

Angiogenesis-, Metastasis- and Signaling Pathway-related Factor Dynamics in Human Colon Cancer Cells Following Interaction with Monocytes

TERUKO HONDA¹, ISAMU YAMAMOTO¹ and HIROYUKI INAGAWA²

¹*Department of Medical Technology, School of Life and Environmental Science, Azabu University, Sagamihara, Kanagawa, Japan;*

²*Department of Integrated and Holistic Immunology, Faculty of Medicine, Kagawa University, Kida-gun, Kagawa, Japan*

Abstract. *Background: In tumors, monocytes differentiate into tumor-associated macrophages following interaction with cancer cells. We have previously reported that angiogenesis- and chemotaxis-related factors are associated with human monocyte differentiation following interaction with colon cancer cells. However, the exact nature of factors remains unknown. We investigated factors associated with differentiation of human colon cancer cells following interaction with monocytes. Materials and Methods: The human colon cancer cell line DLD-1 was co-cultured with the human monocyte cell line THP-1. mRNA expression was analyzed by quantitative real-time PCR. Results: Expression of interleukin-1 β , matrix metalloproteinase (MMP)-1, MMP-2, and MMP-3 increased in human colon cancer cells after co-culture with monocytes. Conversely, the expression of monocyte chemotactic protein-1, tumor necrosis factor- α , and signal transducer and activator of transcription-3 did not increase. Conclusion: Differentiation of human colon cancer cells following interaction with monocytes may be associated with angiogenesis and metastasis but not chemotaxis and signaling pathways. Thus, angiogenesis- and metastasis-related factors associated with differentiation of human colon cancer cells may constitute important targets for colon cancer therapy.*

Monocytes originate from the bone marrow. They develop and are released into the peripheral circulation. They then

enter and populate tissues as macrophages. Subpopulations of macrophages exist, and are defined by anatomical location and functional phenotype (1, 2).

Recently, it was established that most solid tumors were abundantly populated with macrophages, termed tumor-associated macrophages. Tumor-associated macrophages are considered to exert antitumor effects and promote malignant progression (3-11). Several reports describe an association between macrophage number and prognosis in various types of human cancers. Extensive tumor-associated macrophage infiltration is associated with poor prognosis in breast, cervical, and bladder cancer; conversely, their presence is associated with a good prognosis in stomach, lung, and colorectal cancer (4, 6, 7, 12). These discrepancies suggest that monocytes differentiate into tumor-associated macrophages following interaction with cancer cells and that their character differs depending on the cancer type. We previously reported that angiogenesis- and chemotaxis-related factors were associated with differentiation of human monocytes into tumor-associated macrophages following interaction with colon cancer cells (13). However, the nature of these factors remains unknown. We examined changes in mRNA expression of human colon cancer cells following interaction with monocytes to identify factors associated with their differentiation.

Materials and Methods

Cells. DLD-1 cells obtained from the Japan Health Sciences Foundation (Tokyo, Japan), and THP-1 cells obtained from DS Pharma Biomedical (Osaka, Japan) were cultured in a 5% CO₂ atmosphere at 37°C in RPMI-1640 medium (Wako Pure Chemical Industries, Ltd., Osaka, Japan) containing 10% fetal calf serum supplemented with 100 units/ml each of penicillin and streptomycin (Wako Pure Chemical Industries, Ltd.).

Cell co-culture. DLD-1 and THP-1 cells were co-cultured using a cell culture insert (Becton, Dickinson and Co., Franklin Lakes, NJ,

Correspondence to: Teruko Honda, Department of Medical Technology, School of Life and Environmental Science, Azabu University, 1-17-71 Fuchinobe, Chuo-ku, Sagamihara, Kanagawa 252-5201, Japan. Tel: +81 4275471111, Fax: +81 4275476111, e-mail: hondat@azabu-u.ac.jp

Key Words: Monocyte, colon cancer cell, co-culture, angiogenesis, metastasis, signaling pathway, DLD-1, THP-1 cells.

Table I. Primer sequences for real-time PCR.

