

Review

The Potential of RECK Inducers as Antitumor Agents for Glioma

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Abstract. *Reversion-inducing cysteine-rich protein with Kazal motifs (RECK), a tumor and metastasis suppressor gene, is critical for the regulation of the invasive and metastatic activities of tumor cells. RECK is down-regulated in some malignancies and its expression is positively correlated with survival of patients with cancer. Patients with malignant glioma have poor prognosis. Since RECK expression decreases as the tumor stage progresses from less invasive grade II glioma to invasive glioblastoma multiforme, up-regulation of RECK by natural or synthetic agents might be a valuable therapeutic option for glioma. Histone deacetylase inhibitors and non-steroidal anti-inflammatory drugs have been widely used clinically and demonstrated to increase RECK expression in cancer cells, thus they might be used as RECK inducers. In this article, the functions of RECK and the role of RECK in glioma are reviewed, with emphasis on the potential application of RECK inducers in the treatment of glioma.*

Malignant glioma is the most common primary brain tumor, consisting of grade 3 anaplastic astrocytoma and grade 4 glioblastoma multiforme (GBM) (1). It is histologically heterogeneous and invasive, and patients with malignant glioma have poor prognosis, with the survival depending on the histological grade of the tumor (1-4). The median survival time of patients with GBM is 12-15 months and that

of patients with anaplastic astrocytoma 2-5 years (2-4). The therapies for malignant glioma, including surgery, radiotherapy, and chemotherapy, have not been successful (1, 5). Malignant gliomas cannot be resected completely because of their infiltrative nature, although surgical debulking can reduce the mass effect and provide tissues for diagnosis (4). Postoperative radiotherapy can increase the survival of the patients; however, the majority of the patients have local tumor recurrence (4). Brachytherapy and stereotactic radiosurgery have been used to treat glioma, but there is no clear evidence that they can improve the survival of patients (6, 7). In recent years, concomitant temozolomide and radiotherapy, and biodegradable polymers containing carmustine (Gliadel Wafers; MGI Pharma, Inc., Bloomington, MN, USA) have been demonstrated to improve the survival of patients with glioma, and most patients are able to tolerate these treatments because of their limited systemic toxicities (2, 4, 8, 9). However, the prognosis of patients with malignant glioma is still poor, thus, it is mandatory to develop more effective treatment strategies for these tumor types.

Reversion-inducing cysteine-rich protein with Kazal motifs (RECK). RECK gene, encoding a glycosylphosphatidylinositol-anchored glycoprotein of about 110 kDa with multiple serine protease inhibitor-like motifs, is considered a tumor and metastasis suppressor gene (10). RECK was first identified as a cDNA clone inducing morphological reversion in NIH3T3 cells transformed by the v-K-ras oncogene (11). RECK is expressed ubiquitously in normal human tissues, and is essential for normal tissue development, morphogenesis, and remodeling; stabilization of tissue architecture; cell migration; and dynamic cell-cell interaction (10, 12). In addition, RECK expression plays a role in processes such as angiogenesis, chondrogenesis, and myogenesis (13-16). RECK-deficient mice exhibit reduced extracellular matrix

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(ECM) integrity, such as reduced type I collagen, disrupted basement membranes, cellular disarray, increased tissue fragility, and halted vascular development (10, 16). Furthermore, because of the role of RECK in the regulation of various ECM proteins, altered expression of RECK has been found to be involved in various human disorders, such as rheumatoid arthritis and asthma (17, 18). In contrast to normal tissues, RECK is undetectable in some tumor-derived cell lines and down-regulated in some malignancies such as pancreatic, breast, lung, colorectal, prostate, and gastric cancer, cholangiocarcinoma, ameloblastic tumor, middle ear squamous cell cancer, and osteosarcoma (10, 19-29). RECK expression is also positively correlated with the survival of patients with prostate, lung, pancreatic, breast, stomach, colorectal, hepatocellular cancer, cholangiocarcinoma, neuroblastoma, and osteosarcoma (10, 22, 23, 25, 26, 28-34). Restoration of expression of RECK in tumor cells suppresses tumor angiogenesis, invasion, and metastasis in animal models, and its residual expression level in tumor tissues often correlates with better prognosis (10, 11, 16). All these data suggest that RECK has tumor-suppressing effects.

The antitumor effects of RECK have been associated with its inhibitory effects on matrix metalloproteinases (MMPs) (35). The family of MMPs consists of multiple human zinc-dependent endopeptidases that can degrade cell proteinaceous components of the ECM (36, 37). The ECM is important for creating the cellular environment required during development and morphogenesis; MMPs cleave ECM components, such as collagen, laminin, and fibronectin, as well as non-matrix components, such as growth factors and cell surface receptors (36, 38). In malignant tumors, the remodeling of basement membrane and degradation of ECM are critical steps in tumor development, invasion and metastasis (36). The cancer cells and/or adjacent stromal cells secrete MMPs to degrade the ECM, and facilitate tumor invasion and progression (39, 40). RECK negatively regulates MMP family members, including MMP-2, MMP-9, and membrane type-1 MMP (10). Down-regulation of *RECK* gene expression is strongly associated with high expression of MMP-2 and MMP-9 in various types of cancers (10, 23, 39, 41). RECK inhibits MMP activity through direct suppression of its protease activity, regulation of cellular release, as well as possible sequestration at the cell surface (42). In addition, RECK has also been found to regulate MMP-9, not only post-transcriptionally, but also at the gene expression level (43). Generally, the expressions of RECK and MMPs are inversely correlated (44).

RECK has also been found to be a suppressor of tumor angiogenesis. Angiogenesis is a process by which new blood vessels are formed from the existing vasculature and it plays critical role in tumor growth and metastasis (45, 46). Angiogenesis is stimulated by cytokines and growth factors, such as vascular endothelial growth factor and

thrombospondin (38, 47, 48). These factors act on specific receptors on the endothelial cells, causing endothelial cell proliferation and production of proteolytic enzymes to destroy the matrix, and activate endothelial cell migration and invasion into tissues (49). As stated earlier, RECK affects the expression and activity of MMPs, which are involved in angiogenesis in malignant tumors (38, 50). In addition, angiogenesis in several kinds of cancer cell is suppressed by restored expression of RECK (33). Furthermore, tissue inhibitors of metalloproteinases (TIMPs) inhibit endothelial cell migration through increased RECK expression (51). These data indicate that RECK plays an important role in tumor angiogenesis.

