gonzalez-Munoz A, Clark J, Veijola R, Cubbon R, Chen SL, Rosa R, Cumiskey AM, Serreze DV, Gregory S, Rogers J, Lyons PA, Healy B, Smink LJ, Todd JA, Peterson LB, Wicker LS and Santamaria P: Interleukin-2 gene variation impairs regulatory T-cell function and causes autoimmunity. Nat Genet 39: 329-337, 2007.

Received April 4, 2014 Revised June 19, 2014 Accepted June 20, 2014