Gene	Forward primer	Reverse primer
<i>MCP-1</i>	5'-GCAAGTGTCCCAAAGAAG-3'	5'-GGAGTGAGTGTTCAGTCT-3'
<i>VEGF-A</i>	5'-GCAGCTTGAGTTAAACGAA-3'	5'-CAGTCTTTCCTGGTGAGA-3'
<i>IL-1β</i>	5'-TTACAGTGGCAATGAGGAT-3'	5'-TAGTGGTGGTCGGAGATT-3'
<i>TNF-α</i>	5'-AGGGACCTCTCTCTAATCA-3'	5'-TTGCTACAACATGGGCTA-3'
<i>IL-8</i>	5'-CGCCAACACAGAAATTATTG-3'	5'-GGATACCACAGAGAATGAATT-3'
<i>MMP-1</i>	5'-CTGGCCACAACCTGCCAAATG-3'	5'-CTGTCCCTGAACAGCCCAGTACTTA-3'
<i>MMP-2</i>	5'-TTGACGGTAAGGACGGACT-3'	5'-CTTGCAGTACTCCCCATCG-3'
<i>MMP-3</i>	5'-TGATCCTGCTTTGTCCTTTG-3'	5'-TTCAAGCTTCCTGAGGGATT-3'
<i>MMP-7</i>	5'-GACATCATGATTGGCTTTGC-3'	5'-TCTCCTCCGAGACCTGTCC-3'
<i>MT1-MMP</i>	5'-TCGGCCCCAAAGCAGCAGCT-3'	5'-TTCATGGTGTCTGCATCAGC-3'
<i>NF-κB</i>	5'-ATGGCTTCTATGAGGCTGAG-3'	5'-GTGTTGTTGGTCTGGATGC-3'
<i>STAT3</i>	5'-GAGGACTGAGCATCGAGCA-3'	5'-CATGTGATCTGACACCCTGAA-3'

MCP-1: Monocyte chemotactic protein 1, *VEGF-A*: vascular endothelial growth factor A, *IL-1 β* : interleukin 1 β , *TNF- α* : tumor necrosis factor- α , *IL-8*: interleukin 8, *MMP-1*: matrix metalloproteinase 1, *MMP-2*: matrix metalloproteinase 2, *MMP-3*: matrix metalloproteinase 3, *MMP-7*: matrix metalloproteinase 7, *MT1-MMP*: membrane type 1-matrix metalloproteinase, *NF- κ B*: nuclear factor- κ B, *STAT3*: signal transducer and activator of transcription-3.

USA) with a 0.4- μ m porous membrane to separate the upper and lower chambers. DLD-1 cells were cultured in the upper chamber at 2×10^5 cells/ml, and THP-1 cells were cultured in the lower chamber at 2×10^5 cells/ml. DLD-1 and THP-1 cells were collected 0, 1, 3, and 5 days after co-culture.

RNA extraction. Total RNA extractions from DLD-1 and THP-1 cells were performed using TRIzol Reagent (Invitrogen Corporation, Carlsbad, CA, USA) according to the manufacturer's protocol. RNA was quantified by absorbance at 260 nm. cDNA was synthesized using reverse transcriptase with Oligo(dT)20 (Toyobo Co., Ltd., Osaka, Japan).

Quantitative real-time PCR. mRNA expression was analyzed by quantitative real-time PCR (Model MiniOpticon; Bio-Rad Laboratories, Inc., Hercules, CA, USA) using SsoFast EvaGreen Supermix (Bio-Rad Laboratories, Inc.). After initial heat denaturation at 95°C for 3 min, PCR conditions were set at 95°C for 10 s and 60°C for 30 s for 40 cycles. The primers used are shown in Table I. Relative quantifications were achieved by normalization to the value of the housekeeping gene β -actin. Data are expressed as change (n-fold) in mRNA expression compared with co-culture at day 0.