Regulation of RECK. The regulation of RECK is interesting. The *RECK* gene is a common negative target for oncogenic signals that act on the specificity protein 1 (SP1)-binding site of the *RECK* promoter (12). *RECK* is down-regulated upon cell transformation by trio related transforming gene in ATL tumor cells (TGAT), human epidermal growth factor receptor 2 (HER-2/neu), and rat sarcoma (RAS) oncoproteins (52-55). TGAT oncoprotein inhibits RECK through 15 amino acids on its C-terminal (55). HER-2/neu induces the binding of SP proteins and histone deacetylase 1 to the *RECK* promoter to repress RECK, and activates the extracellular signal-regulated kinase (ERK) signaling pathway (54). RAS inhibits RECK expression *via* histone deacetylation and promoter methylation mechanisms, and acts through inhibition of the SP1 promoter site of the *RECK* gene (52, 53, 56). Activation of RAS signaling, including the rapidly growing fibrosarcoma (RAF)/mitogen-activated protein kinase kinase (MEK)/ERK and MEK kinase (MEKK)/MEK/c-jun-N-terminal kinase (JNK) pathways, can then up-regulate microRNA-21 and suppress RECK expression in tumor cells (52, 54, 57).

Although the main functions of RECK are considered to occur through the inhibition of MMPs as described above, there are reports revealing that RECK might act *via* some mechanisms other than through MMPs (50, 52, 54, 55). The relationship between RECK and neurogenic locus notch homolog protein (NOTCH) signaling has been noted in neural precursor cells, with RECK specifically targeting NOTCH signaling (15). In addition, the upstream regulator of RECK, RAS, can increase the level of NOTCH-1 (58). The *NOTCH* genes encode heterodimeric transmembrane receptors that can be activated by interacting with a family of its ligands (59). They are important in a variety of cellular processes, including proliferation, differentiation, survival, and apoptosis (60). Upon activation, NOTCH is cleaved, releasing intracellular NOTCH (ICN), which translocates into the nucleus (60). ICN associates with transcriptional factors to regulate the expression of target genes; thus, it plays an important role in development and cell growth (60). As for cancer, NOTCH signaling is linked to epigenetic

silencing and cell-cycle control during tumorigenesis in *Drosophila* (61), and dysregulated expression of NOTCH1 has been noted in some tumors, including glioma, lung, colon and pancreatic cancer, and hematopoietic malignancies (60, 62-66). Furthermore, down-regulation of NOTCH-1 reduces pancreatic cell invasion, whereas NOTCH-1 overexpression leads to increased tumor cell invasion (60). These information suggests that RECK might function through NOTCH signaling in cancer cells (67).

There is another family of metalloproteinases, the disintegrins and metalloproteinases or adamalysins (ADAMs), which contain extracellular disintegrin, metalloproteinase, cysteine-rich, epidermal growth factor-like domains, and transmembrane and cytoplasmic regions (37). Some ADAMs (ADAM-10, 28 and 33) have been found to be present in the nervous system and exert effects on neuronal migration (15, 37), and RECK directly interacts with ADAM-10 in neural precursor cells (15, 37). Because ADAMs are considered to be related to cancer progression, the relationship between ADAMs and RECK in cancer cells is interesting and deserves further investigation (37). In addition, transforming growth factor- β (TGF- β) has been implicated in many aspects of cancer progression, including proliferation, infiltrative growth, angiogenesis, and immune suppression (68). TGF- β can stimulate tumor invasion by regulating the activity of MMP, which is also closely related to the function of RECK (69). Moreover, TGF- β signaling in activated pancreatic stellate cells promotes ECM accumulation *via* preservation of the protease-inhibitory activity of RECK (70). Thus RECK might interact with TGF- β and affect the invasiveness and tumor growth of cancer cells.

As a whole, the information about the regulation of RECK is limited and the pathways mentioned above might represent only part of the regulatory mechanisms. More studies are necessary to understand the regulatory mechanisms of RECK.

Role of RECK in glioma. One important characteristic of malignant glioma is invasiveness, which involves tumor cell-ECM interactions and the activities of MMPs (40). No expression of MMP-2 and MMP-9 is present in the normal brain tissue, whereas in glioma, positive staining for these MMPs is significantly elevated, progressively increasing with the degree of malignancy (71). On the other hand, RECK negatively regulates MMP-2, MMP-9, and membrane type-1 MMP (16, 44). RECK expression decreases as the tumor stage progresses from less invasive grade II to invasive GBM (1, 35). Thus, MMPs, especially MMP-2 and MMP-9, are closely related to glioma invasiveness, and RECK, as a potent inhibitor of MMP-2 and MMP-9, is involved in the suppression of the invasiveness of glioma cells (44). Cell migration requires actin polymerization for the formation of motility-associated processes, such as lamellipodia (35). Lamellipodia associated with MMPs mediates proteolysis of

ECM constituents, including fibronectin, laminins, and collagens, in tumor cells and transformed cells (72). In glioma cells, RECK has been found to reduce cell motility and invasion through the regulation of actin cytoskeleton rearrangements and stabilization of focal adhesion (35). Overexpression of RECK inhibits the invasive process through rearrangement of actin filaments, promoting a decrease in migratory ability (35). In addition, RECK expression is increased by 1-50 mM valproic acid (di-n-propylacetic acid, VPA), a mood stabilizer and an antiepileptic drug, in T98G glioma cells in a concentration-dependent manner (73). VPA induces cytotoxicity, apoptosis, suppression of invasiveness and MMP-2 activity in T98G cells, while *RECK* siRNA markedly reverses these effects (73). These data suggest that RECK expression and activity of MMPs play a role in VPA-induced cytotoxicity, apoptosis, and suppression of invasiveness in T98G cells (73).