Results

mRNA expression of angiogenesis- and metastasis-related factors in human colon cancer cells is increased following co-culture with monocytes. mRNA expression in DLD-1 cells after co-culture with THP-1 cells was analyzed by quantitative real-time PCR to identify factors associated with human colon cancer cell differentiation. Factors associated with human monocyte differentiation are known to function in tumor progression (14-19). We previously reported that mRNA expression of angiogenesis-related factors such as vascular endothelial growth factor (*VEGF*)-A and interleukin (*IL*)-8 increased and that of signaling

Table II. mRNA expression in DLD-1 cells following co-culture with THP-1 cells.

Gene	Co-culture day			
	0	1	3	5
<i>MCP-1</i>	1	0.6	0.7	0.6
<i>VEGF-A</i>	1	0.7	6.9	11.8
<i>IL-1β</i>	1	1.2	24.8	59.1
<i>TNF-α</i>	1	2.1	1.7	1.9
<i>IL-8</i>	1	0.6	4.5	1.0
<i>MMP-1</i>	1	3.6	11.5	11.7
<i>MMP-2</i>	1	6.9	4.6	17.7
<i>MMP-3</i>	1	5.6	18.6	22.1
<i>MMP-7</i>	1	4.7	4.0	3.9
<i>MT1-MMP</i>	1	3.0	2.8	4.3
<i>NF-κB</i>	1	1.1	1.8	2.2
<i>STAT3</i>	1	0.3	0.5	0.9

MCP-1: Monocyte chemotactic protein 1, *VEGF-A*: vascular endothelial growth factor A, *IL-1 β* : interleukin 1 β , *TNF- α* : tumor necrosis factor- α , *IL-8*: interleukin 8, *MMP-1*: matrix metalloproteinase 1, *MMP-2*: matrix metalloproteinase 2, *MMP-3*: matrix metalloproteinase 3, *MMP-7*: matrix metalloproteinase 7, *MT1-MMP*: membrane type 1-matrix metalloproteinase, *NF- κ B*: nuclear factor- κ B, *STAT3*: signal transducer and activator of transcription-3.

pathway-related factors such as nuclear factor (*NF*)- κ B did not increase in DLD-1 cells after co-culture with THP-1 cells (Table II) (13). In addition to identifying increased expression of chemotaxis-, angiogenesis-, metastasis-, and signaling pathway-related factors, we quantitatively evaluated mRNA expression in DLD-1 cells co-cultured with THP-1 cells. *IL-1 β* mRNA expression increased 24.8-fold by day 3 and 59.1-fold by day 5; matrix