MicroRNAs (miRs) are small RNAs with 19-23 nucleotides in length, which are found in all mammalian cells. The miRs are incorporated into the RNA-induced silencing complex and target the 3'-untranslated region of specific mRNAs by a seed sequence that is located near the 5' region of the miRNA. The results of miR binding are that the mRNA is silenced or degraded, resulting in reduced expression level of the protein encoded by the targeted mRNA (74). miR-21 has been found to regulate genes involved in various pathways, such as cell death, cell proliferation, stress response and metabolism (1). Elevated miR-21 expression is causally linked to proliferation, apoptosis, and migration of several cancer cell lines, and has been found in various cancer types, including breast, lung, colon, prostate, pancreas, ovary and stomach, and in chronic lymphatic leukemia and glioblastoma (1, 75, 76). In glioma, with progression from less invasive grade II glioma to invasive GBM, the *RECK* and *TIMP* levels decrease, whereas miR-21 expression increases (1). miR-21 is associated with glioma cell survival, migration, and invasiveness, and specific inhibition of miR-21 with antisense oligonucleotides leads to decreased migratory and invasion abilities, elevated RECK and reduced MMP activity in glioma cells (1). U87MG glioma cells transfected with either anti-miR-21 or control oligonucleotide were implanted into nude mice subcutaneously, and the tumors produced from miR-21-inhibited cells demonstrated significantly lower MMP activity than did control tumors (1). These data indicate that miR-21 contributes to glioma malignancy by the down-regulation of MMP inhibitors such as RECK, which leads to activation of MMPs, thus promoting invasiveness of glioma cells (1).

As a whole, the pathway involving miR-21, RECK and MMPs seems to play an important role in the invasiveness and tumor grade of gliomas. Treatment targeting this pathway might provide beneficial effects to patients with malignant glioma.

RECK inducers. From the above, it is seen that RECK can suppress the activity of MMPs, cause apoptosis, and inhibit migration, invasiveness and angiogenesis of cancer cells, including glioma cells. Thus, strategies inducing RECK expression might be used as antitumor therapy for glioma. From the literature, two categories of pharmaceutical agents induce RECK expression: histone deacetylase (HDAC) inhibitors and nonsteroidal anti-inflammatory drugs (NSAIDs).

A. HDAC inhibitors: HDAC inhibitors have been found to exert anti-metastatic and anti-angiogenic activities *in vitro* and *in vivo* (77). VPA inhibits the proliferation of and induces the differentiation of cells in different malignancies including leukemia, lymphoma, teratocarcinoma, neuroblastoma, medulloblastoma, and atypical teratoid/rhabdoid tumor; clinically, it has been used to treat leukemia and some types of solid tumors (78-83). VPA can also suppress glioma cell growth and proliferation and is able to cross the blood-brain barrier; thus it is considered a possible treatment agent for malignant glioma (78, 81). In the literature, there is a trend for longer survival of patients with glioblastomas and seizure who are treated with VPA as compared to patients not treated with VPA (84). Furthermore, patients with GBM receiving VPA gain more benefit from temozolomide/radiotherapy than patients receiving an enzyme-inducing antiepileptic drug or patients not receiving any antiepileptic drug (85). In pediatric patients with high grade glioma, or diffuse pontine glioma, receiving VPA as oral maintenance treatment after fractionated focal radiation and chemotherapy, the VPA treatment was found to be safe and have moderate antitumor efficacy (86).

The mechanisms underlying the antitumor effects of VPA are variable. VPA can increase the DNA-binding of activating protein-1 transcription factor, inhibit glycogen synthase kinase-3 β , down-regulate protein kinase C activity, activate peroxisome proliferator-activated receptors, etc. (87, 88). In addition, VPA can inhibit histone deacetylase, which increases histone acetylation to modulate epigenomic and DNA methylation (88-90). VPA is a RECK inducer, and can cause cytotoxicity, apoptosis, and suppression of invasiveness of glioma cells. The induction of RECK expression might also contribute to the mild antitumor effects on gliomas by VPA noted clinically. Because VPA is often used to treat seizure in patients with glioma, and as it is a RECK-inducing agent, it might be a promising adjuvant therapeutic agent for treatment of glioma.

In addition to VPA, trichostatin A (TSA) and phenylbutyrate are also HDAC inhibitors (52, 73, 77, 91). Inhibition of HDAC by TSA has been shown to prevent tumorigenesis and metastasis (89). Treatment with 100 nM TSA for 48 h reduced MMP-2 mRNA and activity, and potently antagonized the inhibitory action of RAS on RECK in NIH3T3 cells (52, 89). In addition, treatment with 100 nM TSA for 48 h up-regulated *RECK* gene expression *via*

transcriptional activation, and suppressed MMP-2 activity and invasiveness of CL-1 human lung cancer cells (52, 77, 91). Phenylbutyrate, a carboxylic acid HDAC inhibitor, inhibited the anaplastic thyroid cancer cell line ARO from penetrating a matrigel-coated transwell, with concomitant suppression of MMP-7 and stimulation of RECK, without affecting MMP-2 expression (73). These two agents might also be potential anticancer agents for glioma, however, more studies are needed to determine their effects and toxicities.