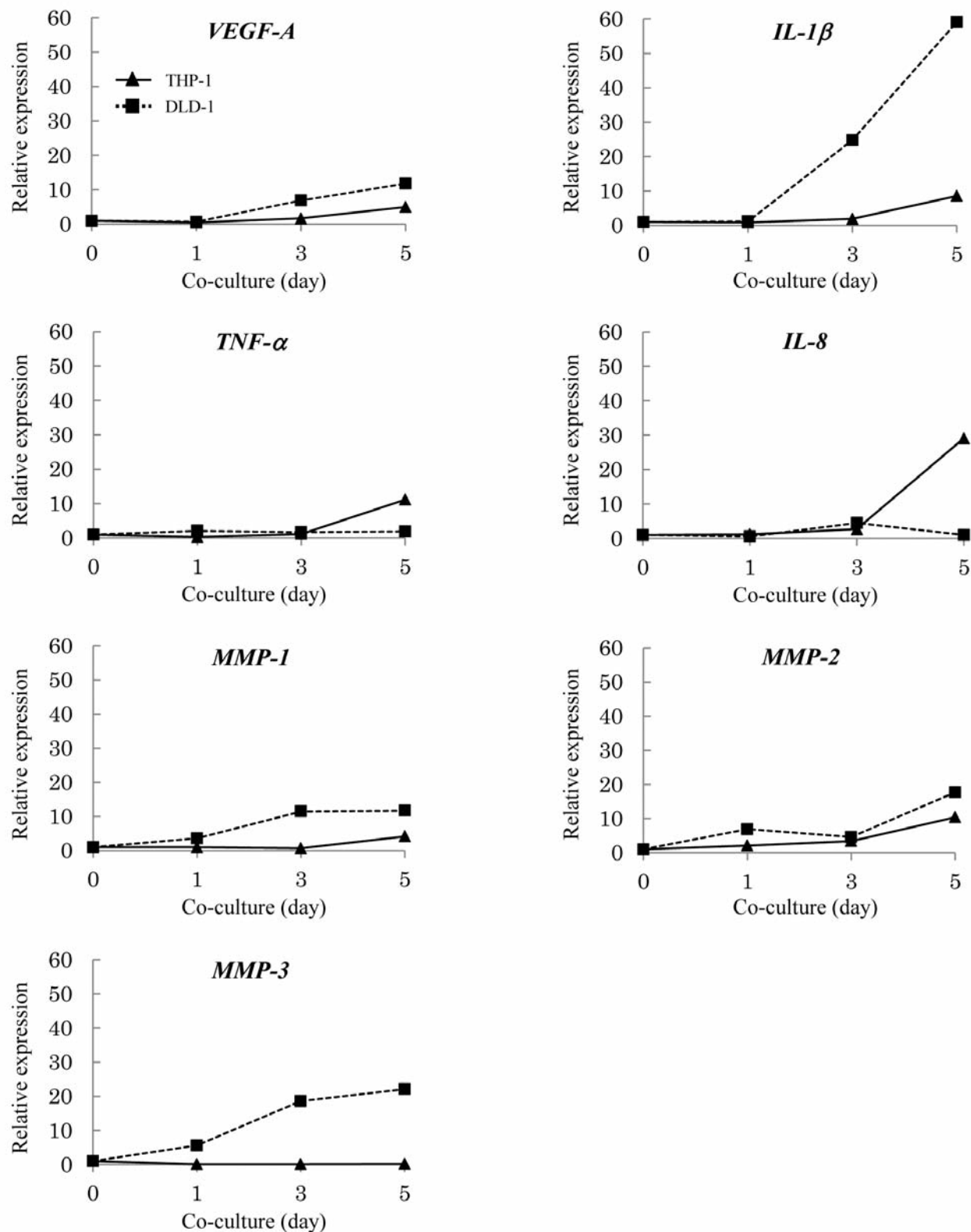


Figure 1. Differential expression of angiogenesis- and metastasis-related factors in DLD-1 and THP-1 cells following co-culture. mRNA expression after co-culture was analyzed by quantitative real-time PCR. Relative quantifications were achieved by normalization to the value of the housekeeping gene β -actin. Data are expressed as change (n-fold) in mRNA expression compared with co-culture at day 0.

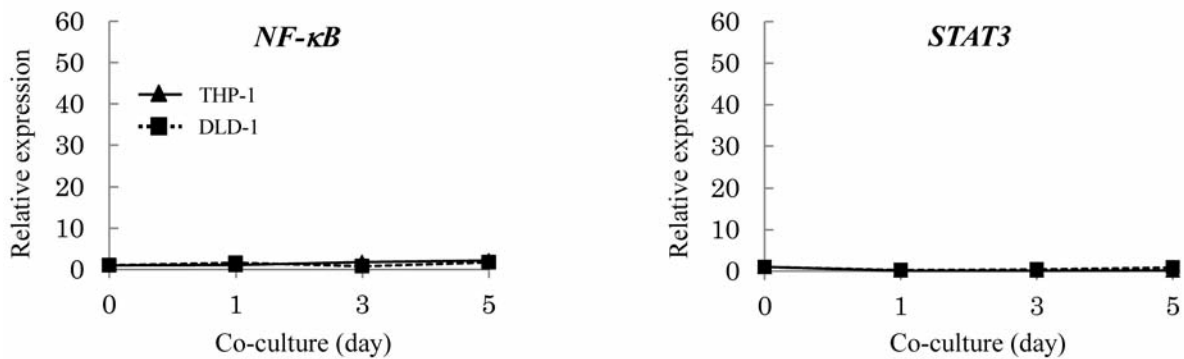


Figure 2. mRNA expression of nuclear factor (NF)- κ B and signal transducer and activator of transcription-3 (STAT3) in DLD-1 and THP-1 cells, following co-culture. mRNA expression after co-culture was analyzed by quantitative real-time PCR. Relative quantifications were achieved by normalization to the value of the housekeeping gene β -actin. Data are expressed as change (n-fold) in mRNA expression compared with co-culture at day 0.