B. Nonsteroidal anti-inflammatory drugs (NSAIDs): NSAIDs are known to exert anti-angiogenic and anti-metastatic activities both *in vitro* and *in vivo* (92). NSAIDs can up-regulate RECK expression in cancer cells (23, 77, 92). Treatment with 500 μ M aspirin [a nonselective cyclooxygenase (COX) inhibitor] for 48 h or 100 μ M NS-398 (a COX-2 inhibitor) for 48 h upregulated *RECK* mRNA and protein levels and suppressed MMP-2 activity in CL-1 human lung cancer cells (77, 92). These effects are considered to be independent of COX-2 inhibition, as prostaglandin E₂ and COX-2 overexpression failed to reverse the effects (77, 92). Treatment with 500 μ M aspirin for 48 h can cause growth suppression, reduction of RAS signaling, including phosphorylation of v-akt murine thymoma (AKT)/ERK/c-JUN, elevation of RECK expression, inhibition of MMP-2 and MMP-9 activity, and suppression of invasiveness of M139 cholangiocarcinoma cells (23). In HeLa cells, aspirin also inhibits ERK and AKT activities by reducing phospho-ERK and phospho-AKT, and induces apoptosis (93). These data suggest that the induction of RECK by aspirin is mediated by the inhibition of the RAS signaling pathway, which is possibly due to the re-activation of transcription initiation at the SP1 promoter site of the *RECK* gene, but not through COX inhibition (23, 56, 77, 92). As a whole, NSAIDs such as aspirin and NS398 might be used as RECK inducers and potential adjuvant therapeutic agents for cancer. Because an inverse association between NSAIDs use and GBM has been noted, NSAIDs might also be effective in the inhibition of GBM development or progression (94). Certainly, whether the RECK expression is induced in gliomas should be affirmed.

Conclusion

The proliferation and invasiveness of cancer are related to many factors, including growth factors, oncogenes, and tumor suppressor genes. Strategies to modulate these factors might be used to treat cancer. Since *RECK* is a tumor and metastasis suppressor gene (11, 50), up-regulation of *RECK* could be a valuable therapeutic approach for therapy of cancer (12). Both natural and synthetic agents have been identified, which enhance RECK expression, including forced expression of RECK, use of mimetics, recombinant peptides, microRNA antagonists, and gene therapy (12). However, the results of most related clinical trials are disappointing, with no evidence

of therapeutic benefits for patients (41, 44). VPA, an HDAC inhibitor and also a RECK inducer, has been shown to exert antitumor effects on gliomas (78, 81, 84-86). Because VPA has been widely used for the treatment of patients with gliomas and seizures, and is often orally administered for several years clinically, it might be used as an adjuvant treatment agent in addition to other treatment strategies such as surgery, radiation and chemotherapy. Because it is difficult to carry out long-term experiments about the effects of VPA on glioma cells, most *in vitro* studies use concentrations of VPA in a higher dose range than what is used clinically (95). Therefore, suitable doses of VPA as a RECK inducer to be used clinically are unknown. Future investigations should include *in vitro* and *in vivo* studies about the effects of long-term and low concentrations of VPA on gliomas, since high concentrations of VPA may cause somnolence, hematotoxicity and hepatotoxicity (86, 95). Long-term daily use of adult-strength aspirin is associated with modest reduction in overall cancer incidence in populations in which colorectal, prostate, and breast cancer are common (96). Nevertheless, clinically, the concentrations of this drug, required to induce RECK expression must be precisely evaluated, as high doses of aspirin may have undesirable side-effects (23). As a whole, further efforts to explore the effects of the induction of RECK expression by HDAC inhibitors, NSAIDs, or other new pharmaceutical agents should be continued. In addition, more studies are necessary to explore the benefits and risks of using HDAC inhibitors and NSAIDs as inducers of RECK expression in the treatment of glioma.