metalloproteinase (MMP)-1 mRNA expression increased 11.5-fold by day 3 and 11.7-fold by day 5; MMP-2 mRNA expression increased 17.7-fold by day 5; and MMP-3 mRNA expression increased 18.6-fold by day 3 and 22.1-fold by day 5. This indicates that mRNA expression of *IL-1 β* , *MMP-1*, *MMP-2*, and *MMP-3* in DLD-1 cells increased after co-culture. Furthermore, the mRNA expression of *IL-1 β* , *MMP-1*, and *MMP-3* in DLD-1 cells increased in a time-dependent manner after co-culture. mRNA expression of *MMP-7* and membrane type 1 (*MT1*)-MMP also demonstrated an increasing trend in DLD-1 cells co-cultured with THP-1 cells (Table II). Conversely, mRNA expression of monocyte chemotactic protein (*MCP-1*), tumor necrosis factor (*TNF*)- α , and signal transducer and activator of transcription-3 (*STAT3*) did not increase until day 5 in DLD-1 cells co-cultured with THP-1 cells (Table II). Therefore, it is clear that mRNA expression in human colon cancer cells changed following interaction with monocytes. Our results suggest that human colon cancer cell differentiation following interaction with monocytes is associated with angiogenesis and metastasis but not chemotaxis and signaling pathways.

mRNA expression pattern of chemotaxis-, angiogenesis-, and metastasis-related factors in human colon cancer cells is different from that in monocytes following co-culture. We examined the mRNA expression patterns of factors associated with cell differentiation in DLD-1 and THP-1 cells after co-culture. In DLD-1 cells, mRNA expression of *VEGF-A*, *IL-1 β* , *MMP-1*, *MMP-2*, and *MMP-3* increased by day 5 after co-culture, whereas that of *TNF- α* and *IL-8* did not increase until day 5 after co-culture. Conversely, in THP-1 cells, mRNA expression of *VEGF-A*, *IL-1 β* , *TNF- α* , *IL-8*, and *MMP-2* increased by day 5 after co-culture, whereas that of *MMP-3* did not increase

until day 5 after co-culture. mRNA expression of *VEGF-A*, *IL-1 β* , *MMP-1*, *MMP-2*, and *MMP-3* increased more in DLD-1 cells after co-culture than in THP-1 cells (Figure 1). It is established that *VEGF-A*, *IL-1 β* , *TNF- α* , and *IL-8* are angiogenesis-related factors and that MMPs are metastasis-related factors. Our results indicate that the alterations in angiogenesis- and metastasis-related factors in human colon cancer cells after co-culture differ from those altered in monocytes. Furthermore, the mRNA expression patterns of angiogenesis- and metastasis-related factors in human colon cancer cells after co-culture differ from those in monocytes. Therefore, our results suggest that the mechanism of human colon cancer cell differentiation following interaction differs from that of monocyte differentiation.

mRNA expression of signaling pathway-related factors is not increased in human colon cancer cells and monocytes following co-culture. We examined the mRNA expression of signaling pathway-related factors in DLD-1 and THP-1 cells to understand their mechanisms of differentiation following interaction. *NF- κ B* and *STAT3* mRNA expressions in DLD-1 and THP-1 cells did not increase up to day 5 after co-culture (Figure 2). This suggests that the mRNA expression of signaling pathway-related factors does not increase in human colon cancer cells and monocytes following interaction.