References

- Gabriely G, Wurdinger T, Kesari S, Esau CC, Burchard J, Linsley PS and Krichevsky AM: MicroRNA 21 promotes glioma invasion by targeting matrix metalloproteinase regulators. *Mol Cell Boil* 28: 5369-5380, 2008.
- Brandsma D and van den Bent MJ: Molecular-targeted therapies and chemotherapy in malignant gliomas. *Curr Opin Oncol* 19: 598-605, 2007.
- Tseng SH, Lin SM, Chen JC, Su YH, Huang HY, Chen CK, Lin PY and Chen Y: Resveratrol suppresses the angiogenesis and tumor growth of gliomas in rats. *Clin Cancer Res* 10: 2190-2202, 2004.
- Wen PY and Kesari S: Malignant gliomas in adults. *N Engl J Med* 359: 492-507, 2008.
- Chen Y, Chen JC and Tseng SH: Tetrandrine suppresses tumor growth and angiogenesis of gliomas in rats. *Int J Cancer* 124: 2260-2269, 2009.
- Selker RG, Shapiro WR, Burger P, Blackwood MS, Arena VC, Gilder JC, Malkin MG, Mealey JJ Jr., Neal JH, Olson J, Robertson JT, Barnett GH, Bloomfield S, Albright R, Hochberg FH, Hiesiger E, Green S and Brain Tumor Cooperative Group: The Brain Tumor Cooperative Group NIH Trial 87-01: a randomized comparison of surgery, external radiotherapy, and carmustine versus surgery, interstitial radiotherapy boost, external radiation therapy, and carmustine. *Neurosurgery* 51: 343-355, 2002.
- Tsao MN, Mehta MP, Whelan TJ, Morris DE, Hayman JA, Flickinger JC, Mills M, Rogers CL and Souhami L: The American Society for Therapeutic Radiology and Oncology (ASTRO) evidence-based review of the role of radiosurgery for malignant glioma. *Int J Radiat Oncol Biol Phys* 63: 47-55, 2005.
- Hegi ME, Diserens AC, Gorlia T, Hamou MF, de Tribolet N, Weller M, Kros JM, Hainfellner JA, Mason W, Mariani L, Bromberg JE, Hau P, Mirimanoff RO, Cairncross JG, Janzer RC and Stupp R: *MGMT* gene silencing and benefit from temozolomide in glioblastoma. *N Engl J Med* 352: 997-1003, 2005.
- Yamanaka R and Saya H: Molecularly targeted therapies for glioma. *Ann Neurol* 66: 717-729, 2009.
- Noda M and Takahashi C: Recklessness as a hallmark of aggressive cancer. *Cancer Sci* 98: 1659-1665, 2007.
- Takahashi C, Sheng Z, Horan TP, Kitayama H, Maki M, Hitomi K, Kitaura Y, Takai S, Sasahara RM, Horimoto A, Ikawa Y, Ratzkin BJ, Arakawa T and Noda M: Regulation of matrix metalloproteinase-9 and inhibition of tumor invasion by the membrane-anchored glycoprotein RECK. *Proc Natl Acad Sci USA* 95: 13221-13226, 1998.
- Nagini S: RECKing MMP: relevance of reversion-inducing cysteine-rich protein with Kazal motifs as a prognostic marker and therapeutic target for cancer (a review). *Anticancer Agents Med Chem* 2012.
- Echizenya M, Kondo S, Takahashi R, Oh J, Kawashima S, Kitayama H, Takahashi C and Noda M: The membrane-anchored MMP-regulator RECK is a target of myogenic regulatory factors. *Oncogene* 38: 5850-5857, 2005.
- Kondo S, Shukunami C, Morioka Y, Matsumoto N, Takahashi R, Oh J, Atsumi T, Umezawa A, Kudo A, Kitayama H, Hiraki Y and Noda M: Dual effects of the membrane-anchored MMP regulator RECK on chondrogenic differentiation of ATDC5 cells. *J Cell Sci* 120: 849-857, 2007.
- Muraguchi T, Takegami Y, Ohtsuka T, Kitajima S, Chandana EP, Omura A, Miki T, Takahashi R, Matsumoto N, Ludwig A, Noda M and Takahashi C: RECK modulates Notch signaling during cortical neurogenesis by regulating ADAM10 activity. *Nat Neurosci* 10: 838-845, 2007.
- Oh J, Takahashi R, Kondo S, Mizoguchi A, Adachi E, Sasahara RM, Nishimura S, Imamura Y, Kitayama H, Alexander DB, Ide C, Horan TP, Arakawa T, Yoshida H, Nishikawa S, Itoh Y, Seiki M, Itohara S, Takahashi C and Noda M: The membrane-anchored MMP inhibitor RECK is a key regulator of extracellular matrix integrity and angiogenesis. *Cell* 107: 789-800, 2001.
- Paulissen G, Rocks N, Quesada-Calvo F, Gosset P, Gosset P, Foidart JM, Noel A, Louis R and Cataldo DD: Expression of ADAMs and their inhibitors in sputum from patients with asthma. *Mol Med* 12: 171-179, 2006.
- Van Lent PL, Span PN, Sloetjes AW, Sloetjes AW, Radstake TR, van Lieshout AW, Heuvel JJ, Sweep CG and van den Berg WB: Expression and localisation of the new metalloproteinase inhibitor RECK (reversion inducing cysteine-rich protein with Kazal motifs) in inflamed synovial members of patients with rheumatoid arthritis. *Ann Rheum Dis* 64: 368-374, 2005.
- Chung TT, Yeh CB, Li YC, Su SC, Chien MH, Yang SF and Hsieh YH: Effect of RECK gene polymorphisms on hepatocellular carcinoma susceptibility and clinicopathologic features. *PLoS One* 7: e33517, 2012.

- 20 Kumamoto H and Ooya K: Immunohistochemical detection of MT1-MMP, RECK, and emmprin in ameloblastic tumors. *J Oral Pathol med* 35: 345-351, 2006.
- 21 Liu X, Wang W, Chen J, Chen C, Zhou J and Cao L: Expression of reversion-inducing cysteine-rich protein with Kazal motifs and matrix metalloproteinase 9 in middle ear squamous cell carcinoma. *ORL J Otorhinolaryngol Relat Spec* 74: 16-21, 2012.
- 22 Masui T, Doi R, Koshiba T, Fujimoto K, Tsuji S, Nakajima S, Koizumi M, Toyoda E, Tulachan S, Ito D, Kami K, Mori T, Wada M, Noda M and Imamura M: RECK expression in pancreatic cancer: Its correlation with lower invasiveness and better prognosis. *Clin Cancer Res* 9: 1779-1784, 2003.
- 23 Namwat N, Puetkasichonpasutha J, Loilome W, Yongvanit P, Techasen A, Puapairoj A, Sripa B, Tassaneeyakul W, Khuntikeo N and Wongkham S: Down-regulation of reversion-inducing-cysteine-rich protein with Kazal motifs (RECK) is associated with enhanced expression of matrix metalloproteinases and cholangiocarcinoma metastases. *J Gastroenterol* 46: 664-675, 2011.
- 24 Riddick AC, Shukla CJ, Pennington CJ, Bass R, Nuttall RK, Hogan A, Sethia KK, Ellis V, Collins AT, Maitland NJ, Ball RY and Edwards DR: Identification of degradome components associated with prostate cancer progression by expression analysis of human prostatic tissues. *Br J Cancer* 92: 2171-2180, 2005.
- 25 Song SY, Son HJ, Nam E, Rhee JC and Park C: Expression of reversion-inducing-cysteine-rich protein with Kazal motifs as a prognostic indicator in gastric cancer. *Eur J Cancer* 42: 101-108, 2006.
- 26 Span PN, Sweep CG, Manders P, Beex LV, Leppert D and Lindberg RL: Matrix metalloproteinase inhibitor reversion-inducing cysteine-rich protein with Kazal motifs: A prognostic marker for good clinical outcome in human breast carcinoma. *Cancer* 97: 2710-2715, 2003.
- 27 Takenaka K, Ishikawa S, Kawano Y, Yanagihara K, Miyahara R, Otake Y, Morioka Y, Takahashi C, Noda M, Wada H and Tanaka F: Expression of a novel matrix metalloproteinase regulator, RECK, and its clinical significance in resected non-small cell lung cancer. *Eur J Cancer* 40: 1617-1623, 2004.
- 28 Van der Jagt MF, Sweep FC, Waas ET, Hendriks T, Ruers TJ, Merry AH, Wobbes T and Span PN: Correlation of reversion-inducing cysteine-rich protein with kazal motifs (RECK) and extracellular matrix metalloproteinase inducer (EMMPRIN), with MMP-2, MMP-9 and survival in colorectal cancer. *Cancer Lett* 237: 289-297, 2006.
- 29 Xu J, Wu S and Shi X: Expression of matrix metalloproteinase regulator, RECK, and its clinical significance in osteosarcoma. *J Orthop Res* 28: 1621-1625, 2010.
- 30 Chang HC, Cho CY and Hung WC: Down-regulation of RECK by promoter methylation correlates with lymph node metastasis in non-small cell lung cancer. *Cancer Sci* 98: 169-173, 2007.
- 31 Dong Q, Yu D, Yang CM, Jiang B and Zhang H: Expression of the reversion-inducing cysteine-rich protein with Kazal motifs and matrix metalloproteinase-14 in neuroblastoma and the role in tumor metastasis. *J Exp Pathol* 91: 368-373, 2010.
- 32 Furumoto K, Arii S, Mori A, Furuyama H, Gorrin Rivas MJ, Nakao T, Isobe N, Murata T, Takahashi C, Noda M and Imamura M: RECK gene expression in hepatocellular carcinoma: correlation with invasion-related clinicopathological factors and its clinical significance. Reverse-reducing-cysteine-rich protein with Kazal motifs. *Hepatology* 33: 189-195, 2001.
- 33 Noda M, Oh J, Takahashi R, Kondo S, Kitayama H and Takahashi C: RECK: a novel suppressor of malignancy linking oncogenic signaling to extracellular matrix remodeling. *Cancer Metastasis Rev* 22: 167-175, 2003.
- 34 Rabien A, Burkhardt M, Jung M, Fritzsche F, Ringsdorf M, Schick Tanz H, Loening SA, Kristiansen G and Jung K: Decreased RECK expression indicating proteolytic imbalance in prostate cancer is associated with higher tumor aggressiveness and risk of prostate-specific antigen relapse after radical prostatectomy. *Eur Urol* 51: 1259-1266, 2007.
- 35 Silveira Correa TC, Massaro RR, Brohem CA, Taboga SR, Lamers ML, Santos MF and Maria-Engler STM: RECK-mediated inhibition of glioma migration and invasion. *J Cell Biochem* 110: 52-61, 2010.
- 36 Meng N, Li Y, Zhang H and Sun X-F: RECK, a novel matrix metalloproteinase regulator. *Histol Histopathol* 23: 1003-1010, 2008.
- 37 Nuttall RK, Sampieri CL, Pennington CJ, Gill SE, Schultz GA and Edwards DR: Expression analysis of the entire MMP and TIMP gene families during mouse tissue development. *FEBS Lett* 563: 129-134, 2004.
- 38 Yoon SO, Park SJ, Yun CH and Chung AS: Roles of matrix metalloproteinases in tumor metastasis and angiogenesis. *J Biochem Mol Biol* 36: 128-137, 2003.
- 39 Itoh Y and Nagase H: Matrix metalloproteinases in cancer. *Essays Biochem* 38: 21-36, 2002.
- 40 Nakada N, Nakada S, Demuth T, Tran NL, Hoelzinger DB and Berens ME: Molecular targets of glioma invasion. *Cell Mol Life Sci* 64: 458-478, 2007.
- 41 Coussens LM, Fingleton B and Matrisian LM: Matrix metalloproteinase inhibitors and cancer: Trials and tribulations. *Science* 295: 2387-2392, 2002.
- 42 Welm B, Mott J and Werb Z: Development biology: Vasculogenesis is a wreck without RECK. *Curr Biol* 12: R209-R211, 2002.
- 43 Takagi S, Simizu S and Osada H: RECK negatively regulates matrix metalloproteinase-9 transcription. *Cancer Res* 69: 1502-1508, 2009.
- 44 Silveira Correa TC, Brohem CA, Winnischofer SMB, da Silva Cardeal LB, Sasahara RM, Taboga SR, Sogayar MC and Maria-Engler SS: Down-regulation of the RECK-tumor- and metastasis-suppressor gene in glioma invasiveness. *J Cell Biochem* 99: 256-267, 2006.
- 45 Buchler P, Gazdhar A, Schubert M, Giese N, Reber HA, Hines OJ, Giese T, Ceyhan GO, Müller M, Büchler MW and Friess H: The Notch signaling pathway is related to neurovascular progression of pancreatic cancer. *Ann Surg* 242: 791-801, 2005.
- 46 Chesler L, Goldenberg DD, Seales IT, Satchi-Fainaro R, Grimmer M, Collins R, Struett C, Nguyen KN, Kim G, Tihan T, Bao Y, Brekken RA, Bergers G, Folkman J and Weiss WA: Malignant progression and blockade of angiogenesis in a murine transgenic model of neuroblastoma. *Cancer Res* 67: 9435-9442, 2007.
- 47 Ribatti D, Marimpietri D, Pastorino F, Brignole C, Nico B, Vacca A and Ponzoni M: Angiogenesis in neuroblastoma. *Ann NY Acad Sci* 1028: 133-142, 2004.
- 48 Watnick RS, Cheng YN, Rangarajan A, Ince TA and Weinberg RA: Ras modulates Myc activity to repress thrombospondin-1 expression and increase tumor angiogenesis. *Cancer Cell* 33: 219-231, 2003.