Reportedly, *NF- κ B* and *STAT3* are expressed in many solid tumors and are activated by inflammatory cytokines, such as *TNF- α* . Therefore, we examined *TNF- α* mRNA expression in DLD-1 cells after co-culture. *TNF- α* mRNA expression in DLD-1 cells did not increase until day 5 after co-culture (Figure 1). This suggests that the mechanism of cancer cell differentiation following interaction with monocytes differs depending on cancer type.

Discussion

We showed that mRNA expression of *VEGF-A*, *IL-1 β* , *MMP-1*, *MMP-2*, and *MMP-3* increased in human colon cancer cells following co-culture with monocytes. Moreover, we demonstrated that mRNA expression of *VEGF-A*, *IL-1 β* , *TNF- α* , *IL-8*, and *MMP-2* increased in human monocytes following co-culture with colon cancer cells. These results indicate that mRNA expression changes following co-culture in both human monocytes and colon cancer cells. Furthermore, our results imply that human colon cancer cell differentiation following interaction is associated with angiogenesis and metastasis but not chemotaxis and signaling pathways. Therefore, human colon cancer cells may promote angiogenesis and metastasis in tumor tissues following interaction with monocytes. Thus, factors associated with human colon cancer cell differentiation may constitute important targets for colon cancer therapy.

Our results also indicate that the alterations in angiogenesis- and metastasis-related factors in human colon cancer cells following interaction differ from those altered in monocytes. In addition, we showed that the mRNA expression patterns of angiogenesis- and metastasis-related factors in human colon cancer cells following interaction differ from those in monocytes. Therefore, the mechanism of differentiation of human colon cancer cells following interaction may differ from that of monocytes. Thus, our results suggest that a synergistic effect, resulting from the interaction between human colon cancer cells and monocytes, promotes the progression of tumors to malignancy.

mRNA expression of *MMP-1*, *MMP-2*, and *MMP-3* in human colon cancer cells and that of *MMP-2* in monocytes increases following interaction. It is established that MMPs are associated with the invasive properties of cancer cells (19). Therefore, it is possible that inhibition of *MMP* expression in human colon cancer cells and monocytes correlates with suppression of tumor infiltration and metastasis. Thus, inhibitors of *MMP* expression may become targets for colon cancer therapy.

It is known that activated NF- κ B and STAT3 are required for the development of colitis-associated cancer. Indeed, NF- κ B and STAT3 overexpression at tumor sites may be involved in the differentiation of monocytes into tumor-associated macrophages (20-26). NF- κ B and STAT3 are known to be activated by inflammatory cytokines, such as TNF- α . However, this study indicated that NF- κ B and STAT3 mRNA expression in DLD-1 and THP-1 cells did not increase until day 5 after co-culture. We also showed that TNF- α mRNA expression did not increase in DLD-1 cells until day 5 after co-culture. Extensive tumor-associated macrophage infiltration reportedly correlates with poor prognosis in breast, cervical, and bladder cancer, whereas the presence of tumor-associated macrophages reportedly

correlates with a good prognosis in stomach, lung, and colorectal cancer (4, 6, 7, 12). Thus, our results suggest that the mechanism of cancer cell differentiation following interaction with monocytes differs depending on cancer type.