- 49 Caltagirone S, Rossi C, Poggi A, Ranelletti FO, Natali PG, Brunetti M, Aiello FB and Piantelli M: Flavonoid apigenin and quercetin inhibit melanoma growth and metastatic potential. *Int J Cancer* 87: 595-600, 2000.
- 50 Sasahara RM, Brochado SM, Takahashi C, Oh J, Maria-Engler SS, Granjeiro JM, Noda M and Sogayar MC: Transcriptional control of the RECK metastasis/angiogenesis-suppressor gene. *Cancer Detect Prev* 26: 435-443, 2002.
- 51 Oh J, Seo DW, Diaz T, Wei B, Ward Y, Ray JM, Morioka Y, Shi S, Kitayama H, Takahashi C, Noda M and Stetler-Stevenson WG: Tissue inhibitor of metalloproteinase 2 inhibits endothelial cell migration through increased expression of RECK. *Cancer Res* 64: 9062-9069, 2004.
- 52 Chang HC, Liu LT and Hung WC: Involvement of histone deacetylase in RAS-induced down-regulation of the metastasis suppressor *RECK*. *Cell Signal* 16: 675-679, 2004.
- 53 Chang HC, Cho CY and Hung WC: Silencing of the metastasis suppressor RECK by RAS oncogene is mediated by DNA methyltransferase 3b-induced promoter methylation. *Cancer Res* 66: 8413-8420, 2006.
- 54 Hsu MC, Chang HC and Hung WC: HER-2/neu represses the metastasis suppressor RECK via ERK and Sp transcription factors to promote cell invasion. *J Biol Chem* 281: 4718-4725, 2006.
- 55 Mori T, Moriuchi R, Okazaki E, Yamada K and Katamine S: TGAT oncoprotein functions as an inhibitor of RECK by association of the unique C-terminal region. *Biochem Biophys Res Commun* 355: 937-943, 2007.
- 56 Sasahara RM, Takahashi C and Noda M: Involvement of SP1 site in RAS-mediated down-regulation of the *RECK* metastasis suppressor gene. *Biochem Biophys Res Commun* 264: 668-675, 1999.
- 57 Loayaz-Puch F, Yoshida Y, Matsuzaki T, Takahashi C, Kitayama H and Noda M: Hypoxia and RAS-signaling pathways converge on, and co-operatively down-regulate, the RECK tumor-suppressor protein through microRNAs. *Oncogene* 29: 2638-2648, 2010.
- 58 Weijzen S, Rizzo P, Braid M, Vaishnav R, Jonkheer SM, Zlobin A, Osborne BA, Gottipati S, Aster JC, Hahn WC, Rudolf M, Siziopikou K, Kast WM and Miele L: Activation of NOTCH signaling maintains the neoplastic phenotype in human RAS-transformed cells. *Nat Med* 8: 979-986, 2002.
- 59 Aster JC, Xu L, Karnell FG, Patriub V, Pui JC and Pear WS: Essential roles for ankyrin repeat and transactivation domains in induction of T-cell leukemia by NOTCH1. *Mol Cell Biol* 20: 7505-7515, 2000.
- 60 Wang Z, Banerjee S, Li Y, Rahman KM, Zhang Y and Sarkar FH: Down-regulation of NOTCH-1 inhibits invasion by inactivation of nuclear-factor- κ B, vascular endothelial growth factor, and matrix metalloproteinase-9 in pancreatic cancer cell. *Cancer Res* 66: 2778-2784, 2006.
- 61 Ferres-Marco D, Gutierrez-Garcia I, Vallejo DM, Bolivar J, Gutierrez-Aviño FJ and Dominguez M: Epigenetic silencers and NOTCH collaborate to promote malignant tumors by Rb silencing. *Nature* 439: 430-436, 2006.
- 62 Allenspach E, Maillard I, Aster J, Patriub V, Pui JC and Pear WS: NOTCH signaling in cancer. *Cancer Biol Ther* 1: 466-476, 2002.
- 63 Axelson H: The NOTCH signaling cascade in neuroblastoma: role of the basic helix-loop-helix proteins HASH-1 and HES-1. *Cancer Lett* 204: 171-178, 2004.
- 64 Kanamori M, Kawaguchi T, Nigro JM, Feuerstein BG, Berger MS, Miele L and Pieper RO: Contribution of NOTCH signaling activation to human glioblastoma multiforme. *J Neurosurg* 106: 417-427, 2007.
- 65 Purow BW, Haque RM, Noel MW, Su Q, Burdick MJ, Lee J, Sundaresan T, Pastorino S, Park JK, Mikolaenko I, Maric D, Eberhart CG and Fine HA: Expression of NOTCH-1 and its ligands, DELTA-like-1 and GAGGED-1, is critical for glioma cell survival and proliferation. *Cancer Res* 65: 2353-2363, 2005.
- 66 Zhang XP, Zheng G, Zou L, Liu HL, Hou LH, Zhou P, Yin DD, Zheng QJ, Liang L, Zhang SZ, Feng L, Yao LB, Yang AG, Han H and Chen JY: NOTCH activation promotes cell proliferation and the formation of neural stem cell-like colonies in human glioma cells. *Mol Cell Biochem* 307: 101-108, 2008.
- 67 Ohishi K, Katayama N, Shiku H, Varnum-Finney B and Bernstein ID: NOTCH signalling in hematopoiesis. *Semin Cell Dev Biol* 14: 143-150, 2003.
- 68 Fogarty MP, Kessler JD and Wechsler-Reya RJ: Morphing into cancer: The role of developmental signaling pathways in brain tumor formation. *J Neurobiol* 64: 458-475, 2005.
- 69 Dziembowska M, Danilkiewicz M, Wesolowska A, Zupanska A, Chouaib S and Kaminska B: Cross-talk SMAD and p38 MAPK signaling in transforming growth factor β signal transduction in human glioblastoma cells. *Biochem Biophys Res Commun* 354: 1101-1106, 2007.
- 70 Masamune A and Shimosegawa T: Signal transduction in pancreatic stellate cells. *J Gastroenterol* 44: 249-260, 2009.
- 71 Wang M, Wang T, Liu S, Yoshida D and Teramoto A: The expression of matrix metalloproteinase-2 and -9 in human gliomas of different pathological grades. *Brain Tumor Pathol* 20: 65-72, 2003.
- 72 Small JV, Auinger S, Memethova M, Koestler S, Goldie KN, Hoenger A and Resch GP: Unraveling the structure of the lamellipodium. *J Microsc* 231: 479-485, 2008.
- 73 Chen Y, Tsai YH and Tseng SH: Valproic acid affected the survival and invasiveness of human glioma cells through diverse mechanisms. *J Neuro-Oncol* 2012.
- 74 Bartel DP: MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 116: 281-297, 2004.
- 75 Chan JA, Krichevsky AM and Kosik KS: MicroRNA-21 is an antiapoptotic factor in human glioblastoma cells. *Cancer Res* 65: 6029-6033, 2005.
- 76 Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M, Prueitt RL, Yanaihara N, Lanza G, Scarpa A, Vecchione A, Negrini M, Harris CC and Croce CM: A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci USA* 103: 2257-2261, 2006.
- 77 Liu LT, Chang HC, Chiang LC and Hung WC: Histone deacetylase inhibitor up-regulates RECK to inhibit MMP-2 activation and cancer cell invasion. *Cancer Res* 63: 3069-3072, 2003.
- 78 Duenas-Gonzalez A, Gandelaria M, Perez-Plascencia-Cardenas E, Perez-Cardenas E, de la Cruz-Hernandez E and Herrera LA: Valproic acid as epigenetic cancer drug: preclinical, clinical and transcriptional effects on solid tumors. *Cancer Treat Rev* 34: 206-222, 2008.
- 79 Furchert SE, Lanvers-Kaminsky C, Juurgens H, Jung M, Loidl A and Fruhwald MC: Inhibitors of histone deacetylase as potential therapeutic tools for high-risk embryonal tumors of the nervous system of childhood. *Int J Cancer* 120: 1787-1794, 2007.