References

- Gordon S and Taylor PR: Monocyte and macrophage heterogeneity. *Nat Rev Immunol* 5: 953-964, 2005.
- Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M and Ley K: Development of monocytes, macrophages, and dendritic cells. *Science* 327: 656-661, 2010.
- Burke B and Lewis CE: *The Macrophage*. Oxford University Press, Oxford, 2002.
- Bingle L, Brown NJ and Lewis CE: The role of tumour-associated macrophages in tumour progression: Implications for new anticancer therapies. *J Pathol* 196: 254-265, 2002.
- Mantovani A, Schioppa T, Biswas SK, Marchesi F, Allavena P and Sica A: Tumor-associated macrophages and dendritic cells as prototypic type II polarized myeloid populations. *Tumori* 89: 459-468, 2003.
- Pollard JW: Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* 4: 71-78, 2004.
- Lewis CE and Pollard JW: Distinct role of macrophages in different tumor microenvironments. *Cancer Res* 15: 605-612, 2006.
- Gabrilovich DI and Nagaraj S: Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 9: 162-174, 2009.
- Qian BZ and Pollard JW: Macrophage diversity enhances tumor progression and metastasis. *Cell* 141: 39-51, 2010.
- Condeelis J and Pollard JW: Macrophages: Obligate partners for tumor cell migration, invasion, and metastasis. *Cell* 124: 263-266, 2006.
- Murray PJ and Wynn TA: Protective and pathogenic functions of macrophage subsets. *Nat Rev Immunol* 11: 723-737, 2011.
- Heusinkveld M and van der Burg SH: Identification and manipulation of tumor associated macrophages in human cancers. *J Transl Med* 9: 216-228, 2011.
- Honda T, Inagawa H and Yamamoto I: Differential expression of mRNA in human monocytes following interaction with human colon cancer cells. *Anticancer Res* 31: 2493-2498, 2011.
- Ferrara N and Davis-Smyth T: The biology of vascular endothelial growth factor. *Endocr Rev* 18: 4-25, 1997.
- Kwon HC, Kim SH, Oh SY, Lee S, Kwon KA, Choi HJ, Park KJ, Kim HJ and Roh MS: Clinicopathological significance of p53, hypoxia-inducible factor 1 α , and vascular endothelial growth factor expression in colorectal cancer. *Anticancer Res* 30: 4163-4168, 2010.
- Claesson-Welsh L and Welsh M: VEGFA and tumour angiogenesis. *J Intern Med* 273: 114-127, 2013.
- Apte RN, Dotan S, Elkabets M, White MR, Reich E, Carmi Y, Song X, Dvorkin T, Krelin Y and Voronov E: The involvement of IL-1 in tumorigenesis, tumor invasiveness, metastasis and tumor-host interactions. *Cancer Metastasis Rev* 25: 387-408, 2006.
- Carmi Y, Voronov E, Dotan S, Lahat N, Rahat MA, Fogel M, Huszar M, White MR, Dinarello CA and Apte RN: The role of macrophage-derived IL-1 in induction and maintenance of angiogenesis. *J Immunol* 183: 4705-4014, 2009.

- 19 Folgueras AR, Pendás AM, Sánchez LM and López-Otín C: Matrix metalloproteinases in cancer: From new functions to improved inhibition strategies. *Int J Dev Biol* 48: 411-424, 2004.
- 20 Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF and Karin M: IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* 118: 285-296, 2004.
- 21 Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, Gutmovich-Pyest E, Urieli-Shoval S, Galun E and Ben-Neriah Y: NF-kappaB functions as a tumour promoter in inflammation-associated cancer. *Nature* 431: 461-466, 2004.
- 22 Yu H, Kortylewski M and Pardoll D: Crosstalk between cancer and immune cells: Role of STAT3 in the tumour microenvironment. *Nat Rev Immunol* 7: 41-51, 2007.
- 23 Hagemann T, Lawrence T, McNeish I, Charles KA, Kulbe H, Thompson RG, Robinson SC and Balkwill FR: "Re-educating" tumor-associated macrophages by targeting NF-kappaB. *J Exp Med* 205: 1261-1268, 2008.
- 24 Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, Scheller J, Rose-John S, Cheroutre H, Eckmann L and Karin M: IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell* 15: 103-113, 2009.
- 25 Lee H, Deng J, Kujawski M, Yang C, Liu Y, Herrmann A, Kortylewski M, Horne D, Somlo G, Forman S, Jove R and Yu H: STAT3-induced S1PR1 expression is crucial for persistent STAT3 activation in tumors. *Nat Med* 16: 1421-1428, 2010.
- 26 Qian WF, Guan WX, Gao Y, Tan JF, Qiao ZM, Huang H, Xia CL: Inhibition of STAT3 by RNA interference suppresses angiogenesis in colorectal carcinoma. *Braz J Med Biol Res* 44: 1222-1230, 2011.

Received April 4, 2013

Revised June 3, 2013

Accepted June 4, 2013