- 80 Garcia-Manero G, Kantarjian HM, Sanchez-Gonzalez B, Yang H, Rosner G, Verstovsek S, Rytting M, Wierda WG, Ravandi F, Koller C, Xiao L, Faderl S, Estrov Z, Cortes J, O'Brien S, Estey E, Bueso-Ramos C, Fiorentino J, Jabbour E and Issa JP: Phase 1/2 study of the combination of 5-aza-2'-deoxycytidine with valproic acid in patients with leukemia. *Blood* 108: 3271-3279, 2006.
- 81 Michaelis M, Doerr HW and Cinatl J Jr.: Valproic acid as anticancer drug. *Curr Pharmacol Design* 13: 3378-3393, 2007.
- 82 Munster P, Marchion D, Bicaku E, Schmitt M, Lee JH, DeConti R, Simon G, Fishman M, Minton S, Garrett C, Chiappori A, Lush R, Sullivan D and Daud A: Phase 1 trial of histone deacetylase inhibition by valproic acid followed by the topoisomerase II inhibitor epirubicin in advanced solid tumors: a clinical and translational study. *J Clin Oncol* 25: 1979-1985, 2007.
- 83 Yang Q, Tian Y, Liu S, Zeine R, Chlenski A, Salwen HR, Henkin J and Cohn SL: Thrombospondin-1 peptide ABT-510 combined with valproic acid is an effective antiangiogenesis strategy in neuroblastoma. *Cancer Res* 67: 1716-1724, 2007.
- 84 van Breemen MSM, Rijsman RM, Taphoorn MJB, Walchenbach R, Zwinkels H and Vecht CJ: Efficacy of antiepileptic drugs in patients with gliomas and seizures. *J Neurol* 256: 1519-1526, 2009.
- 85 Weller M, Gorlia T, Cairncross JG, van den Bent MJ, Mason W, Belanger K, Brandes AA, Bogdahn U, Macdonald DR, Forsyth P, Rossetti AO, Lacombe D, Mirimanoff RO, Vecht CJ and Stupp R: Prolonged survival with valproic acid use in the EORTC/NCIC temozolomide trial for glioblastoma. *Neurology* 77: 1156-1164, 2011.
- 86 Wolff JE, Kramm C, Kortmann RD, Pietsch T, Rutkowski S, Jorch N, Gnekow A and Driever PH: Valproic acid was well tolerated in heavily pretreated pediatric patients with high-grade glioma. *J Neurooncol* 90: 309-314, 2008.
- 87 Gotfryd K, Skladchikova G, Lepekhin EA, Berezin V, Bock E and Walmod PS: Cell type-specific anticancer properties of valproic acid: independent effects on HDAC activity and ERK1/2 phosphorylation. *BMC Cancer* 10: 383, 2010.
- 88 Stockhausen MT, Sjolund J, Manetopoulos C and Axelsson H: Effects of the histone deacetylase inhibitor valproic acid on NOTCH signaling in human neuroblastoma cells. *Br J Cancer* 92: 751-759, 2005.
- 89 Ailenberg M and Silverman M: Trichostatin A – histone deacetylase inhibitor with clinical therapeutic potential - is a selective and potent inhibitor of gelatinase A expression. *Biochem Biophys Res Comm* 298: 110-115, 2002.
- 90 Hrebackova J, Hrabeta J and Eckschlager T: Valproic acid in the complex therapy of malignant tumors. *Curr Drug Therapy* 11: 361-379, 2010.
- 91 Satoh JI and Kuroda Y: β -Catenin expression in human neural cell lines following exposure to cytokines and growth factors. *Neuropathology* 20: 113-123, 2000.
- 92 Liu LT, Chang HC, Chiang LC and Hung WC: Induction of RECK by nonsteroidal anti-inflammatory drugs in lung cancer cells. *Oncogene* 21: 8347-8350, 2002.
- 93 Xiang S, Sun Z, He Q, Yan F, Wang Y and Zhang J: Aspirin inhibits ERBB2 to induce apoptosis in cervical cancer cells. *Med Oncol* 27: 379-387, 2010.
- 94 Sivak-Sears NR, Schwartzbaum JA, Miike R, Moghadassi M and Wrensch M: Case-control study of use of nonsteroidal anti-inflammatory drugs and glioblastoma multiforme. *Am J Epidemiol* 159: 1131-1139, 2004.
- 95 Van Nifterik KA, Van den Berg J, Slotman BJ, Lafleur MV, Sminia P and Stalpers LJ: Valproic acid sensitizes human glioma cells for temozolomide and γ -radiation. *J Neurooncol* 107: 61-67, 2012.
- 96 Jacobs EJ, Thun MJ, Bain EB, Rodriguez C, Henley SJ and Calle EE: A large cohort study of long-term daily use of adult-strength aspirin and cancer incidence. *J Natl Cancer Inst* 99: 608-615, 2007.

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